

Noble Energy Falklands Limited

Environmental Impact Statement (EIS) for Exploration Drilling Offshore the Falkland Islands

APPENDICES

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Noble Energy Falklands Limited

Environmental Impact Statement (EIS) for Exploration Drilling Offshore the Falkland Islands

APPENDICES

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Appendix A: Action Register

As part of the Environmental Impact Statement (EIS), the potential aspects for each key project activity have been considered and potential impacts on environmental sensitivities have been identified. These impacts have been risk assessed to determine their significance following the methodology outlined in Section 5 of the EIS. Appropriate mitigation measures have been put in place to lower the risk as far as possible.

KEY

Event Type: **P** = Planned, **U** = Unplanned

1-6 = Category of Consequence (refer to Table 5.3 - Definition of Consequence Categories in Section 5.6.2 of the EIS)

A-E = Likelihood (Frequency) of Occurrence (refer to Table 5.4 - Likelihood Categories in Section 5.6.3 of the EIS)



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Appendix B: Drilling Operations Supporting Information

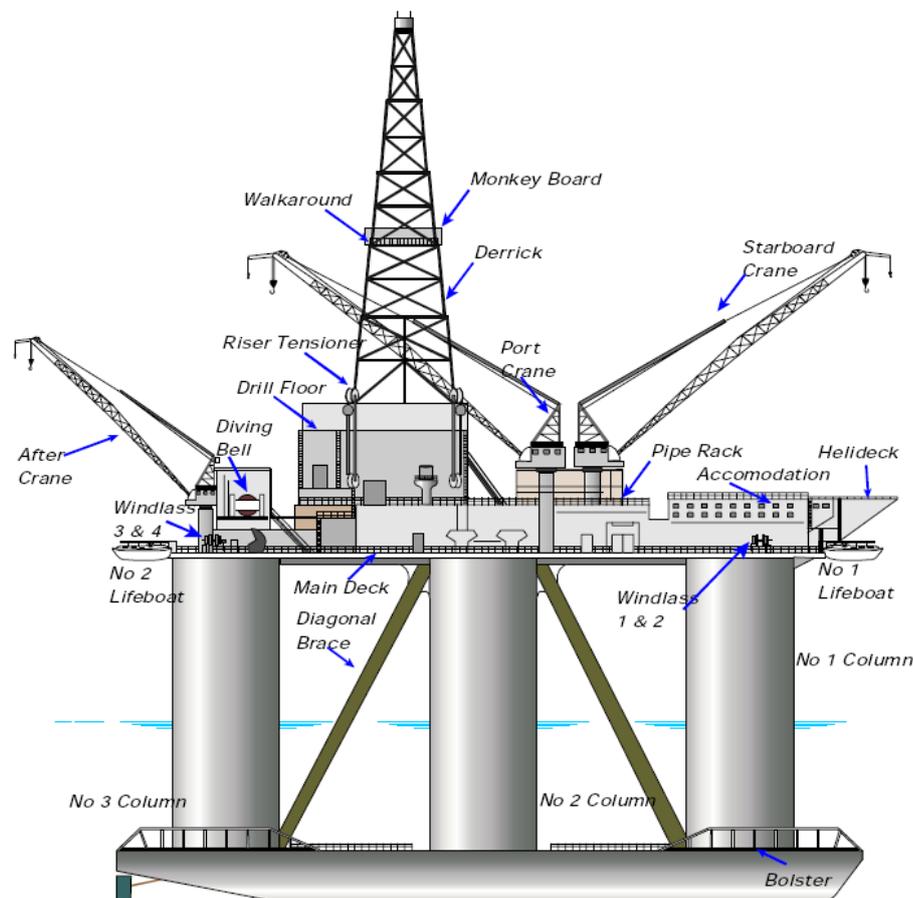
B.1 The Drilling Rig

The exploitation of hydrocarbons requires the construction of a conduit between the surface and the reservoir. This is achieved by the drilling process. Offshore wells are typically drilled by mobile offshore drilling units (MODUs) of which there are three broad designs currently in use: Drill ship;

- Semi-submersible drilling rig, and;
- Jack-up drilling rig.
- The proposed wells will be drilled using a semi-submersible drilling rig.

A semi-submersible rig is a floating unit that is supported primarily on large pontoon structures submerged below the sea surface. The operating decks are typically elevated 30 metres above the pontoons on large steel columns. Semi-submersible rigs are usually anchored to the seabed with six to twelve anchor chains or kept in place by a dynamic positioning (DP) system, which is a computer controlled thruster system used to maintain the rig position. Semi-submersible rigs can be used for drilling, work-over operations or as production platforms, depending on their equipment. Modern semi-submersible rigs - with DP systems - have the capacity to operate in deep water in excess of 1,500 metres. In addition, semi-submersible rigs have great flexibility concerning operating water depth and have the capacity to work in medium water and some shallow water fields. A schematic diagram of a typical semi-submersible drilling rig is given in Figure B.1 below.

Figure B.1: Typical semi-submersible drilling rig layout



To support the drilling operations, the following systems and services are usually located on a semi-submersible rig:

- Bulk Storage – Provided for fuel oil, bulk mud and cement, liquid mud, drill water and potable water;
- Pipe and Materials Storage – Covered storage is provided for sacked material, drilling equipment, spares, etc. and deck storage for drill pipe and casing;
- Helideck – Normally rated for a Sikorsky S-61 helicopter or equivalent;
- Craneage – Two deck cranes provided for loading/off loading equipment/supplies from Offshore Supply Vessels (OSVs);
- Emergency Systems – this includes life saving appliances, fire detection and protection equipment, combustible gas detection systems and life vessels; and
- Environmental Protection – Sewage treatment unit, blow-out preventer (BOP) system, cuttings cleaning equipment, hazardous and non-hazardous drainage systems (which collect rainwater and/or any minor spills and transport them to a drains tank for treatment prior to discharge to sea, or allow transfer to tote tanks for shipment to shore and appropriate disposal).

B.2 Well Construction

Once the rig is on location and secured, drilling operations to reach the potential hydrocarbon reservoir can commence. Well lengths can vary greatly from the order of a few hundred metres to in excess of 10,000 metres.

All rigs have a drilling derrick; a steel tower located over the drill floor which is the area of the rig where the majority of drilling activity is concentrated.

The derrick supports the weight of the drillstring which is screwed together from lengths of hollow drill pipe. Hoisting equipment in the derrick can raise or lower the drillstring. At the bottom of the drillstring is a drill bit, which can vary in size and type (Figure B.2). It is attached to the drill collars, which are heavy pipe-sections that put weight on the bit.

Figure B.2: A range of types of drill bit

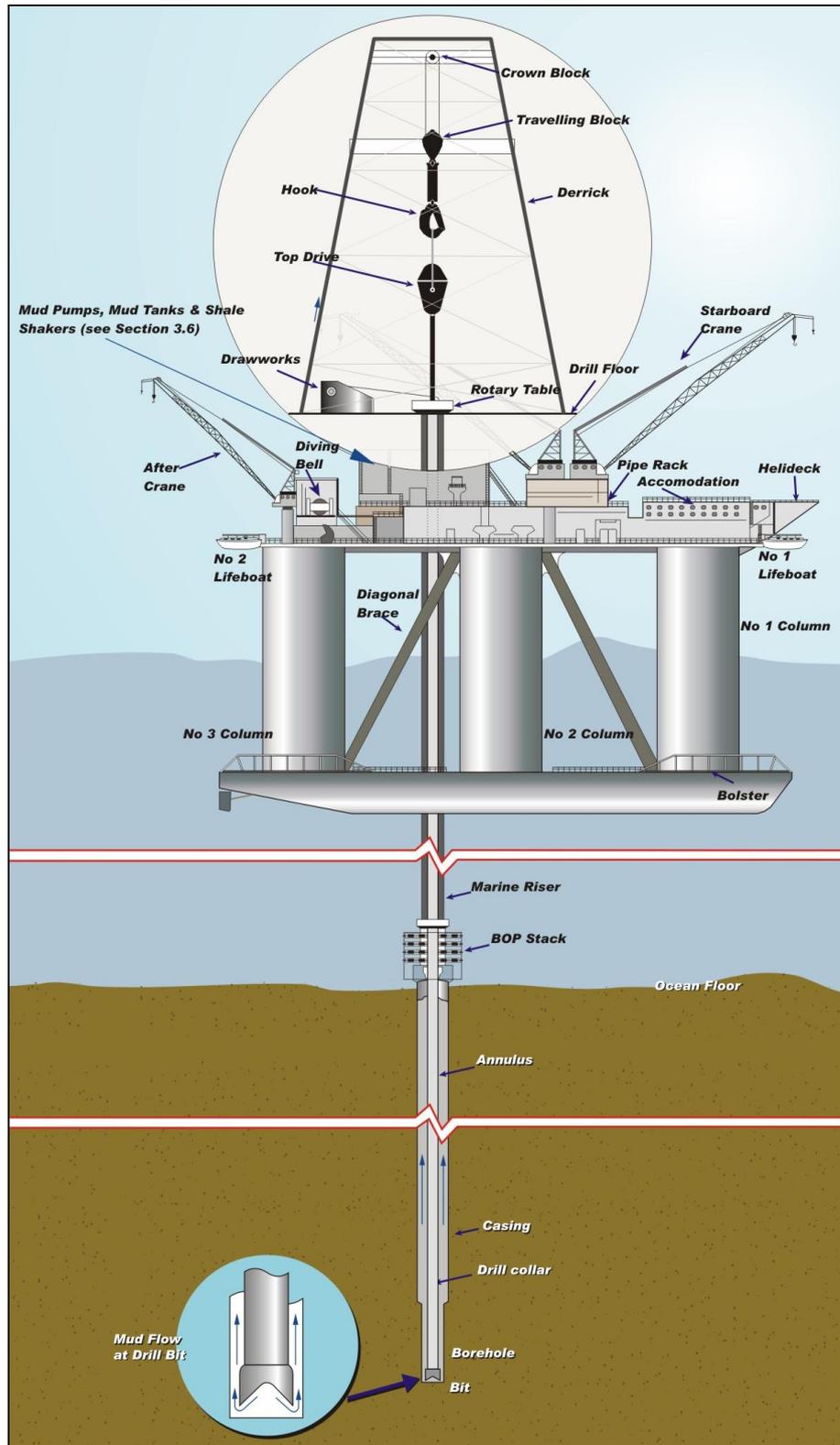


On semi-submersible rigs, a compensator keeps the drillstring stationary while the rig and derrick move as a result of wave motion. The drill bit is rotated either by turning the whole drillstring ("rotary drilling") or by using a downhole turbine which rotates as drilling fluid is pumped through it. In rotary drilling, the rotary motion is imparted to the drillstring by a "rotary table" on the drill floor or more commonly these days a "top drive". This is an electro-hydraulic motor suspended in the top of the derrick. It is attached to the top of the drillstring and imparts torque to it, causing it to rotate.

To add a new section of drill pipe the drillstring is clamped in the drill floor with wedges (slips) and the top drive disconnected. The new joint is screwed into the drillstring suspended in the

drill floor, the top drive connected to the top of the new joint and drilling restarted. The raising and lowering of the top drive and the maintenance of correct tension on the drillstring is controlled by the driller operating the drawworks lever in a control cabin on the drill floor (called the "doghouse"). A schematic of the main components of the conventional rotary drilling process are given in Figure B.3.

Figure B.3: Conventional rotary drilling system



The process of drilling grinds up the rock into small cuttings which are brought to the surface by the drilling fluid (also called "mud"), which is mainly water-based. The mud is pumped continuously down the hollow drillstring while drilling is ongoing. It is forced out of holes at the front of the drill bit (refer to Figure B.2), lubricates and cools the drilling tools, washes up the drill cuttings (small rock chippings) away from the bit and most importantly, balances the pressure of fluids in the rock formations below to prevent blowouts. The drilling mud is continuously circulated down the drill pipe and then pumped back up to the surface via the well annulus (the gap between drillstring and the side of the well bore) with the drilling cuttings. At the surface, the cuttings are removed from the mud, the drilling mud cleaned and then returned to the circulation system. Drilling muds and their use are described in further detail in Section B.4.

In offshore drilling, the first step is to install a wide-diameter conductor pipe (called a Marine Riser) into the seabed to guide the drilling and contain the drilling fluid. On mobile rigs this is drilled into the seabed. The conductor pipe represents the largest diameter of the well profile. As drilling operations continue, completed sections of the well are cased (lined) with steel pipe called 'casing' which is cemented into place. Casing the hole is essential as it not only facilitates drilling mud and cuttings to be re-circulated to the rig, but also seals off the pressure zones and weaker formations thus preventing hole collapse, as well as preventing contamination of potential aquifers by hydrocarbons and drilling materials.

B.3 Well Logging

The well is drilled in sections, each decreasing in diameter. As each well section is drilled, sensors that are integrated into the drill string take measurements whilst the well is being drilled. This process is called Logging Whilst Drilling (LWD). LWD measures and transmits geological parameters (such as rock density, formation pressure and resistivity at the drill bit) while the well is being drilled. Measured data is transmitted to the surface in real time via pressure pulses in the well's mud fluid column. At the end of the section, the drill string is removed from the well and wireline logs deployed into the well to obtain more data on the well characteristics.

B.4 Drilling Mud

During drilling operations, a fluid known as drilling mud is pumped through the drill string down to the drilling bit and once a conductor tube or riser is set in place, is returned to the rig via the space (or annulus) between the drill string and the casing (Figure B.4).

The selection of a type of 'mud' depends primarily on the geology of well to be drilled and the characteristics of the oil and/or gas reservoir in which drilling will take place. Drilling fluid is essential to the drilling operation and five basic properties are usually defined by the well program and monitored during drilling:

- Rheology - A high viscosity fluid is desirable to carry cuttings to surface and suspend weighting agents in the mud (such as barite). However, if viscosity is too high, friction may impede the circulation of the mud causing excessive pump pressure.
- Density - Sufficient hydrostatic pressure is required to prevent the borehole wall from caving in and to keep formation fluid from entering the wellbore. The higher the density of the mud compared to the density of the cuttings, the easier it is to clean the hole and the cuttings will be less inclined to fall through the mud.
- Fluid loss - The aim is to create a low-permeability filter cake to seal between the wellbore and the formation. Control of fluid loss restricts the invasion of the formation by filtrate and minimizes the thickness of filter cake that builds up on the borehole wall, reducing formation damage.
- Solids content - Solids are usually classified as high gravity (HGS) (barite and other weighting agents) or low gravity (LGS) (clays, polymers and bridging materials) deliberately put in the mud, plus drilled solids from dispersed cuttings and ground rock. The amount and type of solids in the mud affect a number of mud properties. A high solids content, particularly LGS, will increase plastic viscosity and gel strength. High-

solids muds have much thicker filter cakes and slower drilling rates. Large particles of sand in the mud cause abrasion on pump parts, tubulars and down-hole motors.

- Chemical properties - The chemical properties of the drilling fluid are central to performance and hole stability. Properties that must be anticipated include the dispersion of formation clays or dissolution of salt formations, the performance of other mud products (for example, polymers are affected by pH and calcium) and corrosion in the well.

The drilling mud also has to function as a lubricant and coolant for the drill bit and string. The ingredients of most drilling muds can be categorised into the following groupings:

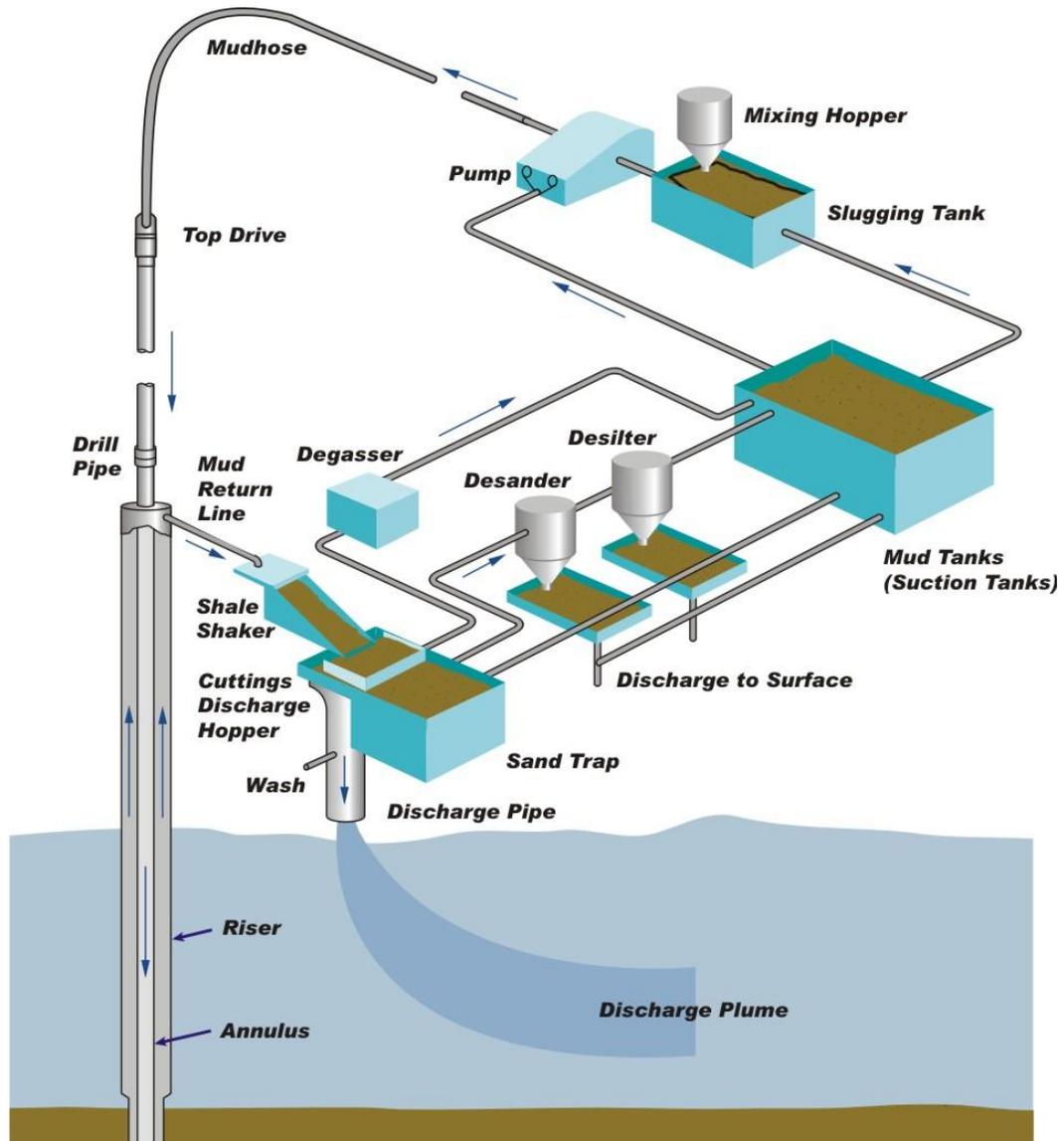
- Water - In water-based mud (WBM) this is the largest component. It may be used in its natural state, or salts may be added to change filtrate reactivity with the formation. Water hardness is usually eliminated through treatment and alkalinity is often controlled.
- Weighting agents - These are added to control formation fluid pressure. The most common is barite.
- Clay - Most commonly, bentonite is used to provide viscosity and create a filter cake on the borehole wall to control fluid loss. Clay is frequently replaced by organic colloids such as biopolymers, cellulose polymers or starch.
- Polymers - These are used to reduce filtration, stabilize clays, flocculate drilled solids and increase cuttings-carrying capacity. Cellulosic, poly-acrylic and natural gum polymers are used in low-solids mud to help maintain hole stability and minimize dispersion of the drill cuttings. Long-chain polymers are adsorbed onto the cuttings, thereby preventing disintegration and dispersion.
- Thinners - These are added to the mud to reduce its resistance to flow and to stifle gel development. They are typically plant tannins, polyphosphates, lignitic materials, lignosulfonates or synthetic polymers.
- Surfactants - These agents serve as emulsifiers, foamers and defoamers, wetting agents, detergents, lubricators and corrosion inhibitors.
- Inorganic chemicals - A wide variety of inorganic chemicals are added to mud to carry out various functions. For example, calcium hydroxide is used in lime mud and calcium chloride in oil based mud (OBM); sodium hydroxide and potassium hydroxide (caustic soda and caustic potash) are used to increase mud pH and solubilize lignite; sodium carbonate (soda ash) to remove hardness, sodium chloride to increase salinity, increase density, prevent hydrate formation and provide inhibition.
- Bridging materials - Calcium carbonate, cellulose fibres, asphalts and gilsonites are added to build up a filter cake on the fractured borehole and help prevent filtrate loss.
- Lost circulation materials - These are used to block large openings in the wellbore. These include walnut shells, mica and mud pills containing high concentrations of xanthum and modified cellulose.
- Specialized chemicals - Scavengers of oxygen, carbon dioxide or hydrogen sulphide are sometimes required, as are biocides and corrosion inhibitors.

Drilling mud is recycled and maintained in good condition throughout the drilling operation. The mud and suspended cuttings are processed on the rig through screens called "shale shakers" to maximise recovery of the mud. The recovered mud is then passed through a desander to remove sand particles and, if necessary, subsequent treatment may be provided by a centrifuge or desilter. This additional equipment removes the fine colloidal solids - the particles too small to be removed by the conventional equipment - which if allowed to build up can make the mud too viscous.

Three major types of mud are typically used in offshore drilling:

- Water based mud (WBM) – water forms the continuous phase of the mud (up to 90 percent by volume);
- Low toxicity oil based mud (LTOBM) – base oils, refined from crude oil, form the continuous phase of the mud, and;
- Synthetic based mud (SBM) – the continuous phase is refined from a number of organic compounds chosen because they act like base oil but are selected to be more biodegradable.

Figure B.4: Typical mud recycling system, once marine riser is in place



B.5 Well Control & Blow out Prevention

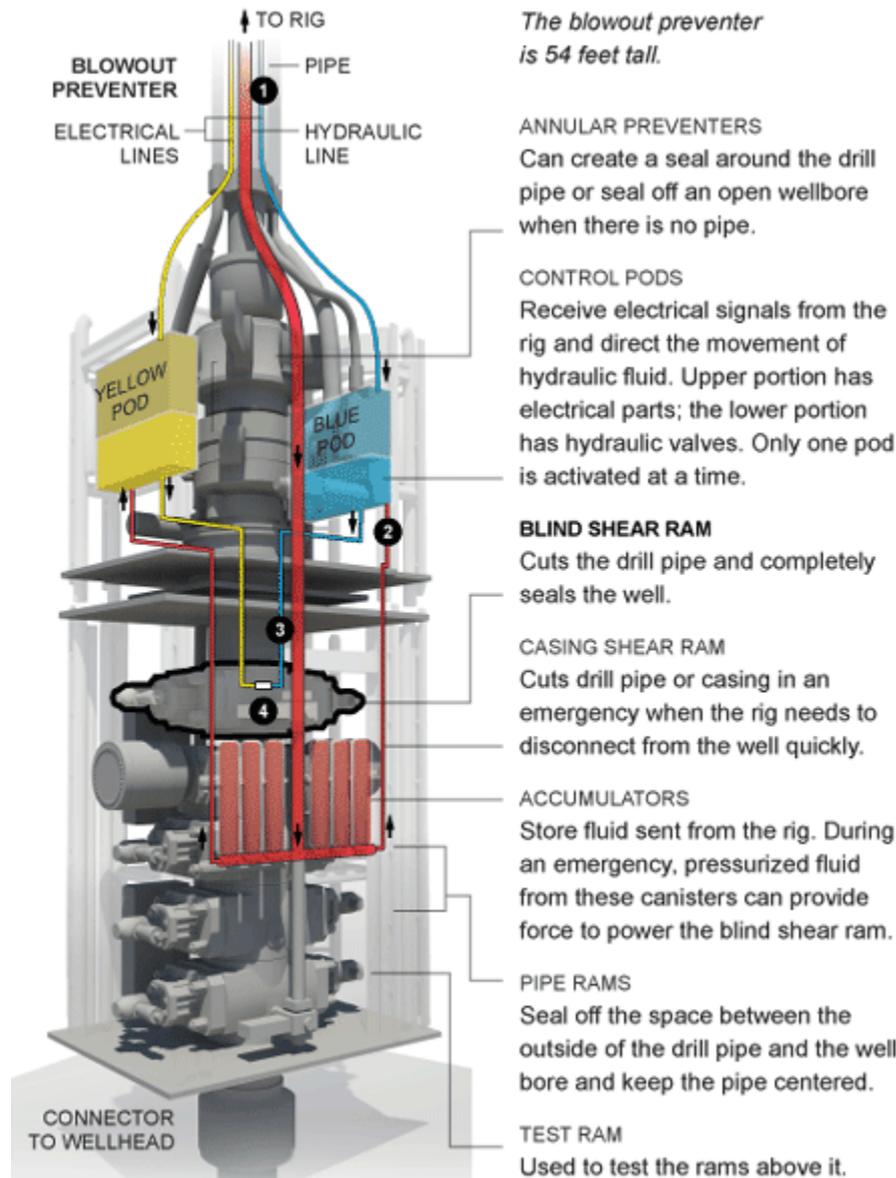
In addition to careful monitoring and control of the fluid system and the installation of casing in each section of the well, a blow-out preventer (BOP) stack, consisting of a series of individual valves (called preventers), is installed on the wellhead at the seafloor after the surface casing has been installed (Figure B.5).

The function of the BOP is to prevent uncontrolled flow from the well by positively closing in the well-bore if non routine flow from the well-bore is detected. The BOP is made up of a series of

hydraulically (or electro-hydraulically) operated gate valves which contain rams (mechanical devices that are used to seal the bore which come in a range of different designs, depending on their intended purpose) which can be functioned in the event of an emergency on the drill rig or down-hole to seal off the wellbore.

The exploration wells are not anticipated to encounter any zones of abnormal pressure during Noble’s exploration drilling programme. The BOP will be rated for pressures well in excess of those expected to be encountered in the well.

Figure B.5: A typical Subsea Blow-Out Preventer (BOP) stack



Appendix C: Offshore Chemical Notification Scheme (OCNS) & Harmonised Mandatory Control Scheme (HMCS) Information

Previously the control of offshore chemical discharges was controlled under the Offshore Chemical Notification Scheme (OCNS). Within the UK, the OCNS has been succeeded by The Offshore Chemicals Regulations 2002, which introduced a new approach to the consideration of chemical use and their discharge; the Harmonised Mandatory Control Scheme (HMCS). The Falkland Islands Government (FIG) aims to follow the example of the UK with regard to offshore chemical use. Both the OCNS and the HMCS are discussed below.

C.1 Offshore Chemical Notification Scheme (OCNS)

The Offshore Chemical Notification Scheme (OCNS) requires that all chemicals used in offshore exploration and production be tested using standard test protocols. Chemicals are then classified based on their biological properties e.g. toxicity and biodegradability. The OCNS scheme was adopted in the UK in 1979 and formed the basis of the Oslo and Paris Commissions (OSPARCOM) Harmonised Offshore Chemical Notification Format (HOCNF) which was established under cover of the Paris Commission Decision 96/3. The objectives of the OCNS and HOCNF are to regulate and manage chemical use by the oil and gas industry and consequently to prevent unacceptable damage to the marine environment through the operational or accidental discharge of chemicals.

The scheme was originally voluntary in the UK and all chemicals were given an OCNS Category ranging from 0 to 4. The system was later altered to harmonise the system with those operated by other countries bordering the North Sea. The HOCNS classifies all chemicals into five groups, A to E, with Category A chemicals being the most toxic and least biodegradable and Category E chemicals considered to be the least harmful to the offshore environment.

In addition to being placed into one of the five HOCNS categories, substances known or expected to cause tainting of fish tissue or substances known or expected to cause endocrine disruption, if lost or discharged, will be identified with a special taint or endocrine disrupter (ED) warning.

Chemicals are categorised on the basis of a series of laboratory tests with particular reference to their ecotoxicological effect, the biodegradability of the chemical and the potential for bioaccumulation in marine species. The ecotoxicological data used to classify the toxicity of chemicals are the results of laboratory tests on aquatic indicator organisms. Acute toxicity is assessed and expressed as either:

- An LC₅₀ – the concentration of the test substance in sea water that causes mortality 50 percent of the test batch; and
- An EC₅₀ – the concentration with a specified sub-lethal effect on 50 percent of the test batch.

The HOCNS grouping for a chemical is determined by comparing the results of toxicity tests for that chemical with the toxicity data given in Table C.1.

Table C.1: HOCNS Grouping Toxicity values (ppm) (Source: CEFAS, 2007)

| HOCNS Grouping | A | B | C | D | E |
|--|-----|---------|------------|---------------|---------|
| Results for aquatic toxicity data (ppm) | <1 | >1-10 | >10-100 | >100-1,000 | >1,000 |
| Results for sediment toxicity data (ppm) | <10 | >10-100 | >100-1,000 | >1,000-10,000 | >10,000 |

Aquatic toxicity - refers to the *Skeletonema costatum* EC₅₀, *Acartia tonsa* LC₅₀, and *Scophthalmus maximus* (juvenile turbot) LC₅₀ test

Sediment toxicity - refers to the *Corophium volutator* LC₅₀ test.

The categorisation also takes into account the chemicals' potential to bio-accumulate and biodegrade and other aspects such as potential endocrine disruption. The bioaccumulation potential and biodegradation rate relates to the fate of a chemical within the marine environment. Bioaccumulation potential describes the net result of uptake, distribution, biodegradation and elimination of a substance within an organism, subsequent to exposure but within the environment. The partition coefficient between octanol and water (expressed as Log Pow) is used as an indication of the potential for a substance to be bioaccumulated. A high value indicates a tendency to accumulate in lipophilic ("oil liking") phases such as the fatty tissues of organisms, suspended particles or sediments. However, because of biodegradation and elimination processes, a high Log P_{ow} does not necessarily imply bioaccumulation will occur. The classification outlined in Table C.2 is generally used to describe bioaccumulation potential.

Table C.2: Classification of Bioaccumulation Potential

| Bioaccumulation Potential | Log P _{ow} |
|---------------------------|---------------------|
| Low | <2 |
| Medium | 2-4 |
| High | >4 |

C.2 Harmonised Mandatory Control Scheme (HMCS)

The OSPAR Decision introducing an HMCS for the use and discharge of chemicals offshore came into force through the Offshore Chemicals Regulations 2002. The regulatory regime requires operators to obtain a permit to use and discharge chemicals in the course of oil and gas exploration and production operations offshore.

The OSPAR Decision and its supporting Recommendations entered into force on 16 January 2001. The Decision requires offshore chemicals to be ranked according to their calculated Hazard Quotients relating to each chemical discharge under standardised platform conditions (HQ = ratio of Predicted Environmental Concentration (PEC) to Predicted No Effect Concentration (PNEC)). It also obliges authorities to use the CHARM "hazard assessment" module as the primary tool for ranking. In the UK this is carried out by a multidisciplinary team at the CEFAS Burnham Laboratory. From this information, operators assess and select their chemical need, calculating PEC:PNECs for actual conditions of use (utilising the CHARM module as appropriate) and bearing in mind the objective of the HMCS to identify substances of concern for substitution and ranking of others to support moves towards the use of less harmful substances. Inorganic chemicals and organic chemicals with functions for which the CHARM model has no algorithms will continue to be ranked using the existing HOCNS hazard groups defined above.

A series of ranked lists are maintained on the CEFAS web site which use a banding system to rank organic chemicals of similar function according to PEC:PNEC "Hazard Quotients" calculated using the CHARM model. The band definitions are given in Table C.3.

Table C.3: Classification of Bioaccumulation Potential

| HQ Band | HQ Value |
|---------|--------------------|
| Gold | $0 > X < 1$ |
| Silver | $1 = < X < 30$ |
| White | $30 = < X < 100$ |
| Blue | $100 = < X < 300$ |
| Orange | $300 = < X < 1000$ |
| Purple | $1000 = < X$ |

The minimum data set of actual values and the parameters used by CEFAS to calculate them are disclosed to chemical suppliers on “templates”. The suppliers then pass these on to operators to enable the calculation of site-specific risk assessments (RQs) for any chemicals they may want to use. Some chemicals are generated and used in-situ on offshore installations (e.g. Sodium Hypochlorite) and don't fall under the remit of any one supplier.

The properties of substances on the OSPAR List of Substances/Preparations Used and Discharged Offshore, Which Pose Little Or No Risk to the Marine Environment (PLONOR) are sufficiently well known that the UK Regulatory Authorities do not require them to be tested. This list is reviewed annually and the notification requirements for these chemicals are given in the PLONOR document.

C.3 Chemicals not Covered by OCNS

The OCNS does not apply to chemicals that might otherwise be used on a ship, helicopter or other offshore structure. Products used solely within domestic accommodation areas - such as additives to potable water systems, paints and other coatings, fuels, lubricants, fire-fighting foams, hydraulic fluids used in cranes and other machinery - are also exempt.

C.4 Substitution Warnings

The substitution of harmful chemicals is an important part of the HMCS. The UK is obliged to implement the policy to replace chemical substances identified as candidates for substitution. Any chemical that contains one or more components that have been recommended for substitution is assigned the chemical label code ‘SUB’.

A chemical may carry a substitution warning for a variety of reasons, which include if the chemical:

- is on the OSPAR List of chemicals for priority action;
- is on the list of chemicals of possible concern;
- is considered by the authorising authority to be of equivalent concern for the marine environment;

Or:

- is inorganic and has a LC_{50} or EC_{50} of less than 1 mg/l

Or:

- has a biodegradation of:

- <20% in OECD 306, marine BODIS or any other accepted marine protocols, or
- <20% in 28 days in freshwater (OECD 301 and 310), or
- if half-life values >60 and 180 days from simulation tests in marine water and sediment, respectively (e.g. OECD 308, 309);

Or meets two of the following three criteria:

- **Biodegradation:**
 - <60% in 28 days in OECD 306, marine BODIS or any other acceptable marine protocol, or in the absence of valid results for such tests (<60% in 28 days (OECD 301B, 301C, 301D, 301F, 310, freshwater BODIS), or
 - <70% in 28 days (OECD 301A, 301E), or
- **Bioaccumulation:**
 - BCF >100 or Log Pow \geq 3 and molecular weight <700, or
 - if the conclusion of a weight-of-evidence expert judgement under Appendix 3 of OSPAR Agreement 2008-5 is negative, or
- **Toxicity:**
 - LC50 <10mg/l or EC50 <10mg/l.

A reliable value of Log P_{ow} cannot be calculated for surfactants and therefore cannot be used to indicate whether a surfactant might bio-accumulate. OSPAR requires regulatory authorities to take a precautionary approach where data are ambiguous or missing. Therefore, substitution warnings are applied to those surfactants that have a molecular weight of <700 and are either:

- Less than 60% or 70% biodegradable in 28 days (according to the test protocol), or
- Have an EC50/LC50 <10 mg/l

unless Cefas is satisfied that other evidence submitted by the product supplier indicates that the substance should not bio-accumulate.

During the risk-assessment process, operators are required to consider the selection of products both in terms of the magnitude of their Risk Quotient (RQ) and the presence of hazardous substances, including candidates for substitution. Operators are required to provide a robust defence for the continued use of products that have a high RQ or contain candidates for substitution.

Chemical suppliers must consider the advice they provide to operators that justifies continued use of any product containing candidates for substitution. In addition, suppliers should consider a managed approach to the replacement of any undesirable components, leading to the reformulation and re-certification of products.

Operators are encouraged to select products without a substitution warning. Therefore, a supplier may wish to seek alternatives at the product-development stage. However, there may be good technical reasons why a particular substance cannot immediately be substituted. The supplier should highlight these to operators so that they can include this information in their justification for the continued use of the product.

Appendix D: Noble Energy Inc. Global Environmental, Health & Safety Management System Elements

GEMS

Global Environmental, Health and Safety Management System

A consistent framework for the management of EHS issues is necessary to protect the environment and the health and safety of our employees and communities. Our GMS incorporates legal requirements and best practices under an umbrella framework consisting of 14 elements:

Prepare

1. Management Commitment and Employee Participation
2. Legal Aspects and Document Control
3. Safe Work and Operating Practices
4. Process Safety and Environmental Information
5. Emergency Preparedness and Community Awareness

Execute

6. Safety and Environmental Training
7. Contractor Safety Management
8. Pre-startup Review
9. Management of Change
10. Risk Assessment and Management

Verify

11. Performance Monitoring and Measuring
12. Incident Reporting, Analysis and Corrective Action
13. Management System Compliance Audit

Perform

14. Operational Integrity and Continual Improvement

Appendix E: Fisheries Statistics Maps

Fisheries statistics for the period 2008 to 2013 have been provided by the Falkland Islands Government (FIG) Department of Natural Resources – Fisheries Department to Noble for the purposes of this ESHIA document. In addition, Vessel Monitoring System (VMS) data have also been provided for the period 2008 to 2012.

Maps for the following species have been produced:

- Southern Blue Whiting (*Micromesistius australis*) (Figure E.1);
- Grenadiers (*Macrouridae*) (Figure E.2);
- Hake (*Merluccius sp.*) (Figure E.3);
- Hoki (*Macruronus magellanicus*) (Figure E.4);
- Rays (*Rajidae*) (Figure E.5);
- Red Cod (*Salilota australis*) (Figure E.6);
- Rock Cod (*Patagonotothen ramsayi*) (Figure E.7);
- Patagonian Toothfish (*Dissostichus eleginoides*) (Figure E.8);
- Kingclip (*Genypterus blacodes*) (Figure E.9);
- Argentine shortfin squid (*Illex argentinus*) (Figure E.10);
- Patagonian Squid (*Doryteuthis gahi*) (Figure E.11);
- Other species (Figure E.12).

Maps for the following VMS positions have been produced:

- VMS Positions for 2008 (Figure E.13);
- VMS Positions for 2009 (Figure E.14);
- VMS Positions for 2010 (Figure E.15);
- VMS Positions for 2011 (Figure E.16);
- VMS Positions for 2012 (Figure E.17);
- VMS Positions – All years and all months (Figure E.18).

The data have been plotted quarterly for each year for both species and VMS position.

Maps of fisheries effort have also been produced:

- Fishing Effort, 2008 (Figure E.19);
- Fishing Effort, 2009 (Figure E.20);
- Fishing Effort, 2010 (Figure E.21);
- Fishing Effort, 2011 (Figure E.22);
- Fishing Effort, 2012 (Figure E.23).

The fisheries catch statistics maps should be used with caution. As the catch maps do not take into account the level of fishing effort, it is important to remember that patterns between areas could be due to differential fishing effort, as levels of fishing effort are highly likely to vary seasonally.

Figure E.1a: Fisheries catch mass (tonnes) for Southern Blue Whiting (*Micromesistius australis*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

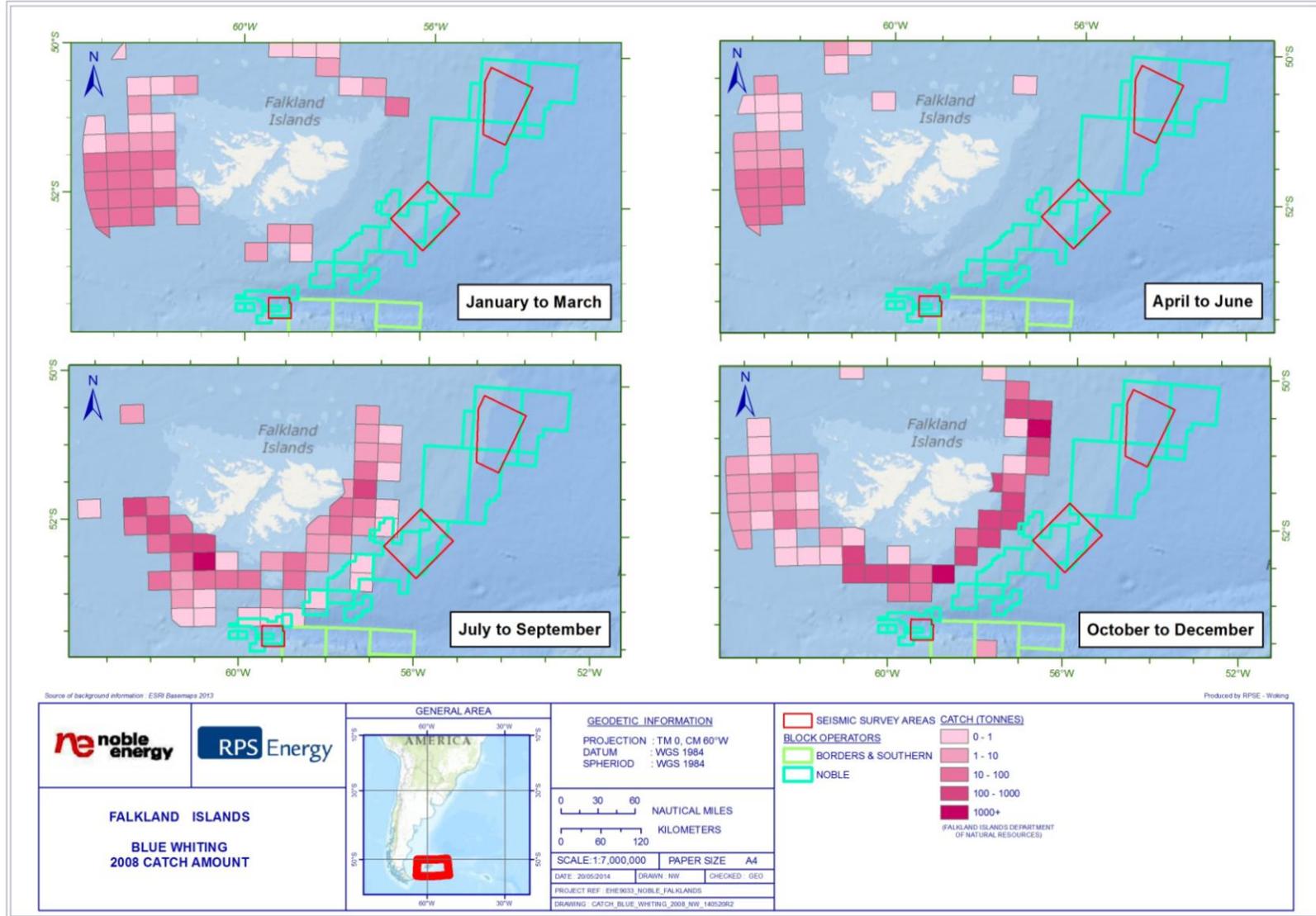


Figure E.1b: Fisheries catch mass (tonnes) for Southern Blue Whiting (*Micromesistius australis*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

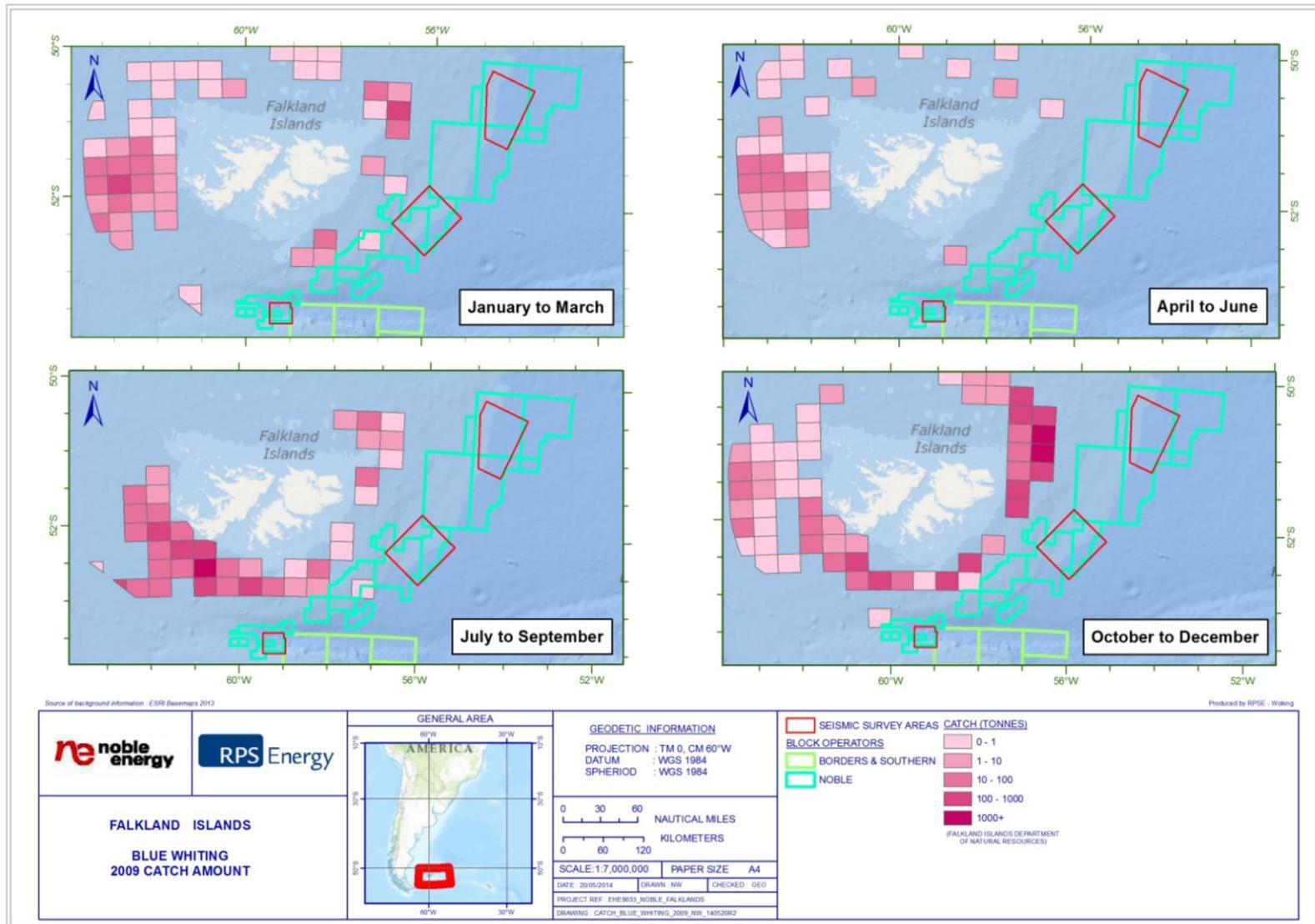


Figure E.1c: Fisheries catch mass (tonnes) for Southern Blue Whiting (*Micromesistius australis*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

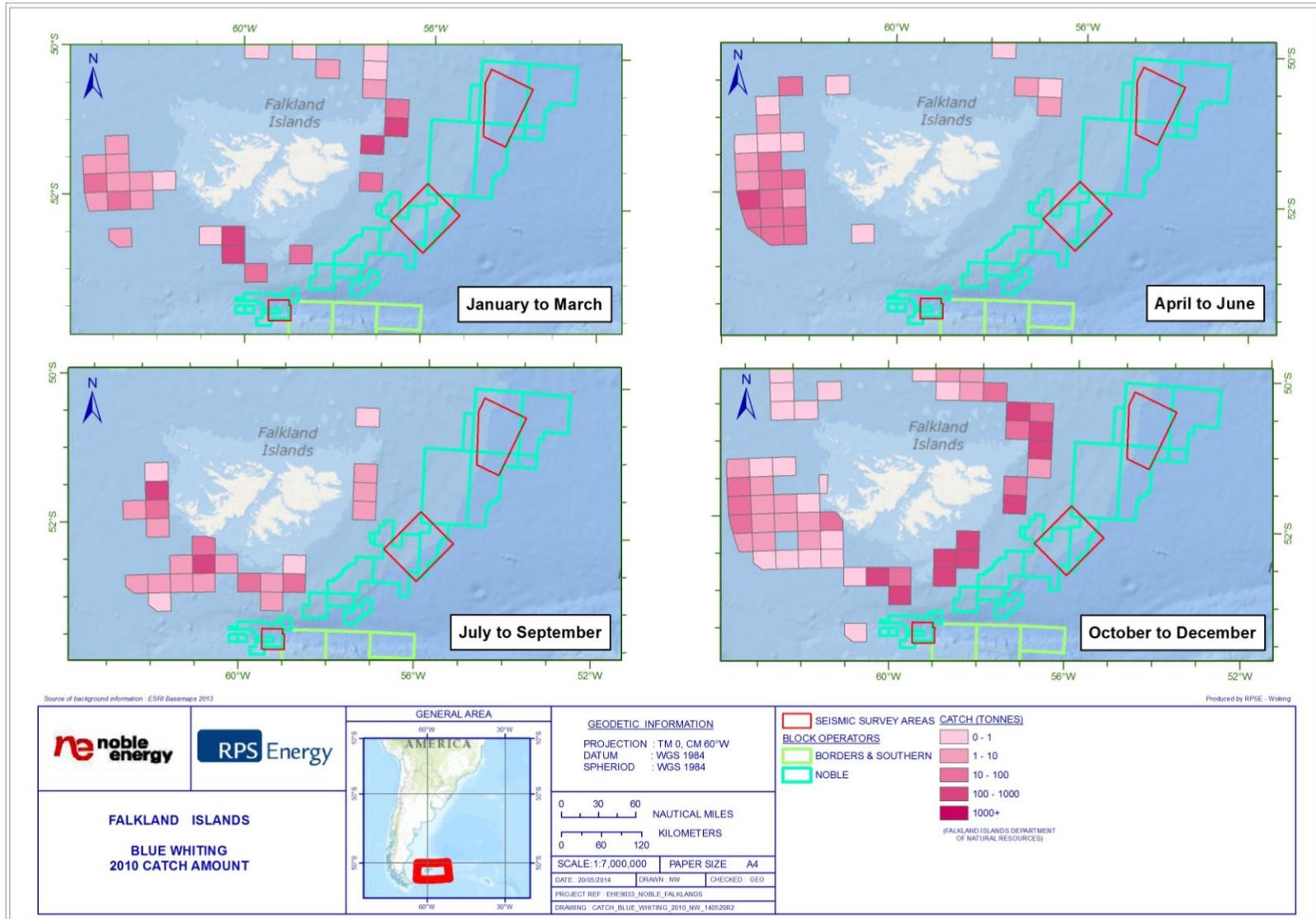


Figure E.1d: Fisheries catch mass (tonnes) for Southern Blue Whiting (*Micromesistius australis*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

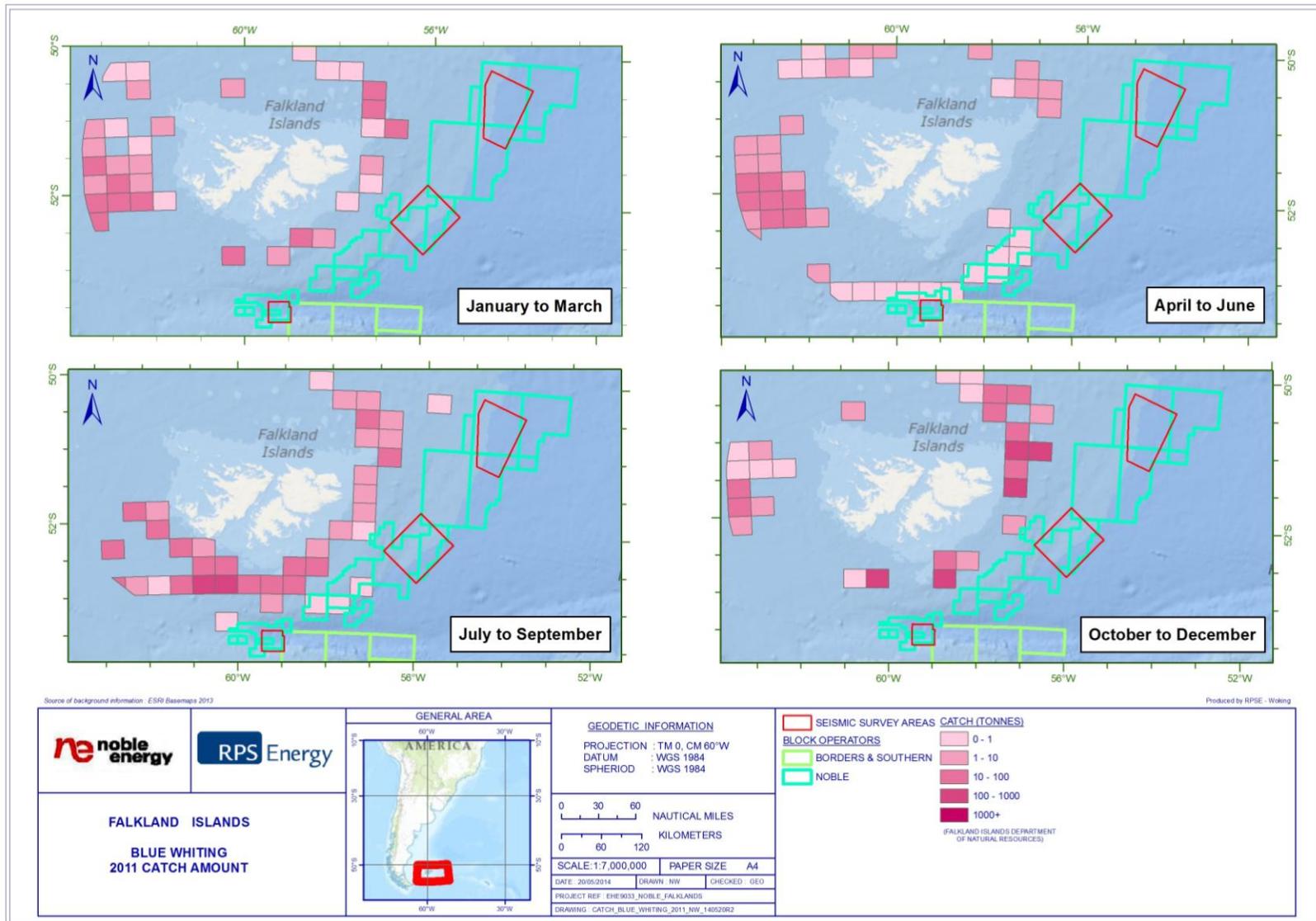


Figure E.1e: Fisheries catch mass (tonnes) for Southern Blue Whiting (*Micromesistius australis*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

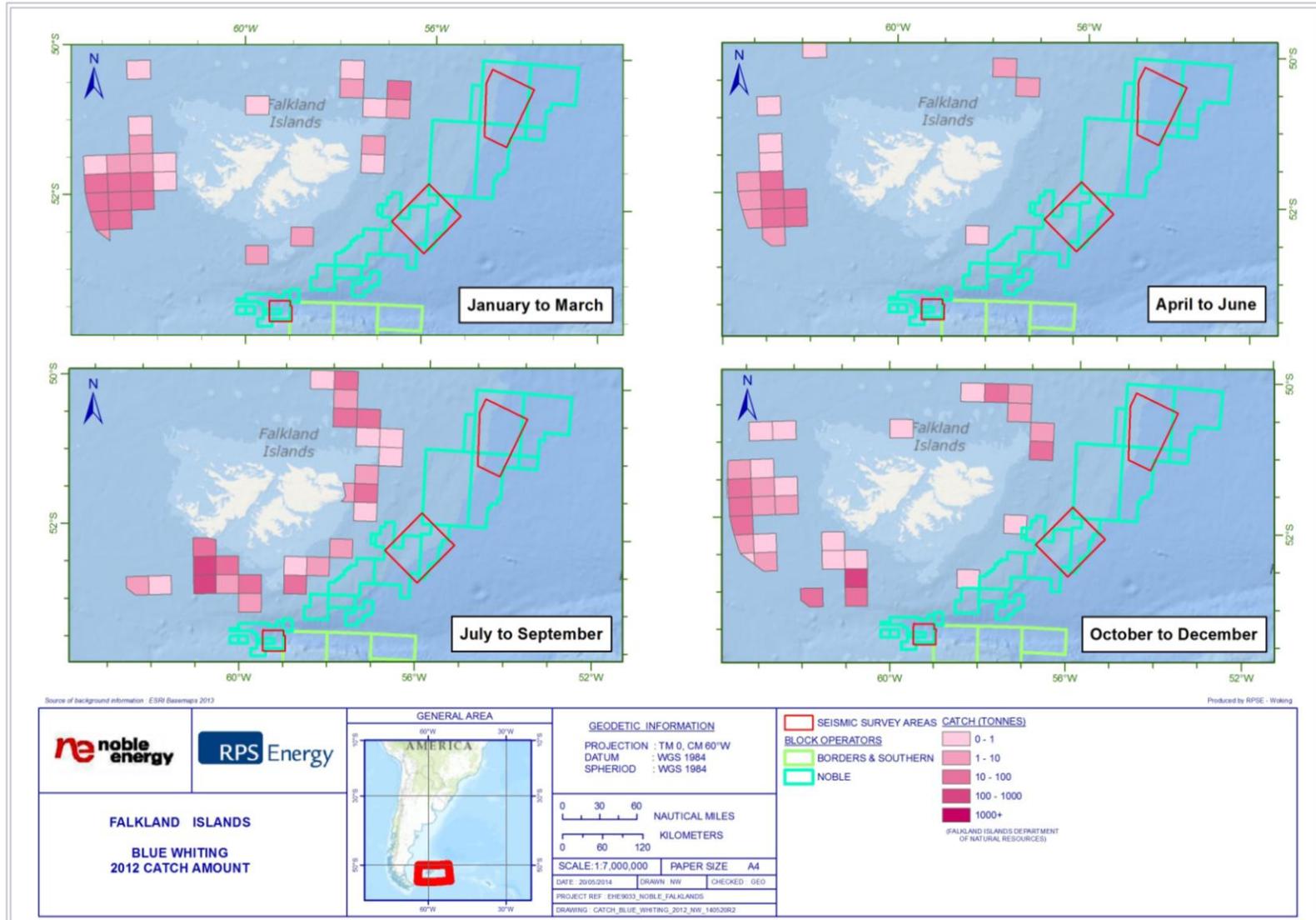


Figure E.1f: Fisheries catch mass (tonnes) for Southern Blue Whiting (*Micromesistius australis*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

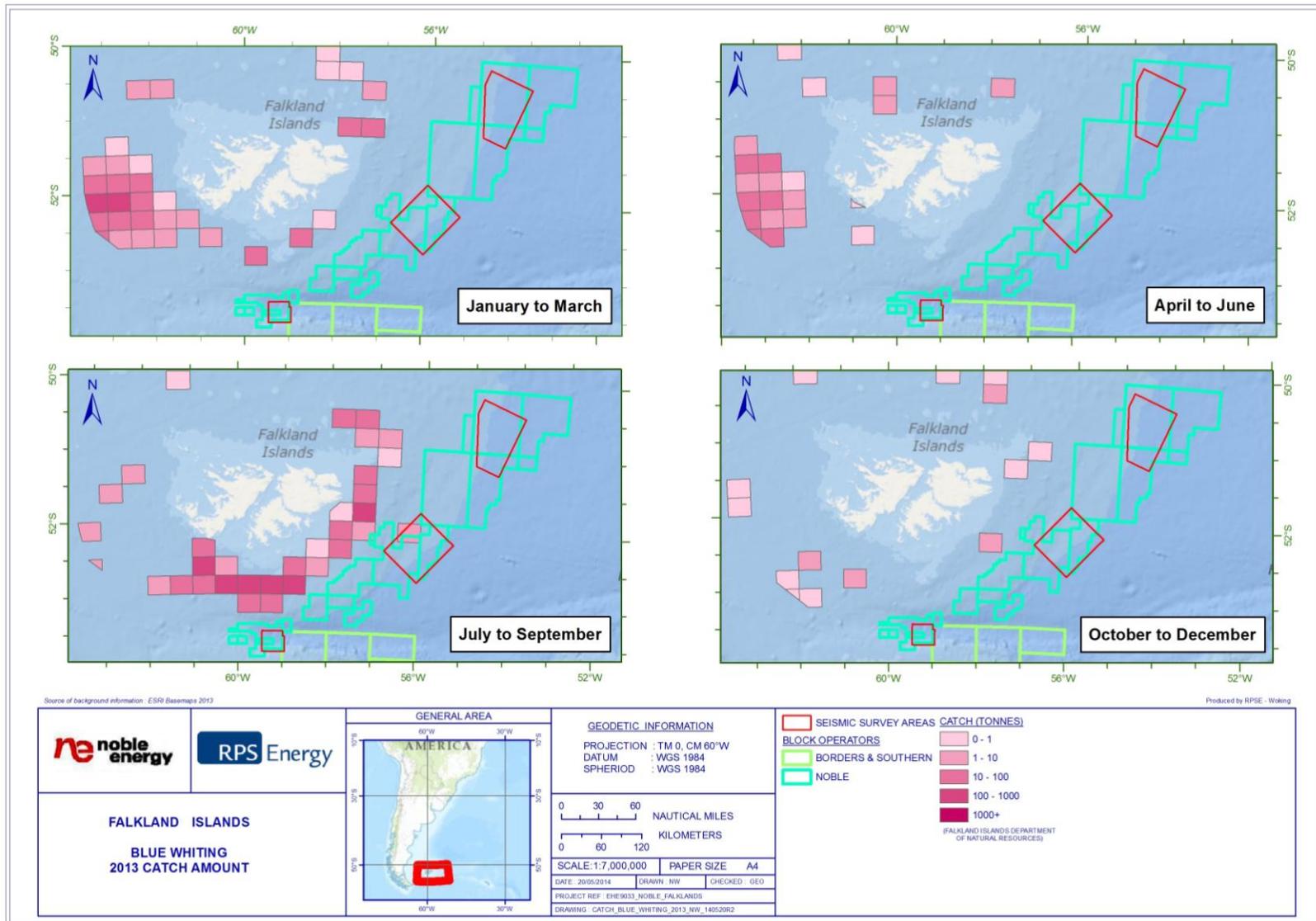


Figure E.2a: Fisheries catch mass (tonnes) for Grenadiers (*Macrouridae*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

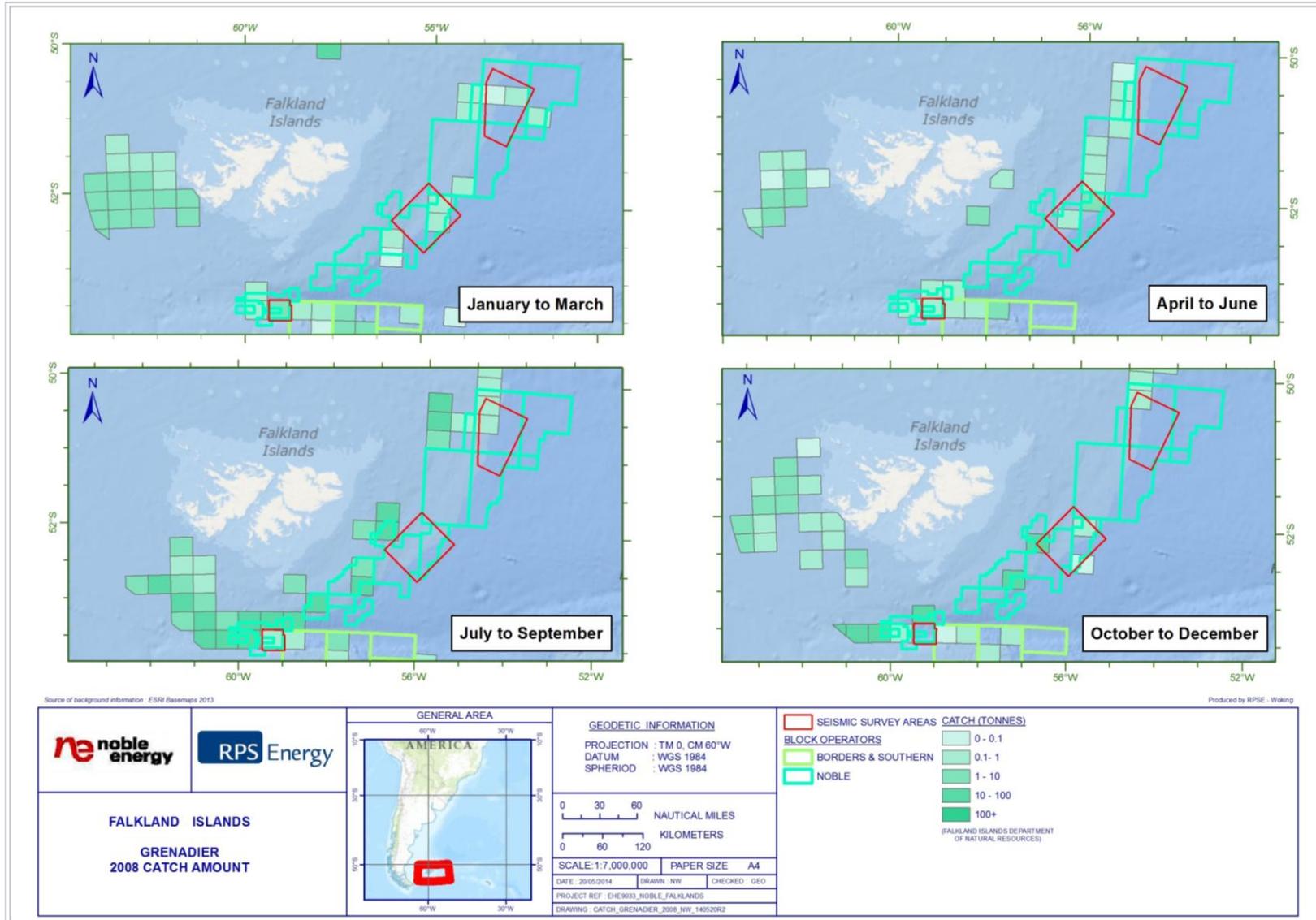


Figure E.2b: Fisheries catch mass (tonnes) for Grenadiers (*Macrouridae*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

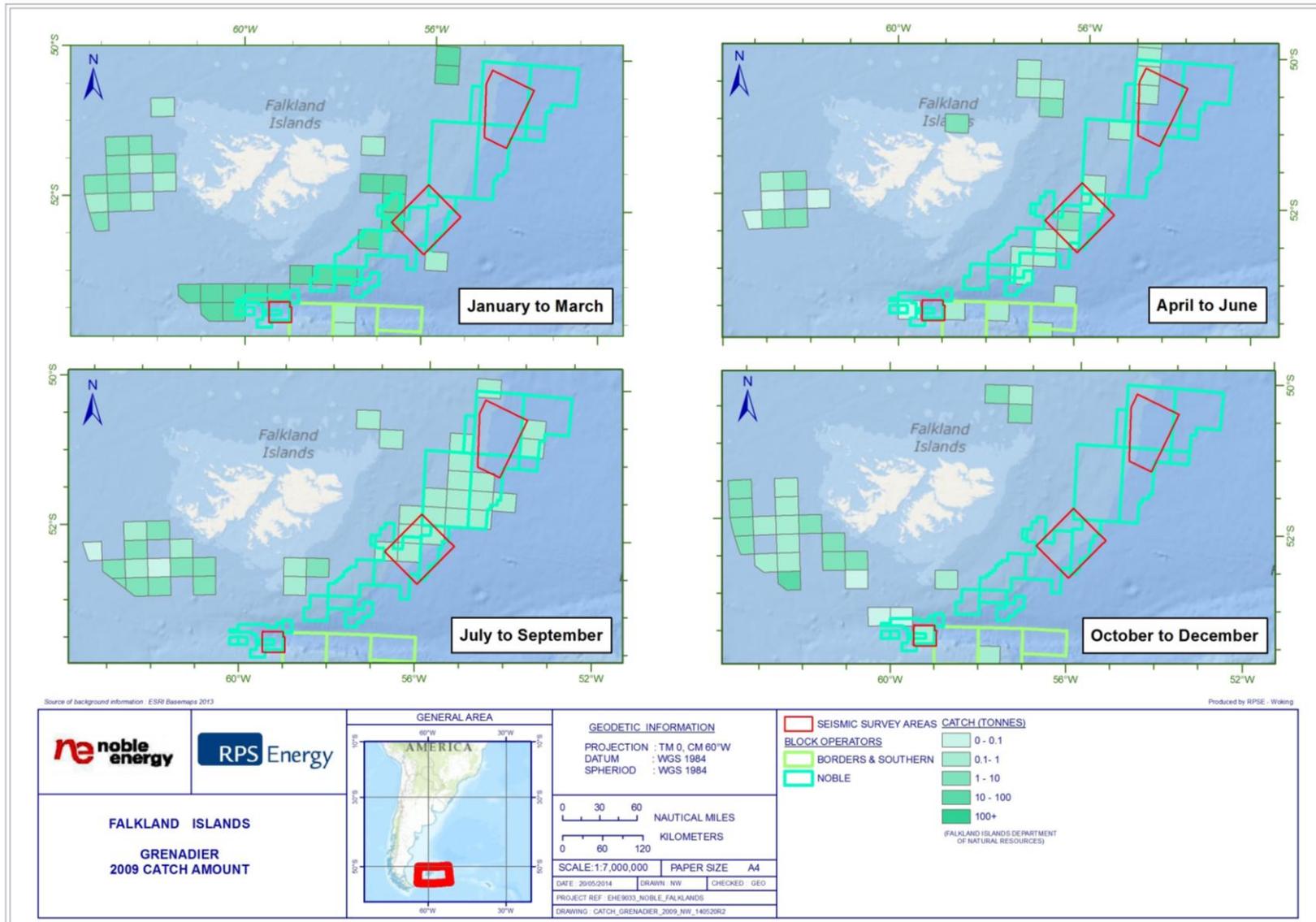


Figure E.2c: Fisheries catch mass (tonnes) for Grenadiers (*Macrouridae*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

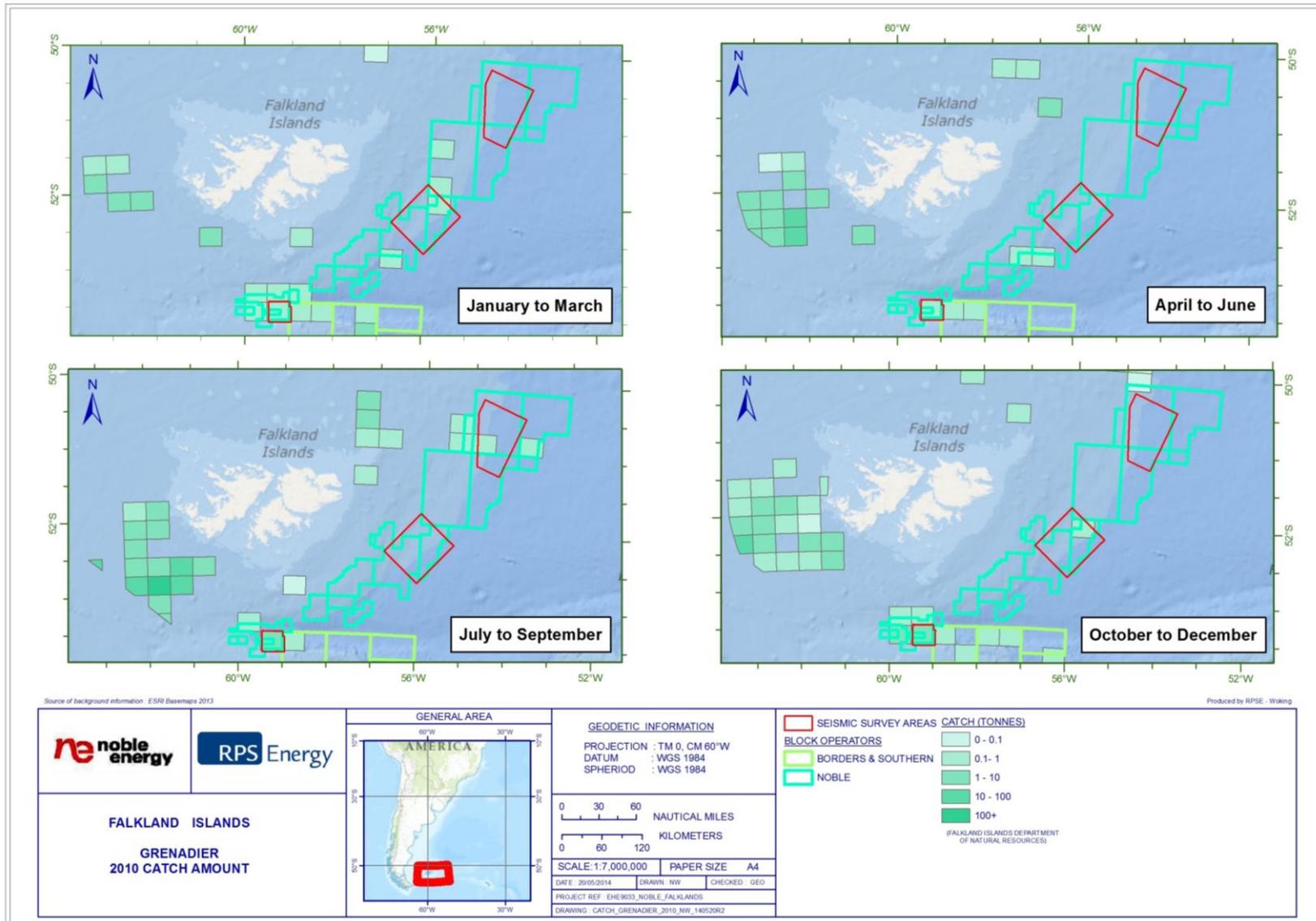


Figure E.2d: Fisheries catch mass (tonnes) for Grenadiers (*Macrouridae*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

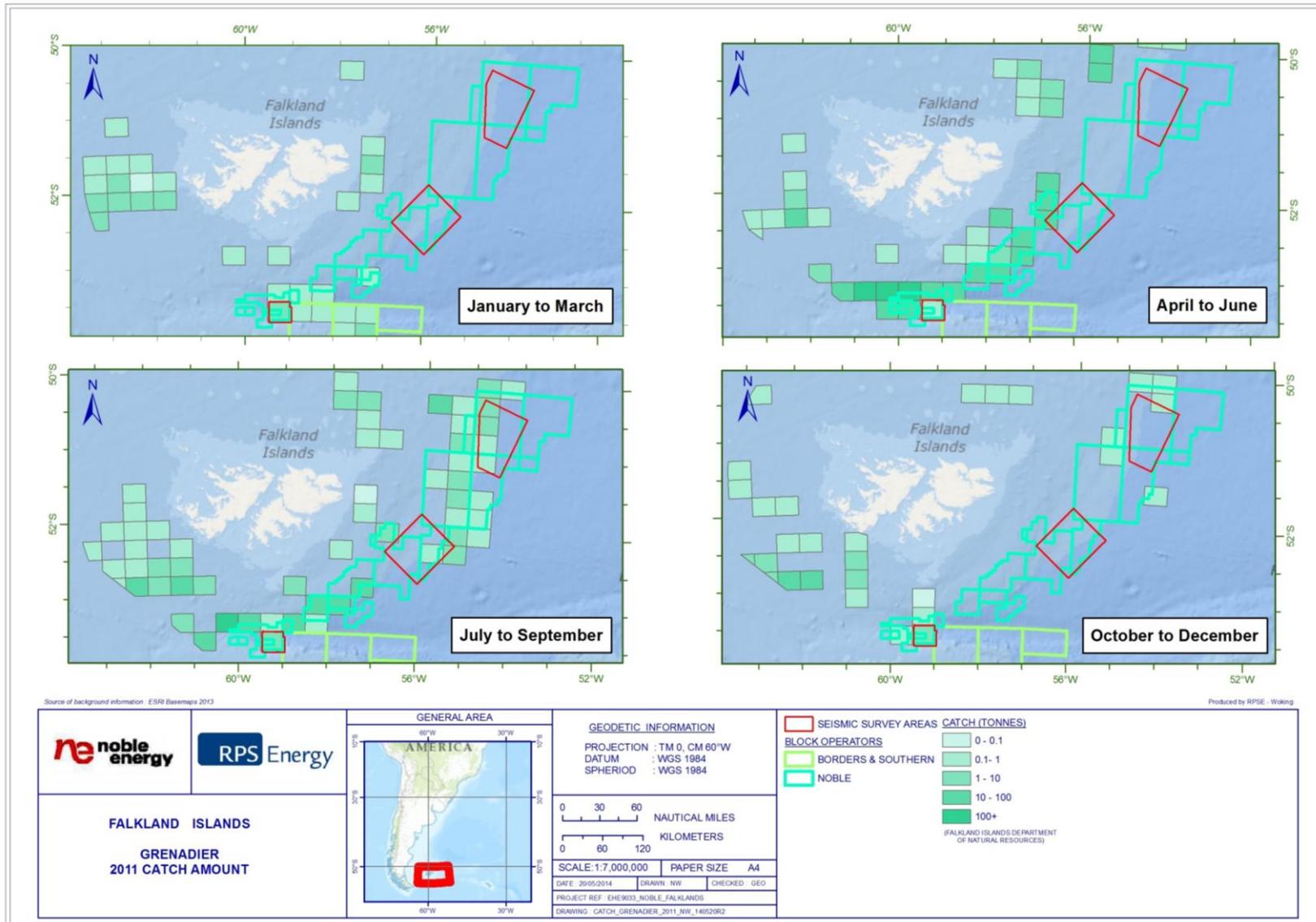


Figure E.2e: Fisheries catch mass (tonnes) for Grenadiers (*Macrouridae*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

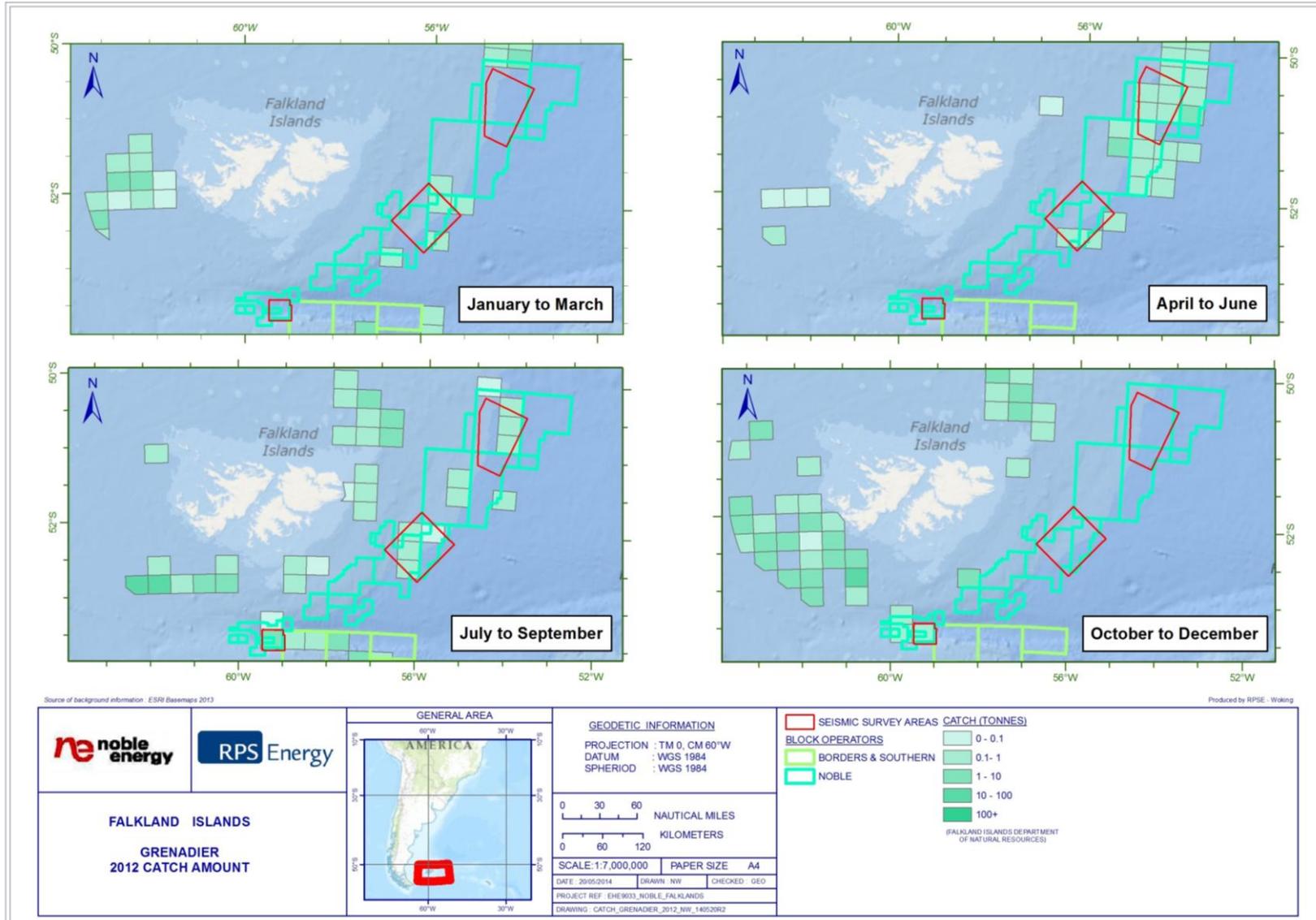


Figure E.2f: Fisheries catch mass (tonnes) for Grenadiers (*Macrouridae*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

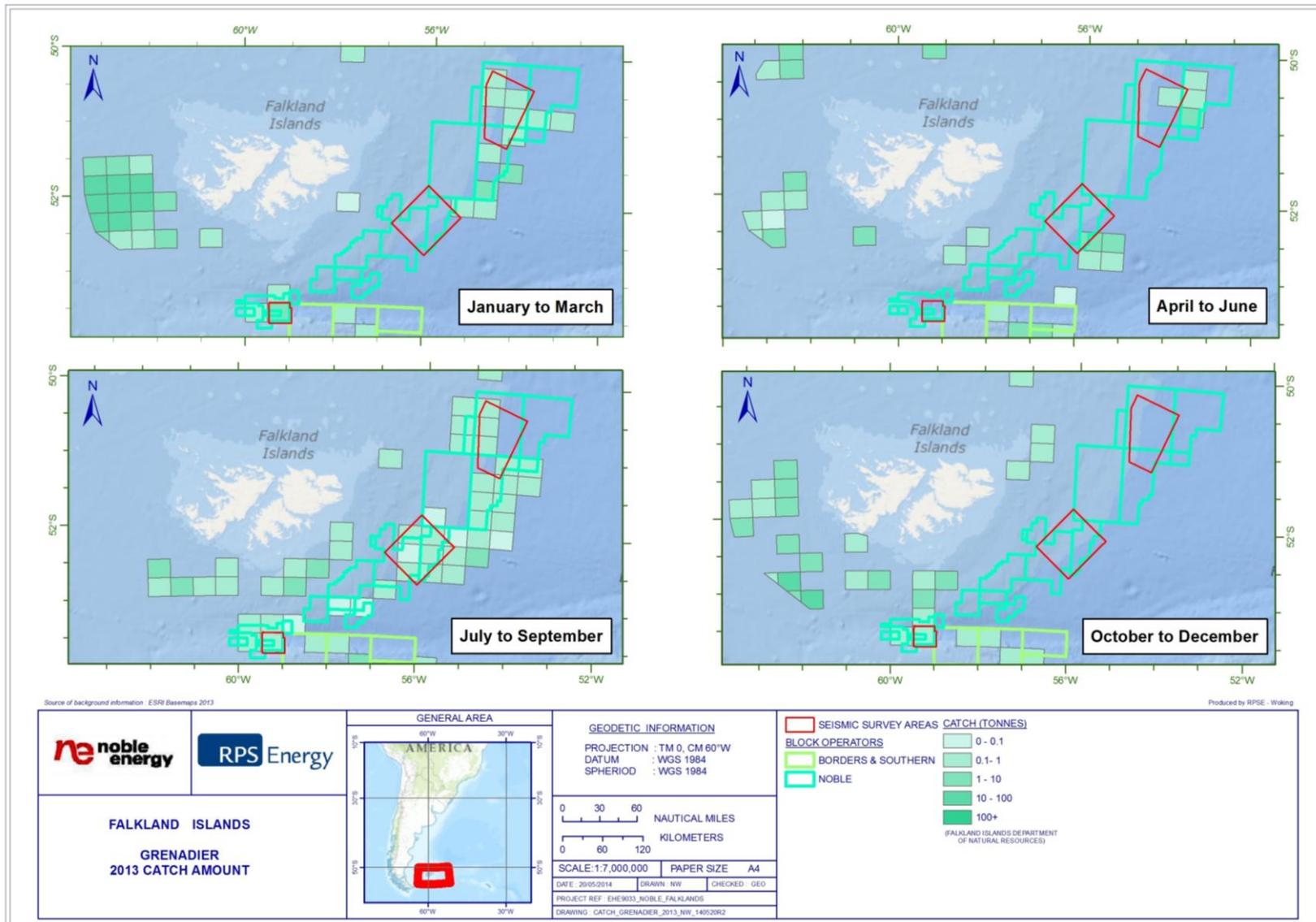


Figure E.3a: Fisheries catch mass (tonnes) for Hake (Merluccius sp.) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

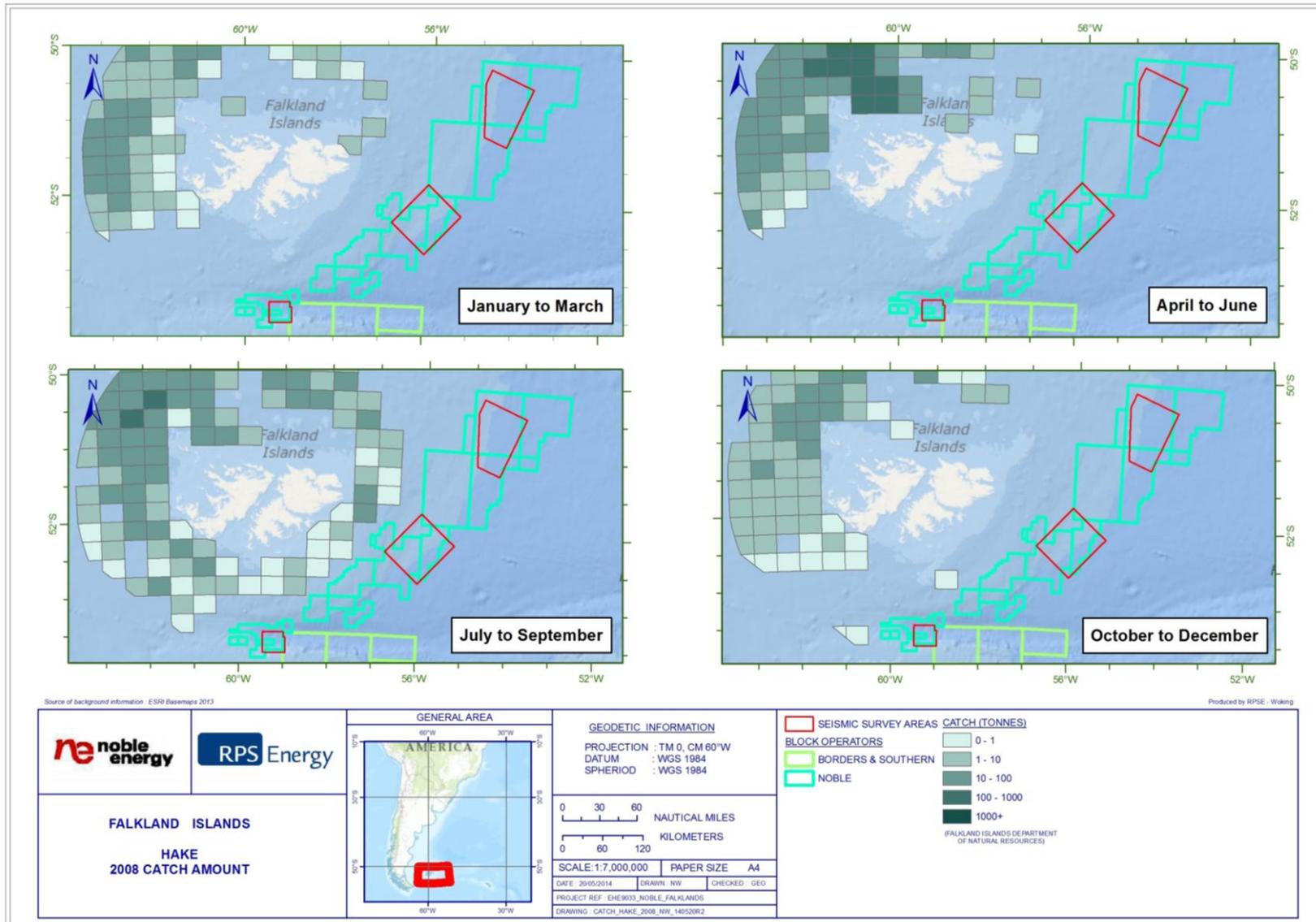


Figure E.3b: Fisheries catch mass (tonnes) for Hake (*Merluccius sp.*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

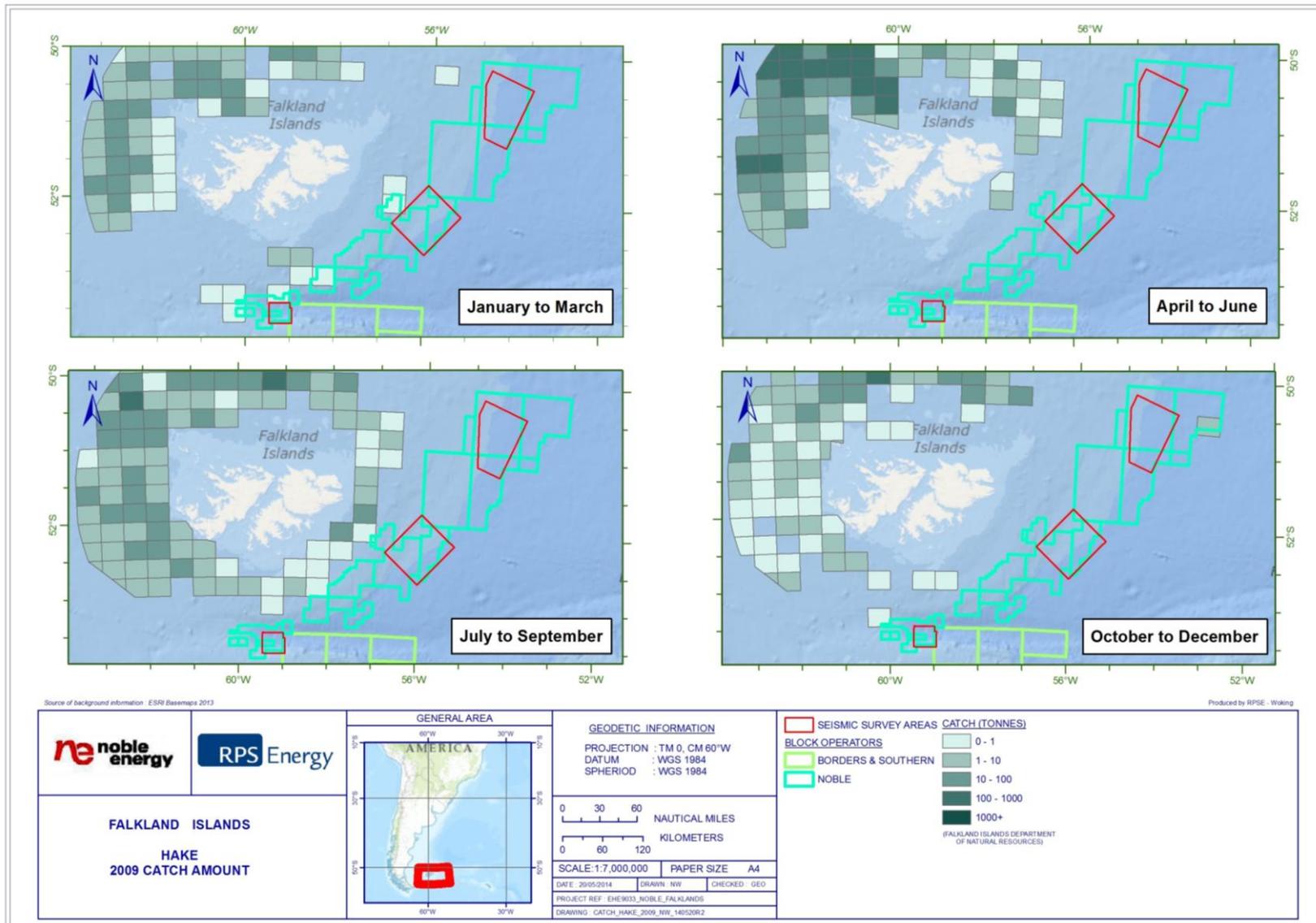


Figure E.3c: Fisheries catch mass (tonnes) for Hake (Merluccius sp.) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

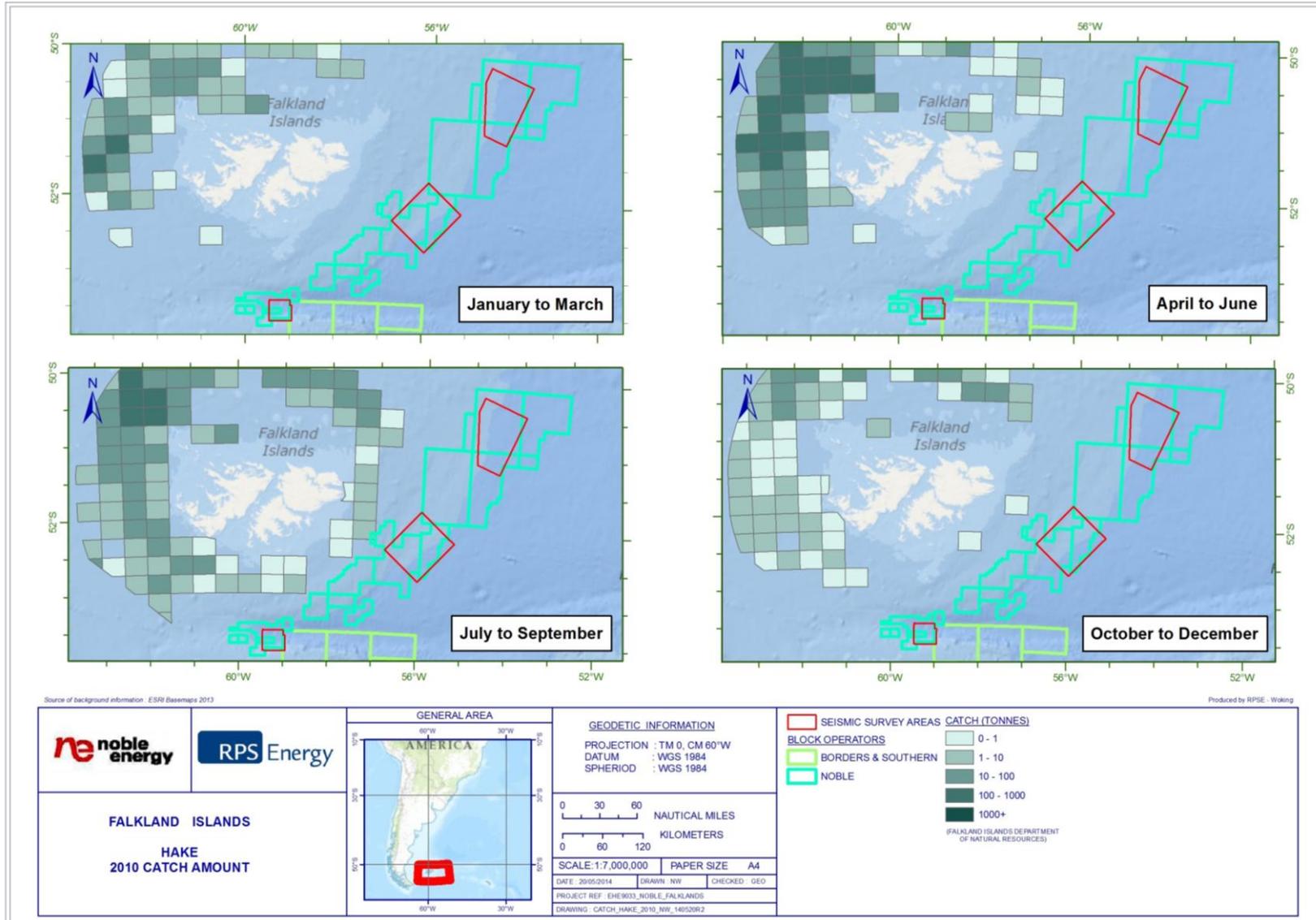


Figure E.3d: Fisheries catch mass (tonnes) for Hake (*Merluccius sp.*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

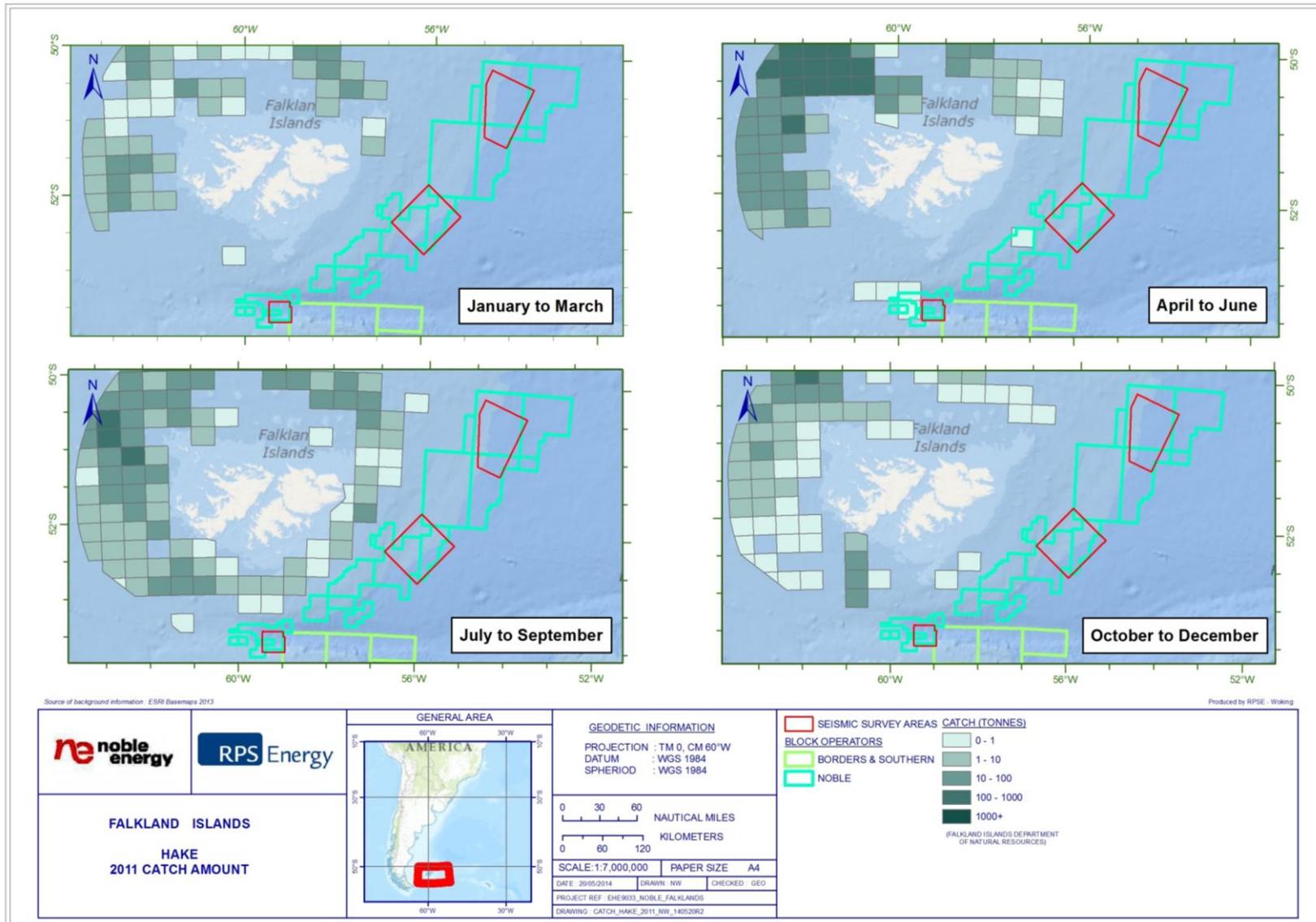


Figure E.3e: Fisheries catch mass (tonnes) for Hake (Merluccius sp.) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

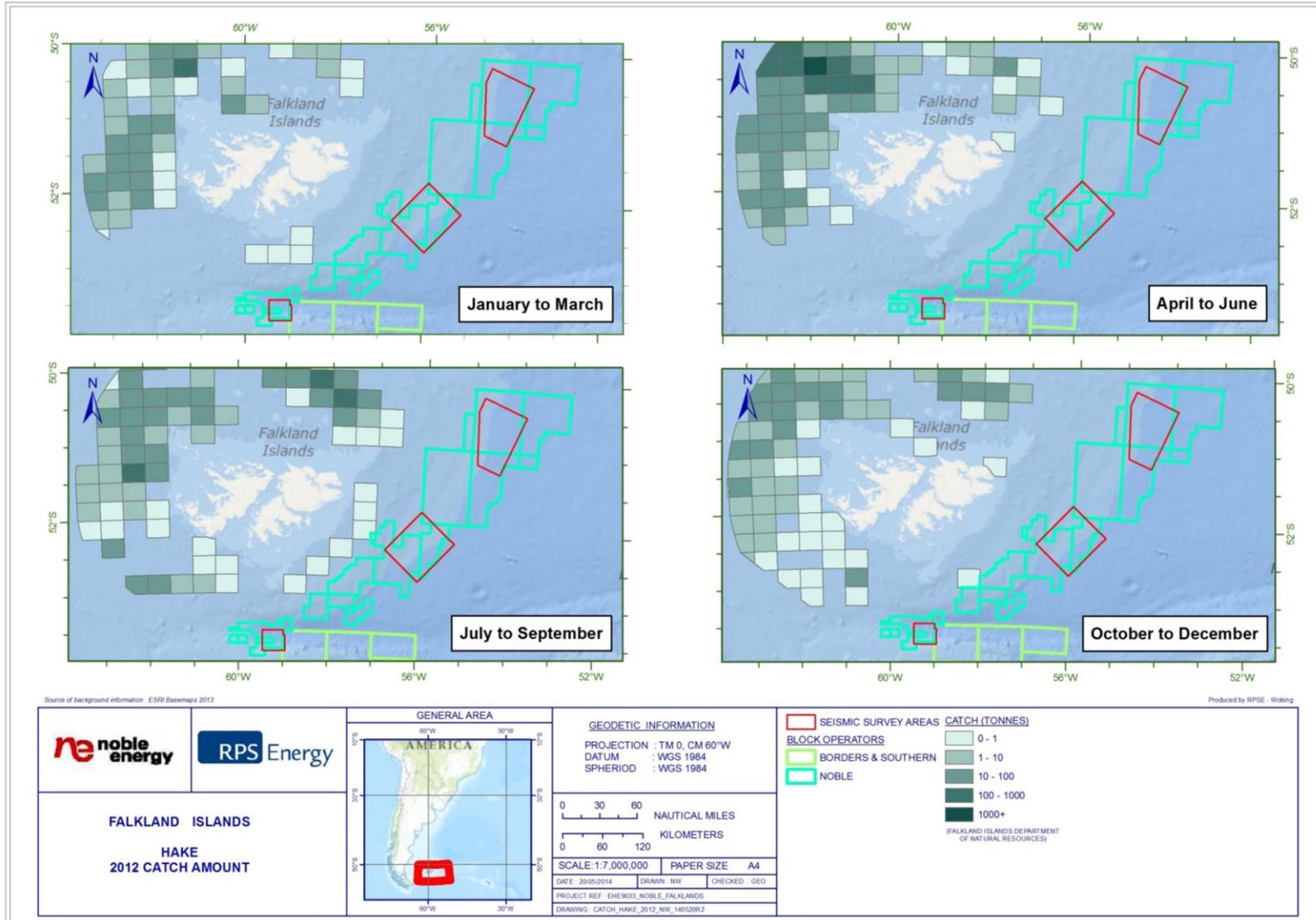


Figure E.3f: Fisheries catch mass (tonnes) for Hake (*Merluccius sp.*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

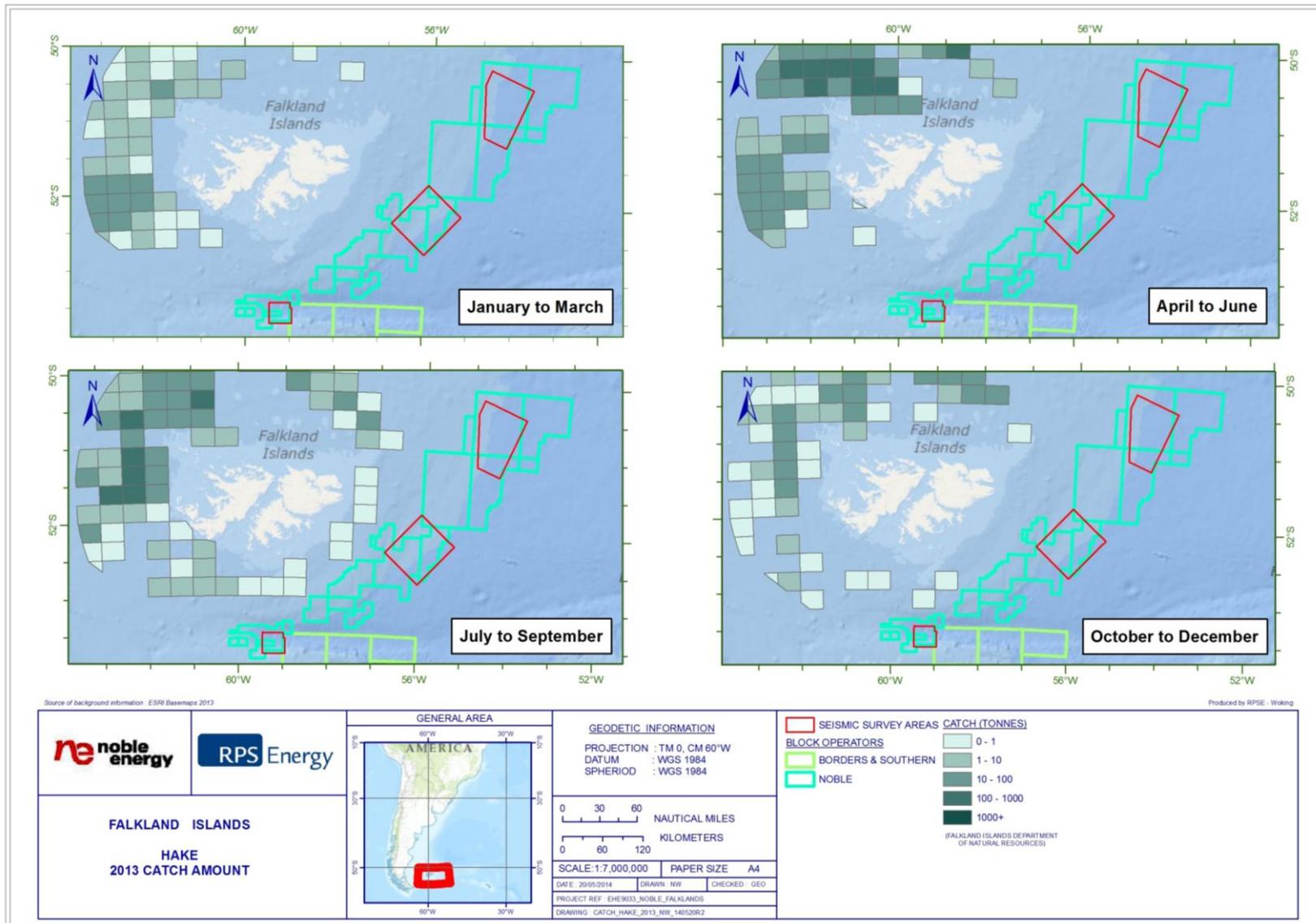


Figure E.4a: Fisheries catch mass (tonnes) for Hoki (*Macrurus magellanicus*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

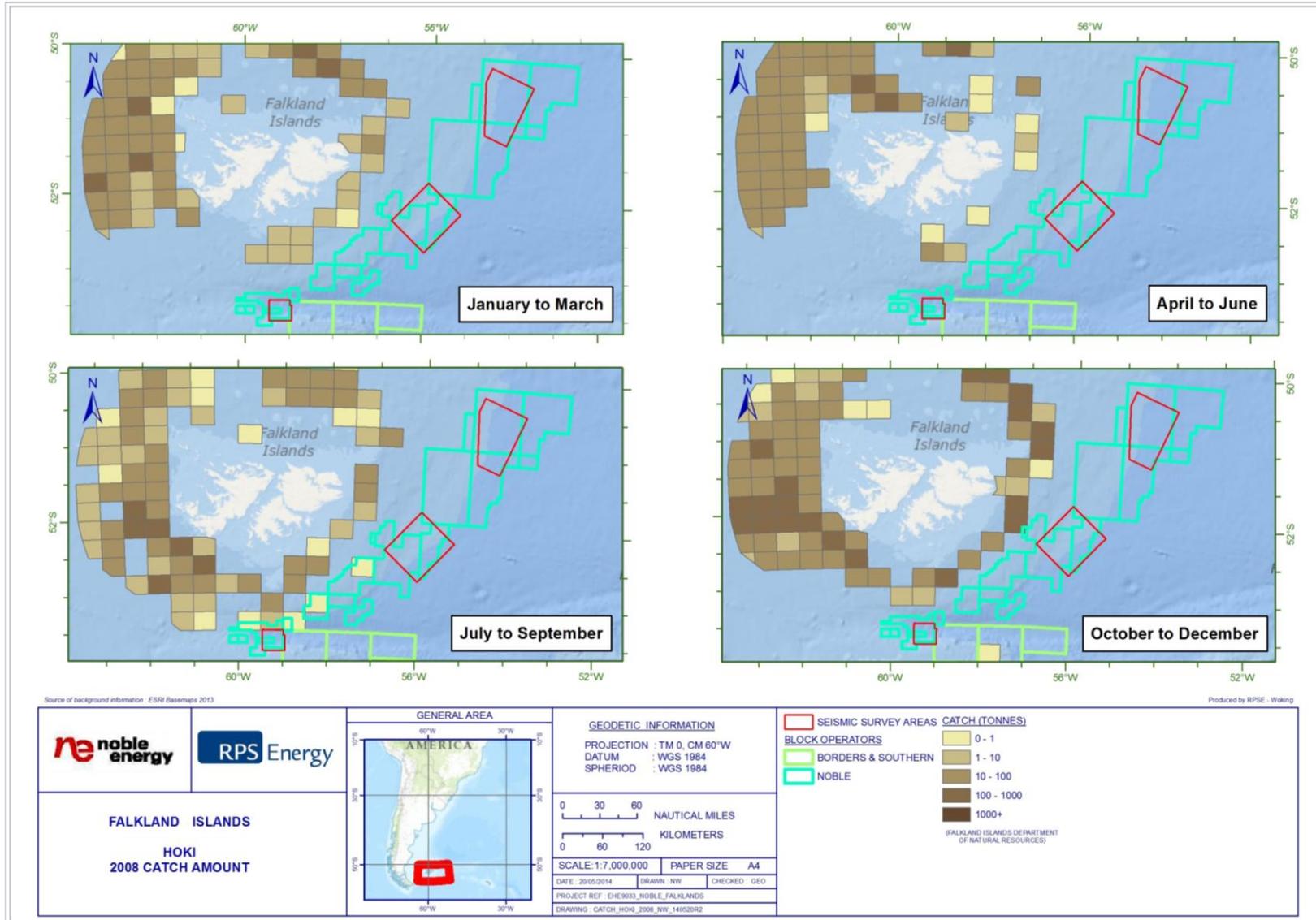


Figure E.4b: Fisheries catch mass (tonnes) for Hoki (*Macruronus magellanicus*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

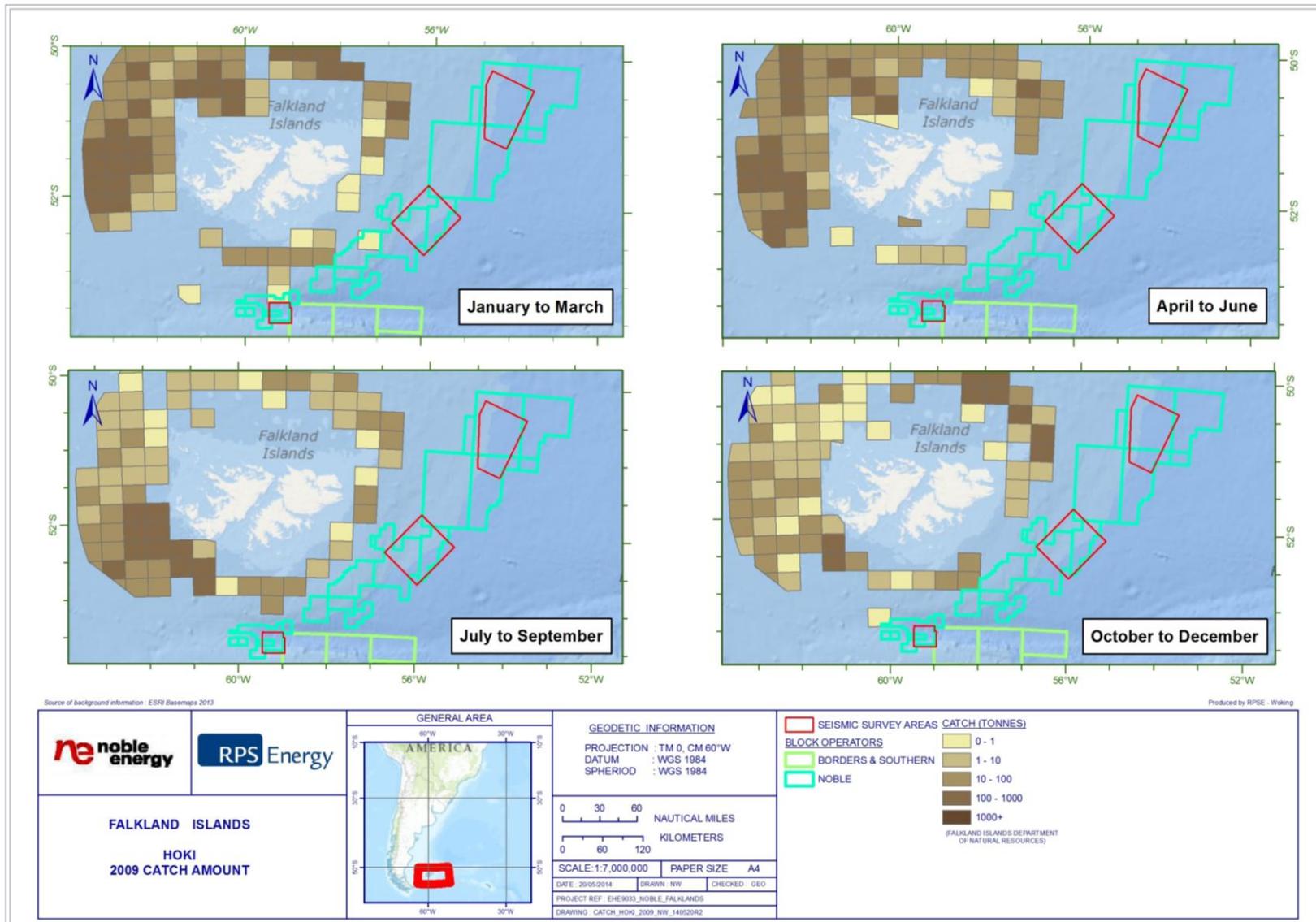


Figure E.4c: Fisheries catch mass (tonnes) for Hoki (*Macrurus magellanicus*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

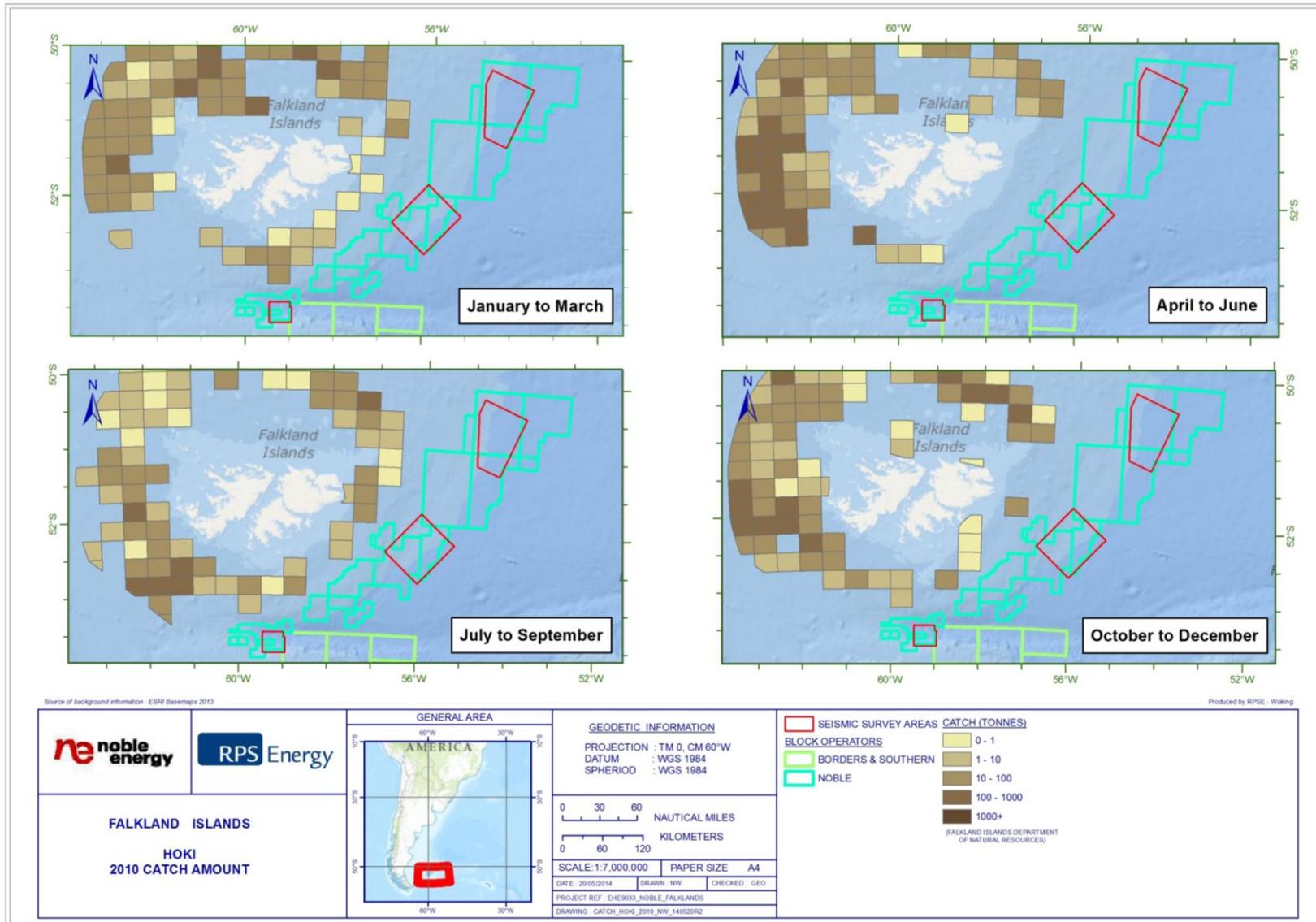


Figure E.4d: Fisheries catch mass (tonnes) for Hoki (*Macruronus magellanicus*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

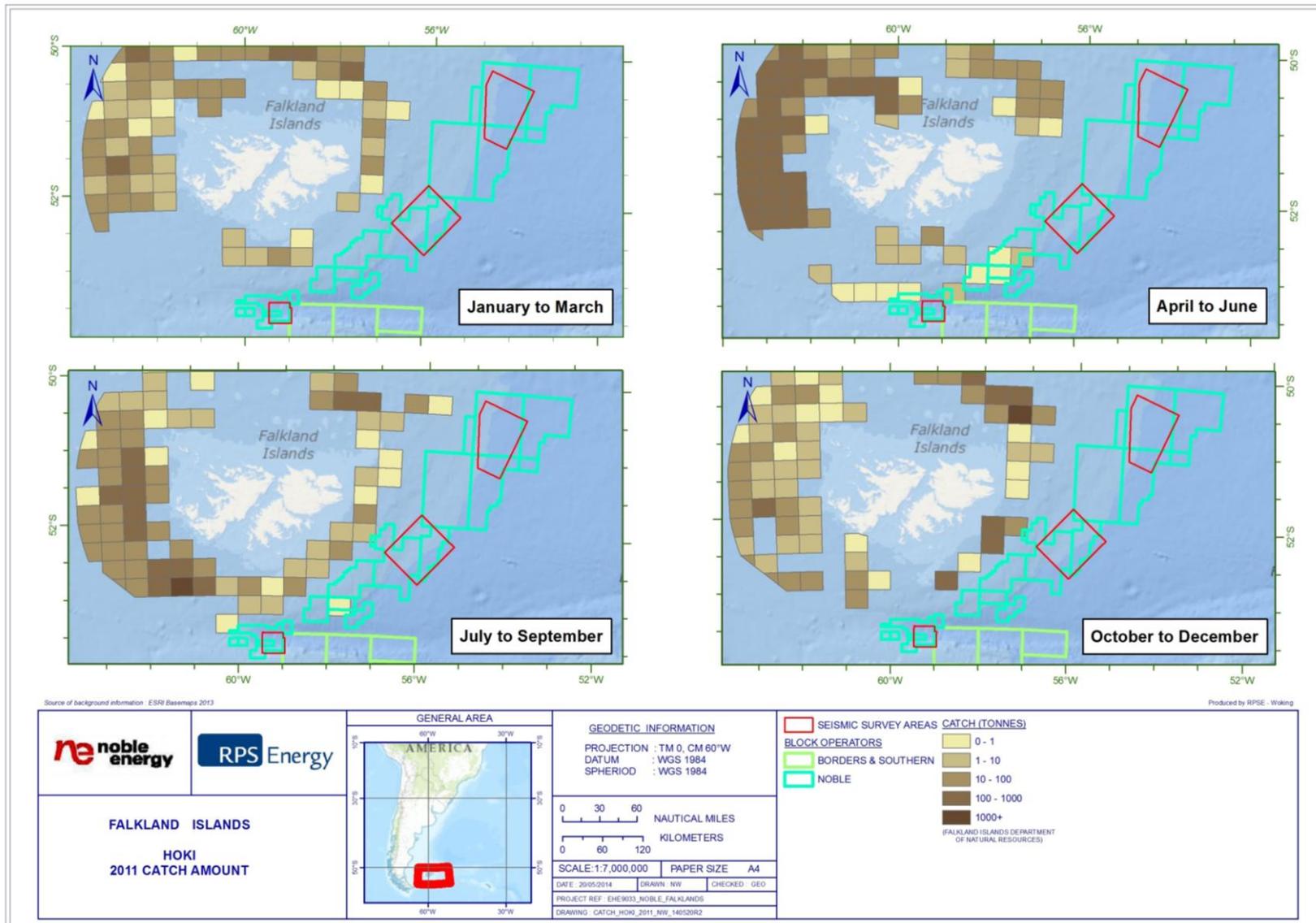


Figure E.4e: Fisheries catch mass (tonnes) for Hoki (*Macrurus magellanicus*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

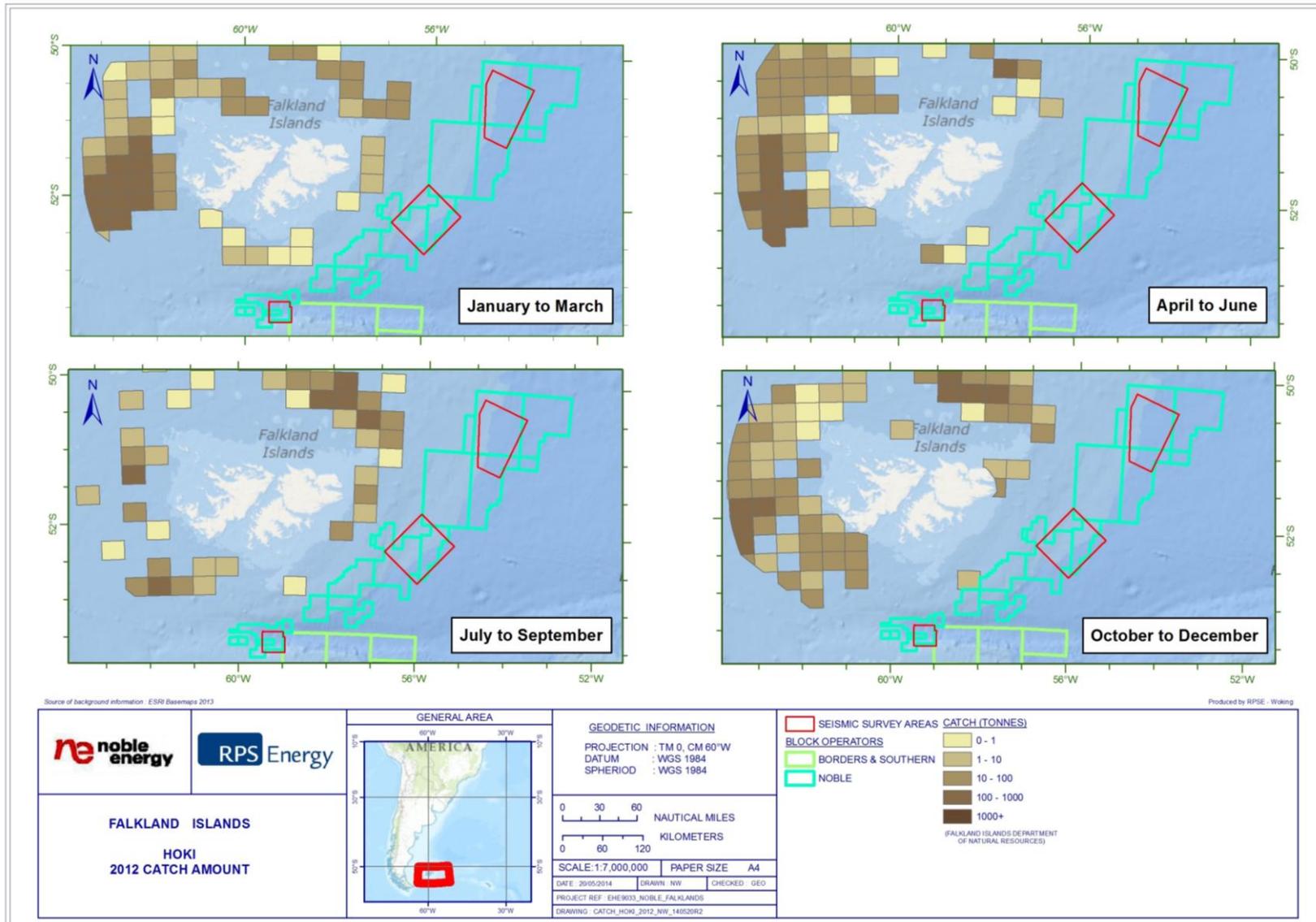


Figure E.4f: Fisheries catch mass (tonnes) for Hoki (*Macrurus magellanicus*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

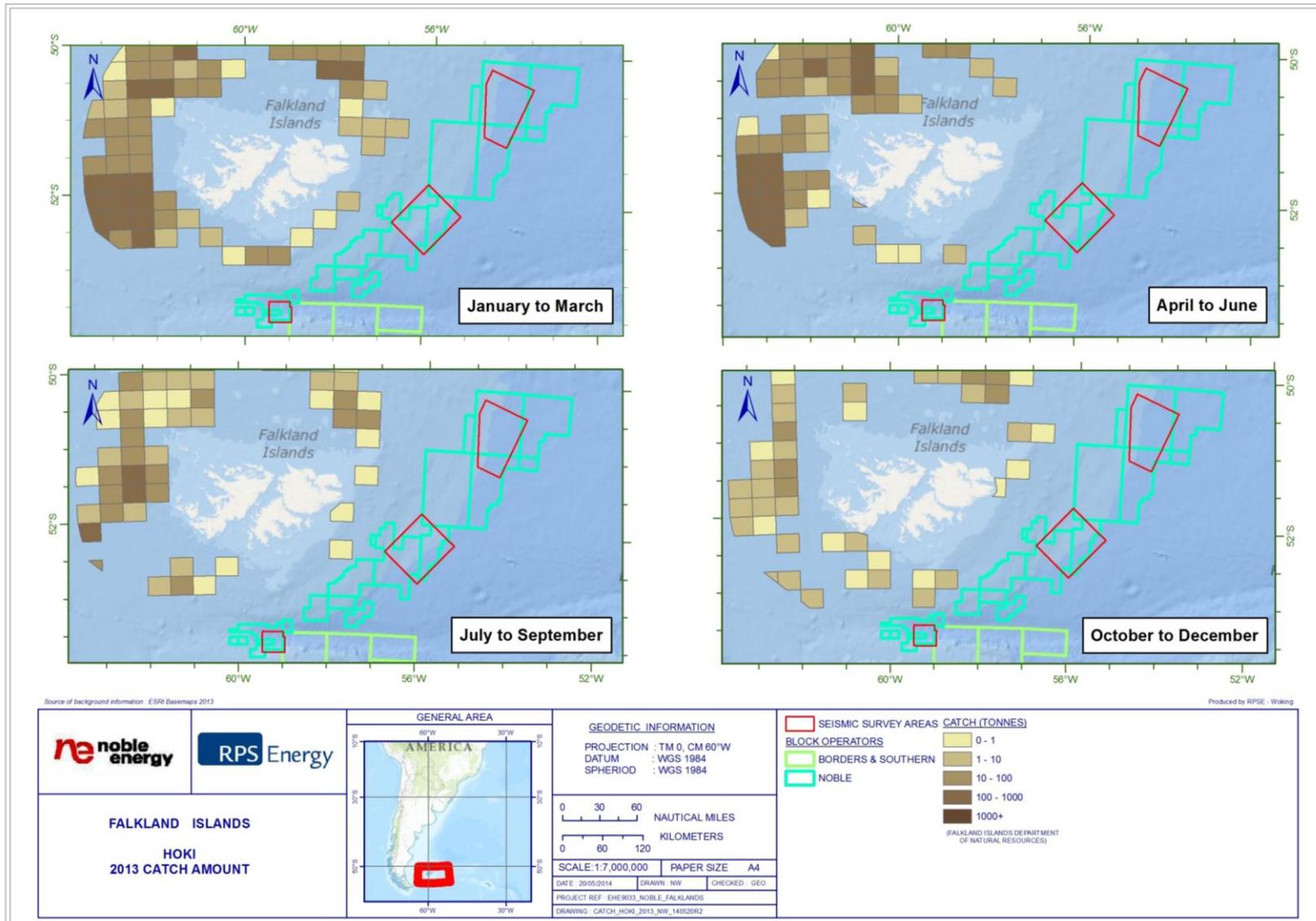


Figure E.5a: Fisheries catch mass (tonnes) for Rays (Rajidae) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

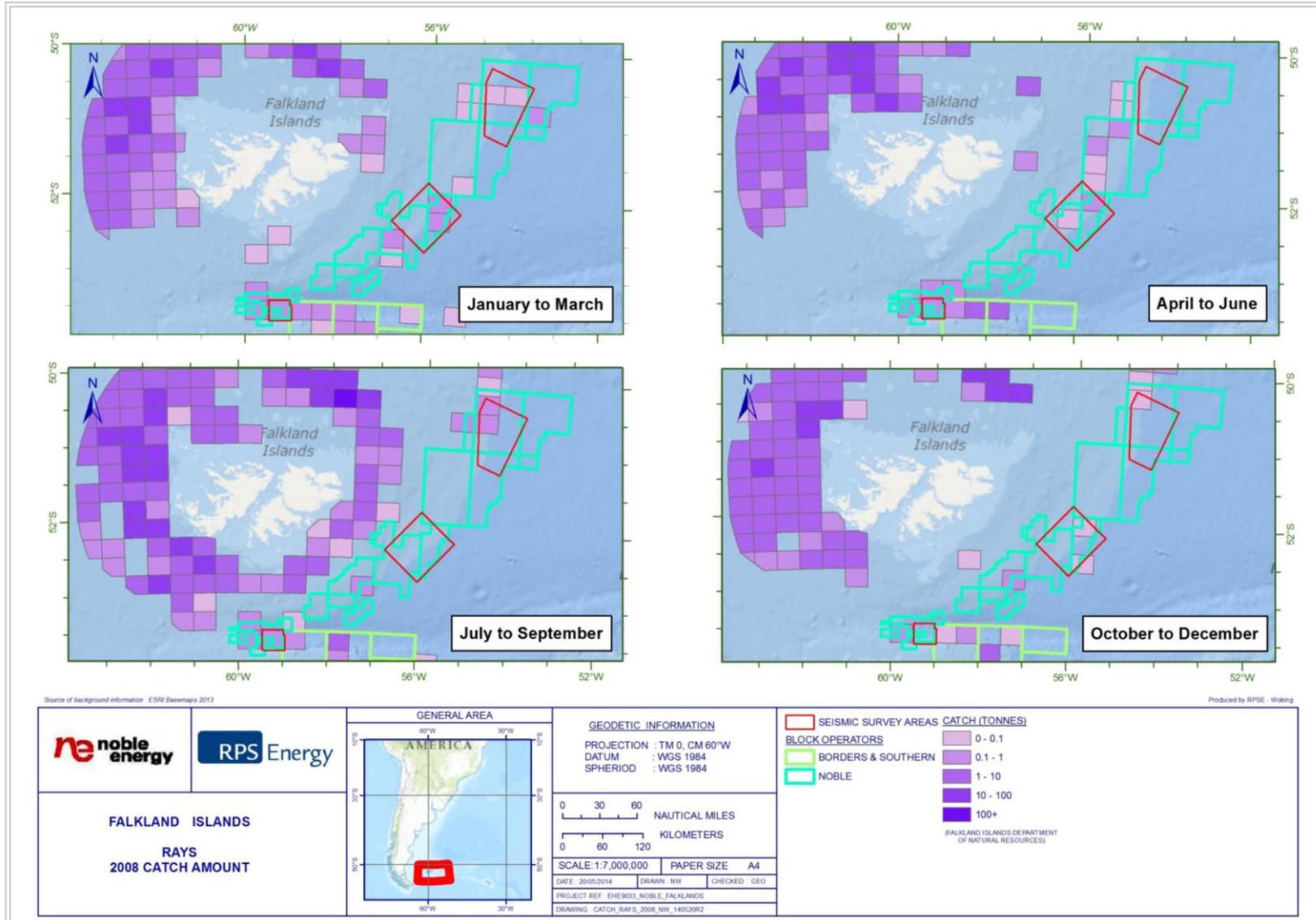


Figure E.5b: Fisheries catch mass (tonnes) for Rays (Rajidae) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

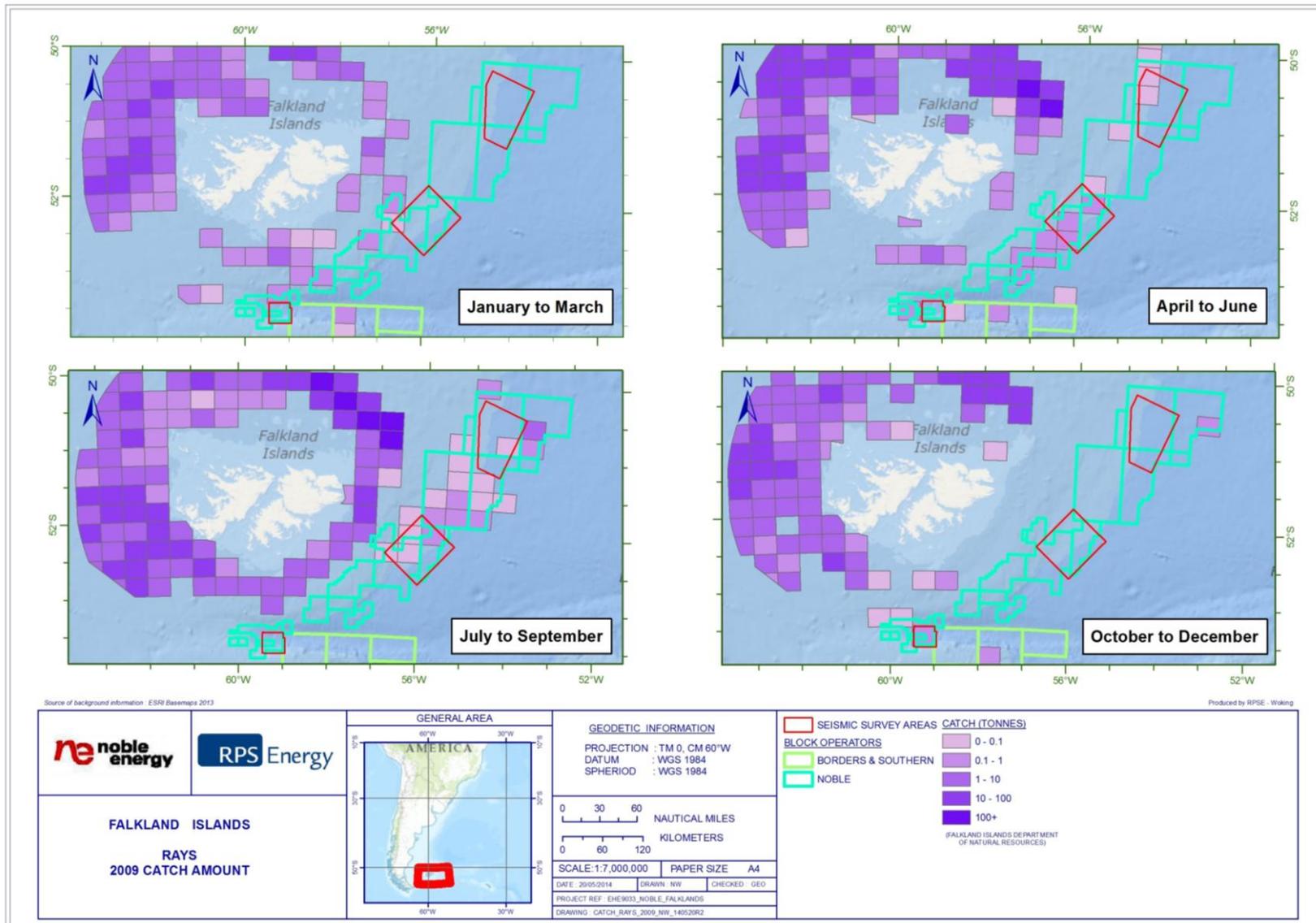


Figure E.5c: Fisheries catch mass (tonnes) for Rays (Rajidae) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

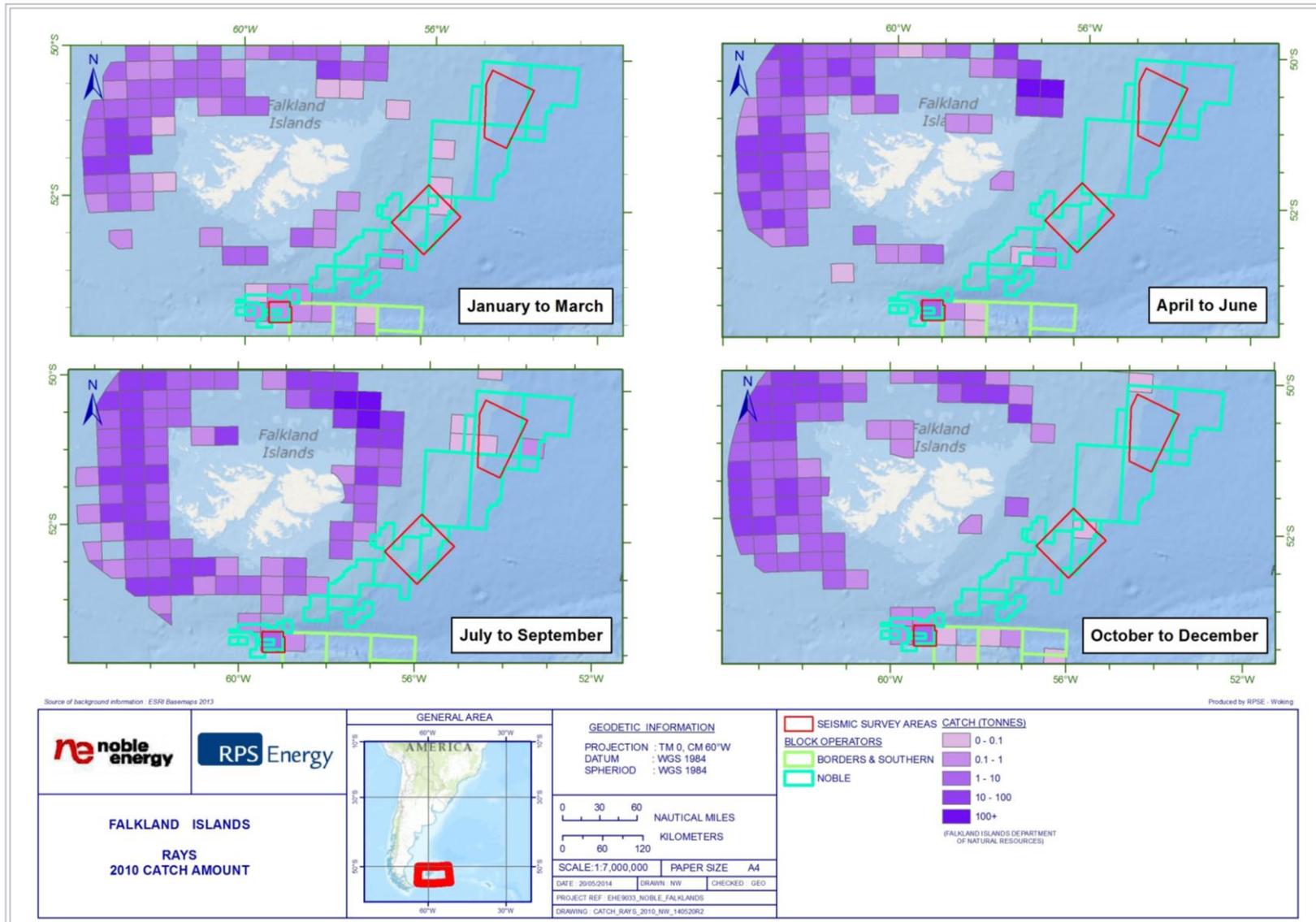


Figure E.5d: Fisheries catch mass (tonnes) for Rays (Rajidae) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

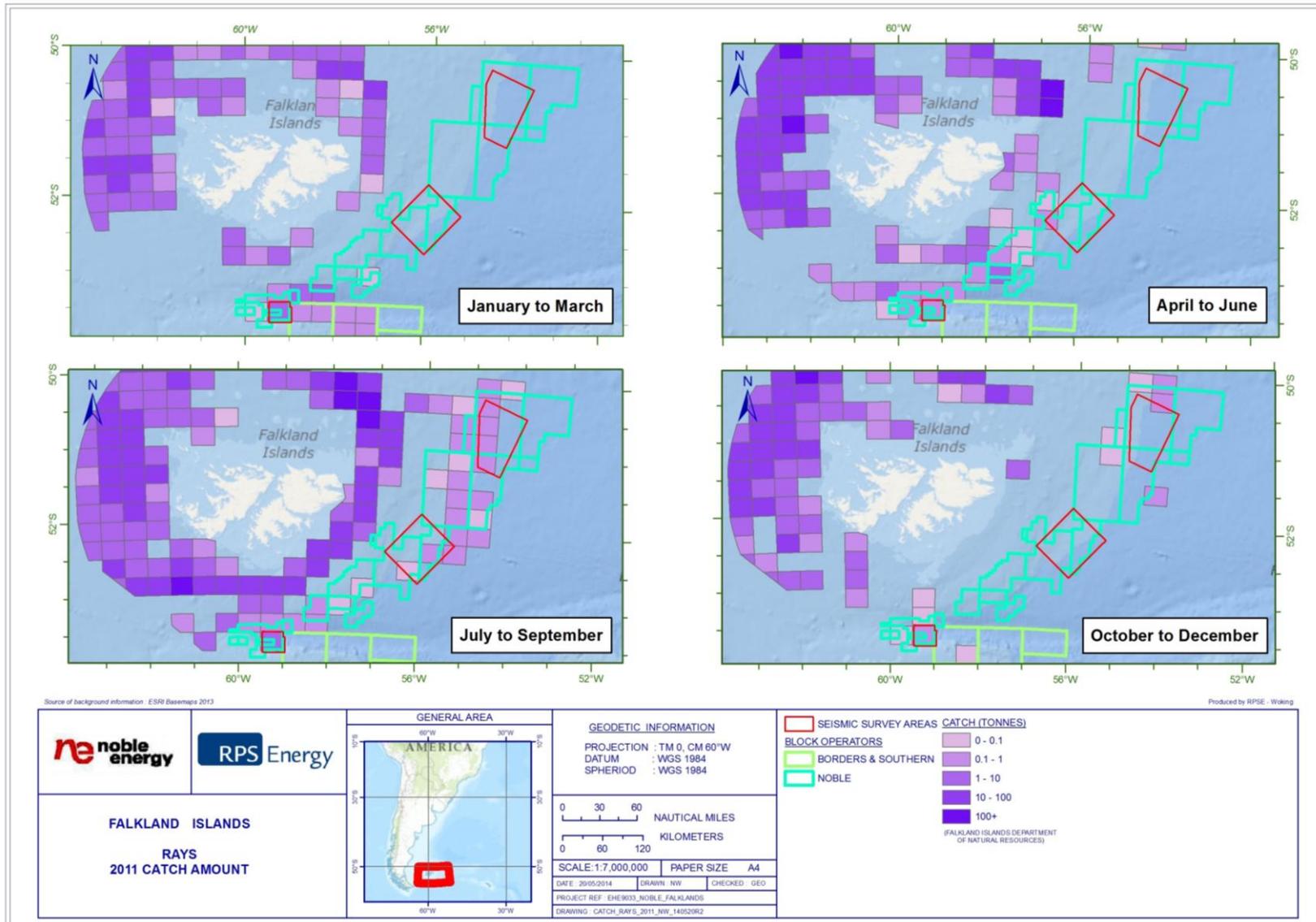


Figure E.5e: Fisheries catch mass (tonnes) for Rays (Rajidae) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

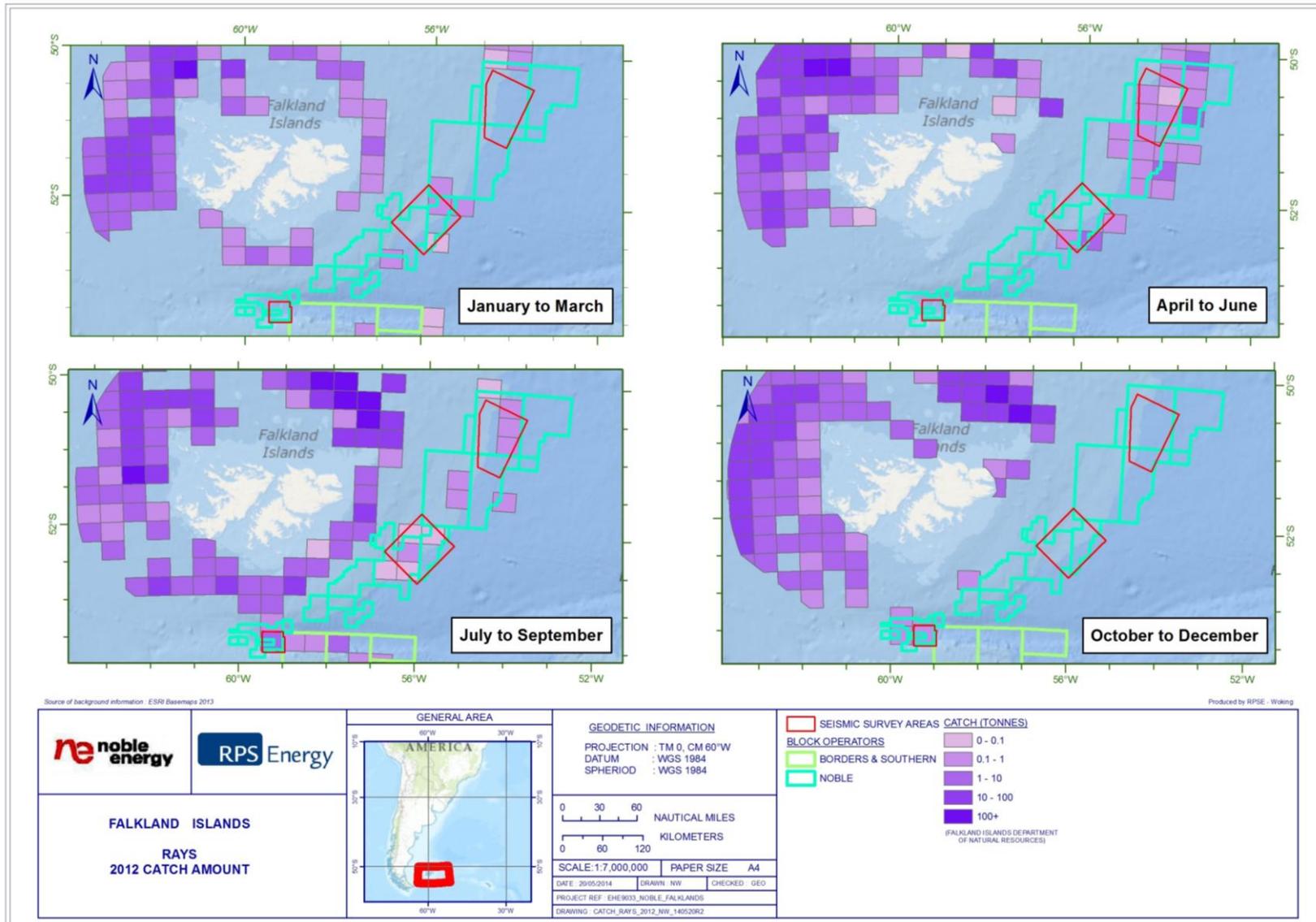


Figure E.5f: Fisheries catch mass (tonnes) for Rays (Rajidae) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

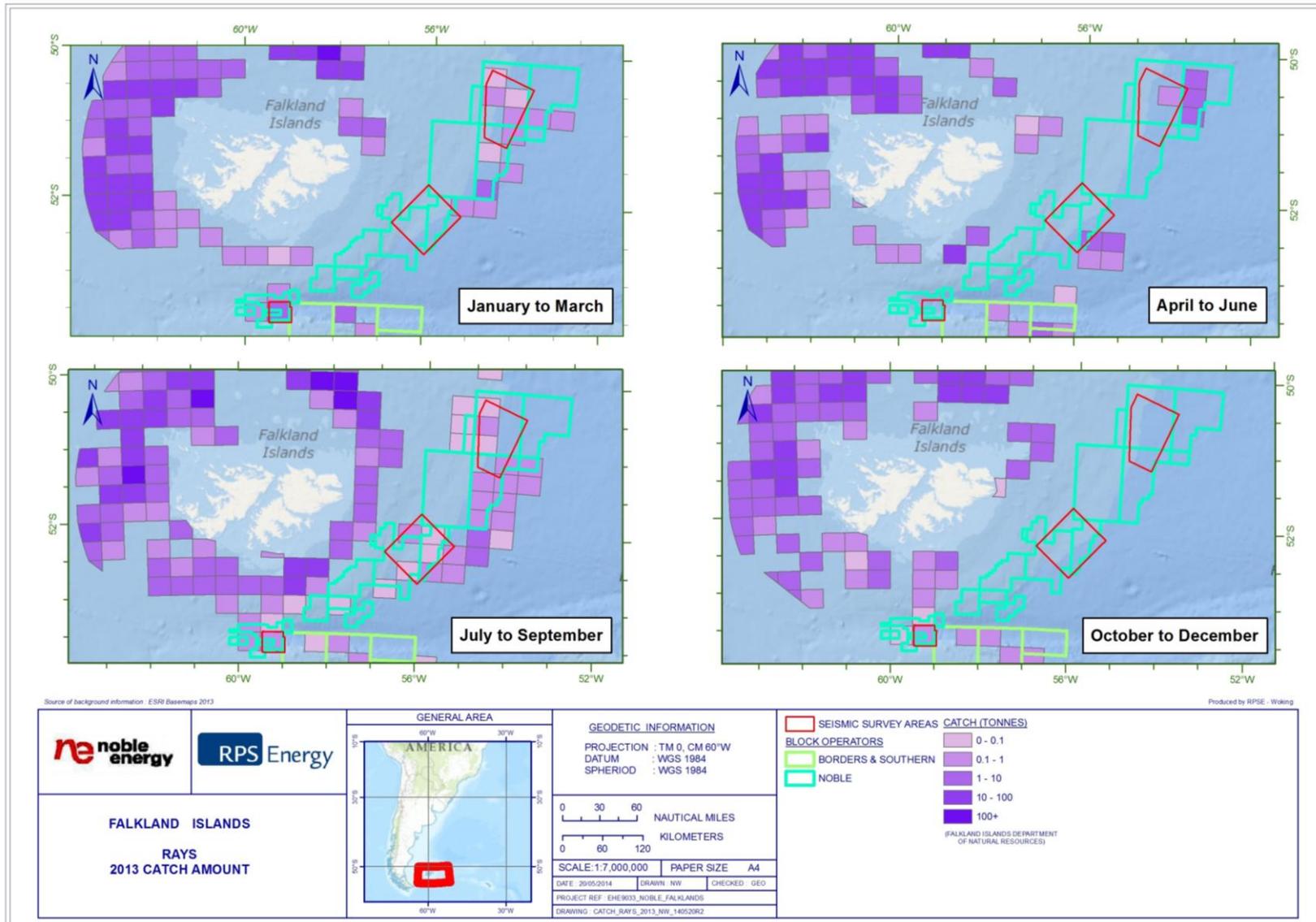


Figure E.6a: Fisheries catch mass (tonnes) for Red Cod (*Salilota australis*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

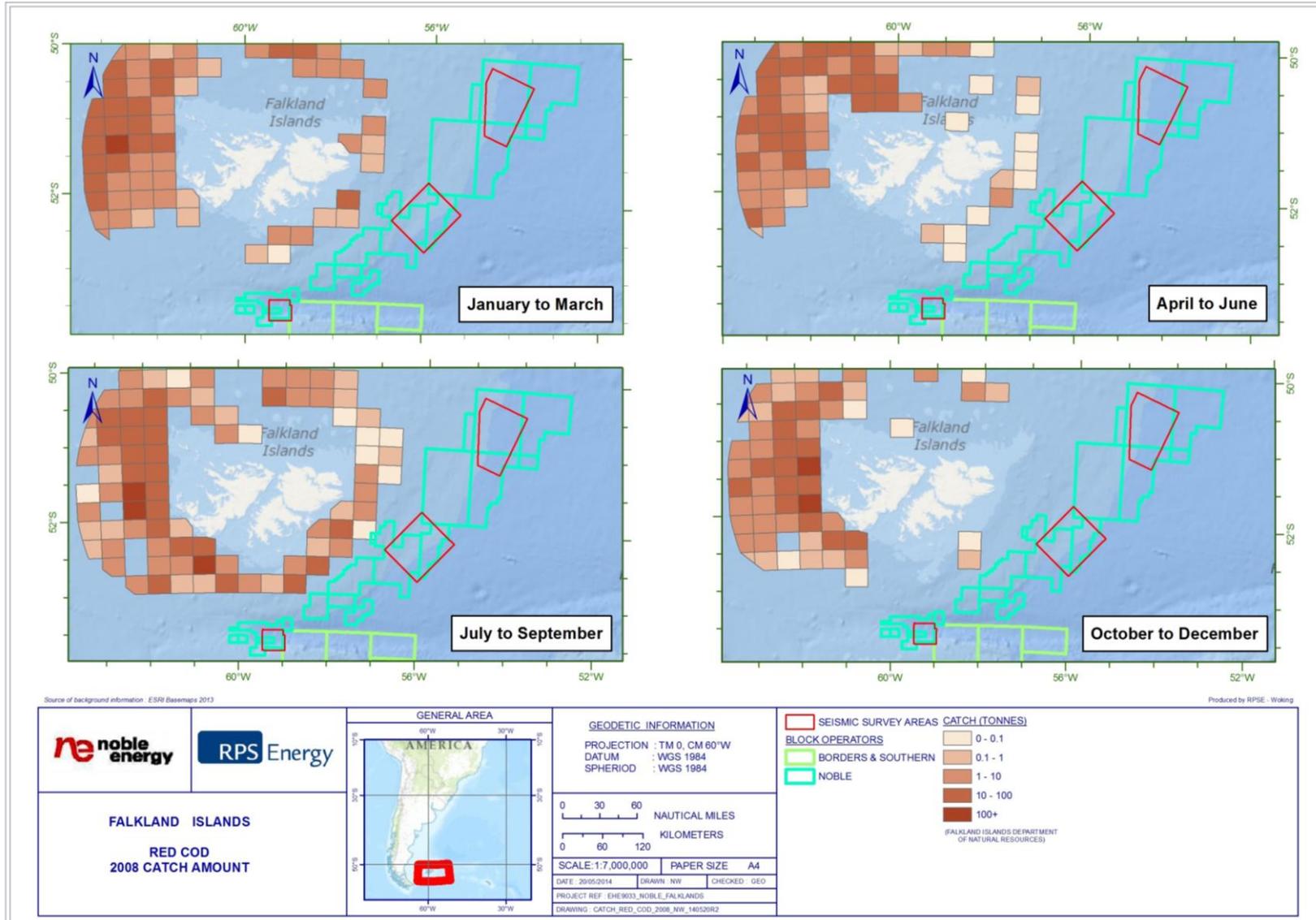


Figure E.6b: Fisheries catch mass (tonnes) for Red Cod (*Salilota australis*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

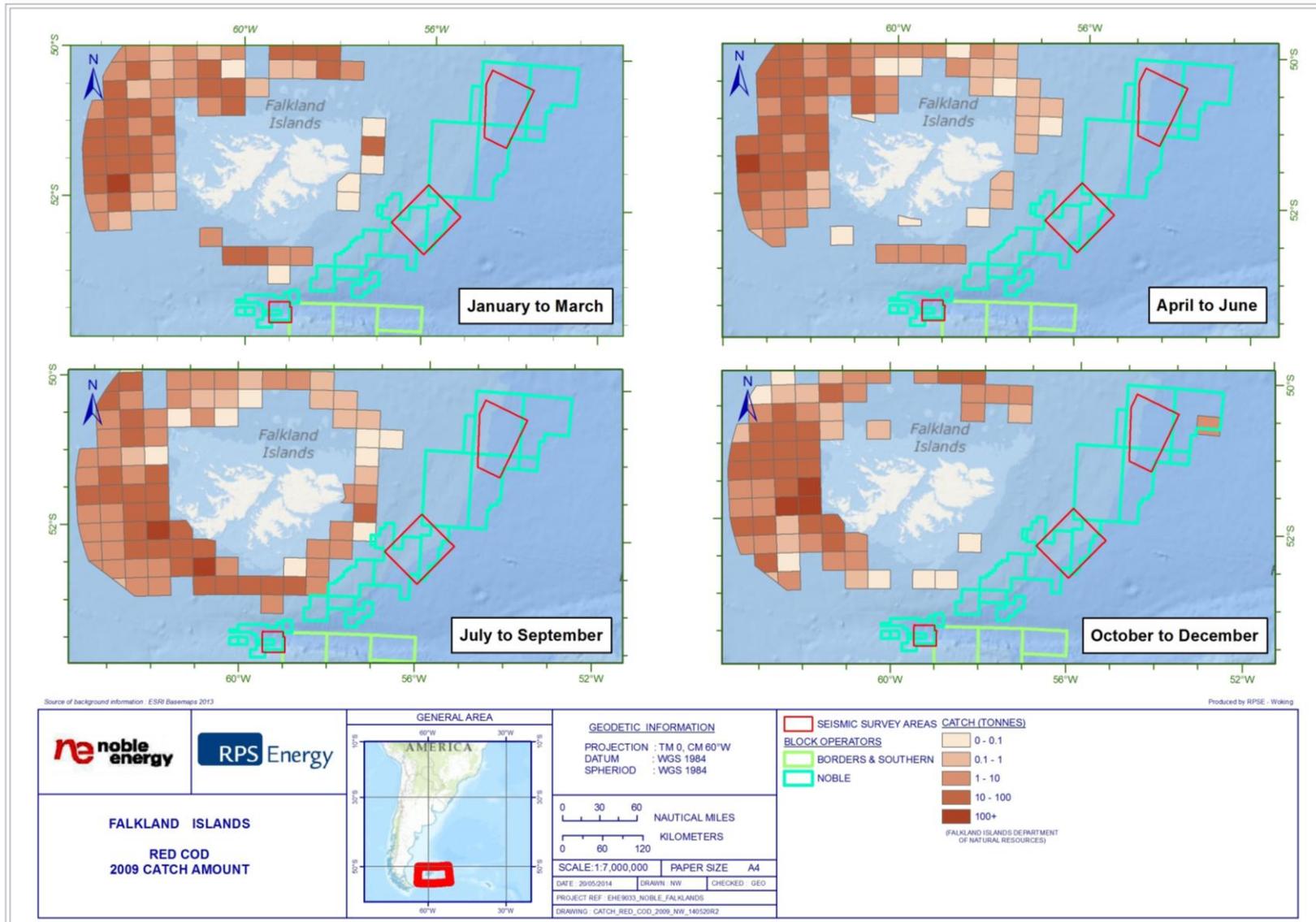


Figure E.6c: Fisheries catch mass (tonnes) for Red Cod (*Salilota australis*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

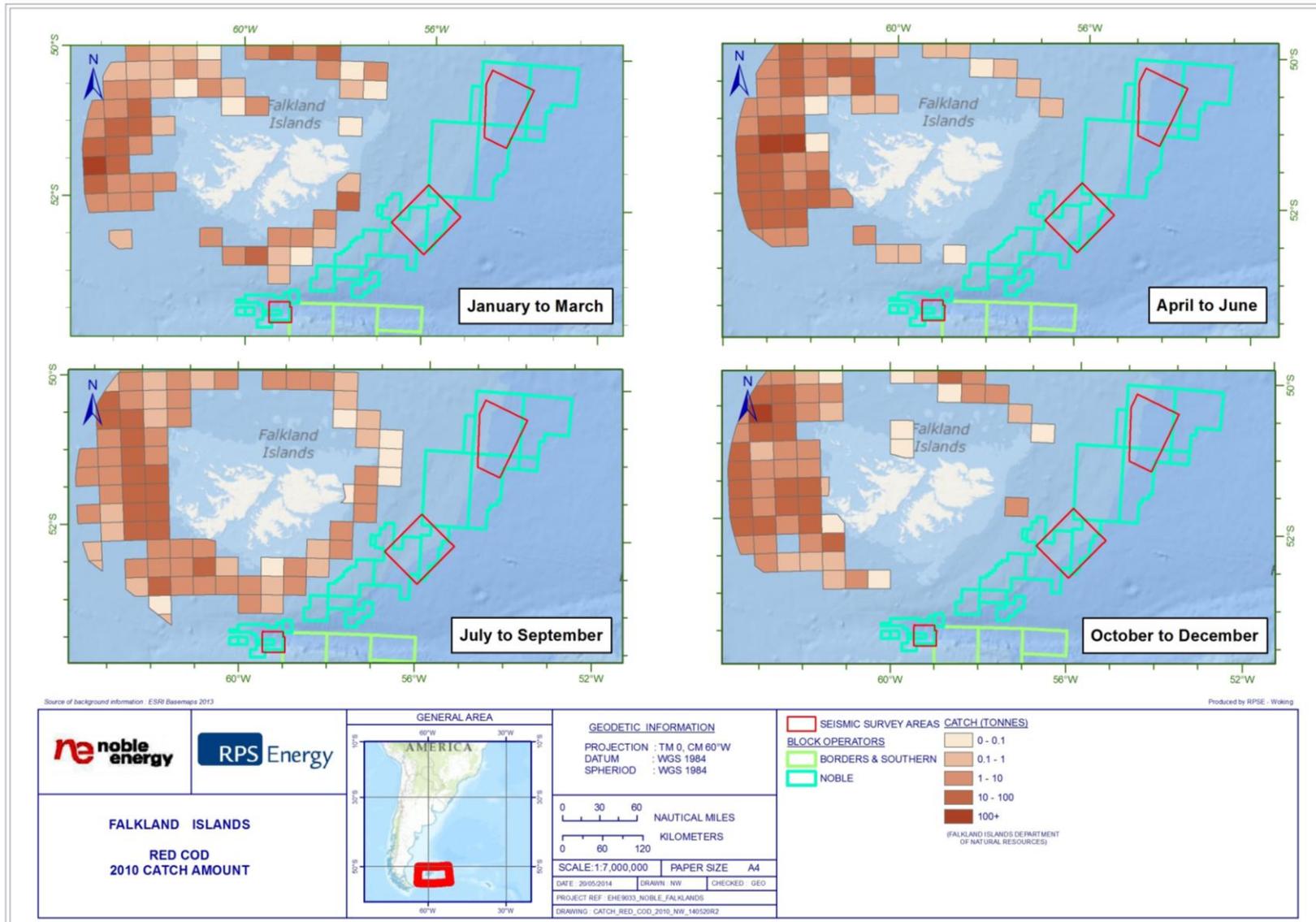


Figure E.6d: Fisheries catch mass (tonnes) for Red Cod (*Salilota australis*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

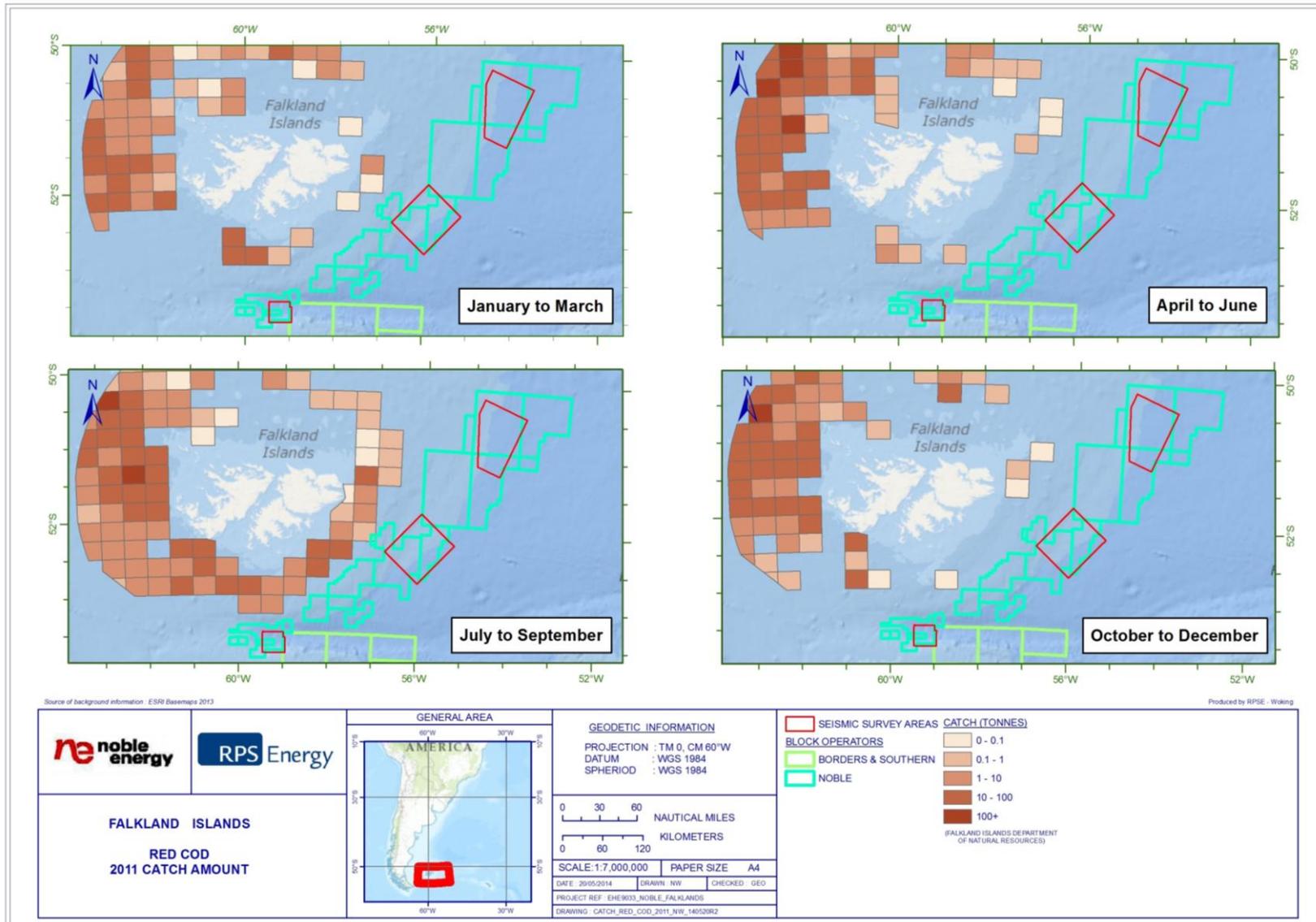


Figure E.6e: Fisheries catch mass (tonnes) for Red Cod (*Salilota australis*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

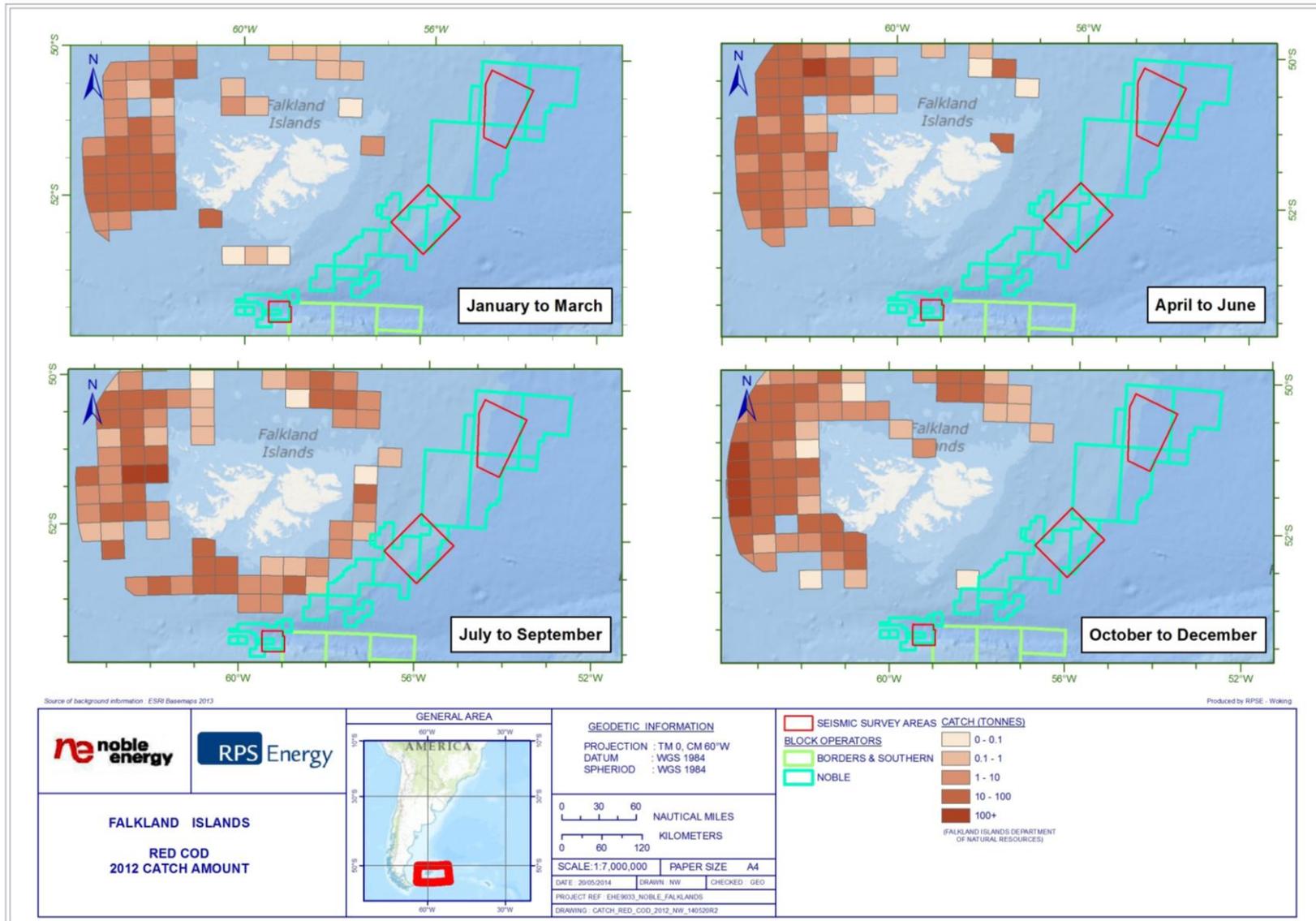


Figure E.6f: Fisheries catch mass (tonnes) for Red Cod (*Salilota australis*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

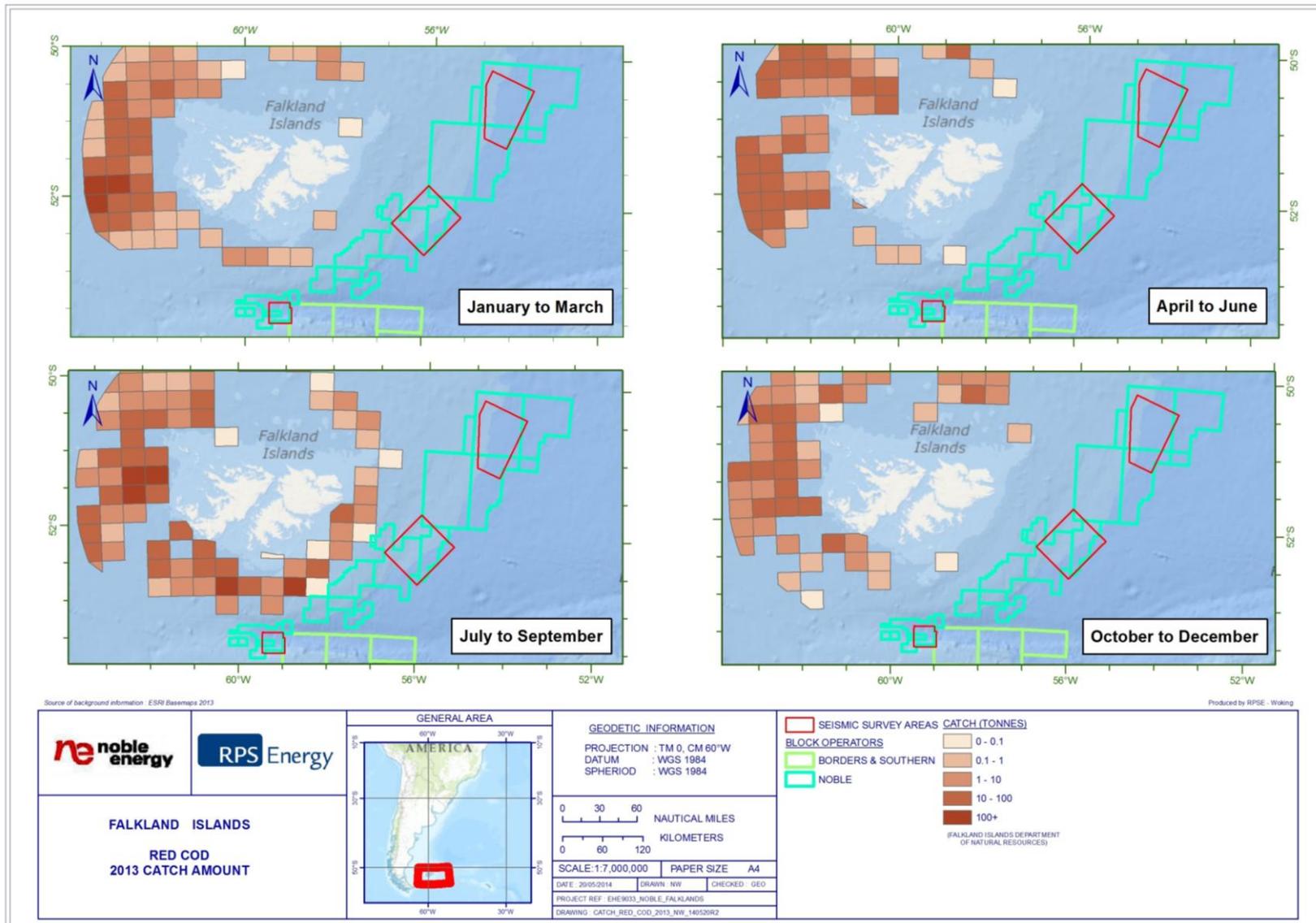


Figure E.7a: Fisheries catch mass (tonnes) for Rock Cod (*Patagonotothen ramsayi*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

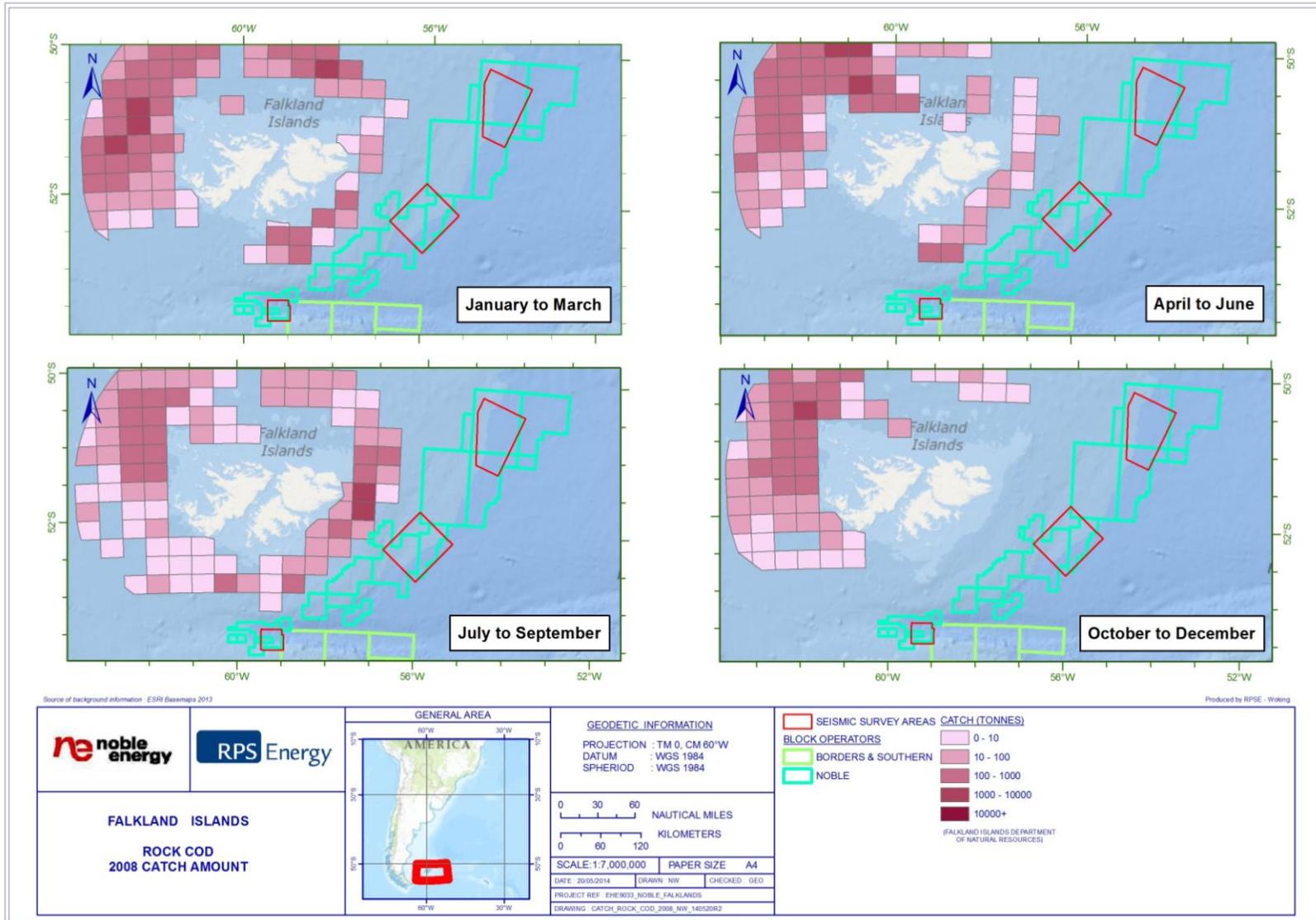


Figure E.7b: Fisheries catch mass (tonnes) for Rock Cod (*Patagonotothen ramsayi*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

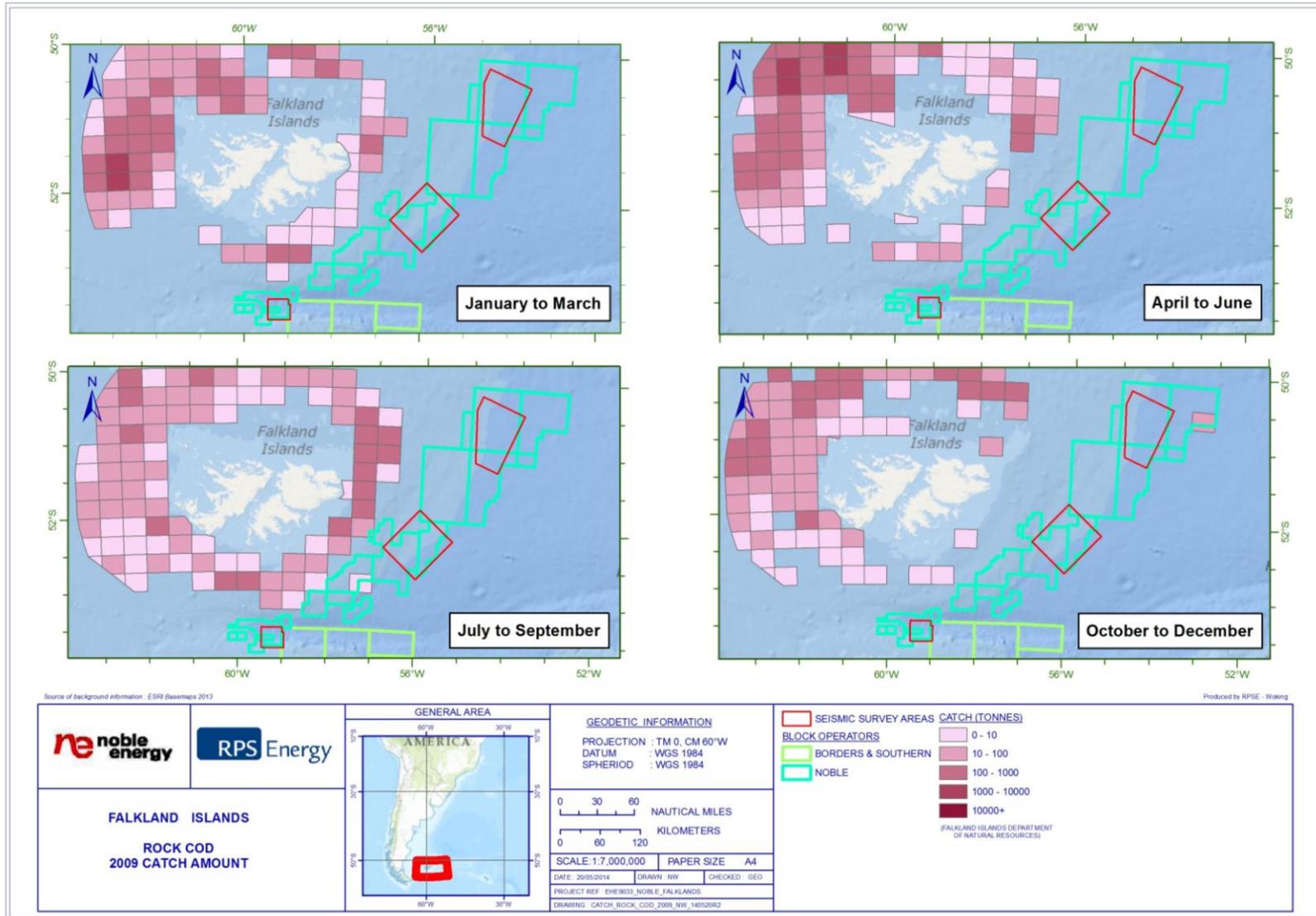


Figure E.7c: Fisheries catch mass (tonnes) for Rock Cod (*Patagonotothen ramsayi*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

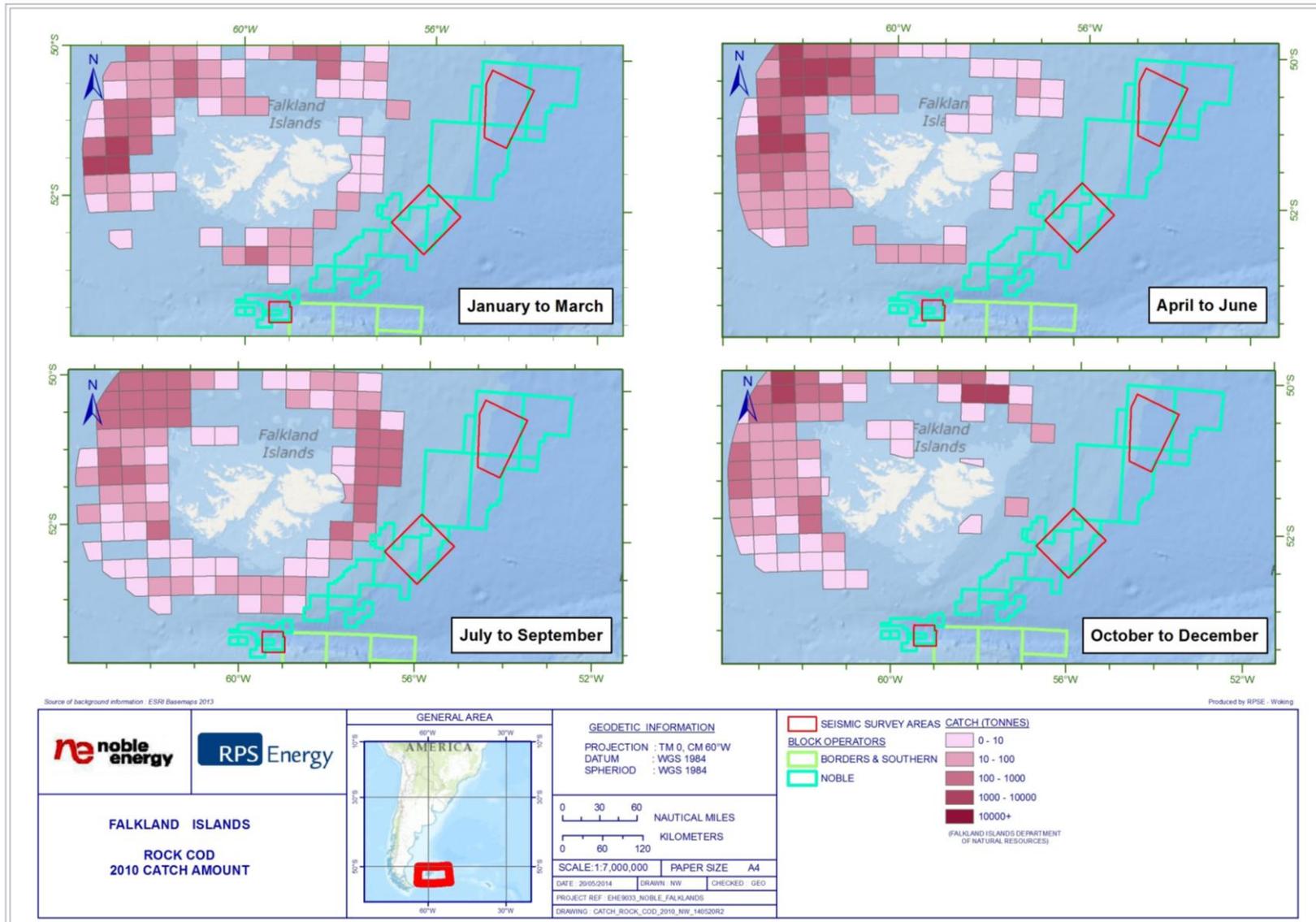


Figure E.7d: Fisheries catch mass (tonnes) for Rock Cod (*Patagonotothen ramsayi*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

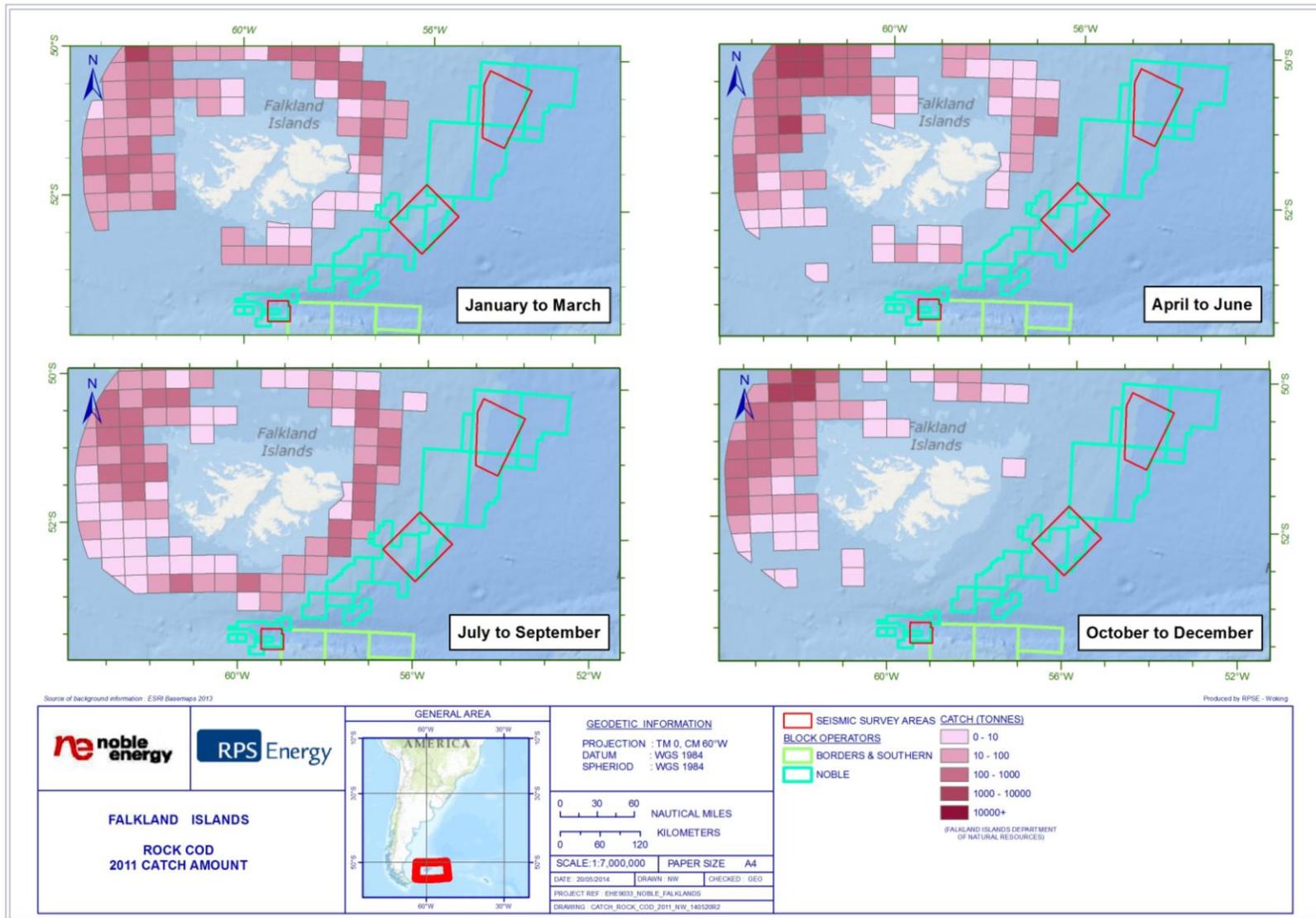


Figure E.7e: Fisheries catch mass (tonnes) for Rock Cod (*Patagonotothen ramsayi*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

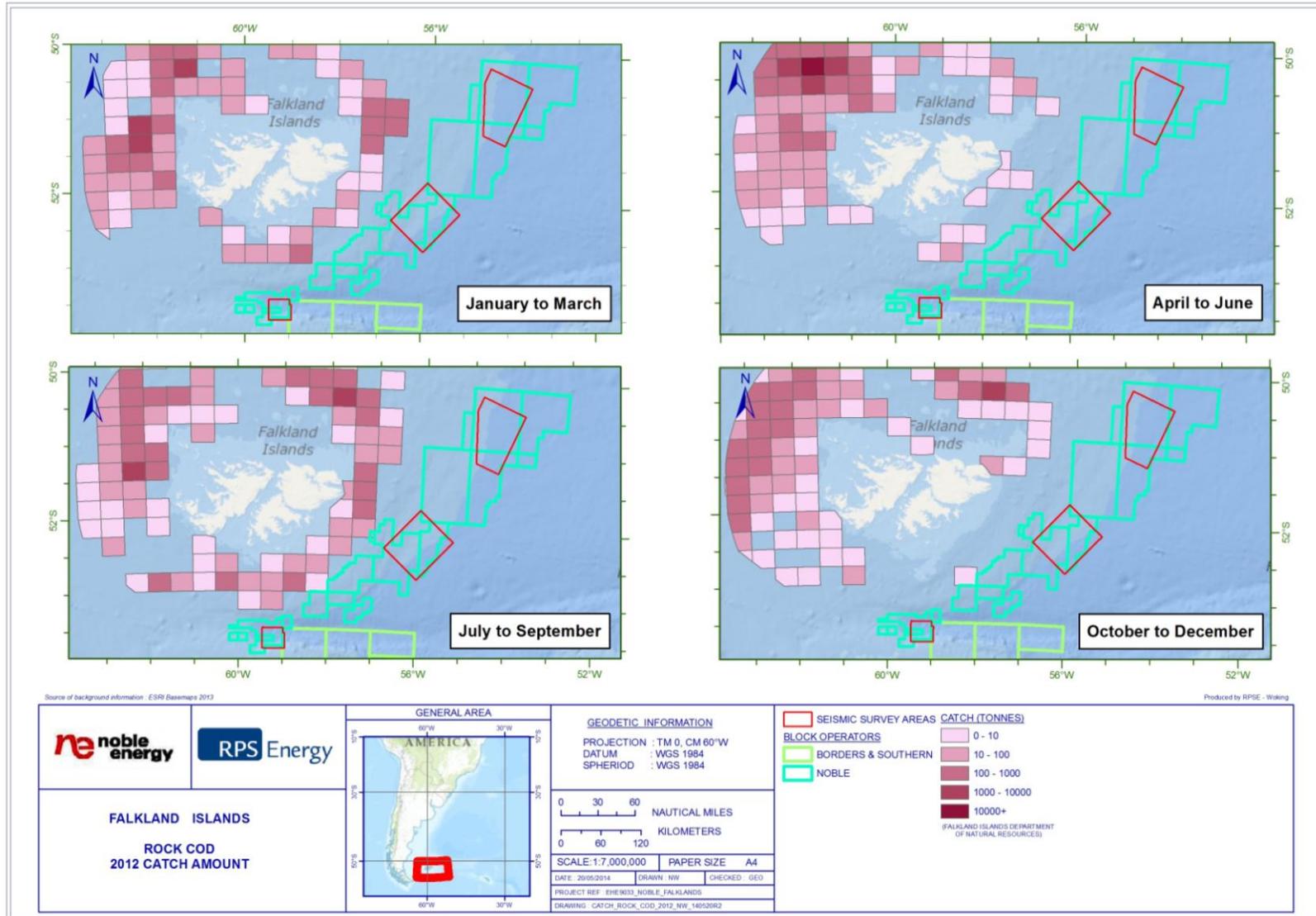


Figure E.7f: Fisheries catch mass (tonnes) for Rock Cod (*Patagonotothen ramsayi*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

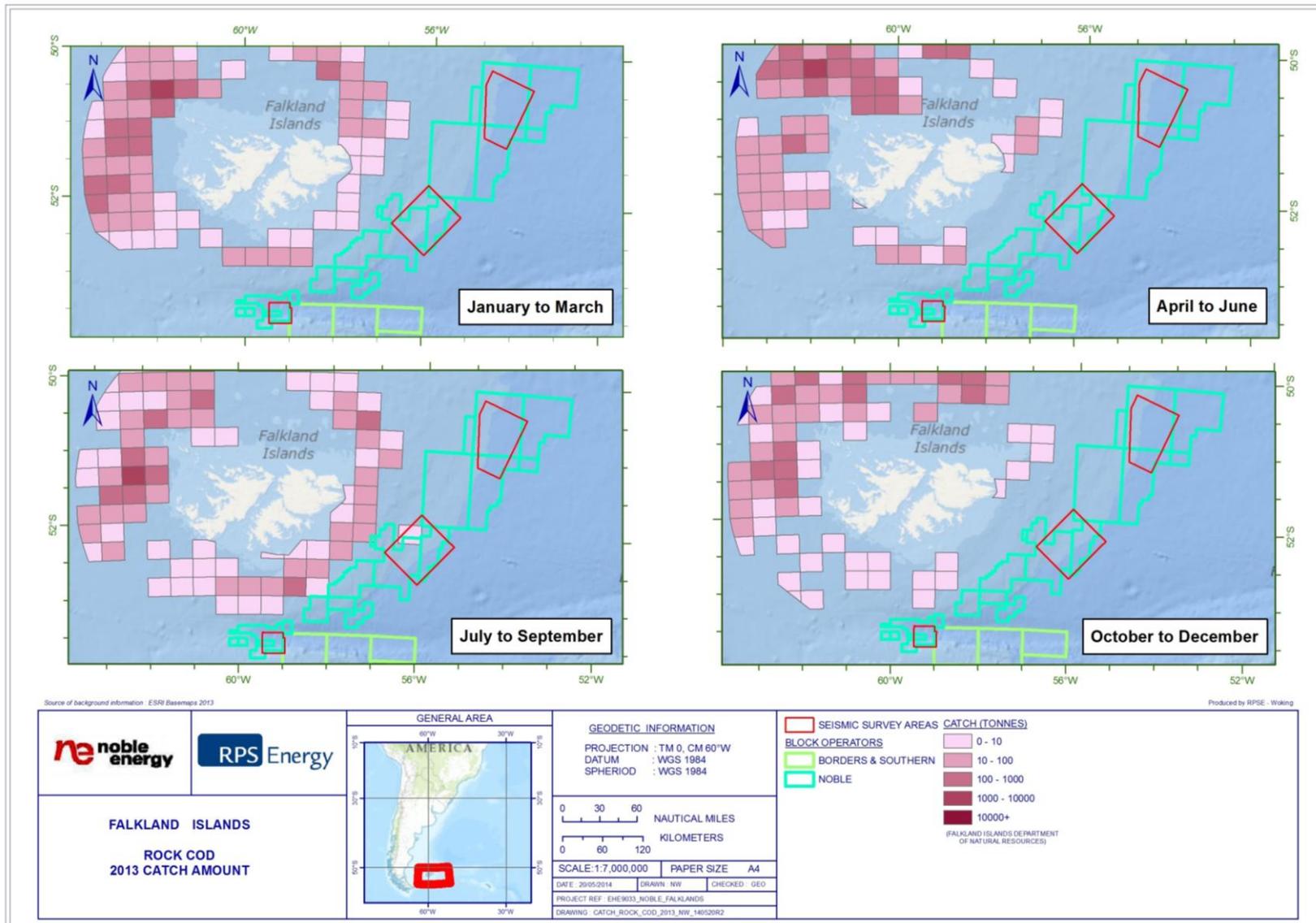


Figure E.8a: Fisheries catch mass (tonnes) for Patagonian Toothfish (*Dissostichus eleginoides*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

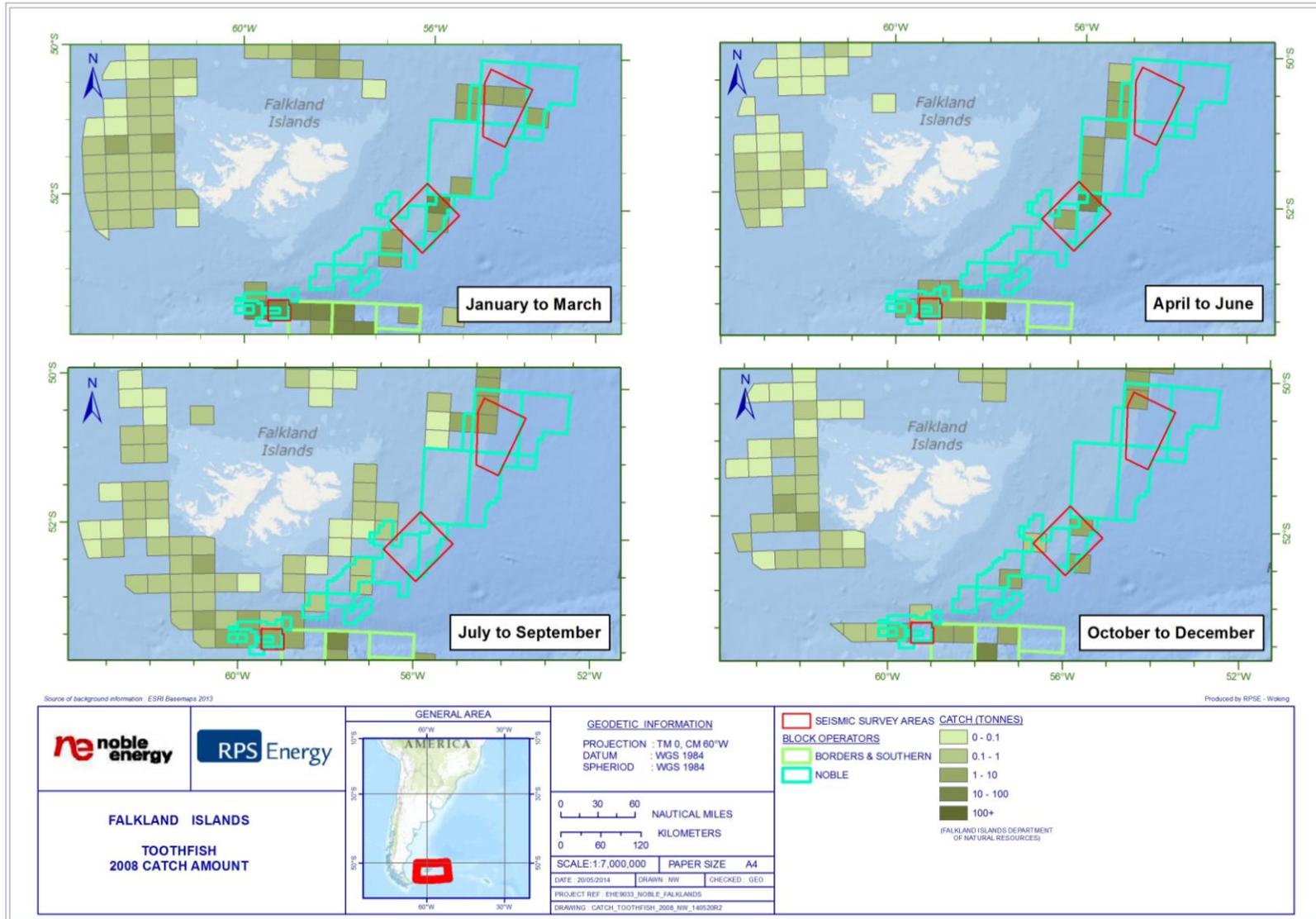


Figure E.8b: Fisheries catch mass (tonnes) for Patagonian Toothfish (*Dissostichus eleginoides*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

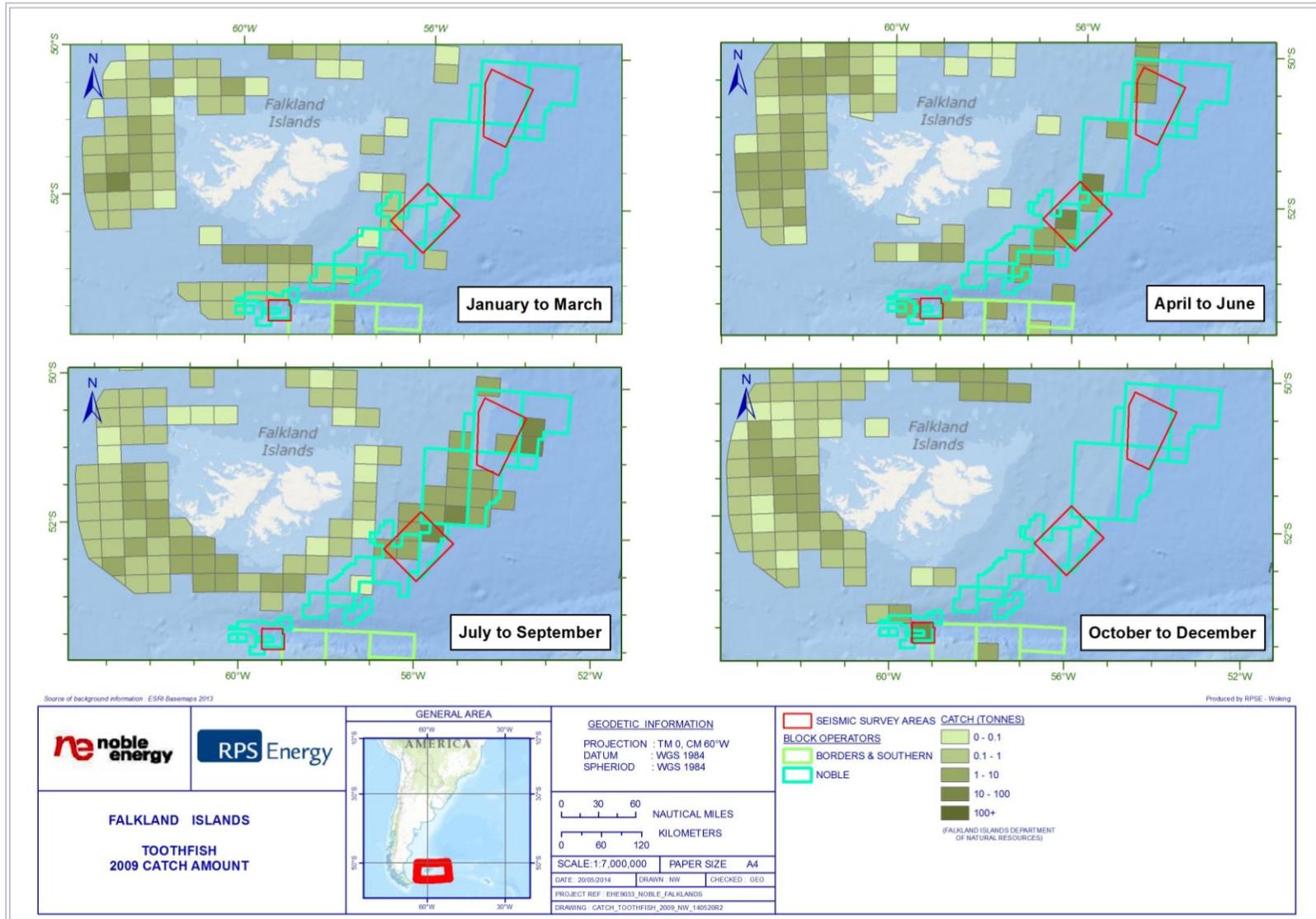


Figure E.8c: Fisheries catch mass (tonnes) for Patagonian Toothfish (*Dissostichus eleginoides*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

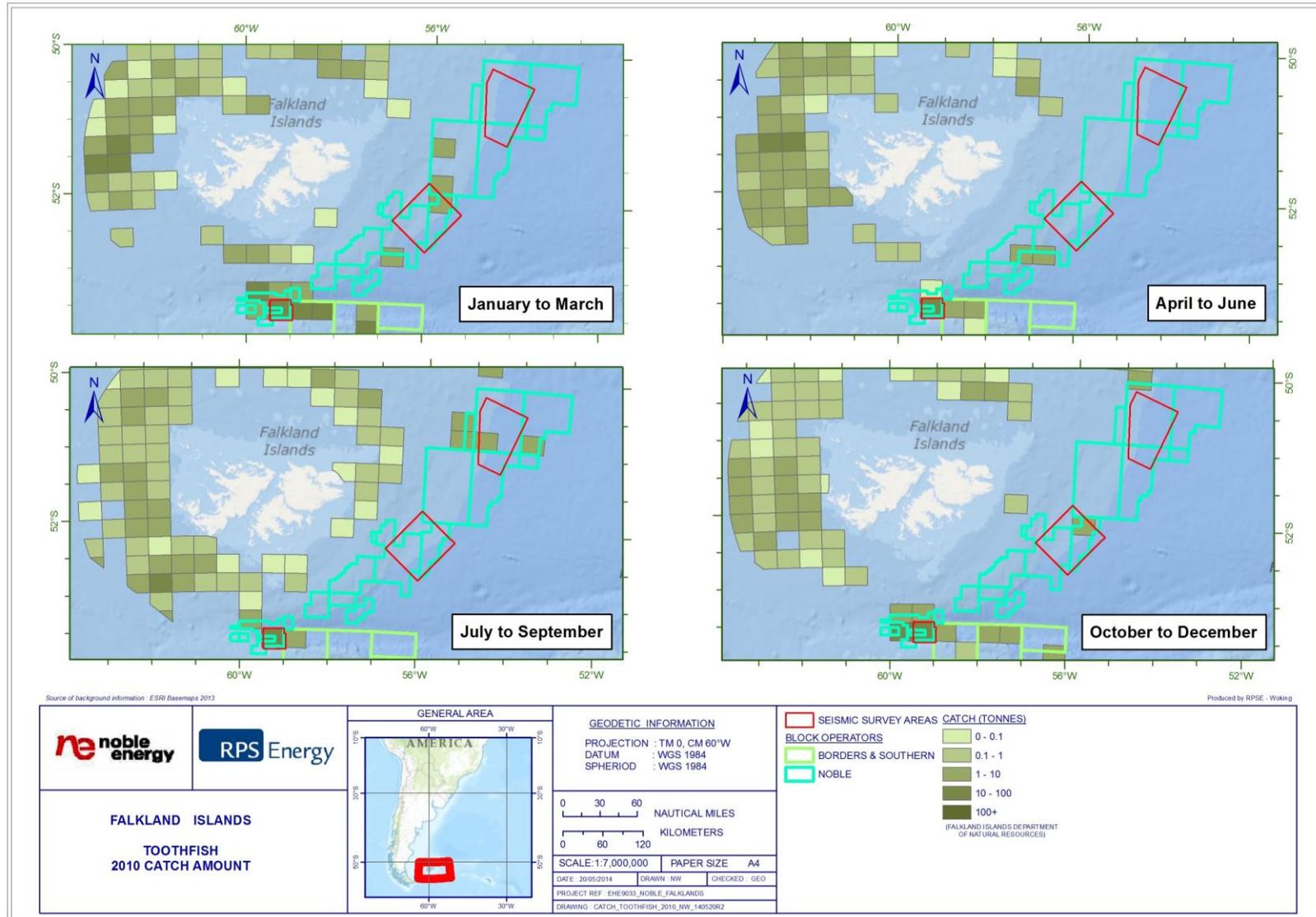


Figure E.8d: Fisheries catch mass (tonnes) for Patagonian Toothfish (*Dissostichus eleginoides*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

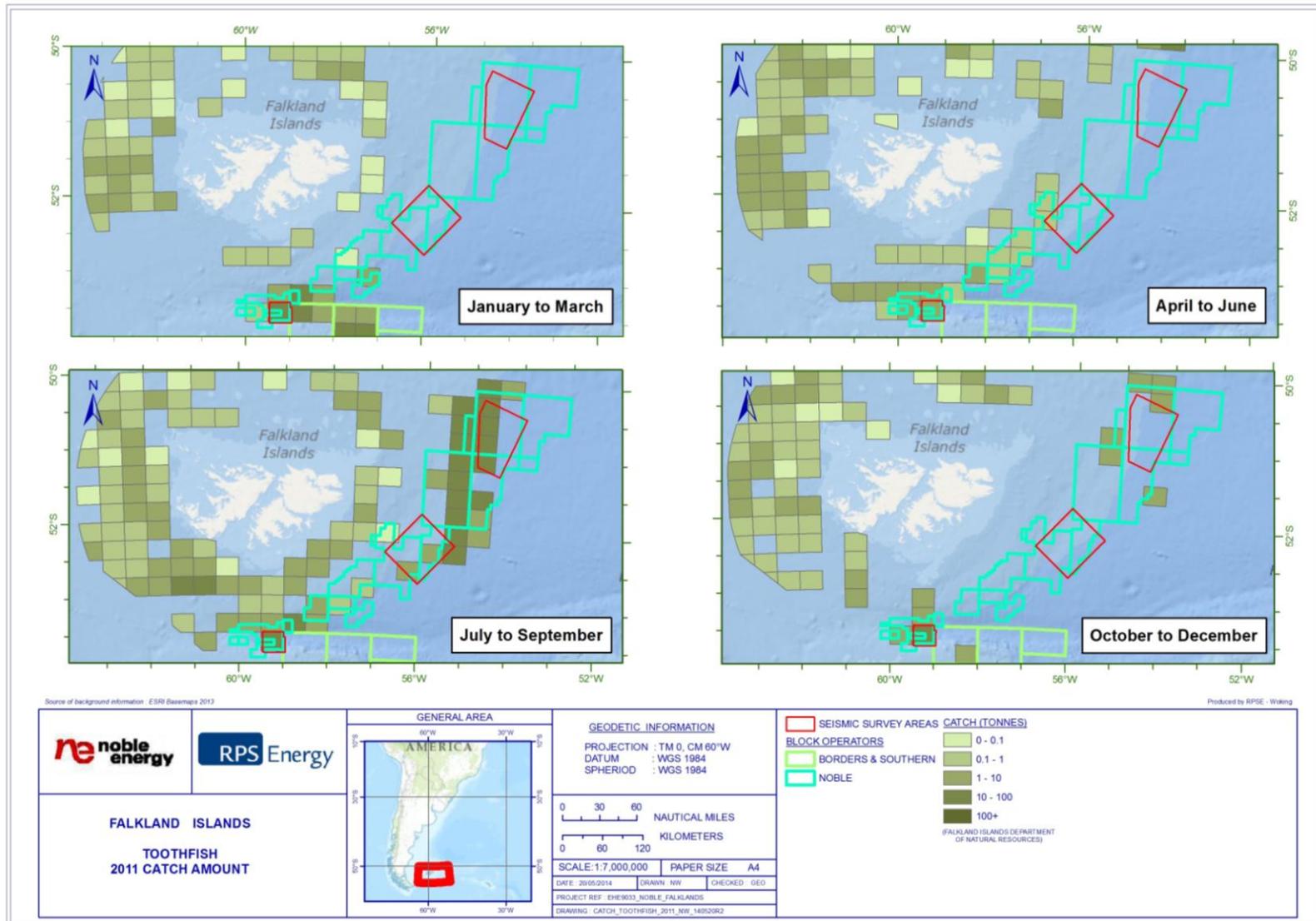


Figure E.8e: Fisheries catch mass (tonnes) for Patagonian Toothfish (*Dissostichus eleginoides*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

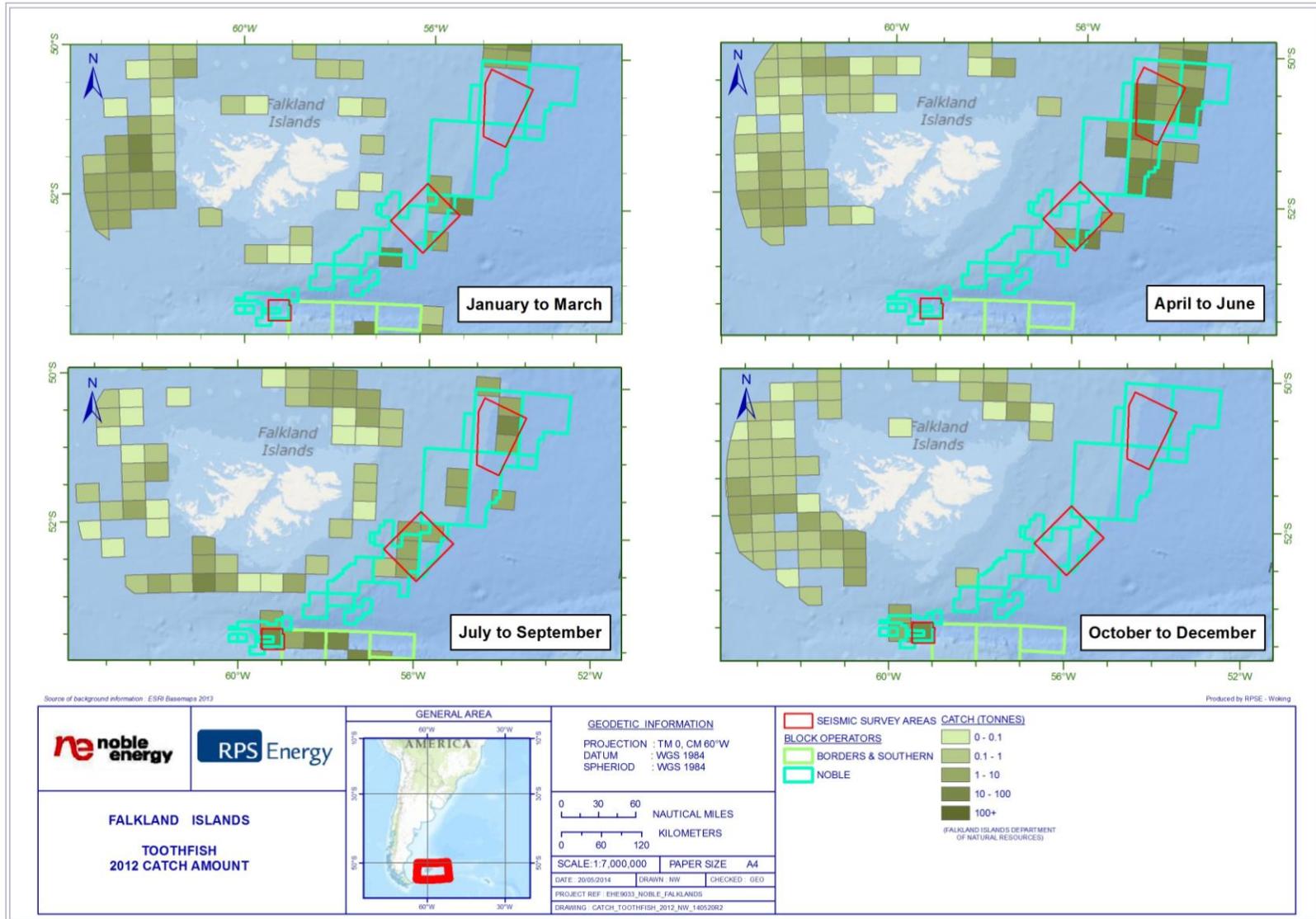


Figure E.8f: Fisheries catch mass (tonnes) for Patagonian Toothfish (*Dissostichus eleginoides*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

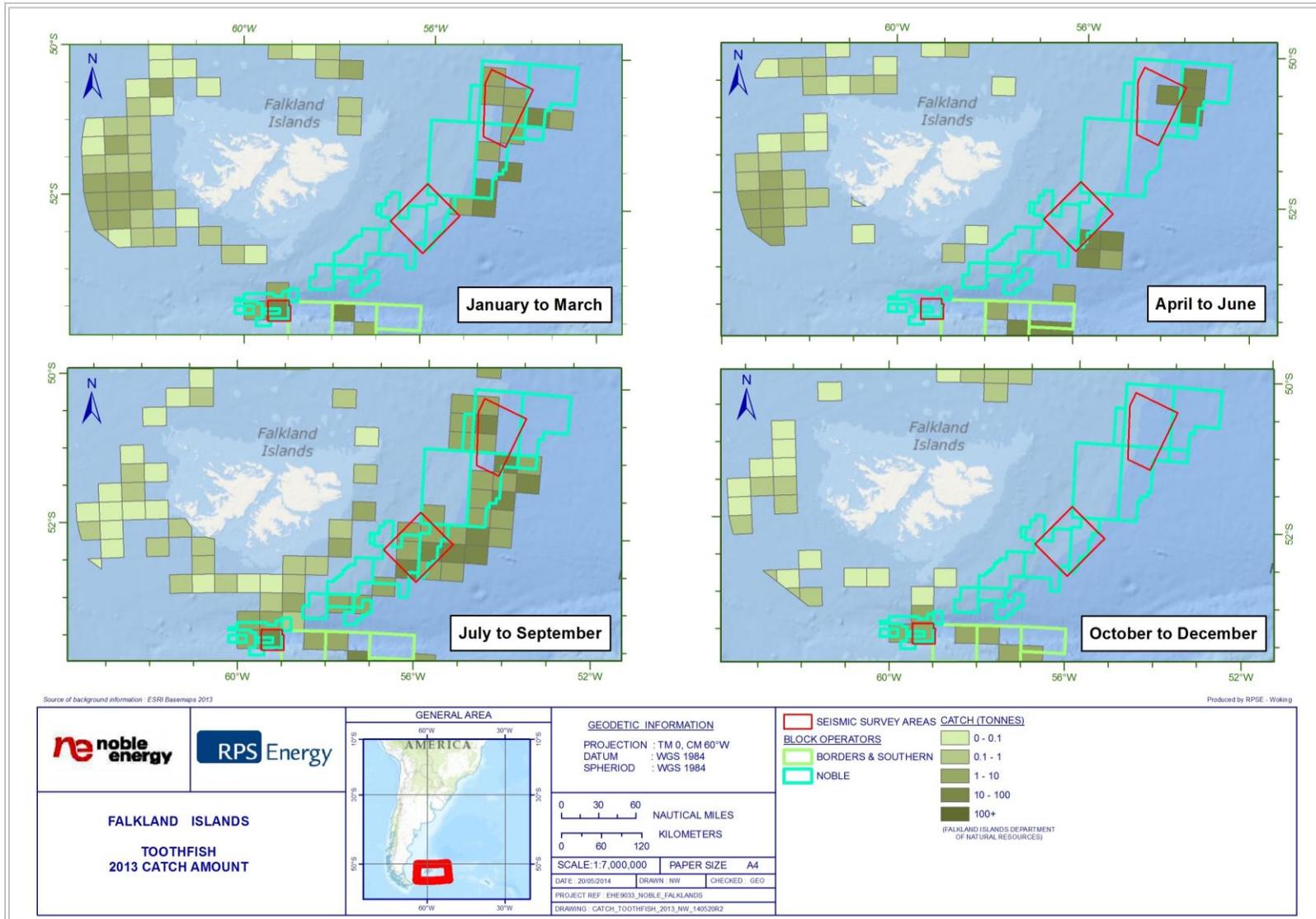


Figure E.9a: Fisheries catch mass (tonnes) for Kingclip (*Genypterus blacodes*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

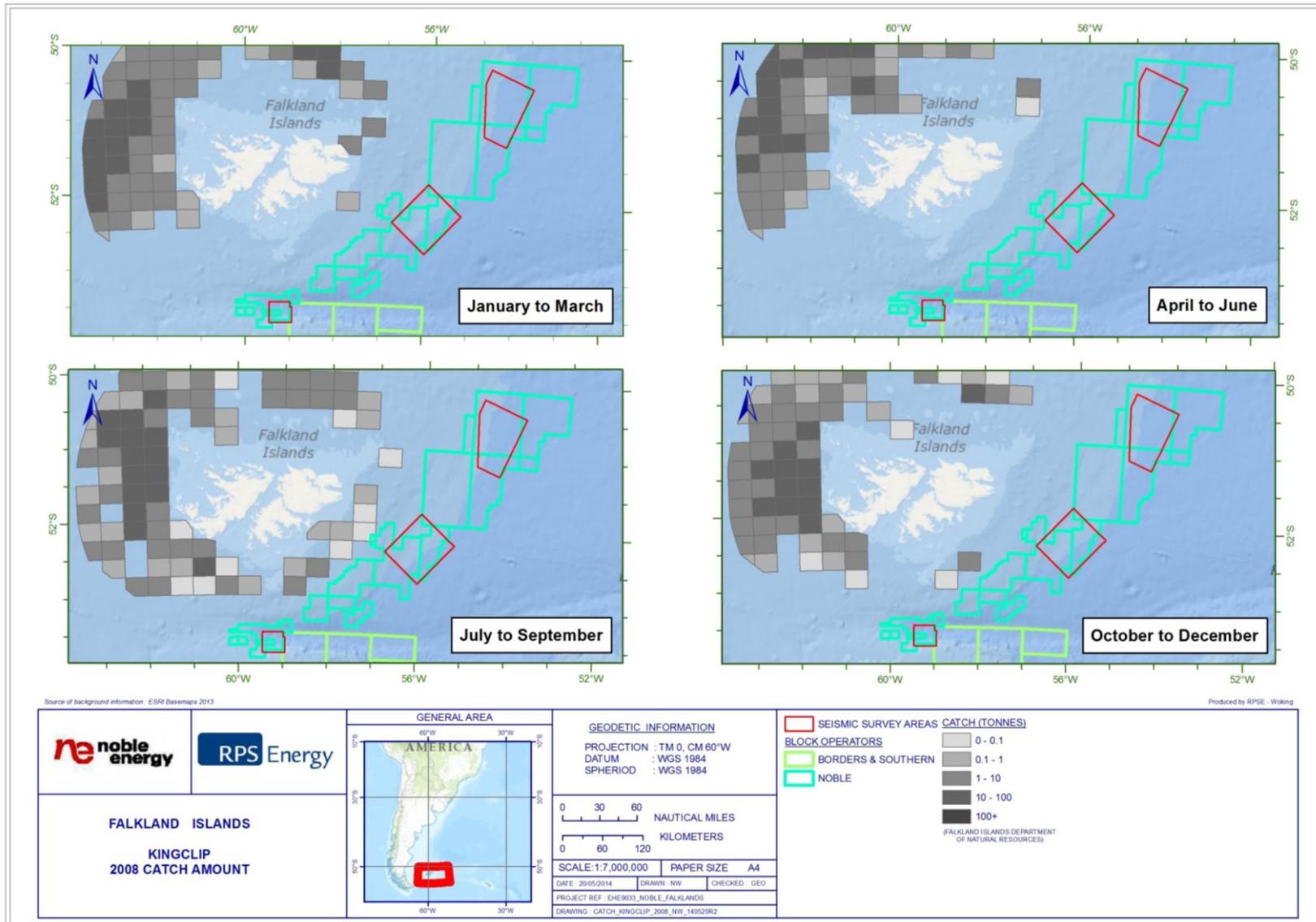


Figure E.9b: Fisheries catch mass (tonnes) for Kingclip (*Genypterus blacodes*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

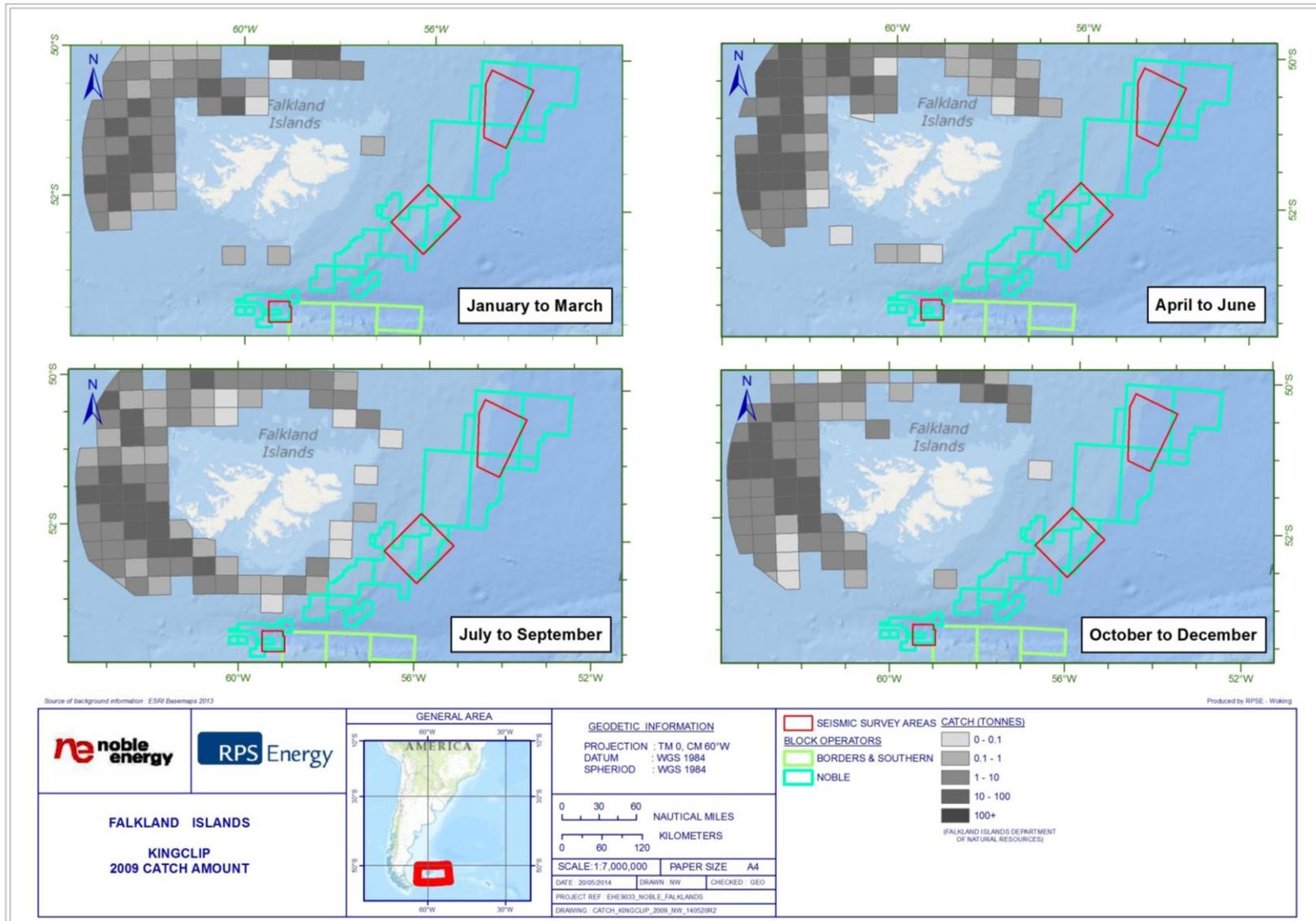


Figure E.9c: Fisheries catch mass (tonnes) for Kingclip (*Genypterus blacodes*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

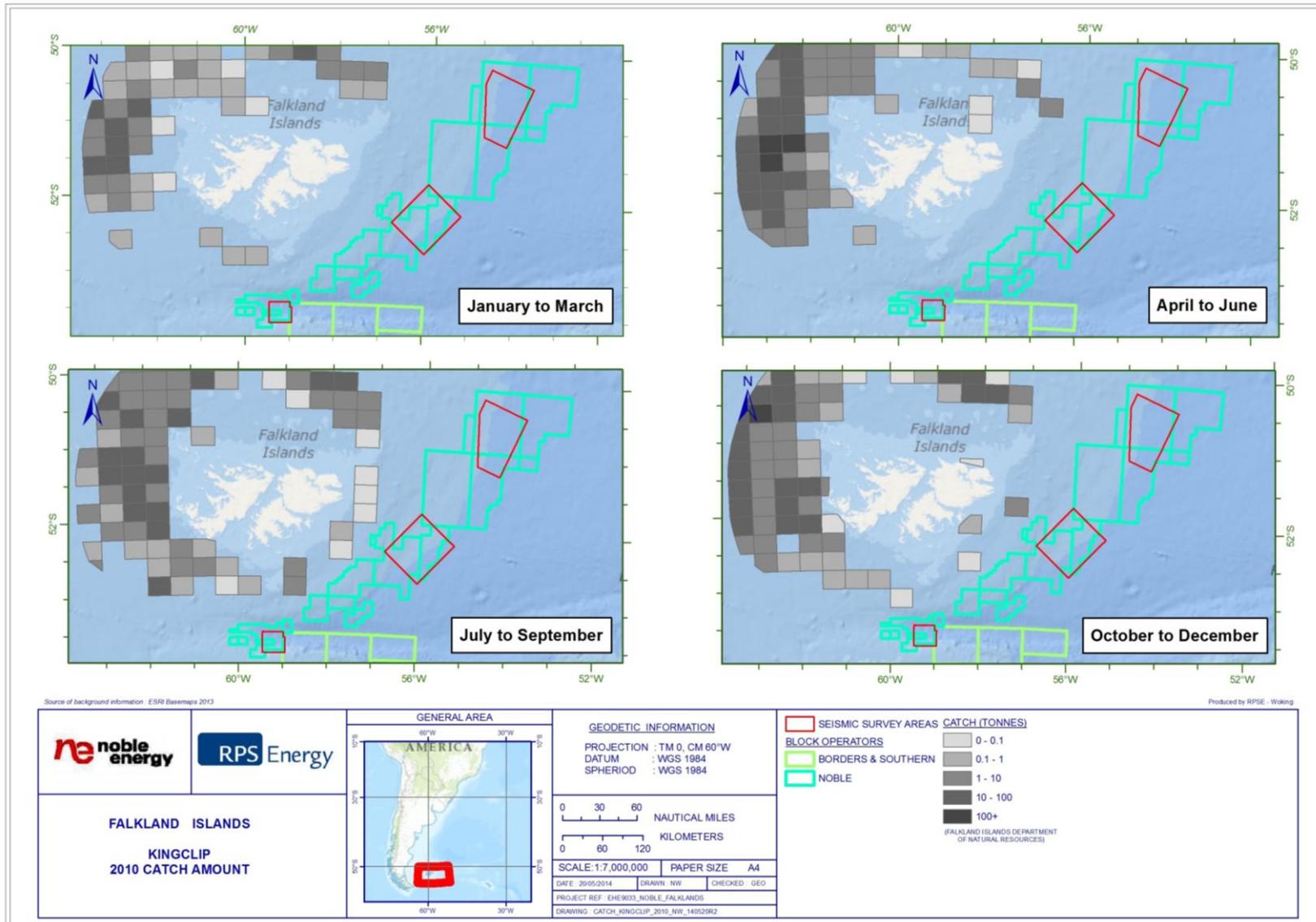


Figure E.9d: Fisheries catch mass (tonnes) for Kingclip (*Genypterus blacodes*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

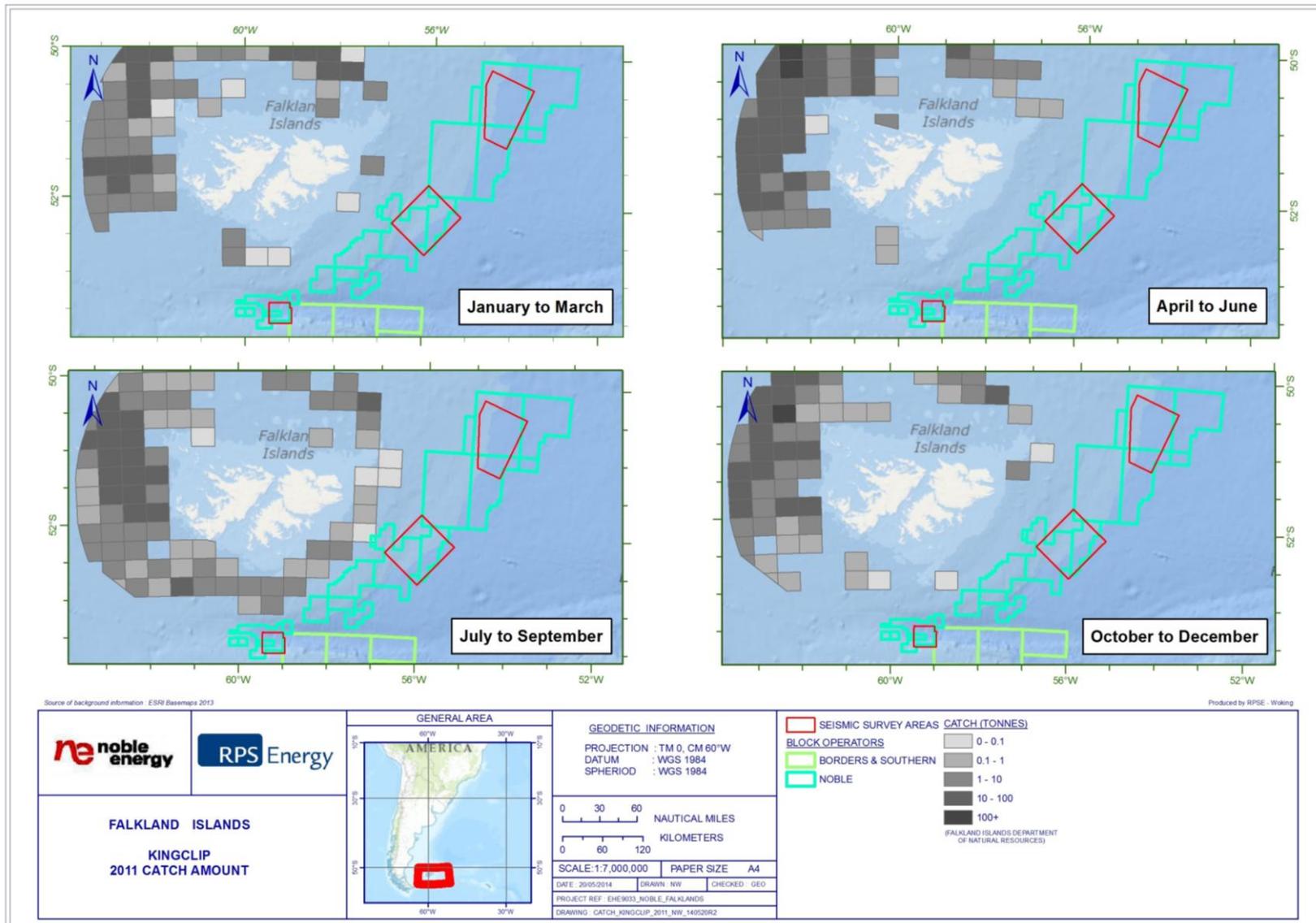


Figure E.9e: Fisheries catch mass (tonnes) for Kingclip (*Genypterus blacodes*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

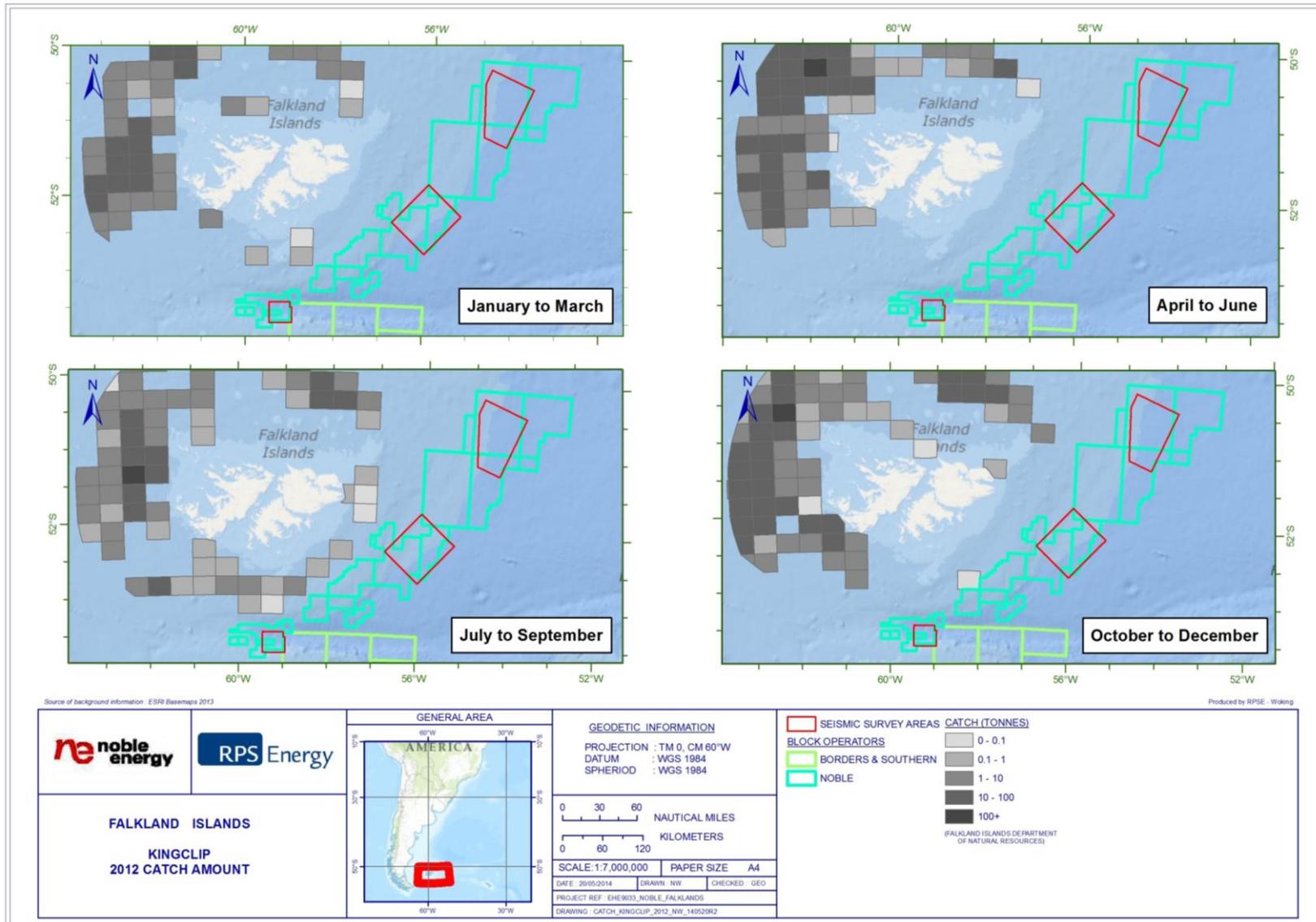


Figure E.9f: Fisheries catch mass (tonnes) for Kingclip (*Genypterus blacodes*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

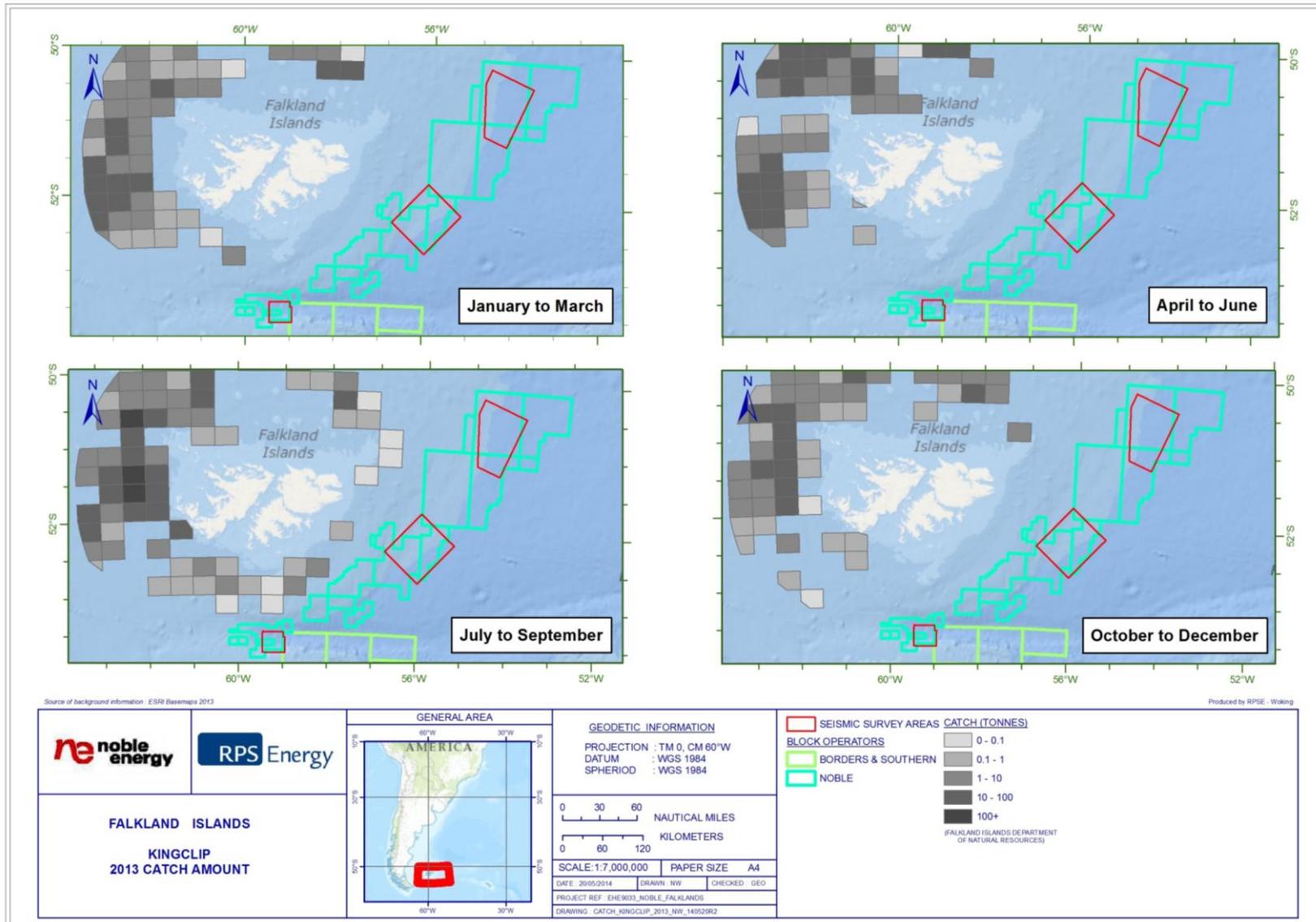


Figure E.10a: Fisheries catch mass (tonnes) for Argentine shortfin squid (*Illex argentinus*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

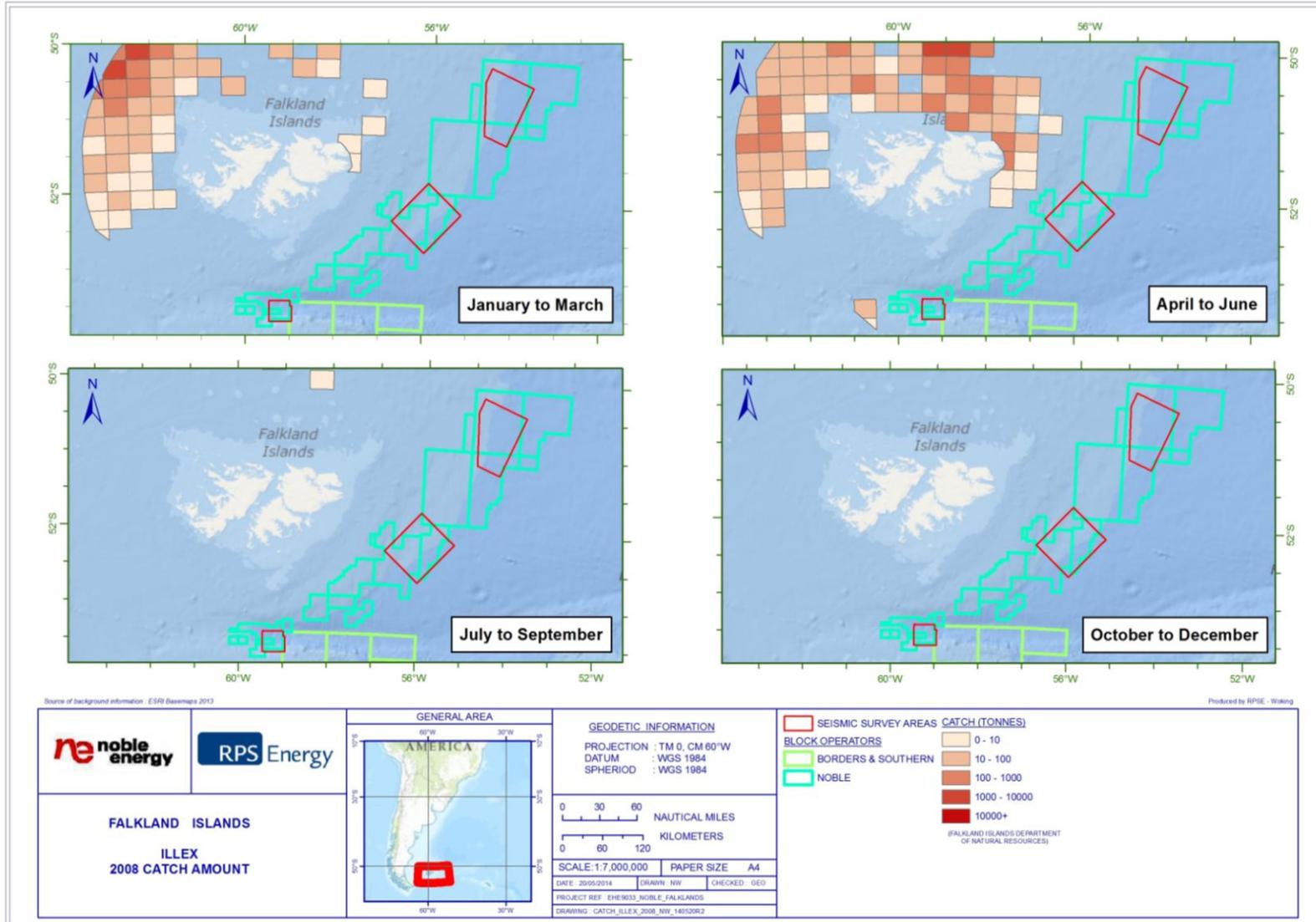


Figure E.10b: Fisheries catch mass (tonnes) for Argentine shortfin squid (*Illex argentinus*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

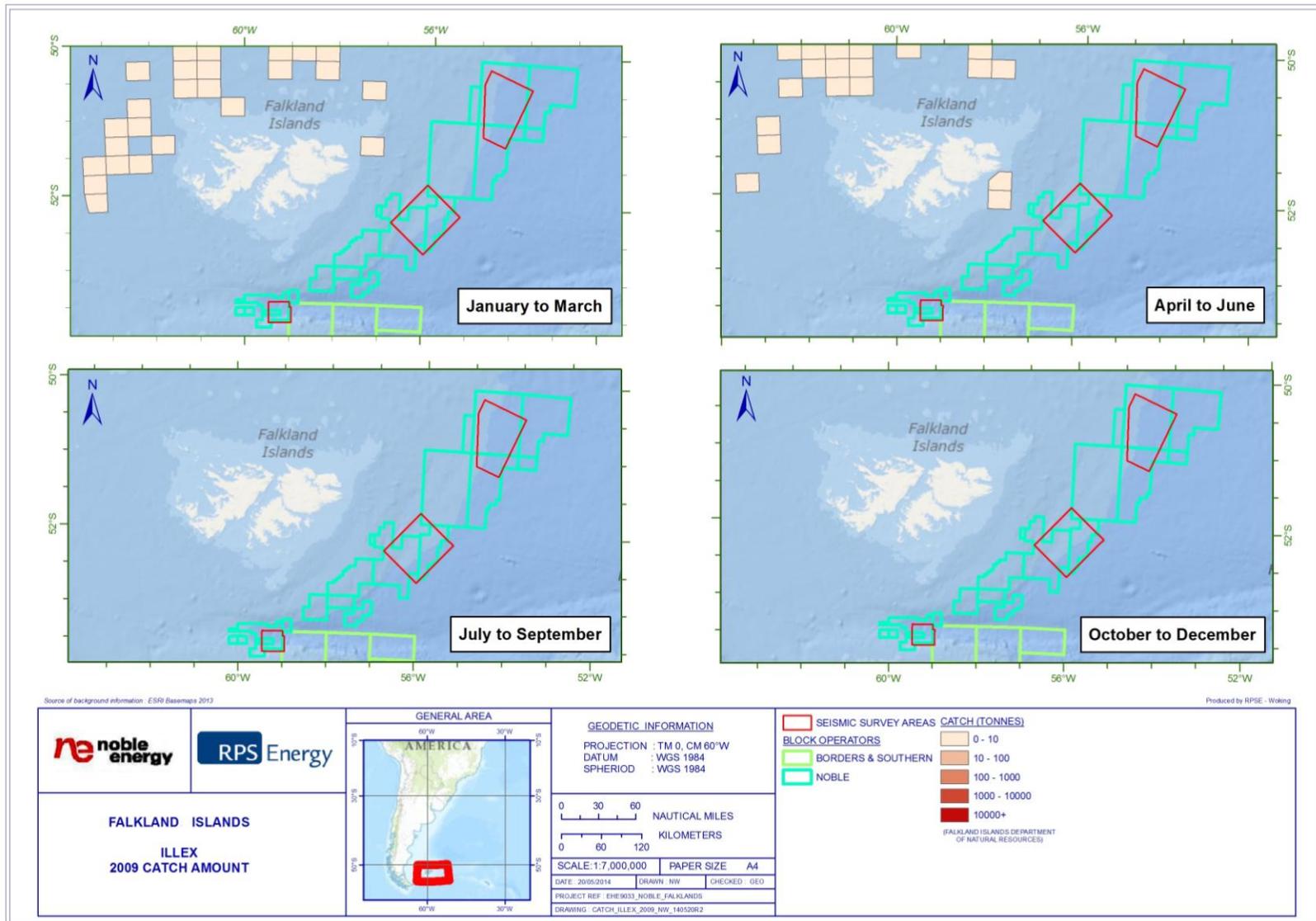


Figure E.10c: Fisheries catch mass (tonnes) for Argentine shortfin squid (*Illex argentinus*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

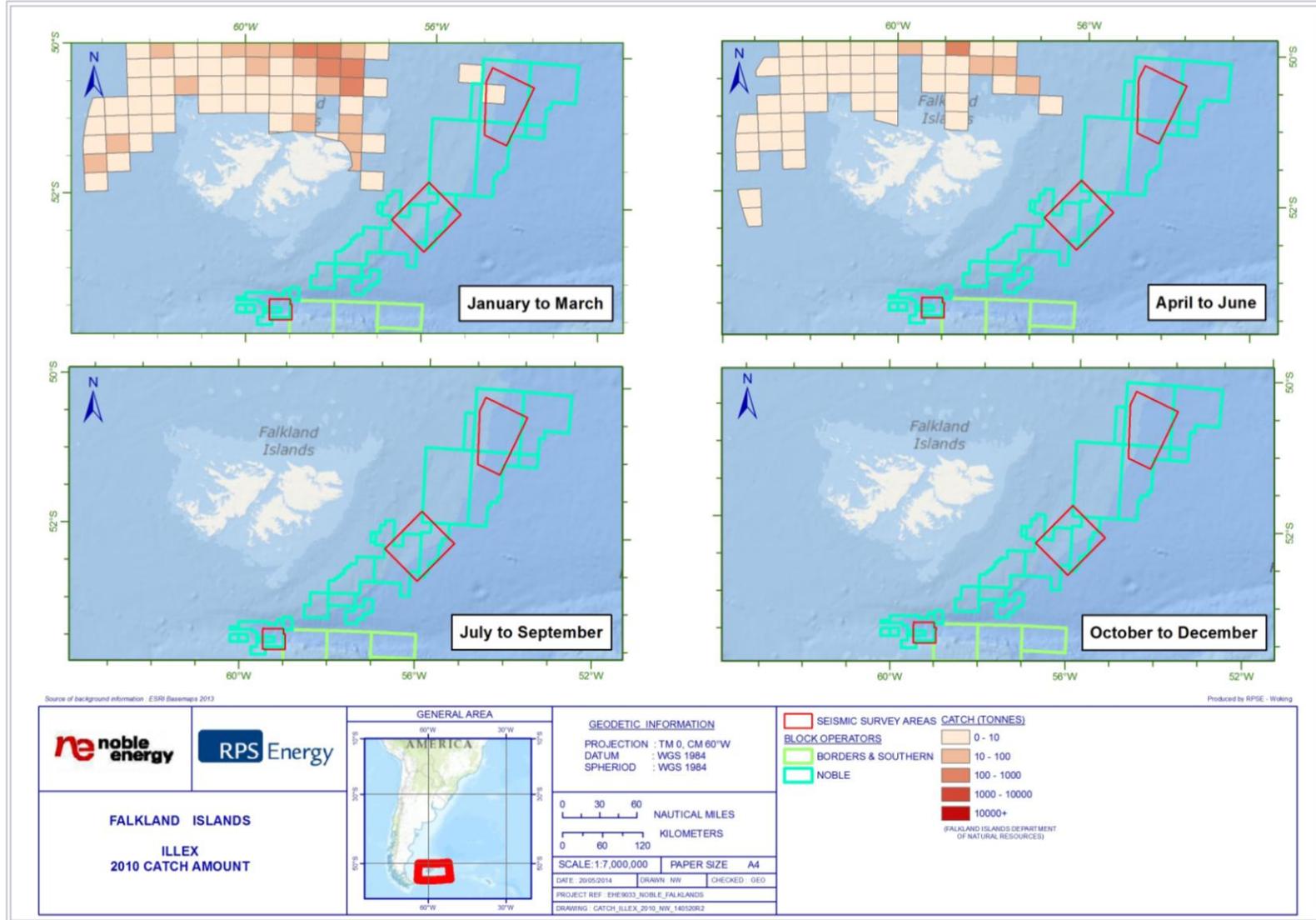


Figure E.10d: Fisheries catch mass (tonnes) for Argentine shortfin squid (*Illex argentinus*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

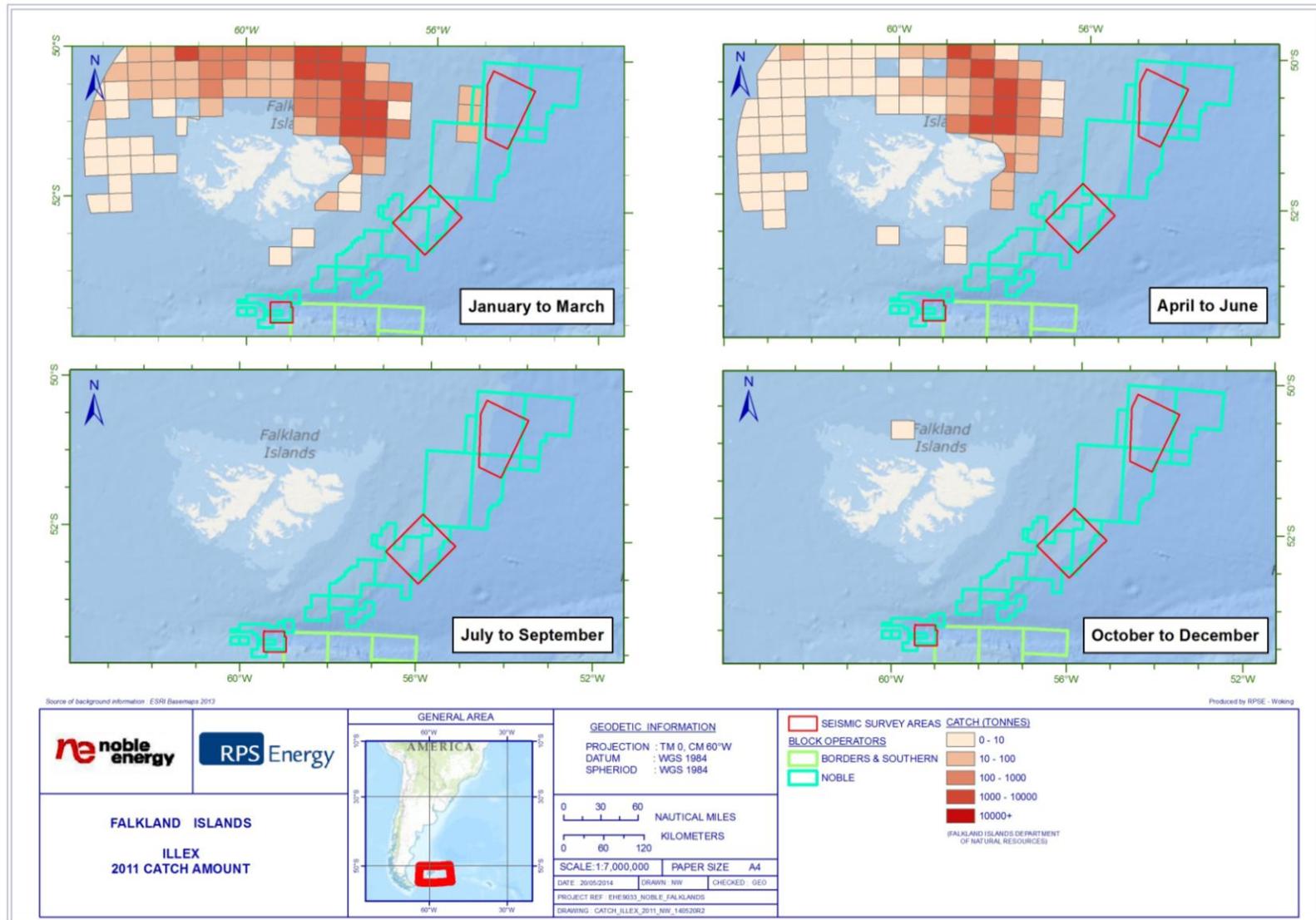


Figure E.10e: Fisheries catch mass (tonnes) for Argentine shortfin squid (*Illex argentinus*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

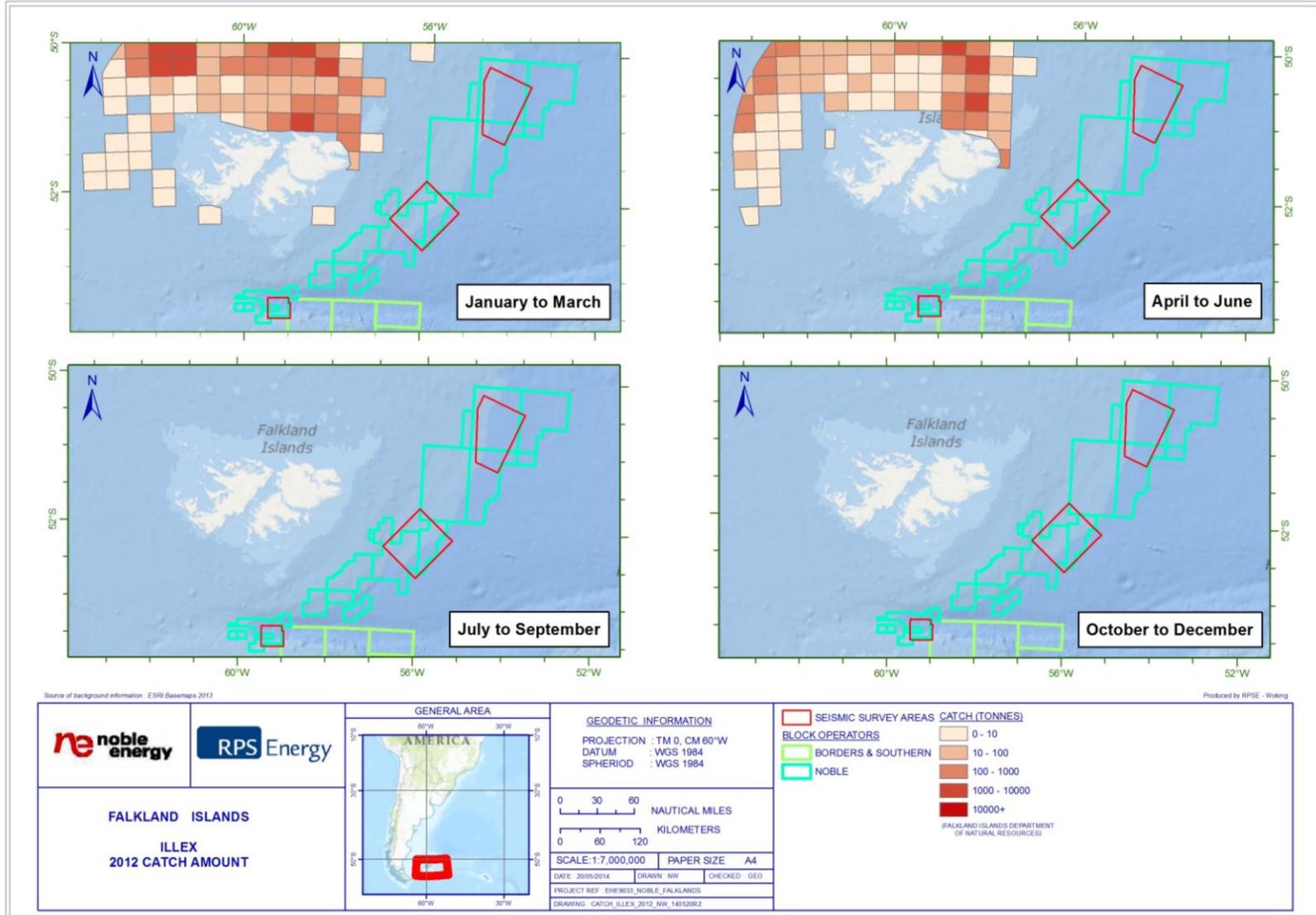


Figure E.10f: Fisheries catch mass (tonnes) for Argentine shortfin squid (*Illex argentinus*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

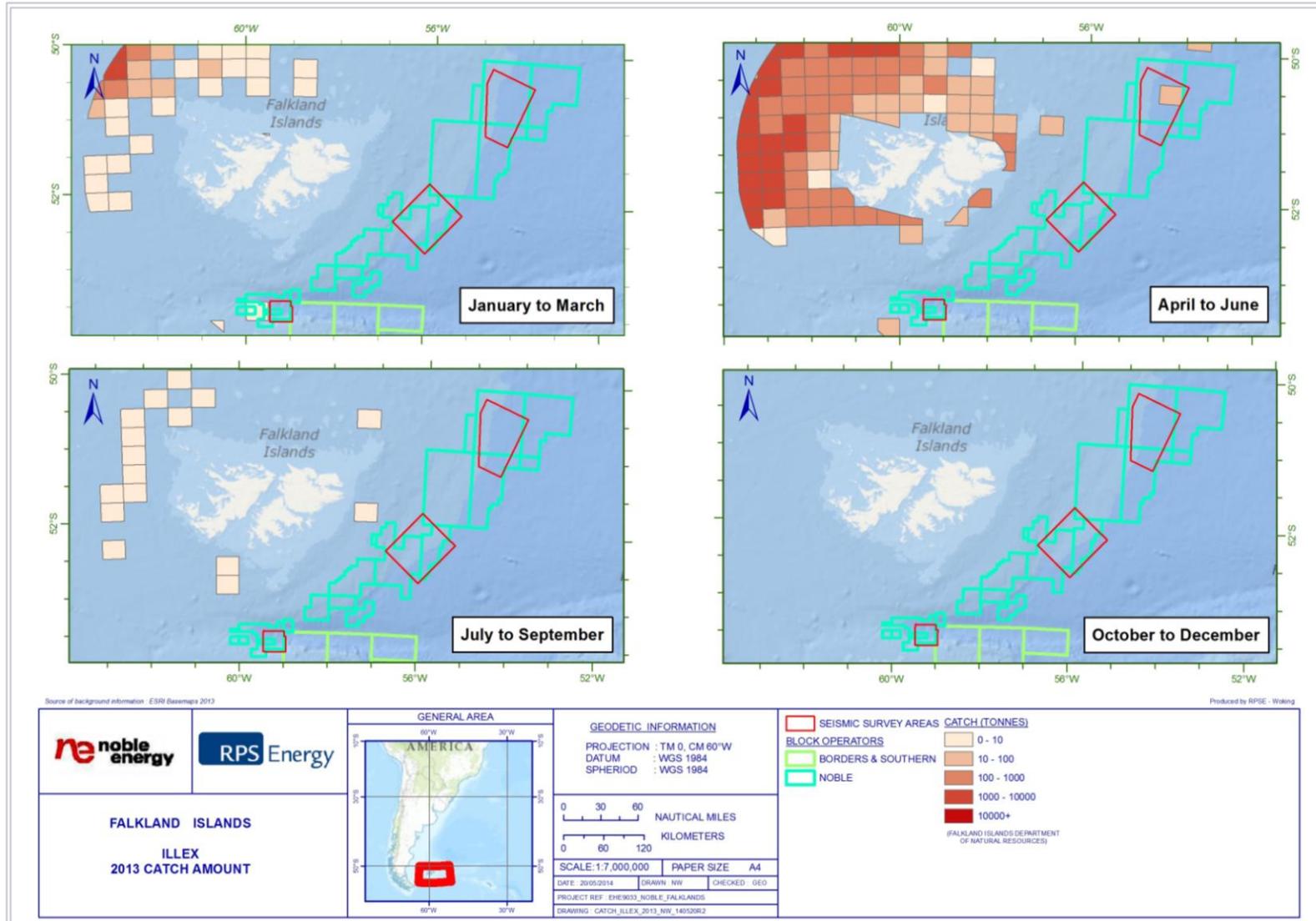


Figure E.11a: Fisheries catch mass (tonnes) for Patagonian Squid (*Doryteuthis gahi*) for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

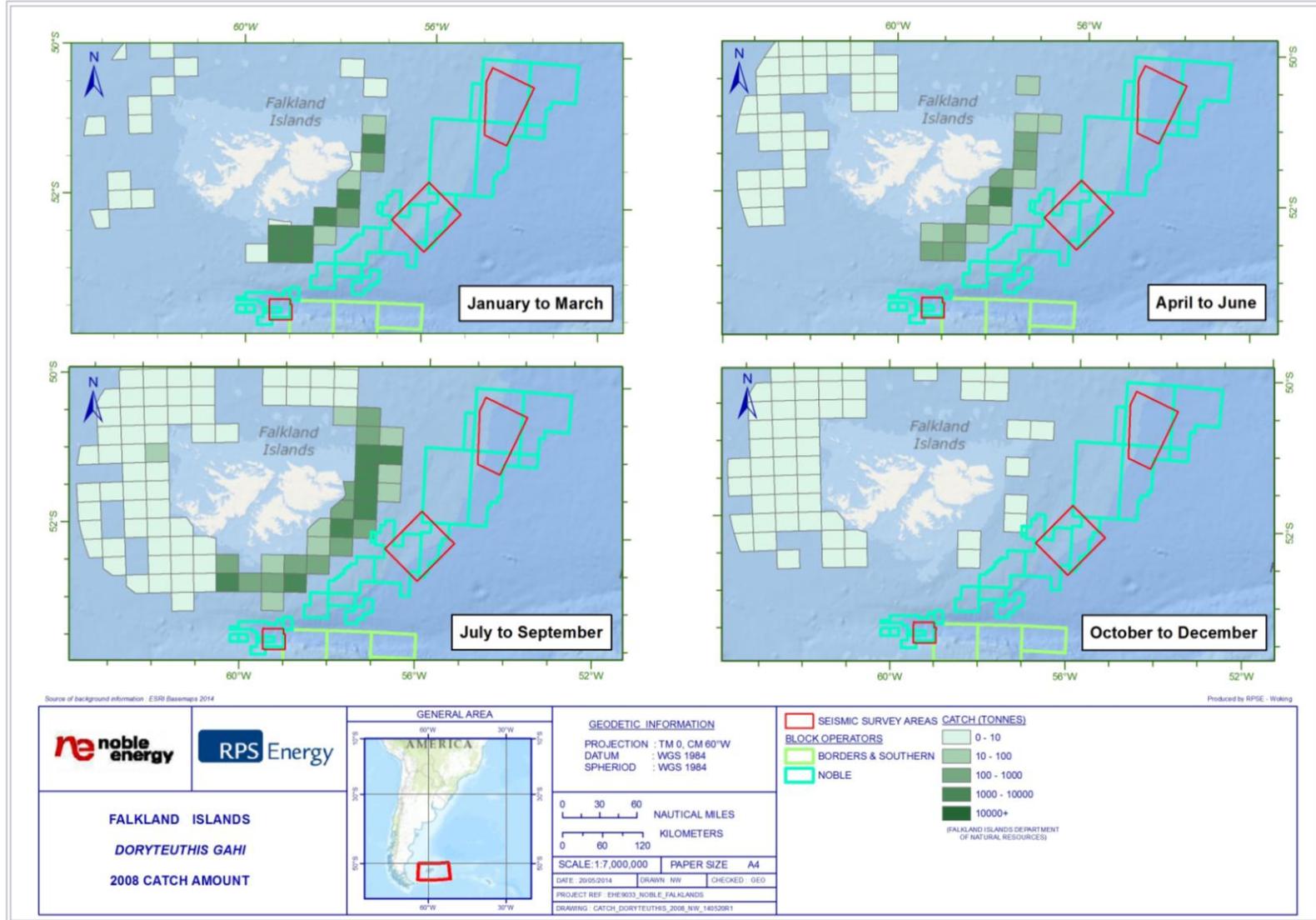


Figure E.11b: Fisheries catch mass (tonnes) for Patagonian Squid (*Doryteuthis gahi*) for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

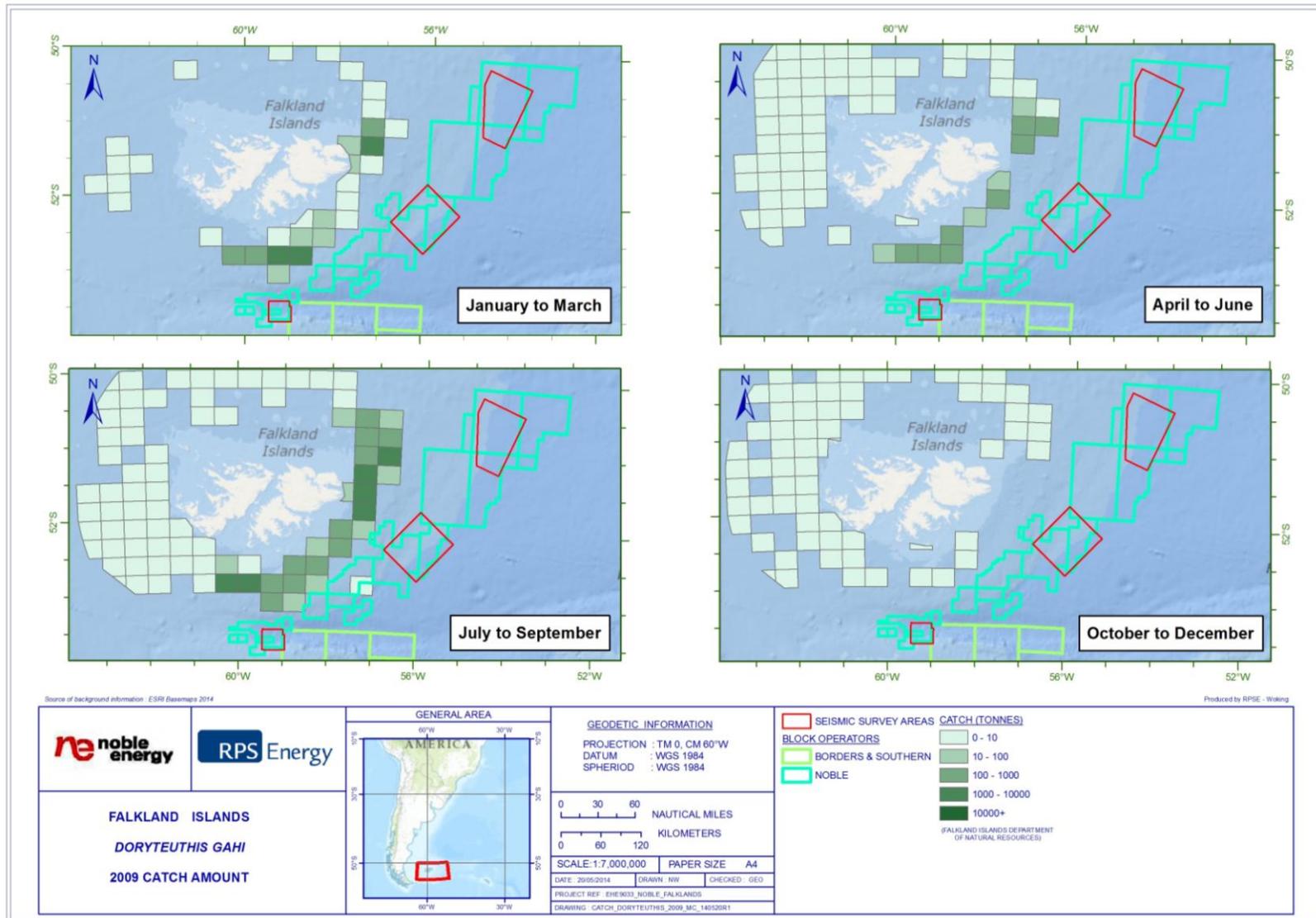


Figure E.11c: Fisheries catch mass (tonnes) for Patagonian Squid (*Doryteuthis gahi*) for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

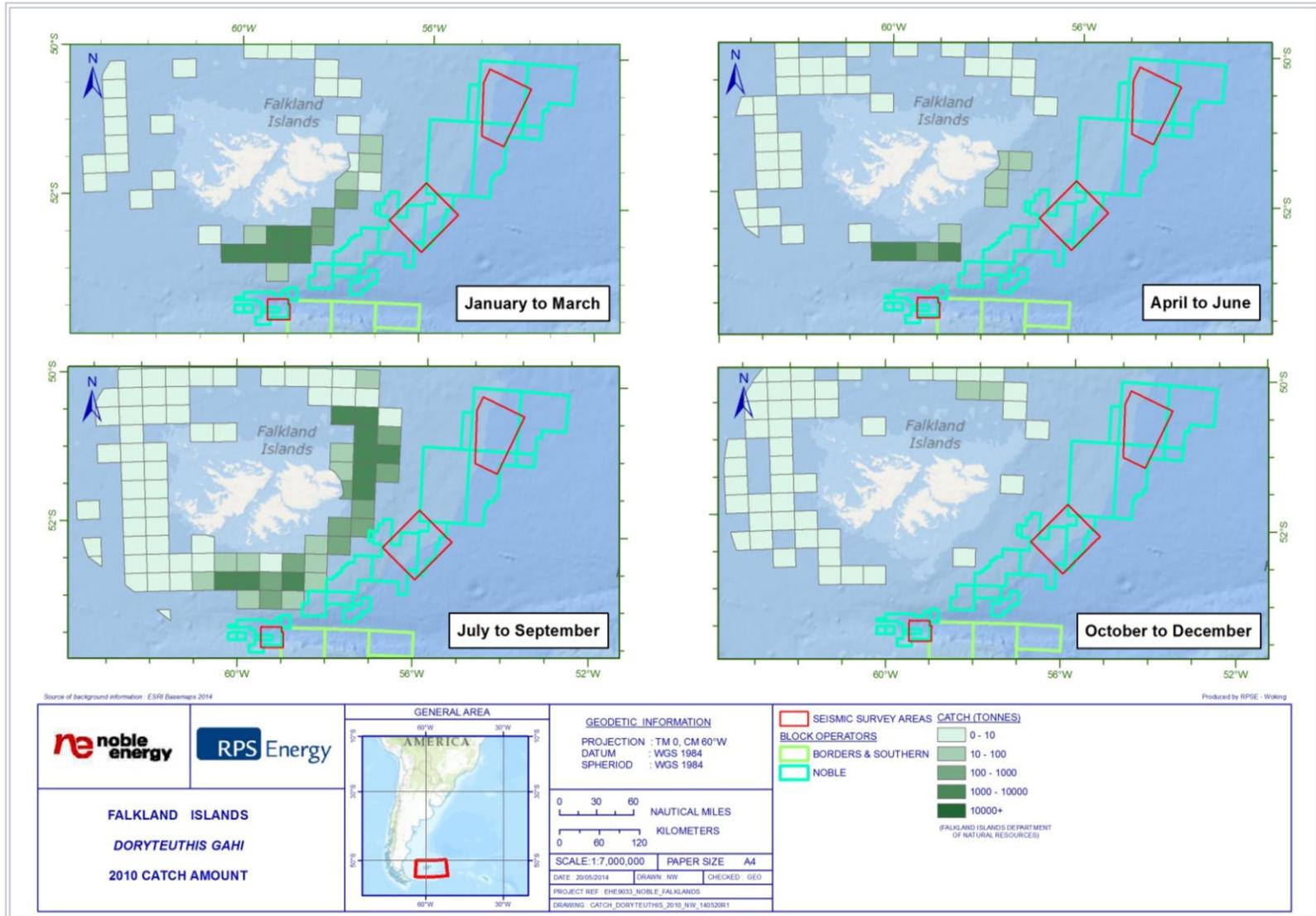


Figure E.11d: Fisheries catch mass (tonnes) for Patagonian Squid (*Doryteuthis gahi*) for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

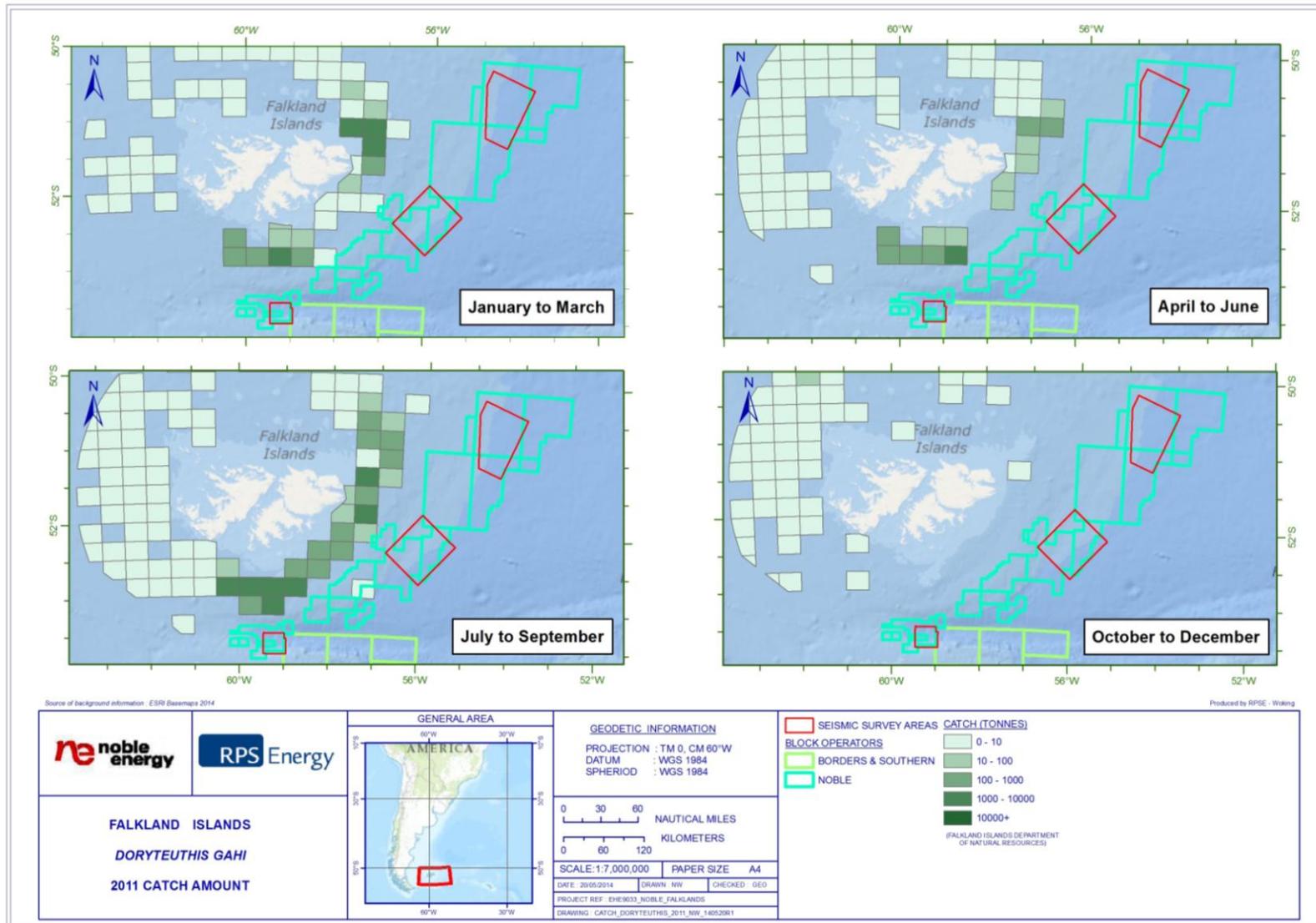


Figure E.11e: Fisheries catch mass (tonnes) for Patagonian Squid (*Doryteuthis gahi*) for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

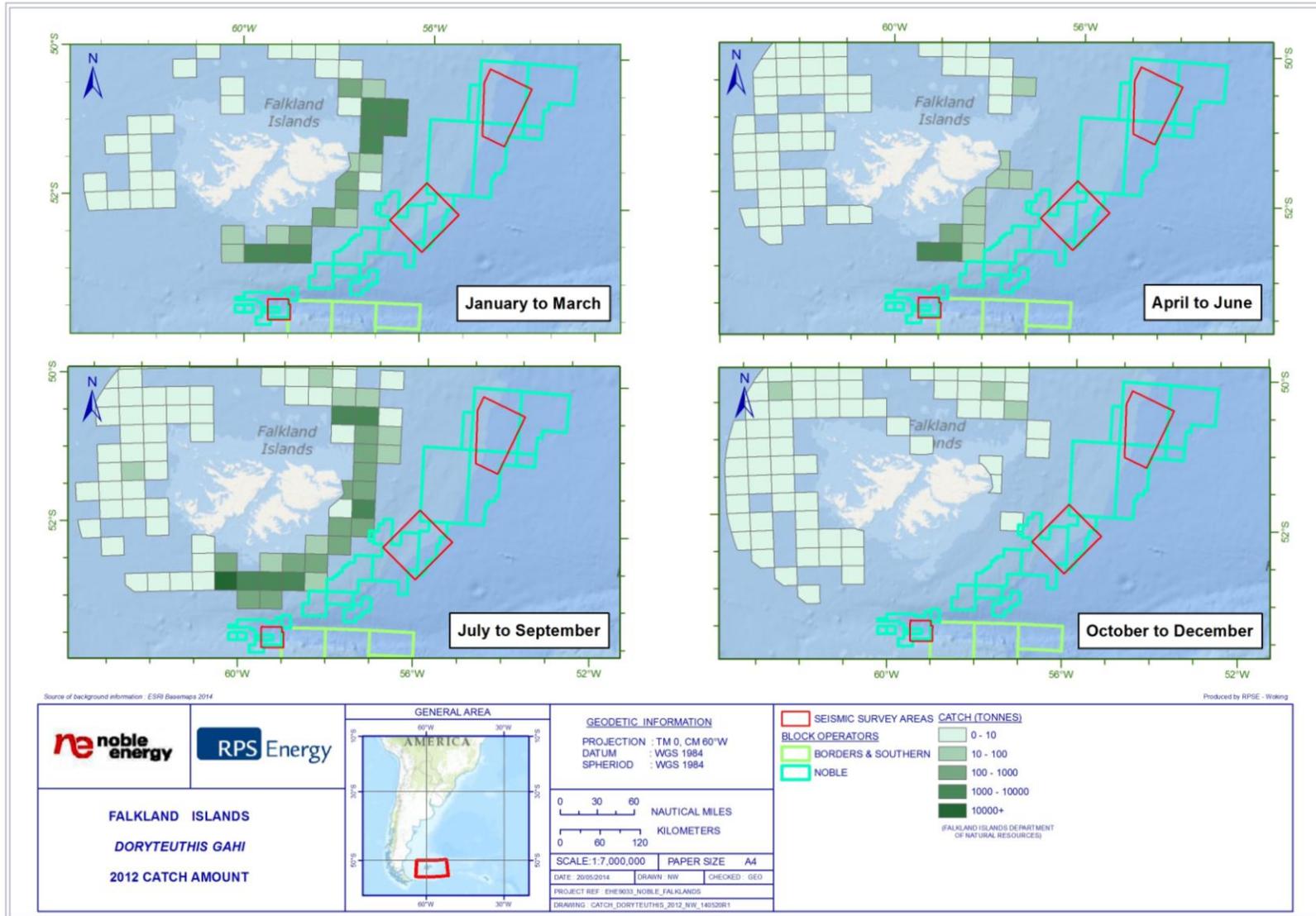


Figure E.11f: Fisheries catch mass (tonnes) for Patagonian Squid (*Doryteuthis gahi*) for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

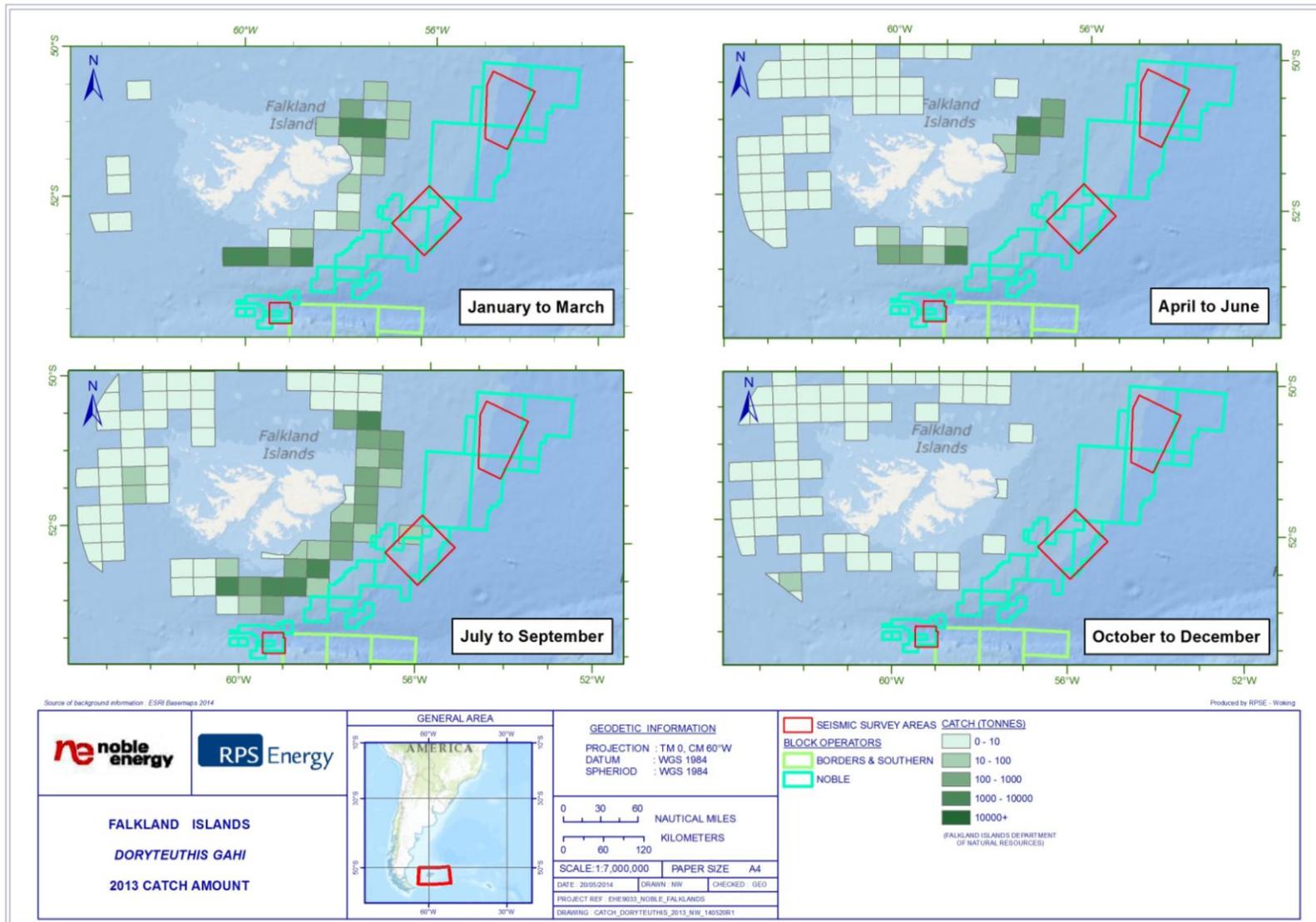


Figure E.12a: Fisheries catch mass (tonnes) for other species for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

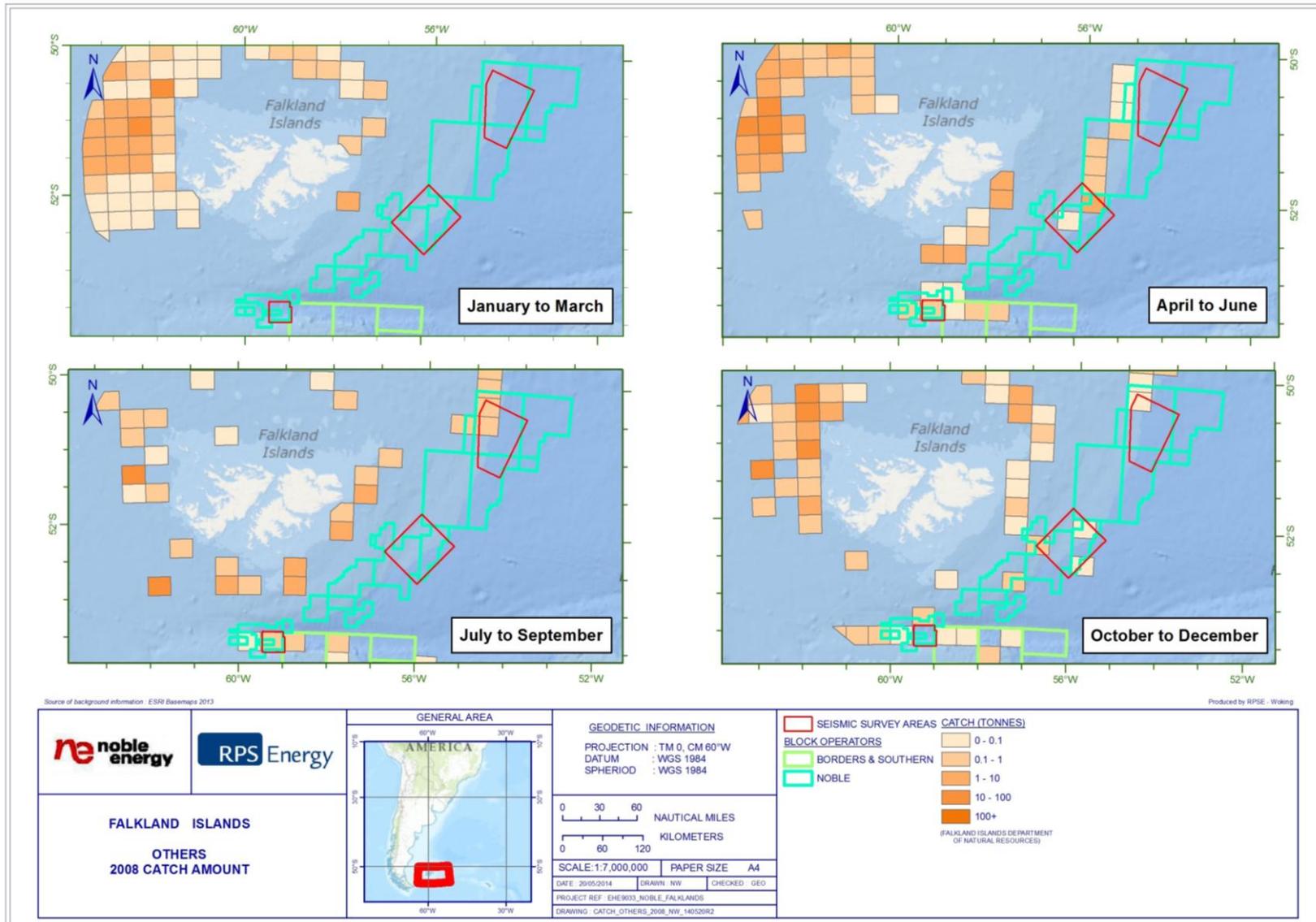


Figure E.12b: Fisheries catch mass (tonnes) for other species for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

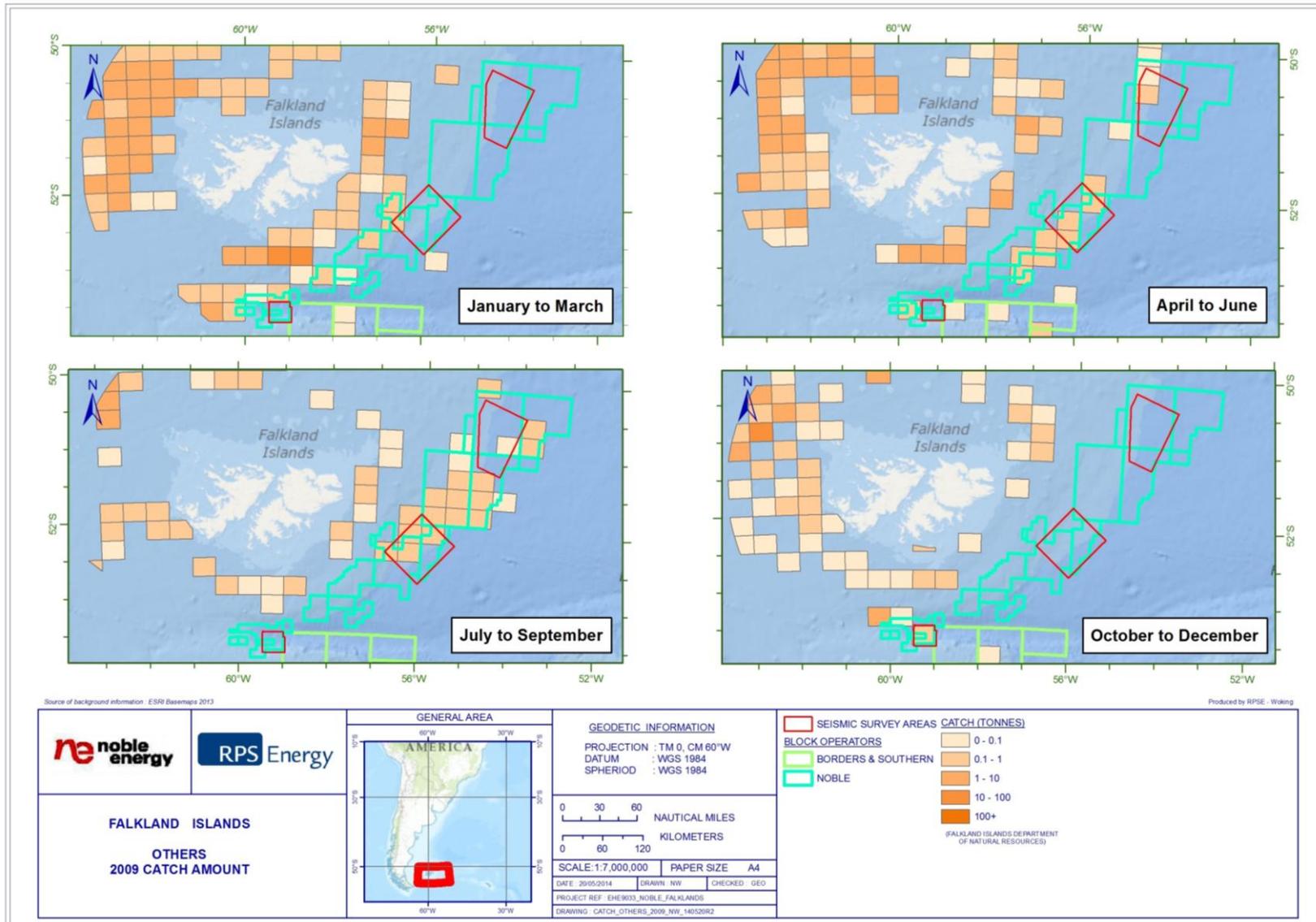


Figure E.12c: Fisheries catch mass (tonnes) for other species for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

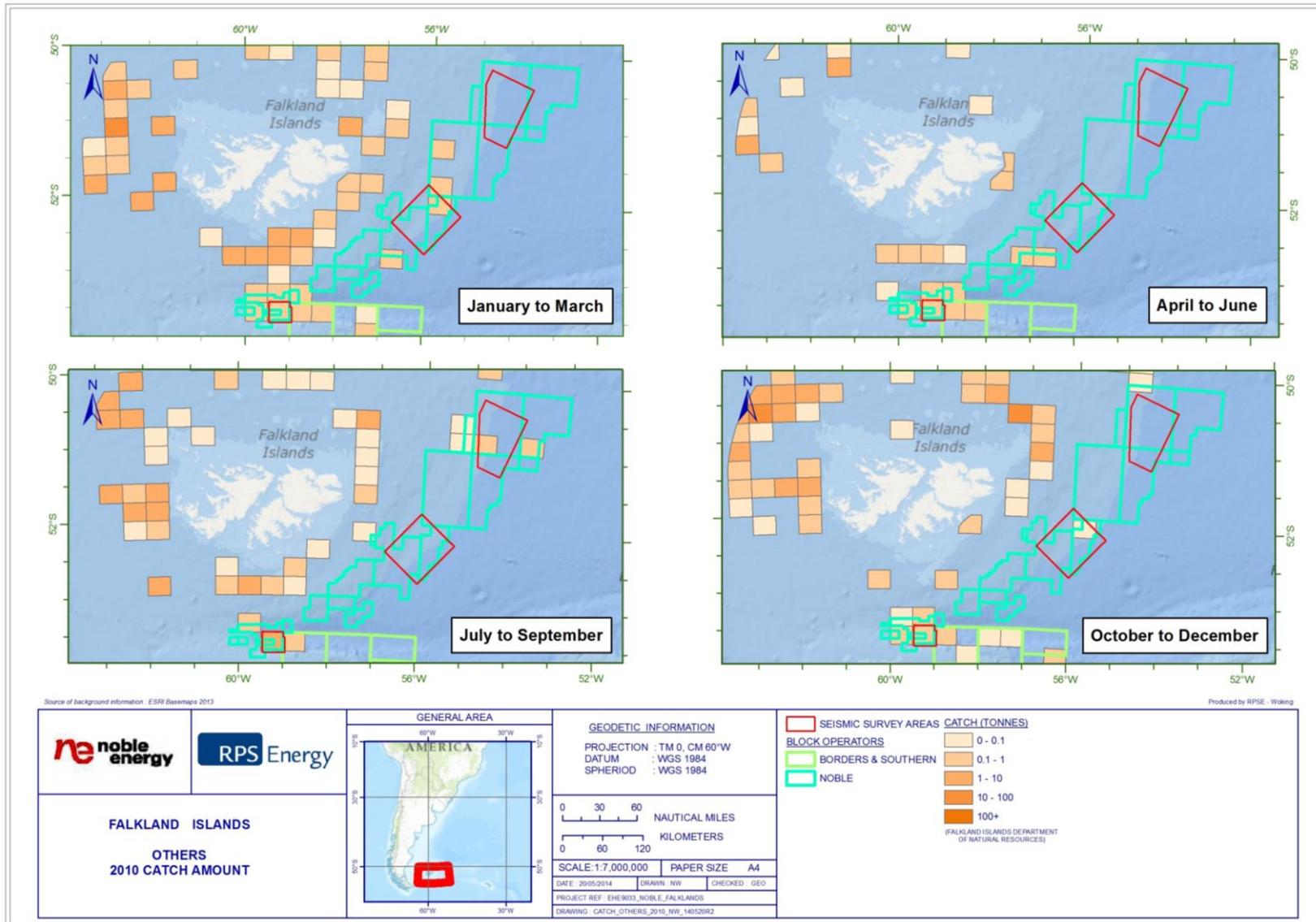


Figure E.12d: Fisheries catch mass (tonnes) for other species for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

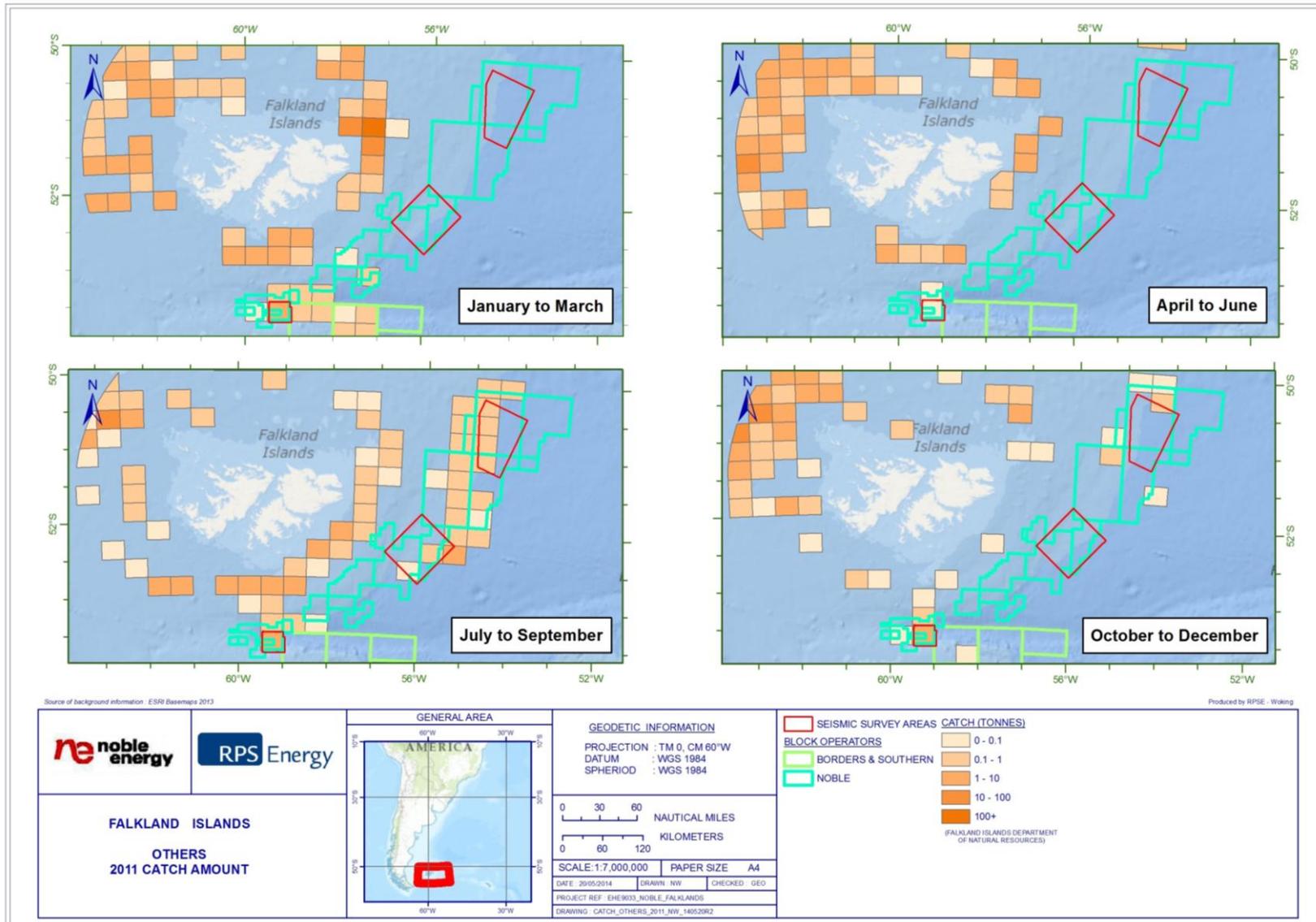


Figure E.12e: Fisheries catch mass (tonnes) for other species for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)

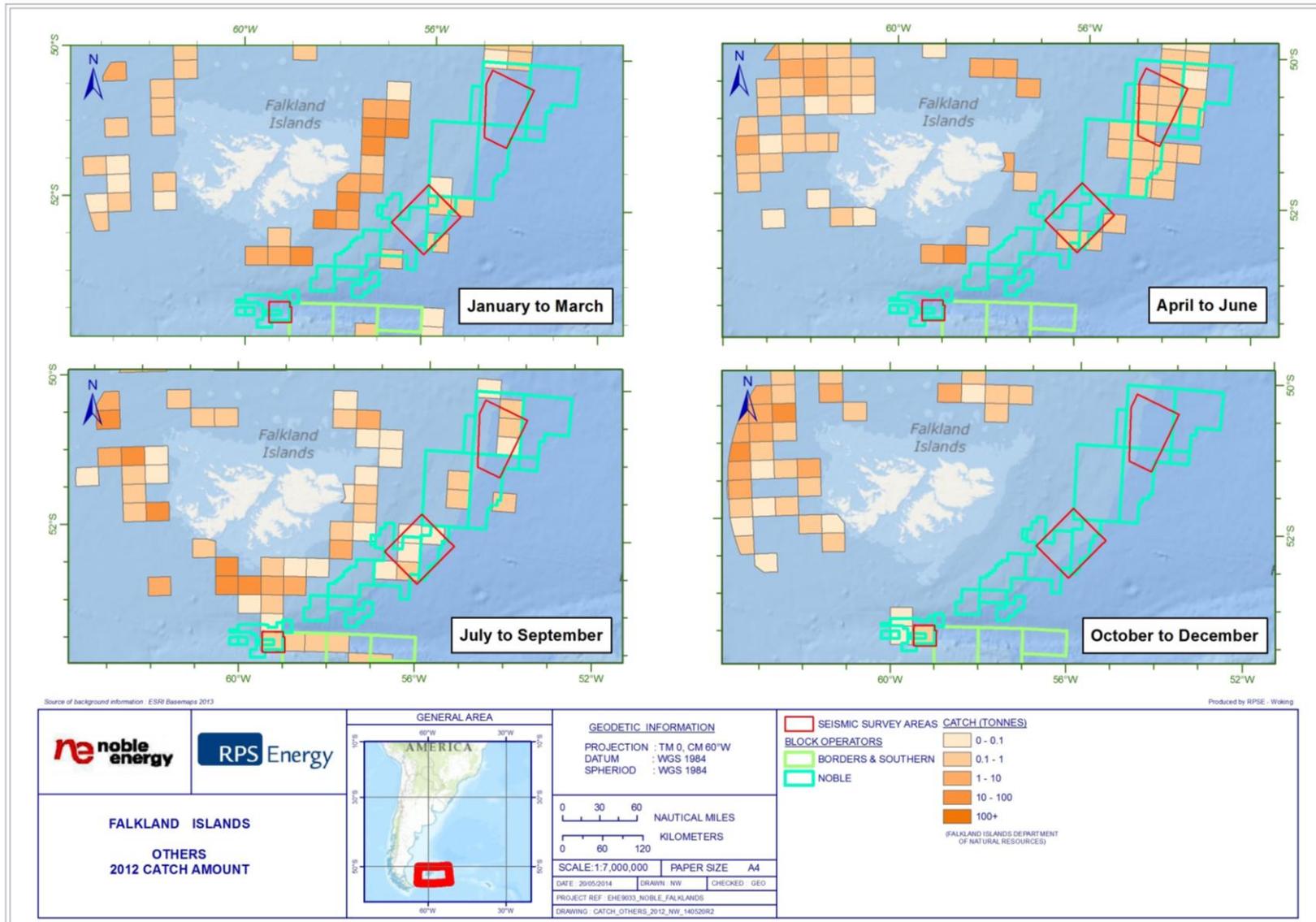


Figure E.12f: Fisheries catch mass (tonnes) for other species for the year 2013 (FIG Department of Natural Resources – Fisheries Department, 2014)

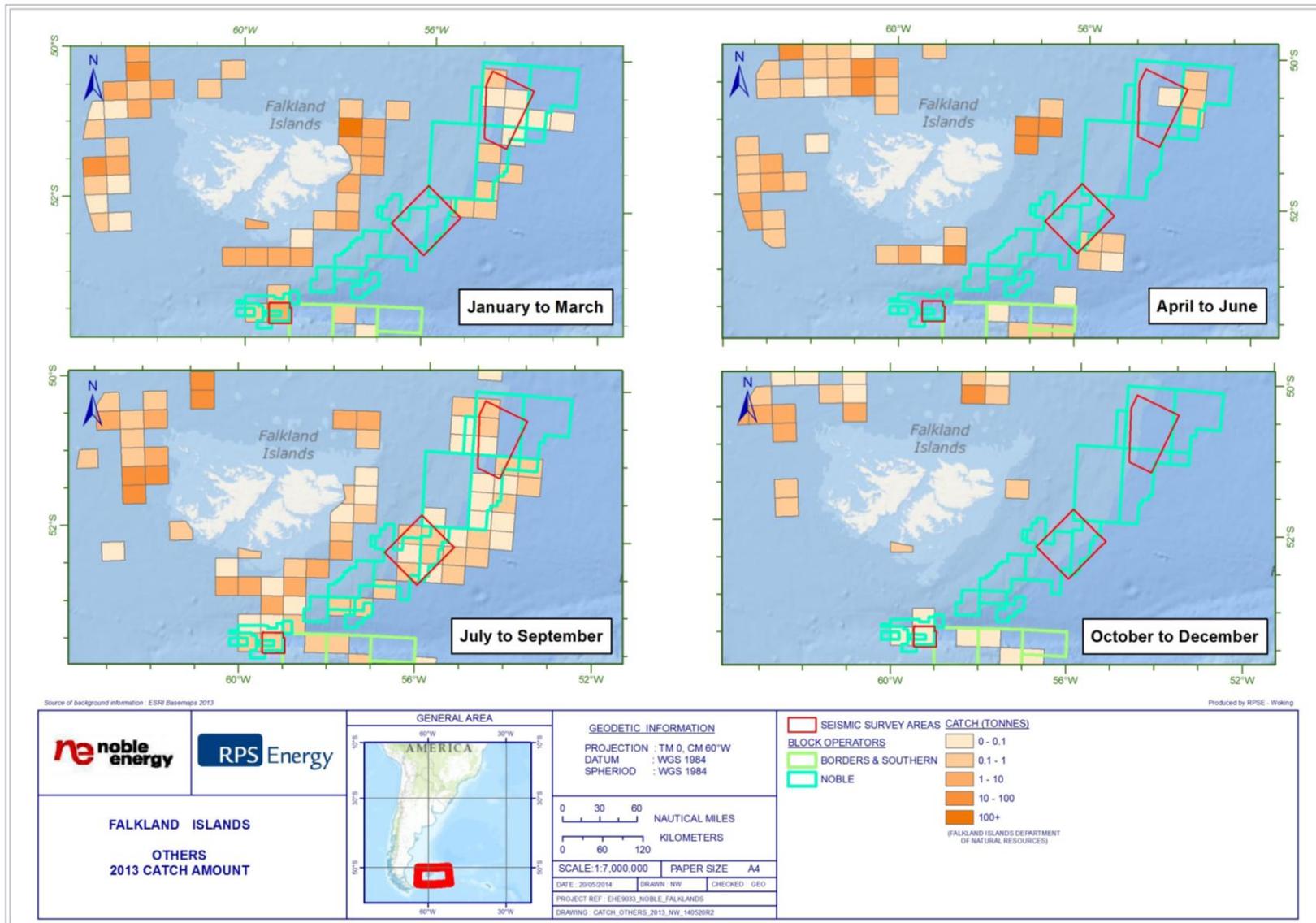


Figure E.13a: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2008 for the months January to March (FIG Department of Natural Resources – Fisheries Department, 2013)

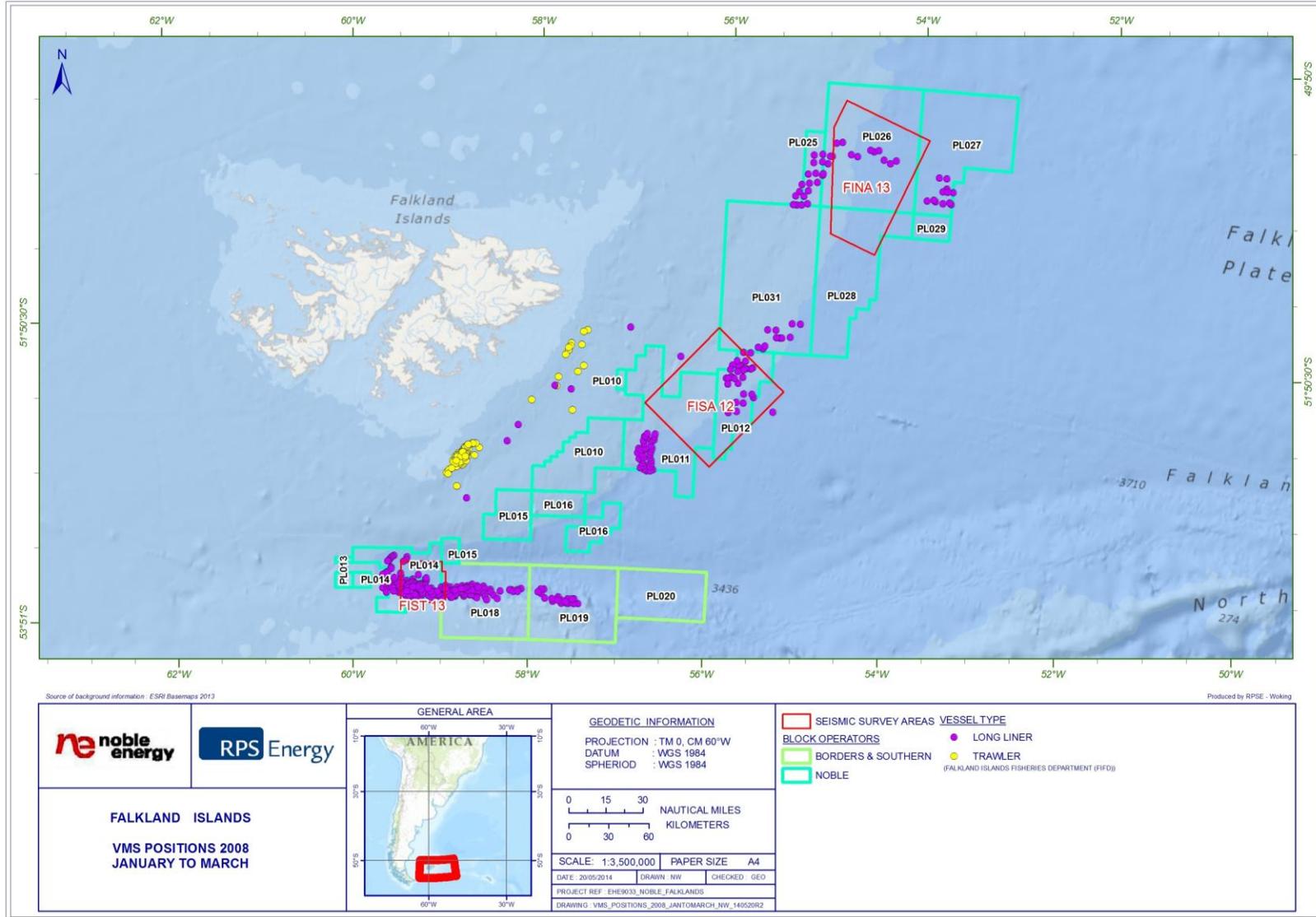


Figure E.13b: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2008 for the months April to June (FIG Department of Natural Resources – Fisheries Department, 2013)

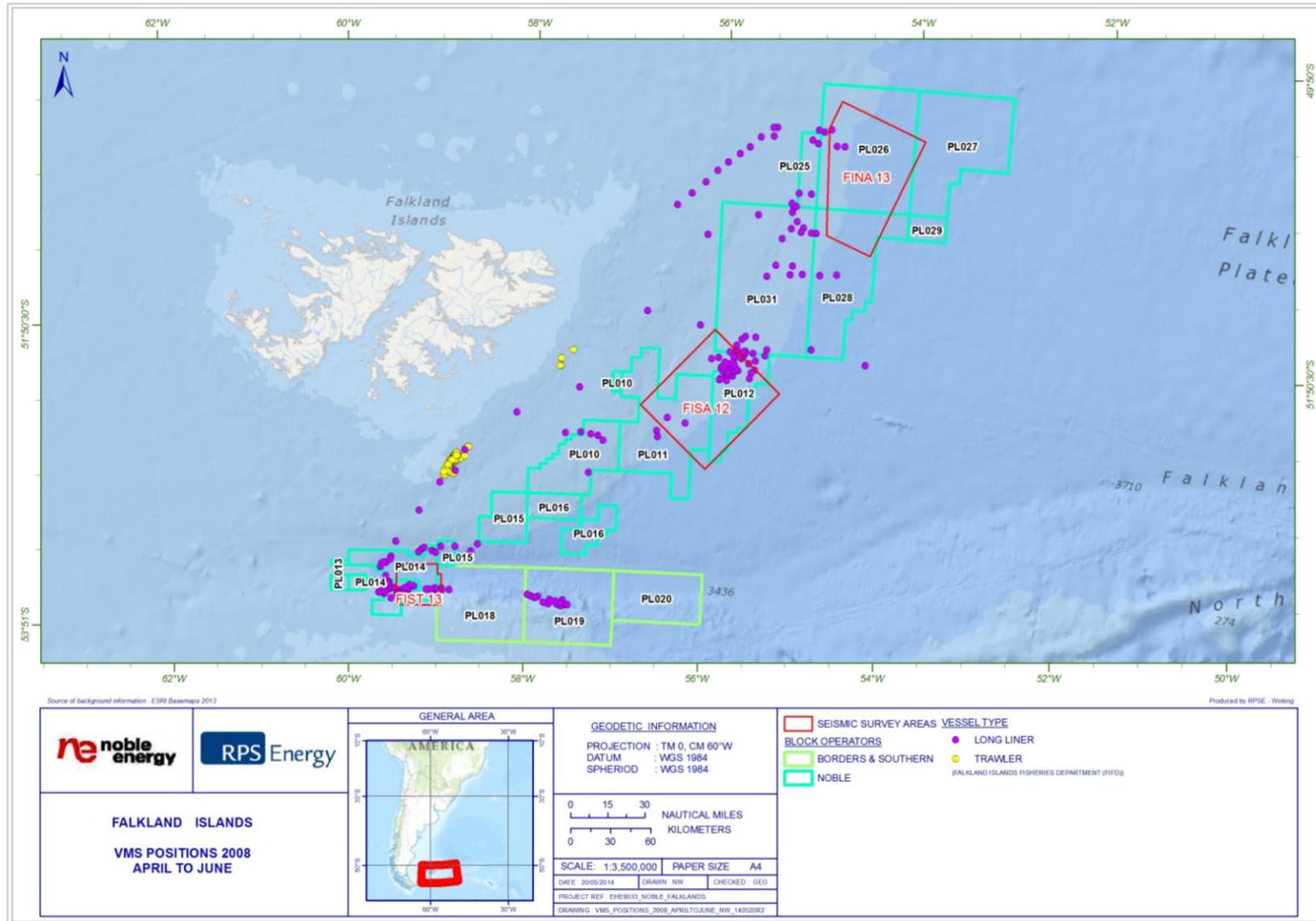


Figure E.13c: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2008 for the months July to September (FIG Department of Natural Resources – Fisheries Department, 2013)

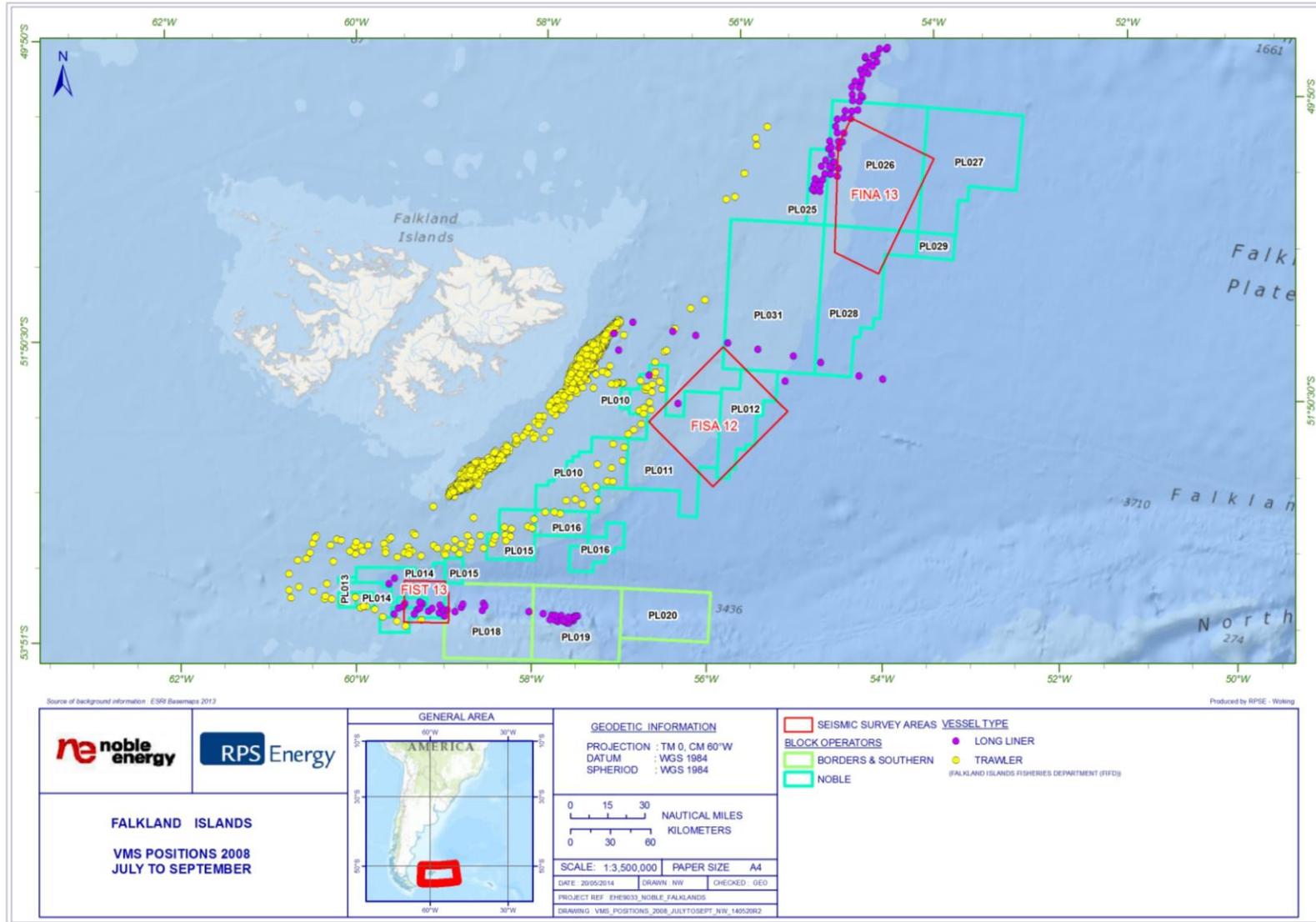


Figure E.13d: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2008 for the months October to December (FIG Department of Natural Resources – Fisheries Department, 2013)

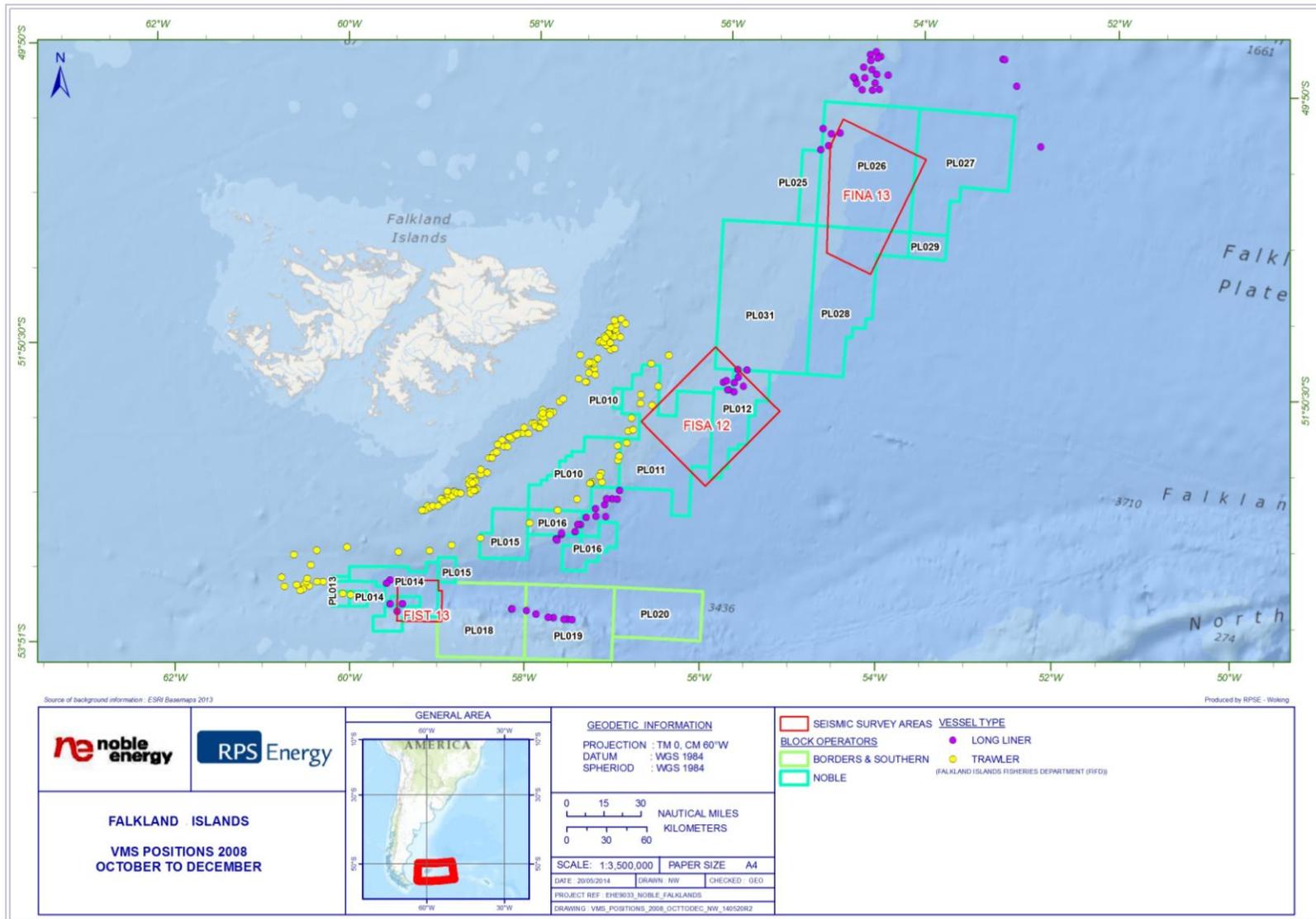


Figure E.13e: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2008, all months (FIG Department of Natural Resources – Fisheries Department, 2013)

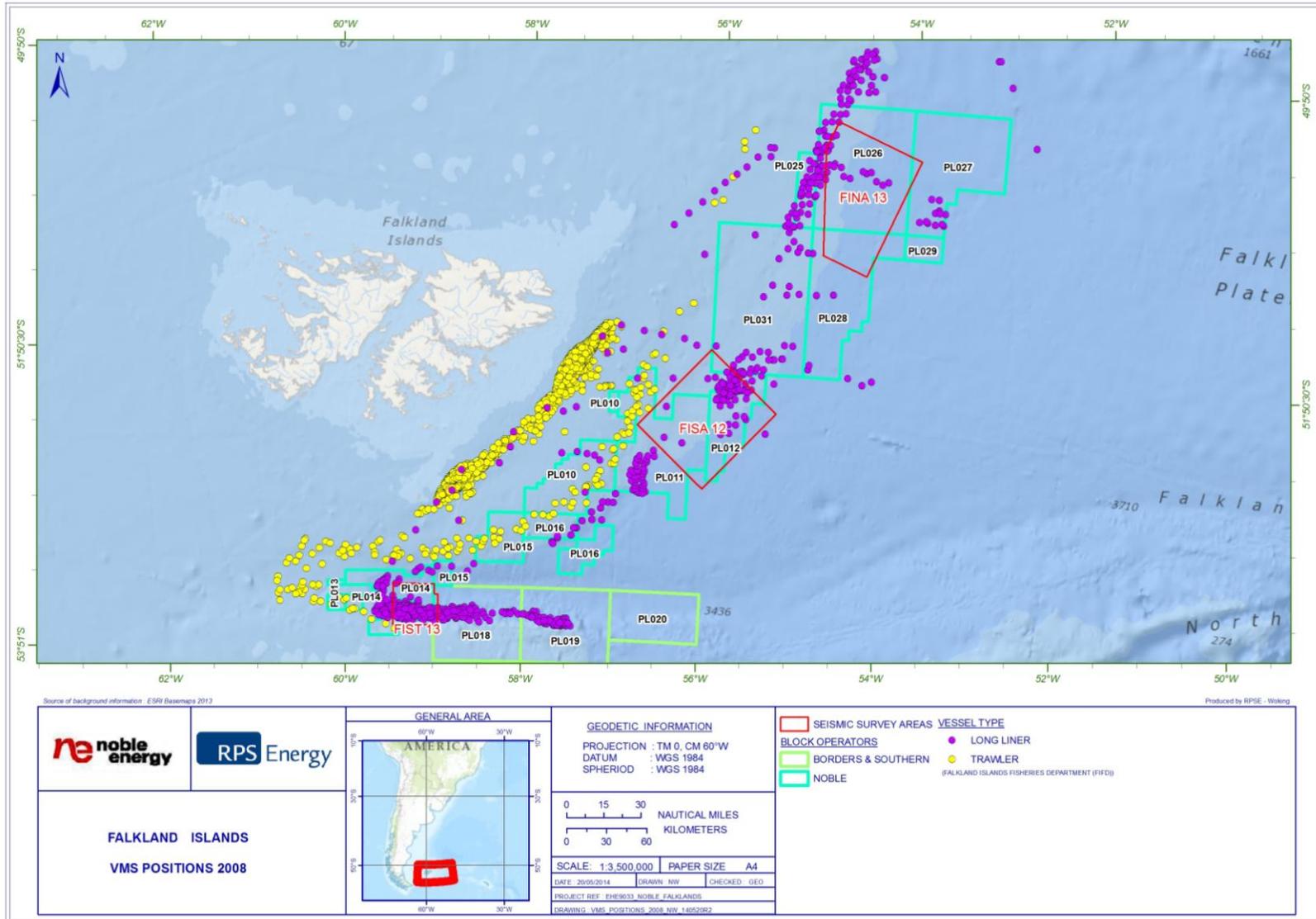


Figure E.14a: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2009 for the months January to March (FIG Department of Natural Resources – Fisheries Department, 2013)

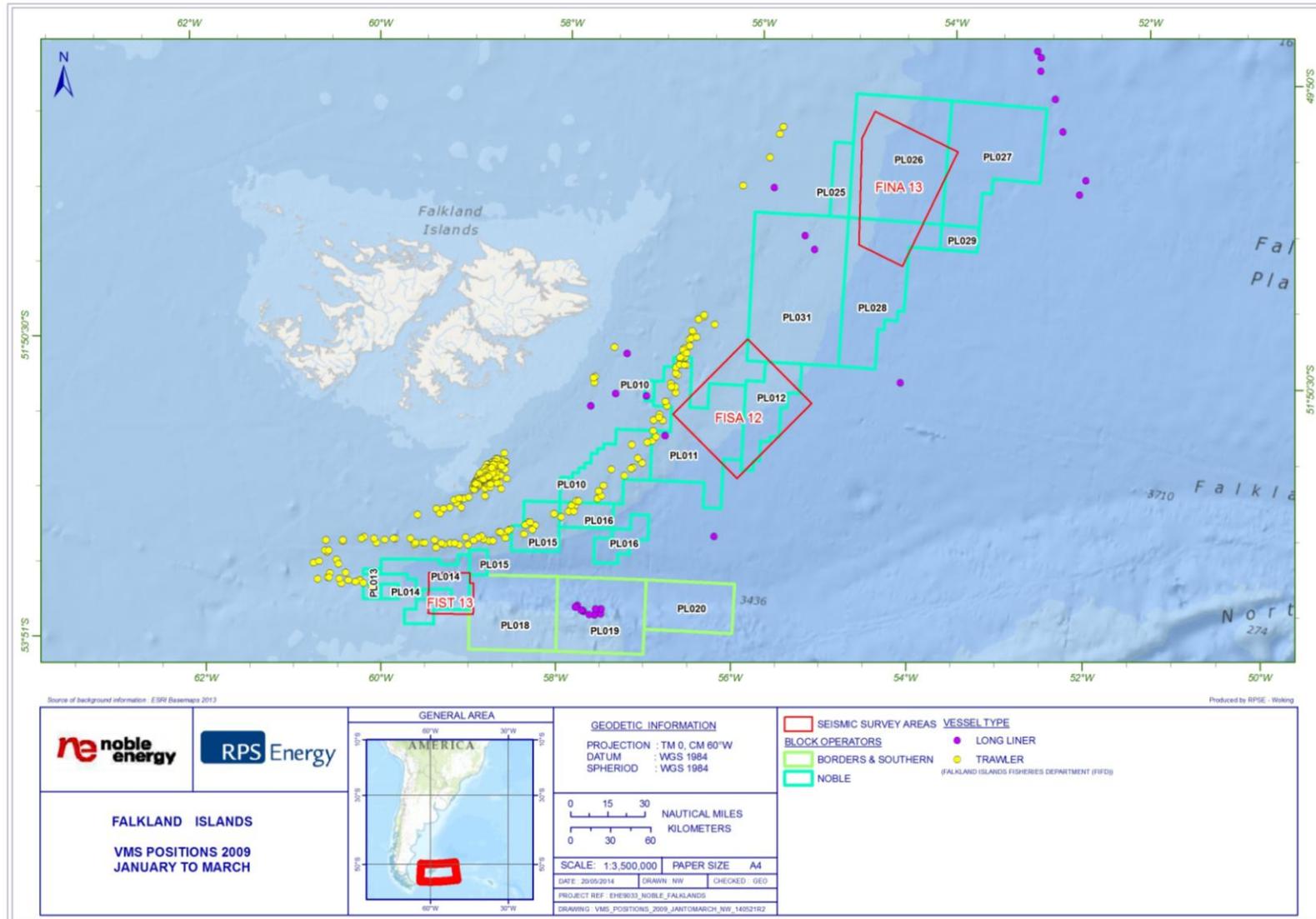


Figure E.14b: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2009 for the months April to June (FIG Department of Natural Resources – Fisheries Department, 2013)

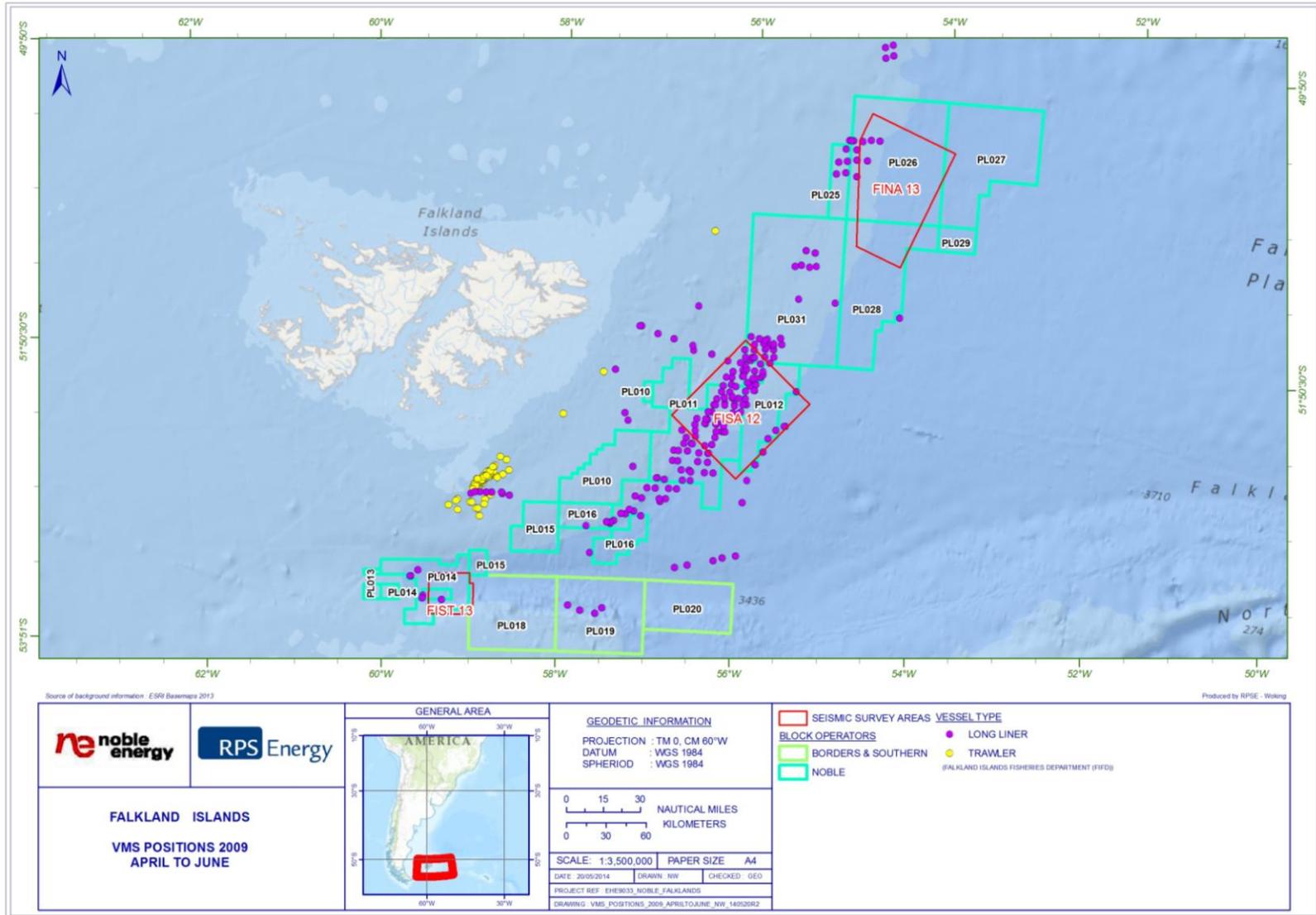


Figure E.14c: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2009 for the months July to September (FIG Department of Natural Resources – Fisheries Department, 2013)

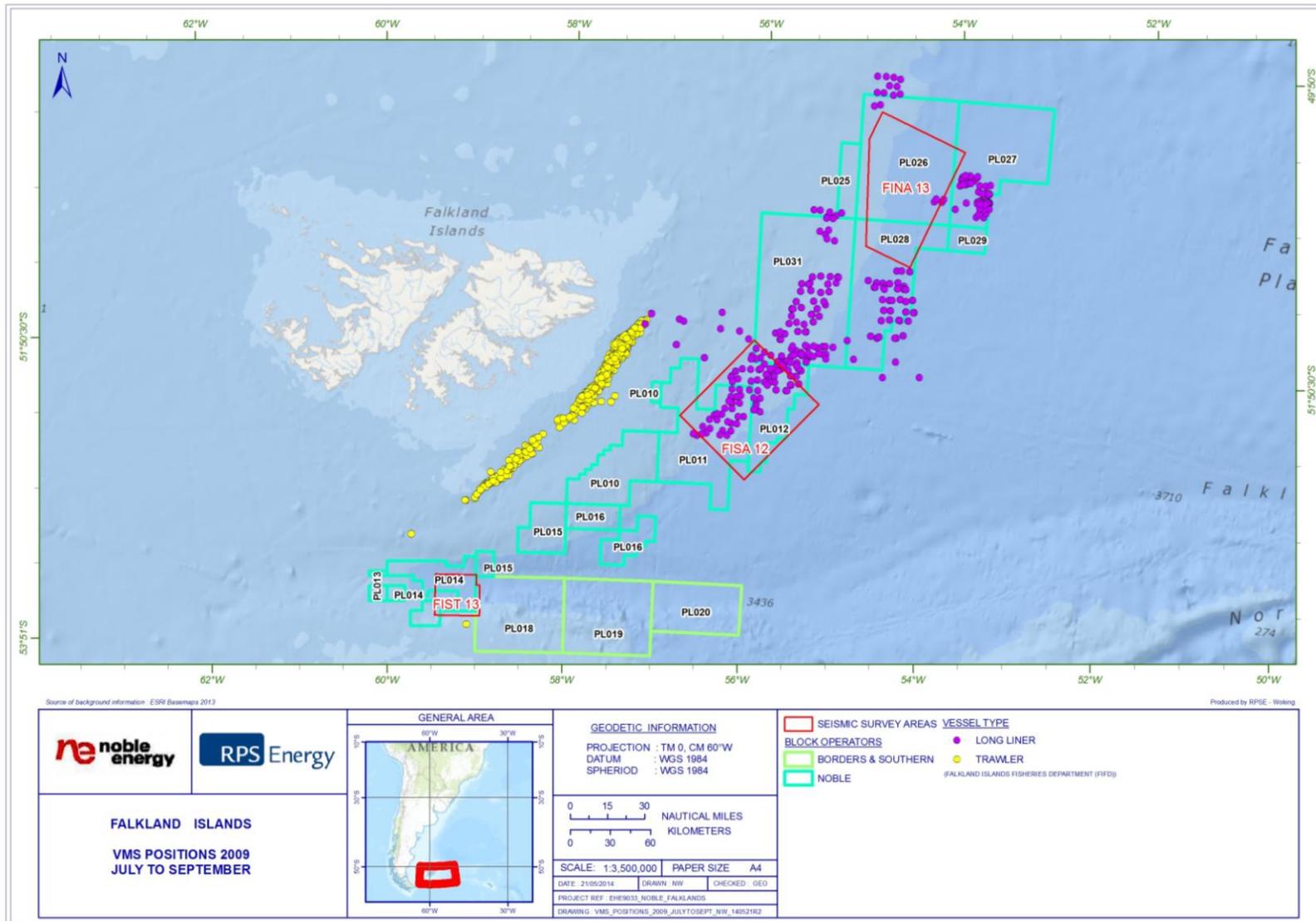


Figure E.14d: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2009 for the months October to December (FIG Department of Natural Resources – Fisheries Department, 2013)

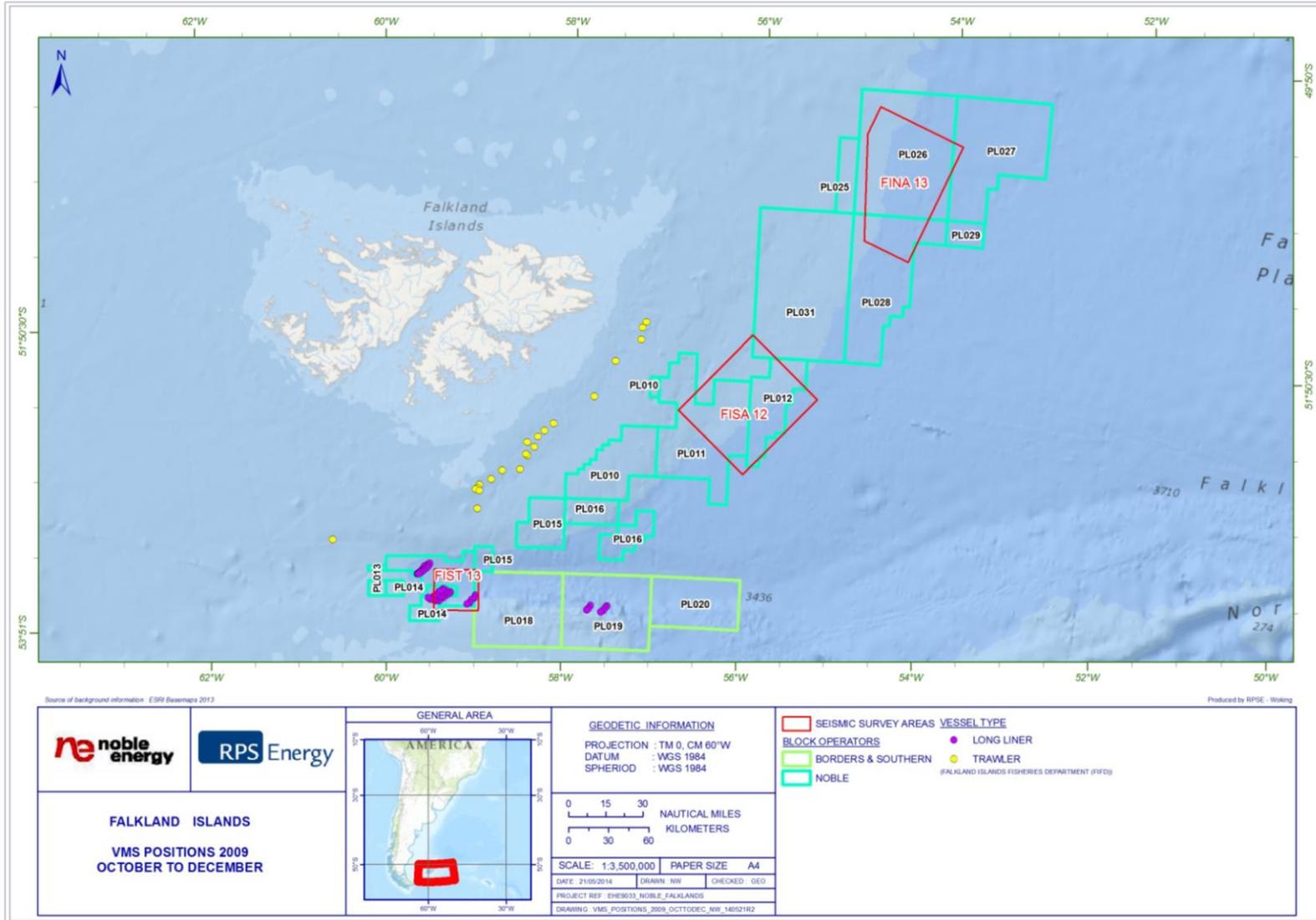


Figure E.14e: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2009, all months (FIG Department of Natural Resources – Fisheries Department, 2013)

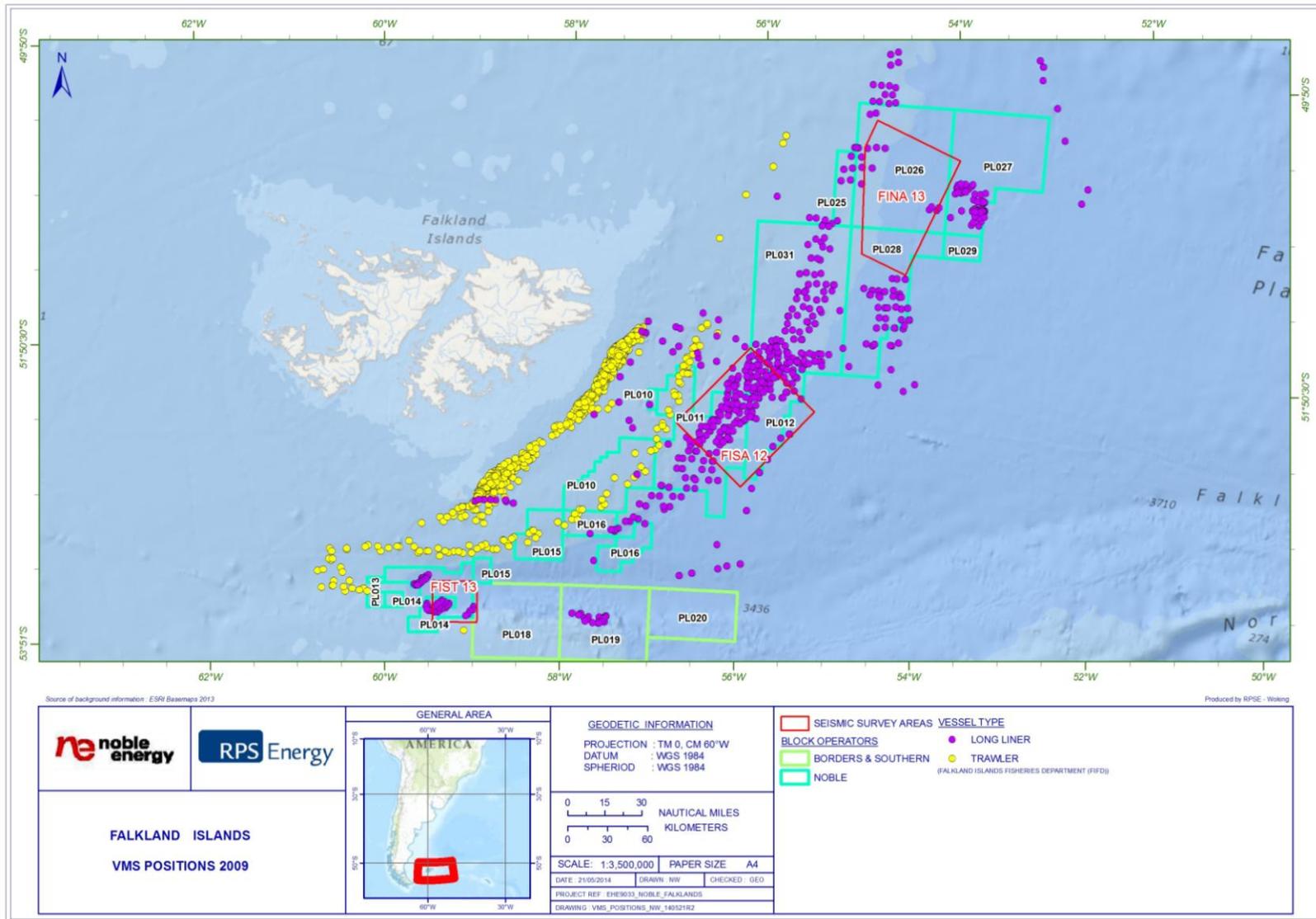


Figure E.15a: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2010 for the months January to March (FIG Department of Natural Resources – Fisheries Department, 2013)

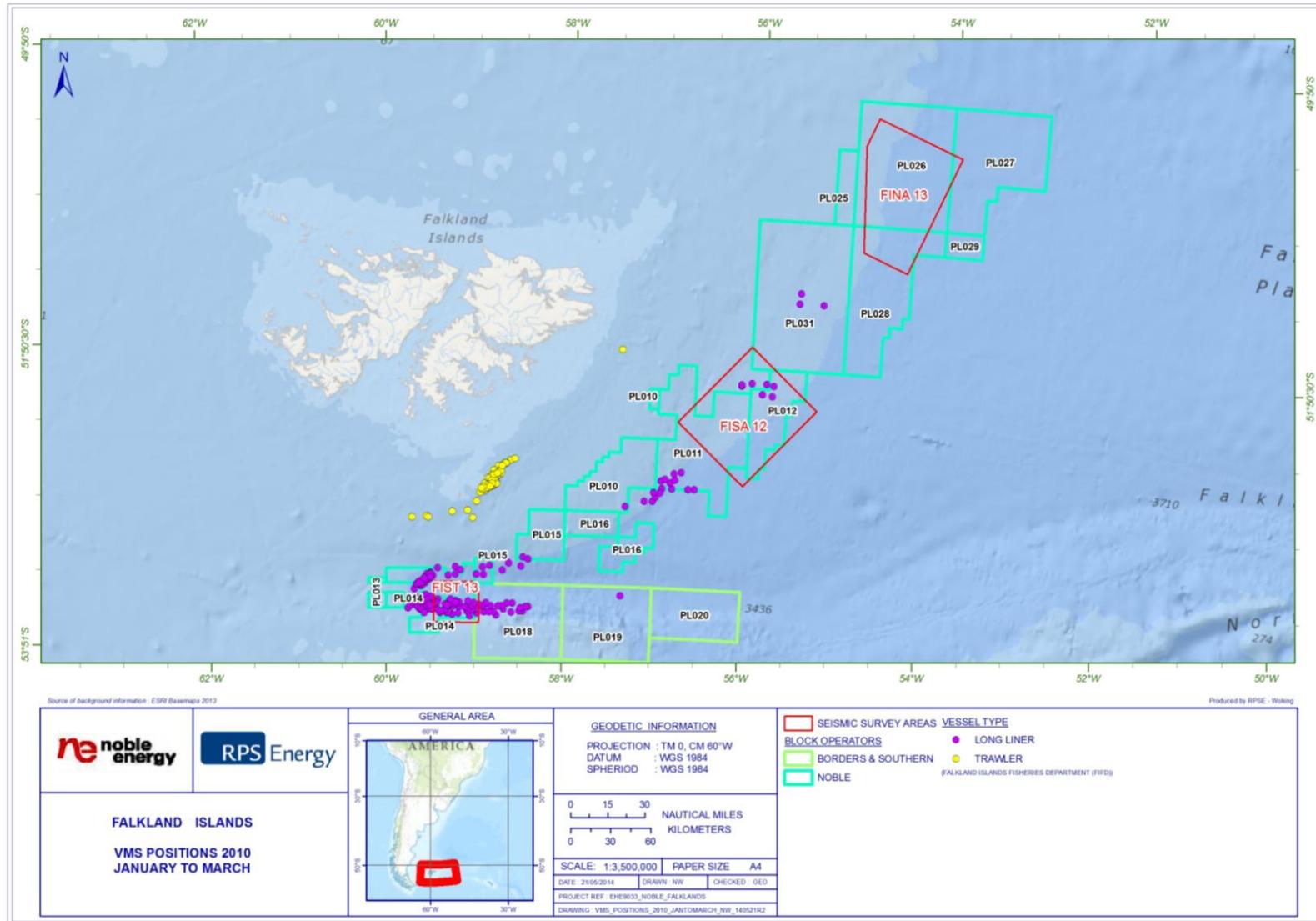


Figure E.15b: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2010 for the months April to June (FIG Department of Natural Resources – Fisheries Department, 2013)

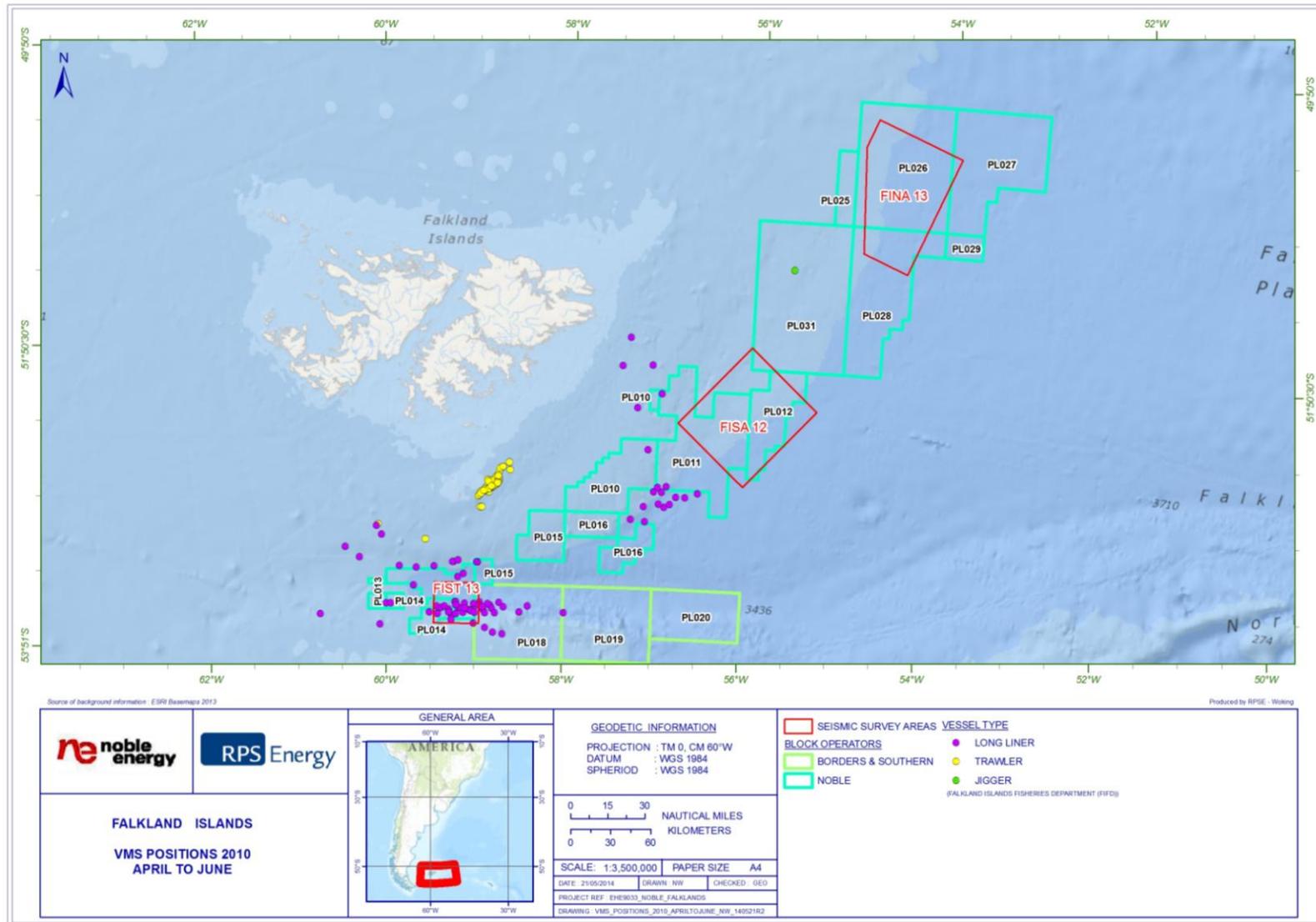


Figure E.15c: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2010 for the months July to September (FIG Department of Natural Resources – Fisheries Department, 2013)

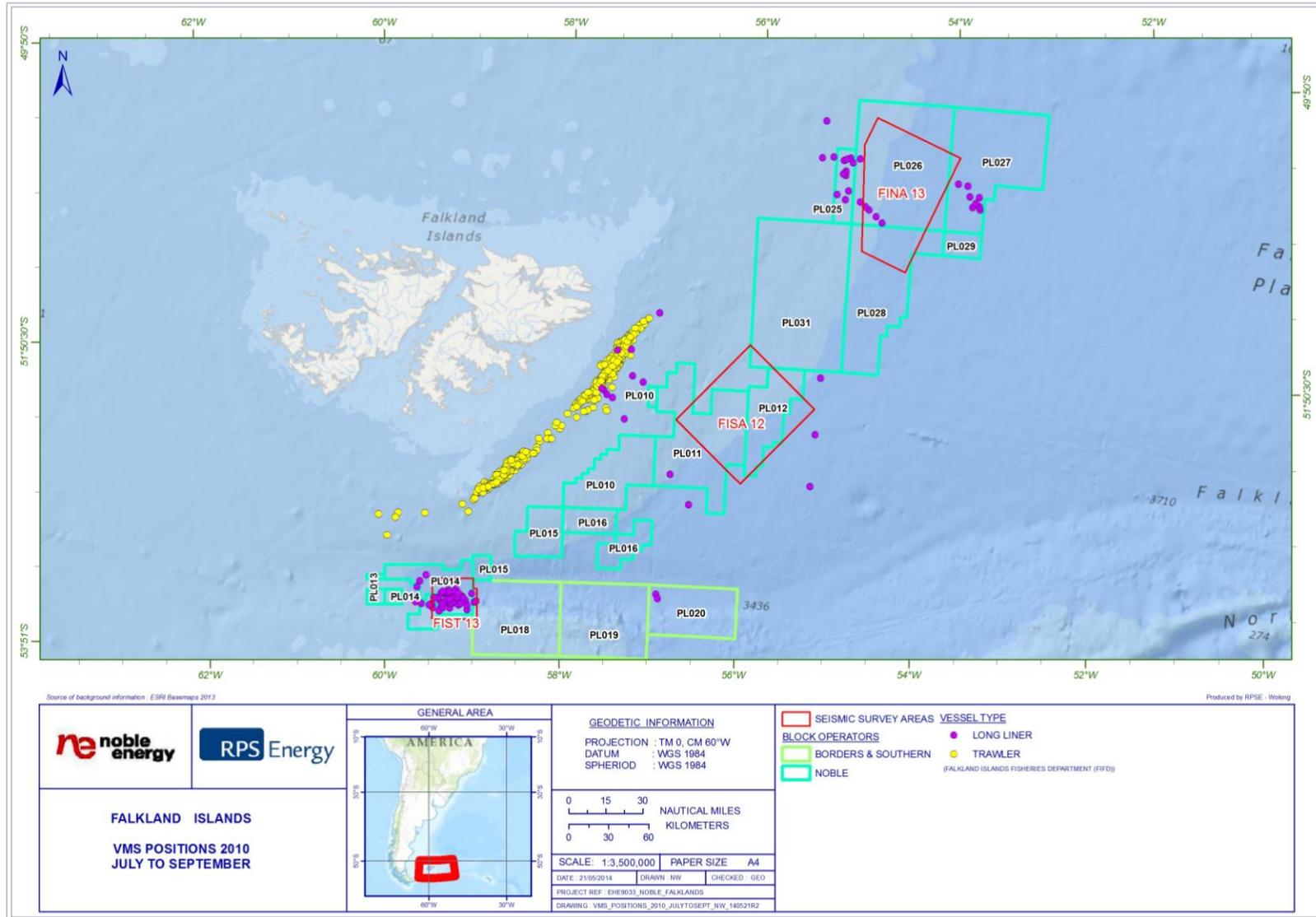


Figure E.15d: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2010 for the months October to December (FIG Department of Natural Resources – Fisheries Department, 2013)

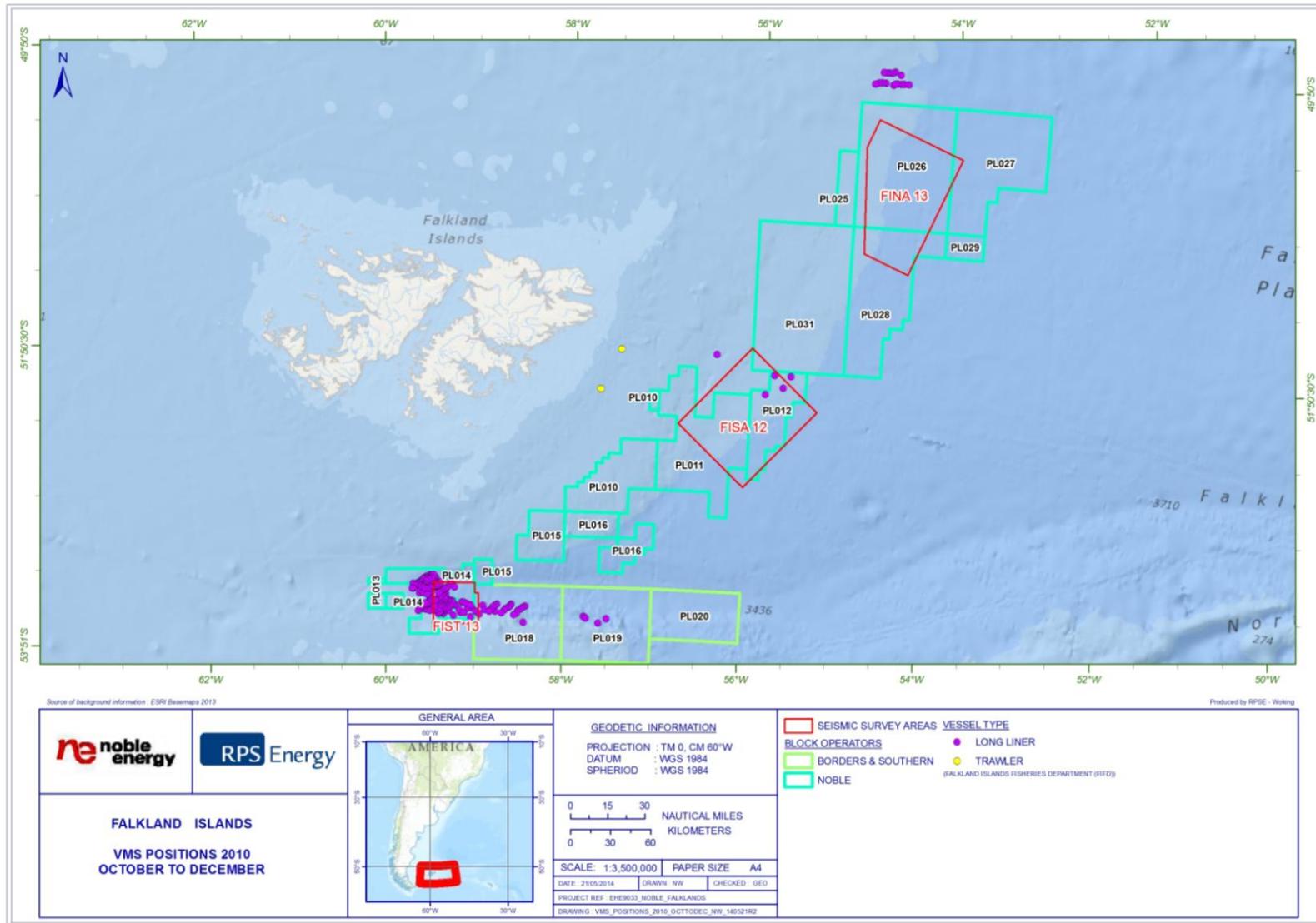


Figure E.15e: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2010, all months (FIG Department of Natural Resources – Fisheries Department, 2013)

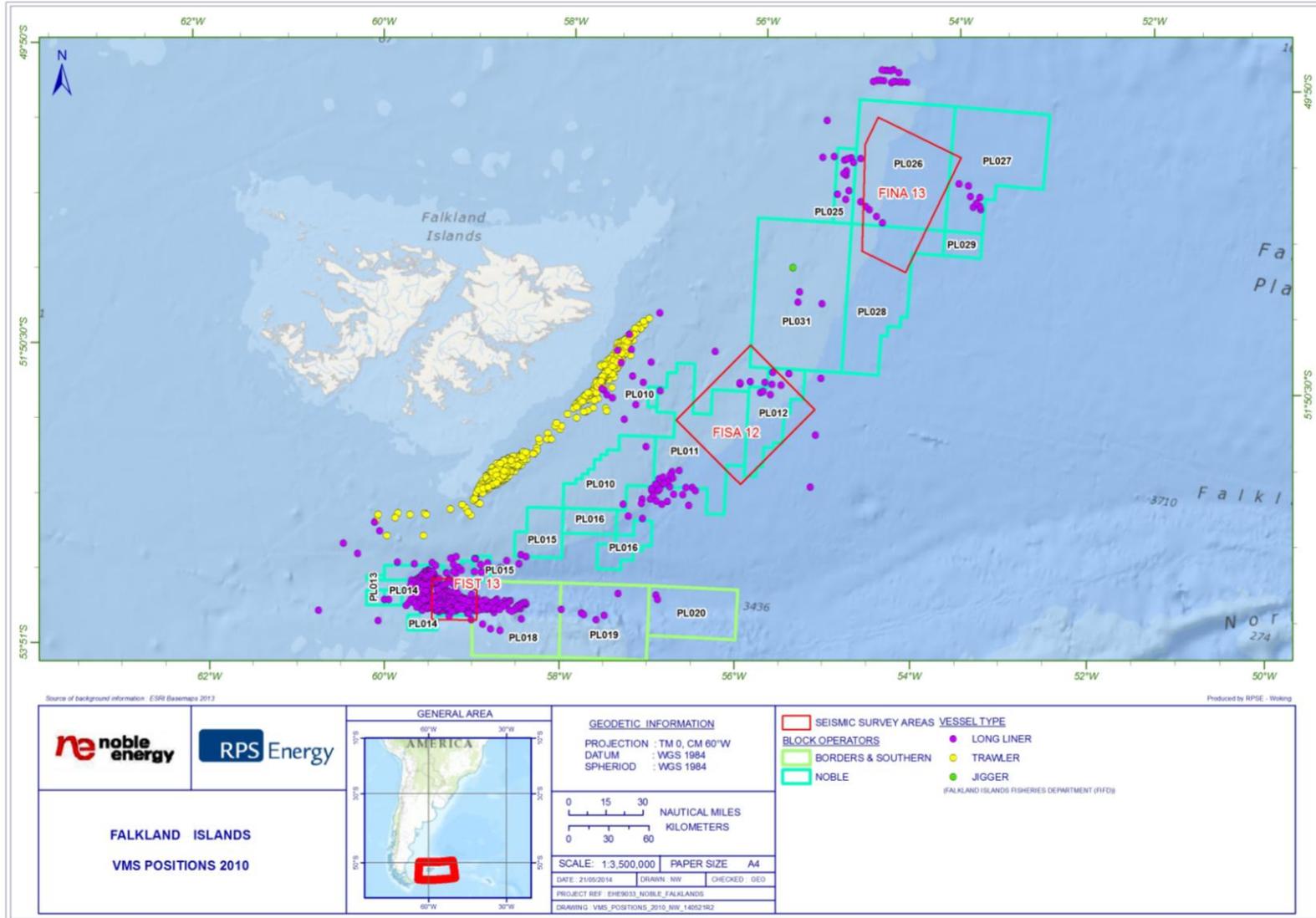


Figure E.16a: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2011 from January to March (FIG Department of Natural Resources – Fisheries Department, 2013)

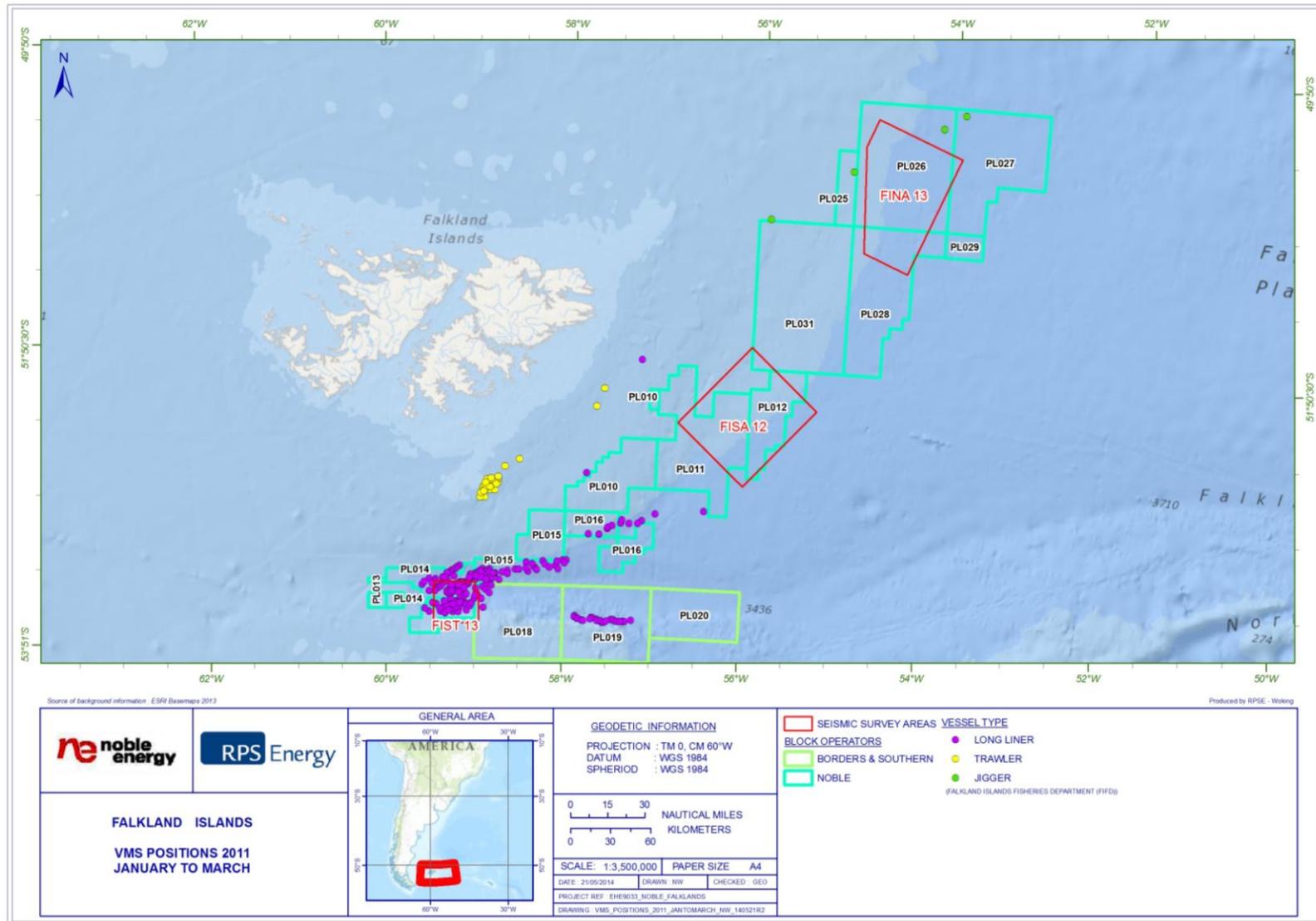


Figure E.16b: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2011 from April to June (FIG Department of Natural Resources – Fisheries Department, 2013)

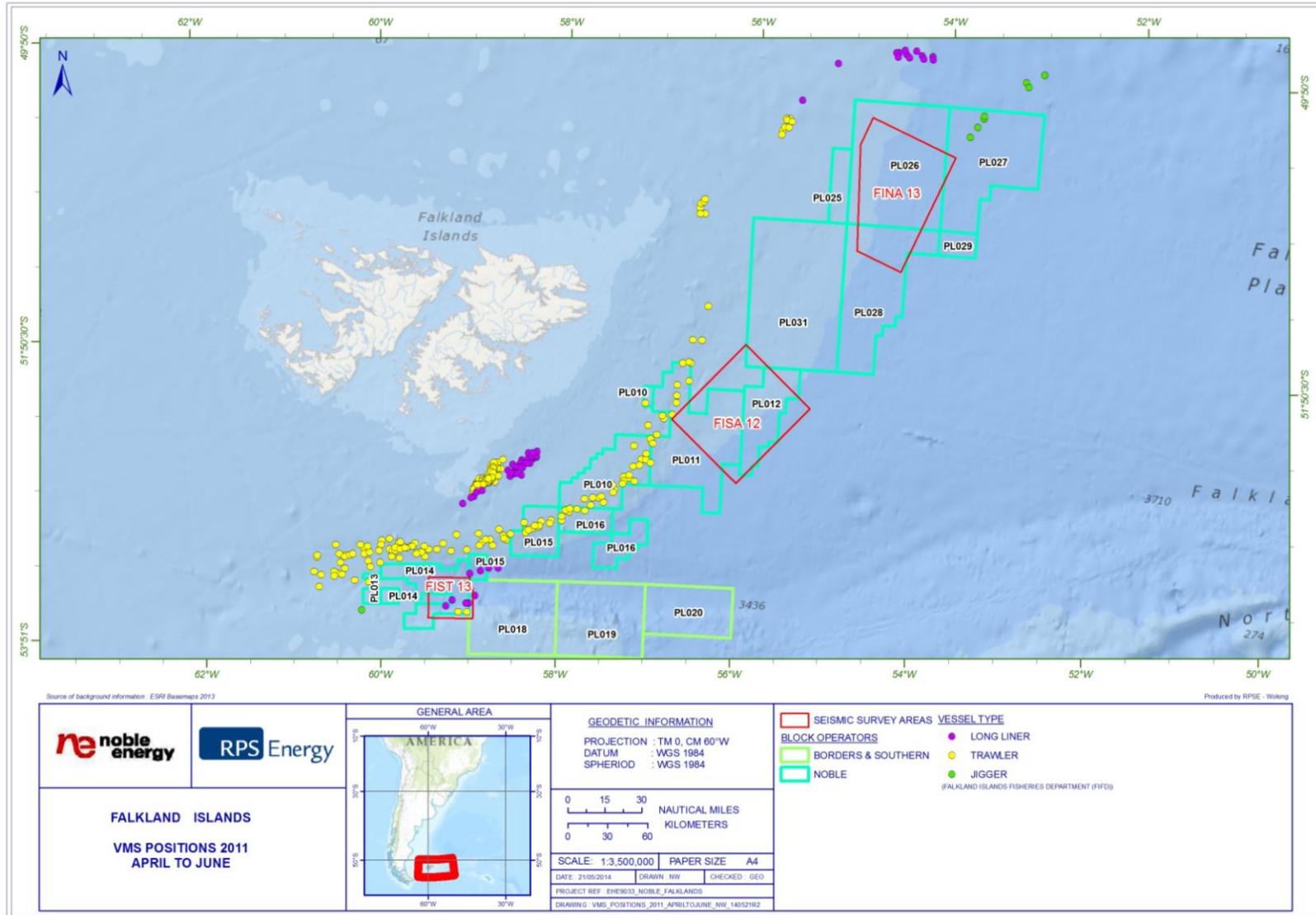


Figure E.16c: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2011 from July to September (FIG Department of Natural Resources – Fisheries Department, 2013)

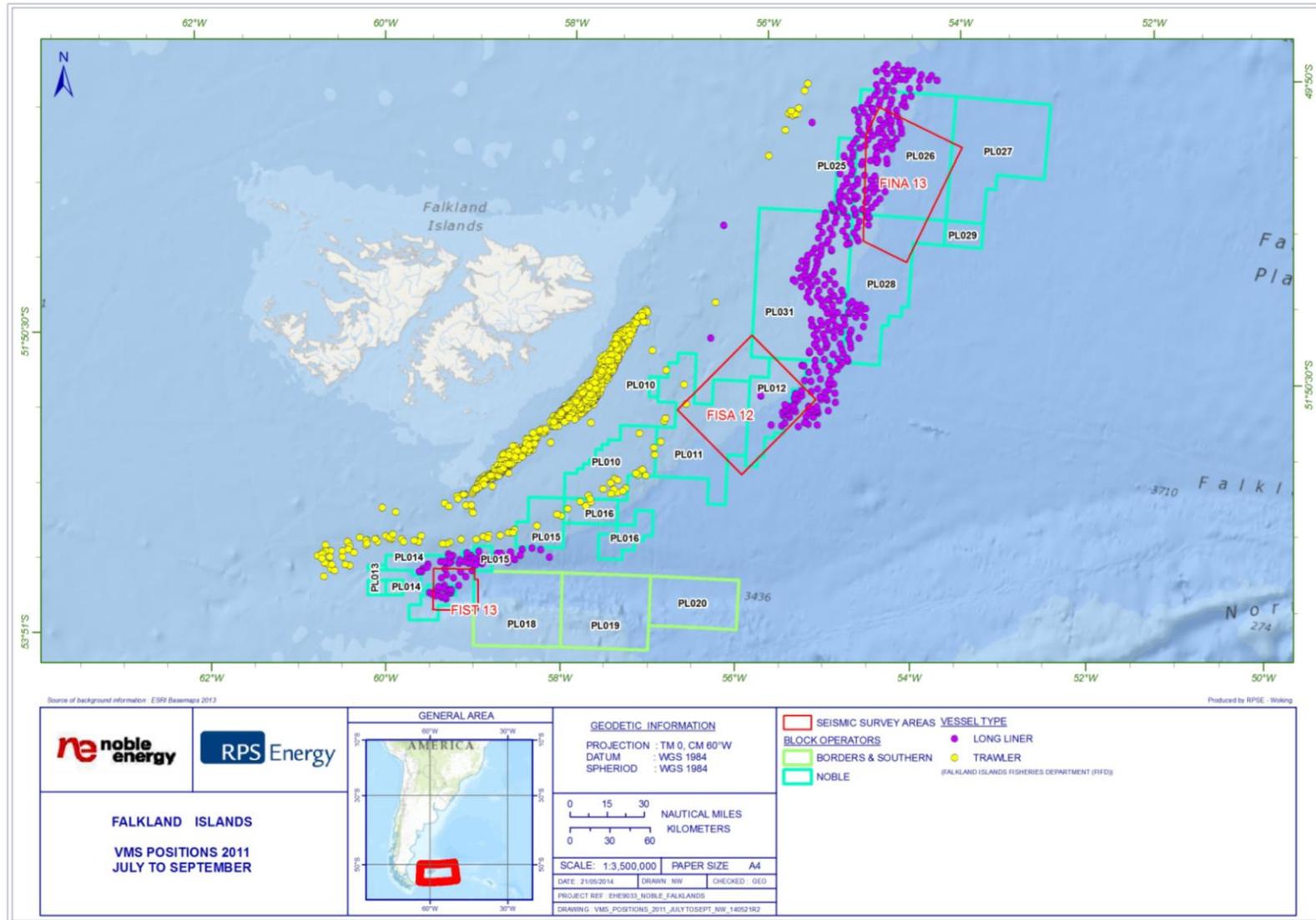


Figure E.16d: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2011 from October to December (FIG Department of Natural Resources – Fisheries Department, 2013)

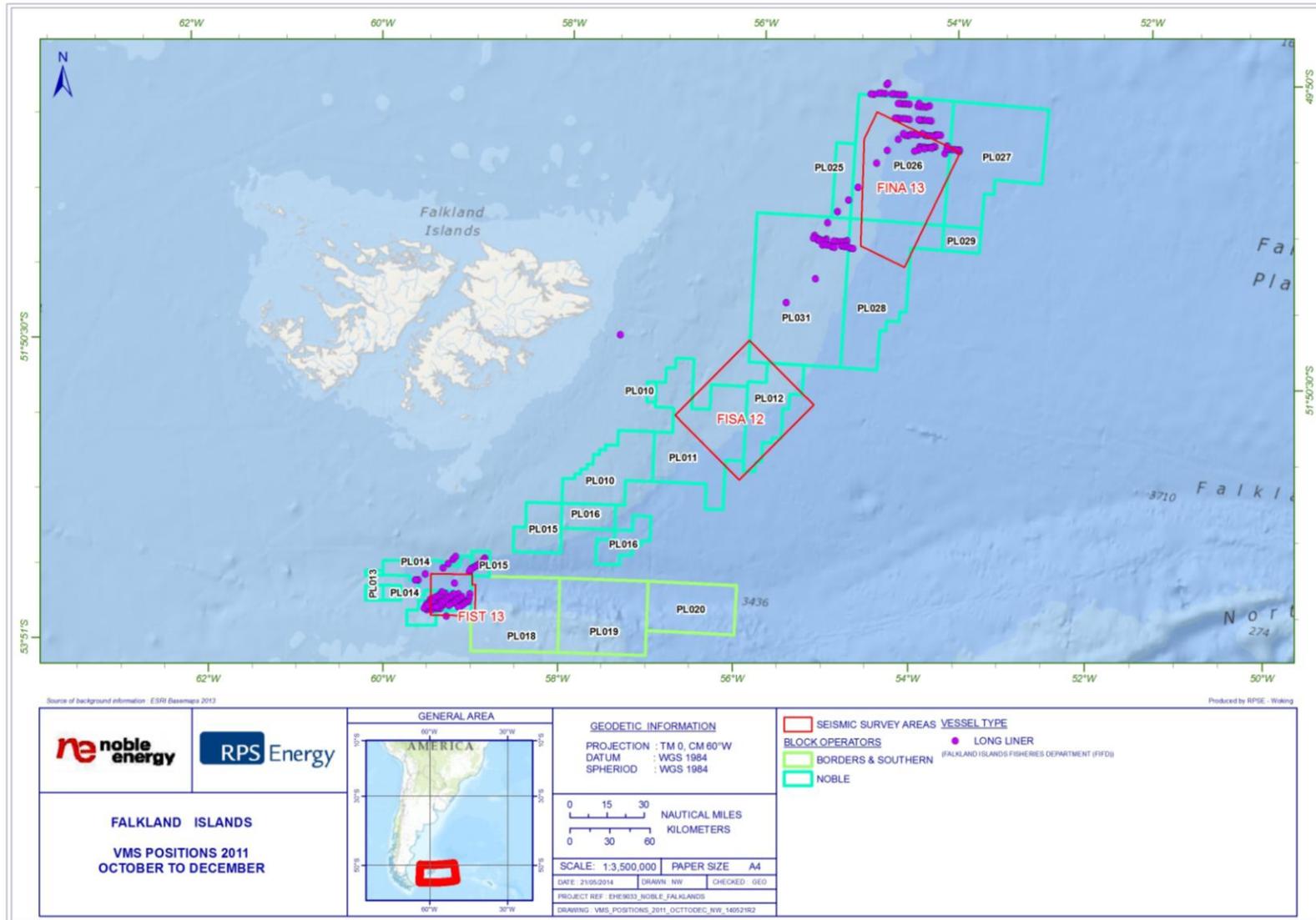


Figure E.16e: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2011, all months (FIG Department of Natural Resources – Fisheries Department, 2013)

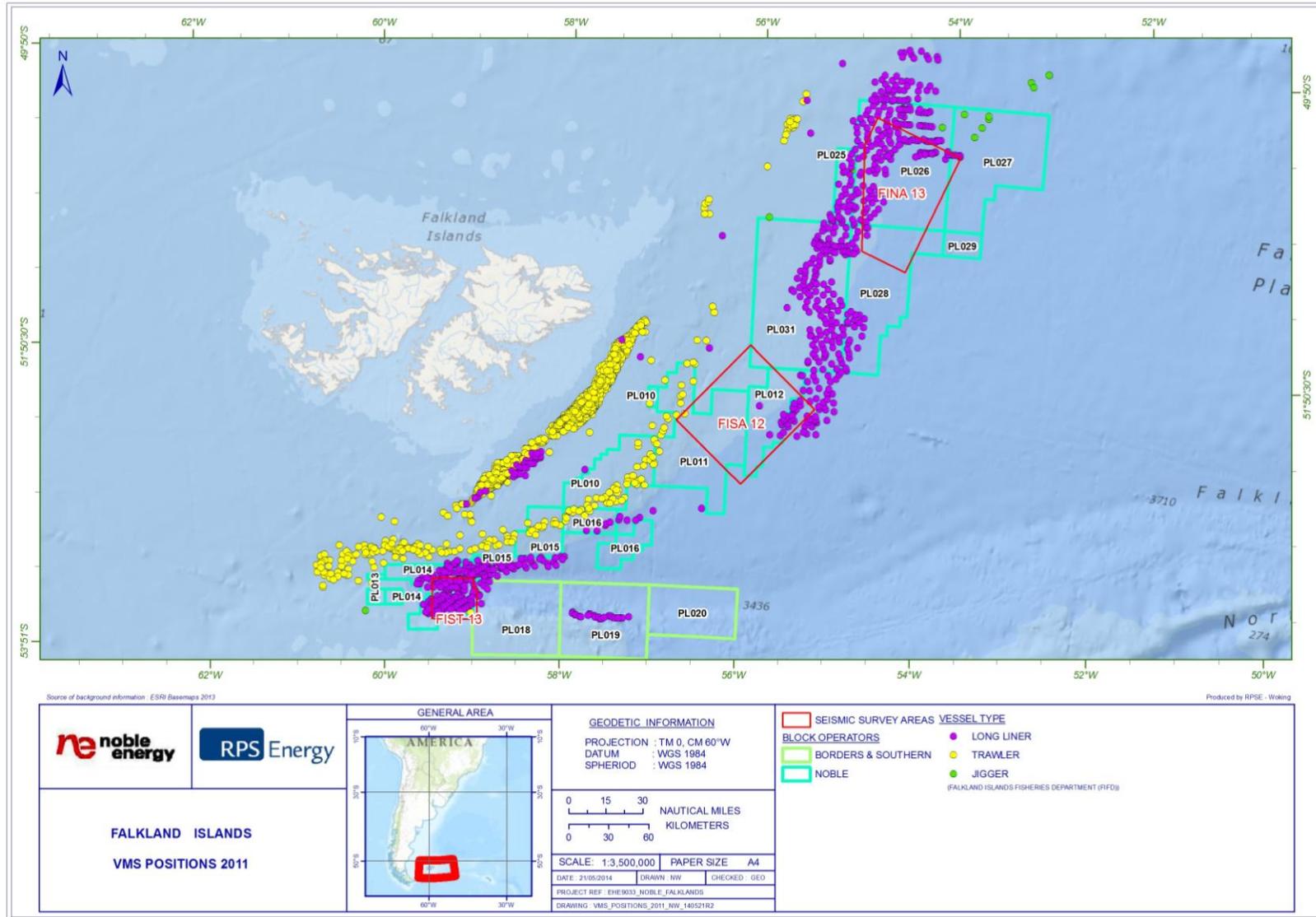


Figure E.17a: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2012 for January to March (FIG Department of Natural Resources – Fisheries Department, 2013)

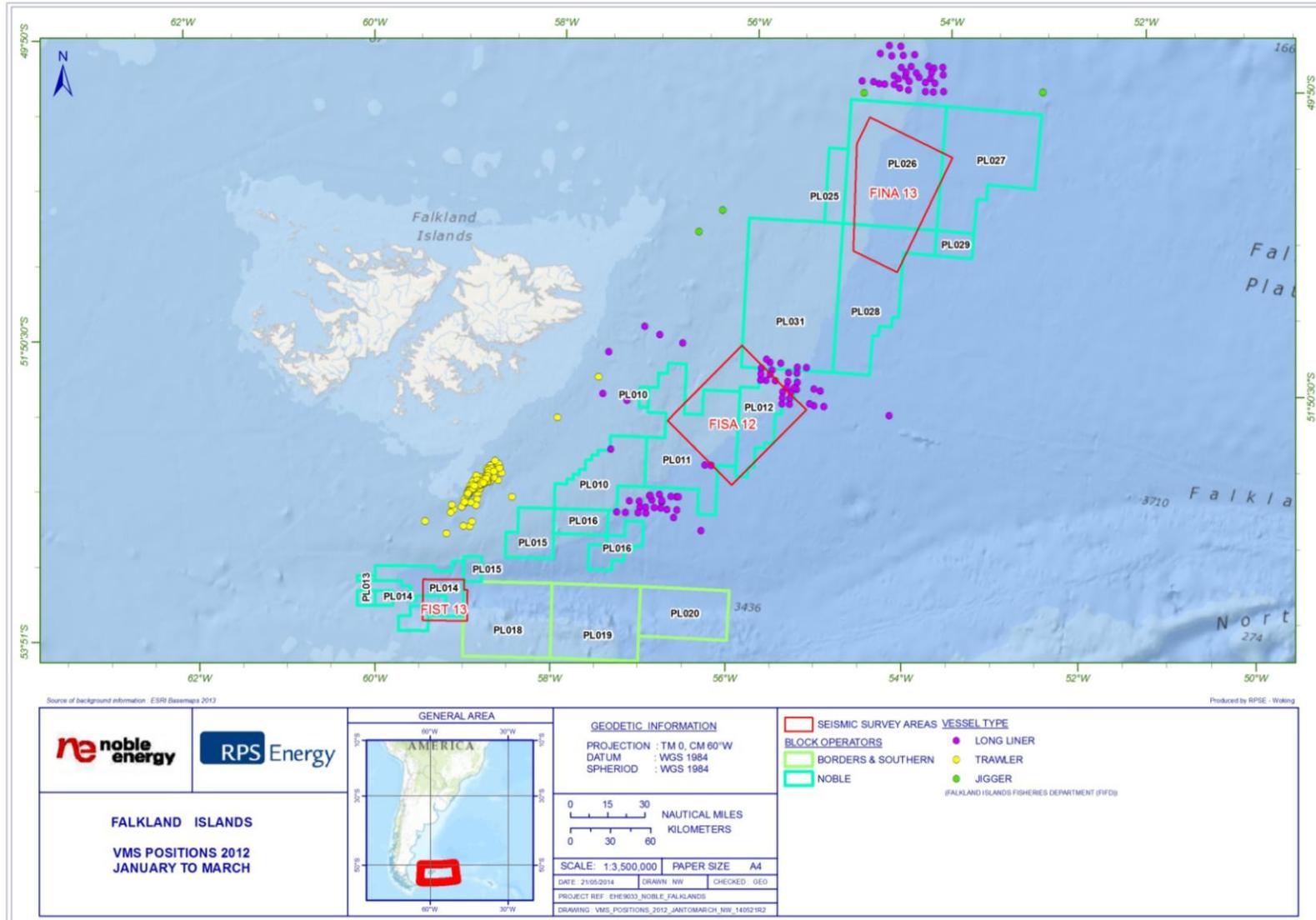


Figure E.17b: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2012 for April to June (FIG Department of Natural Resources – Fisheries Department, 2013)

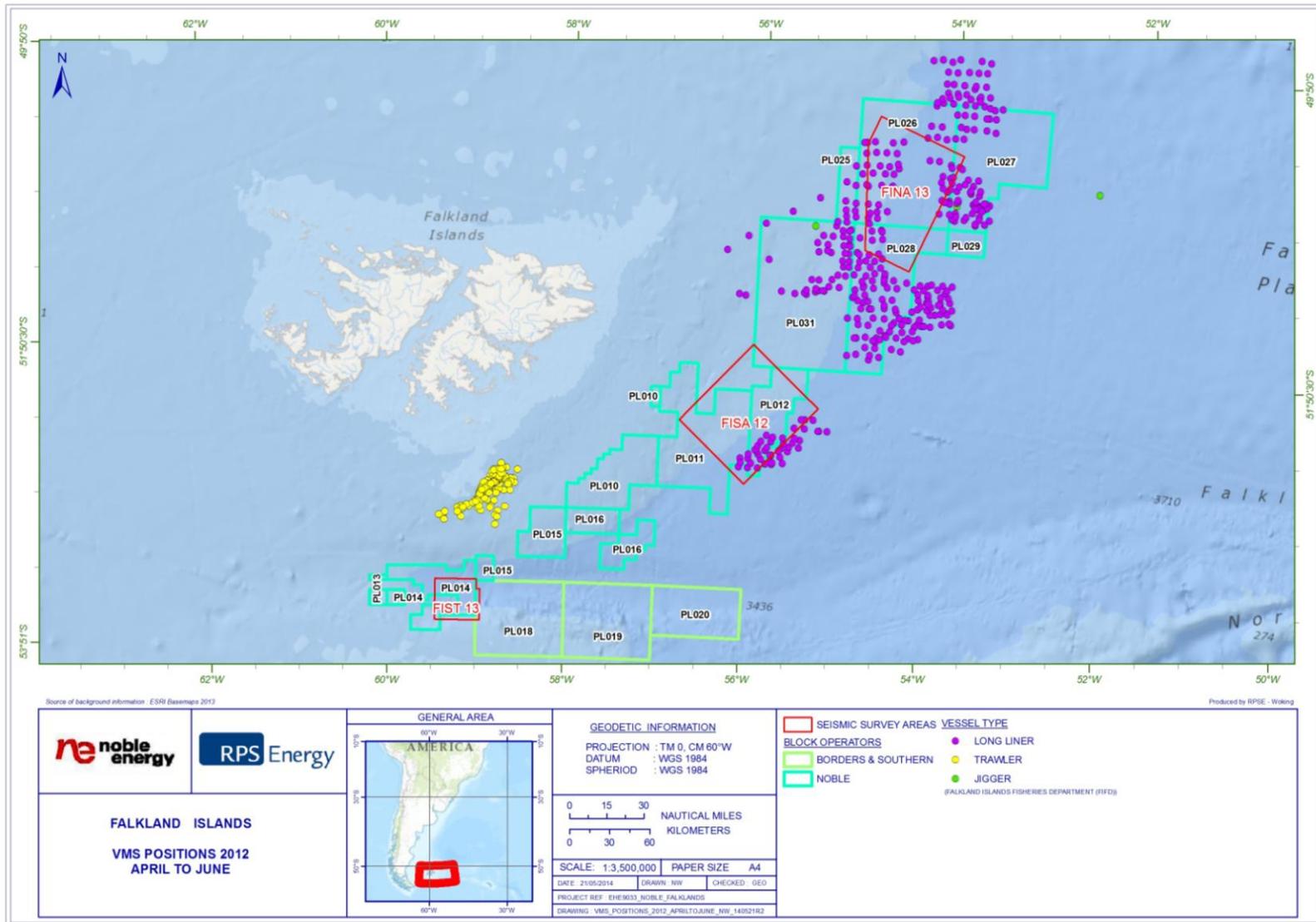


Figure E.17c: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2012 for July to September (FIG Department of Natural Resources – Fisheries Department, 2013)

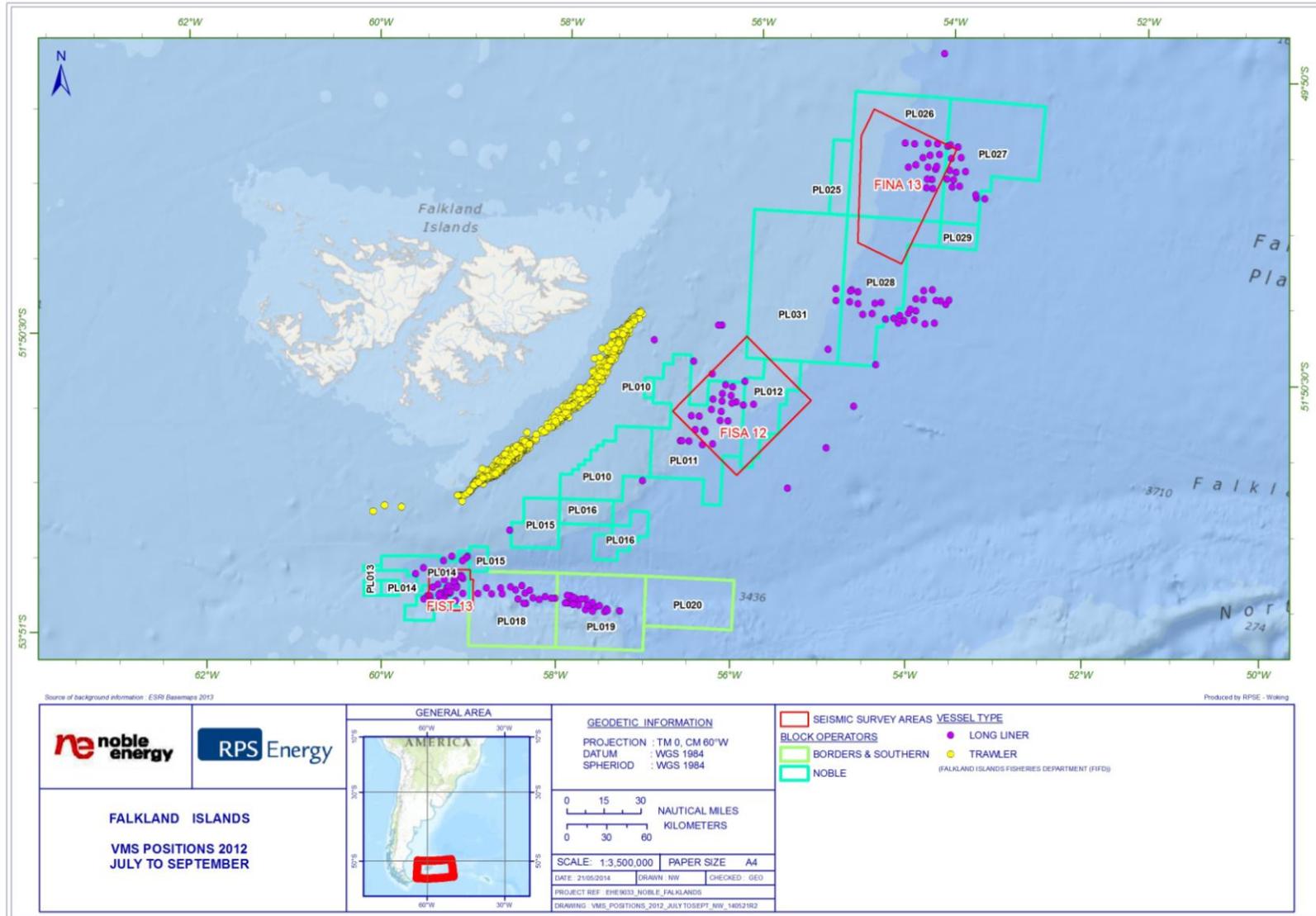


Figure E.17d: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2012 for October to December (FIG Department of Natural Resources – Fisheries Department, 2013)

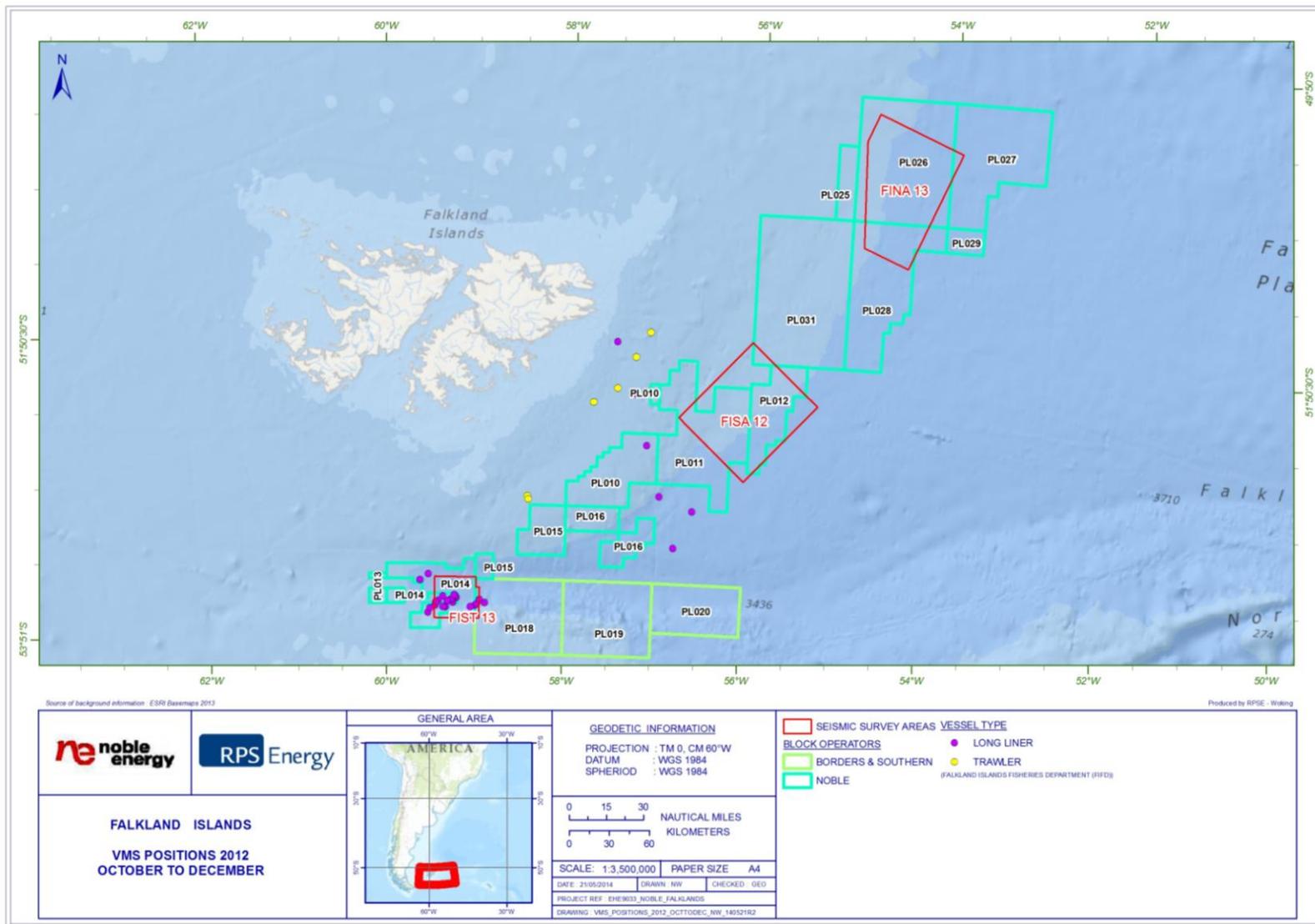


Figure E.17e: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands in 2012, all months (FIG Department of Natural Resources – Fisheries Department, 2013)

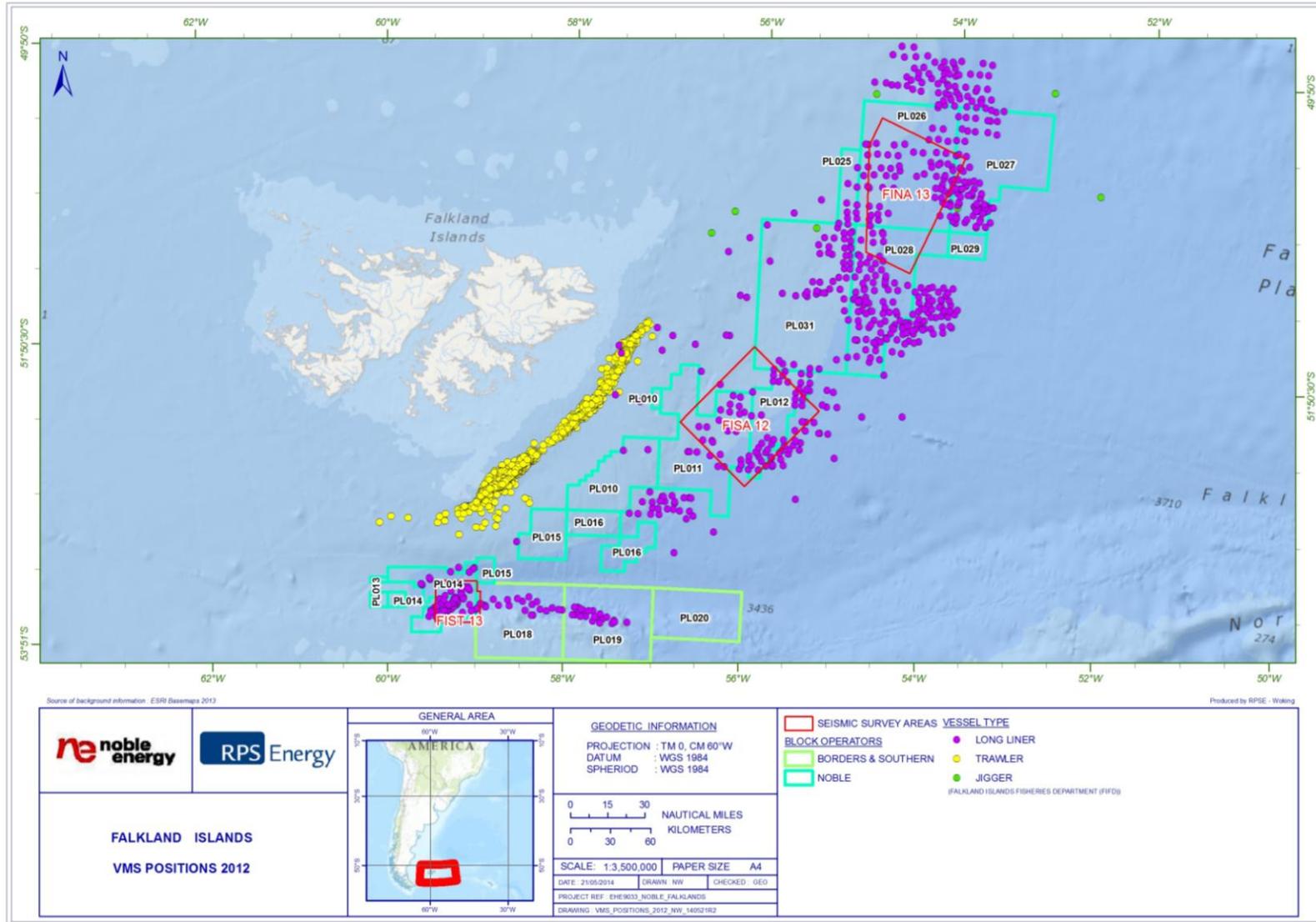


Figure E.18: Fishing Vessel Monitoring System (VMS) data for fishing vessels offshore the Falkland Islands from 2008 to 2012, all months (FIG Department of Natural Resources – Fisheries Department, 2013)

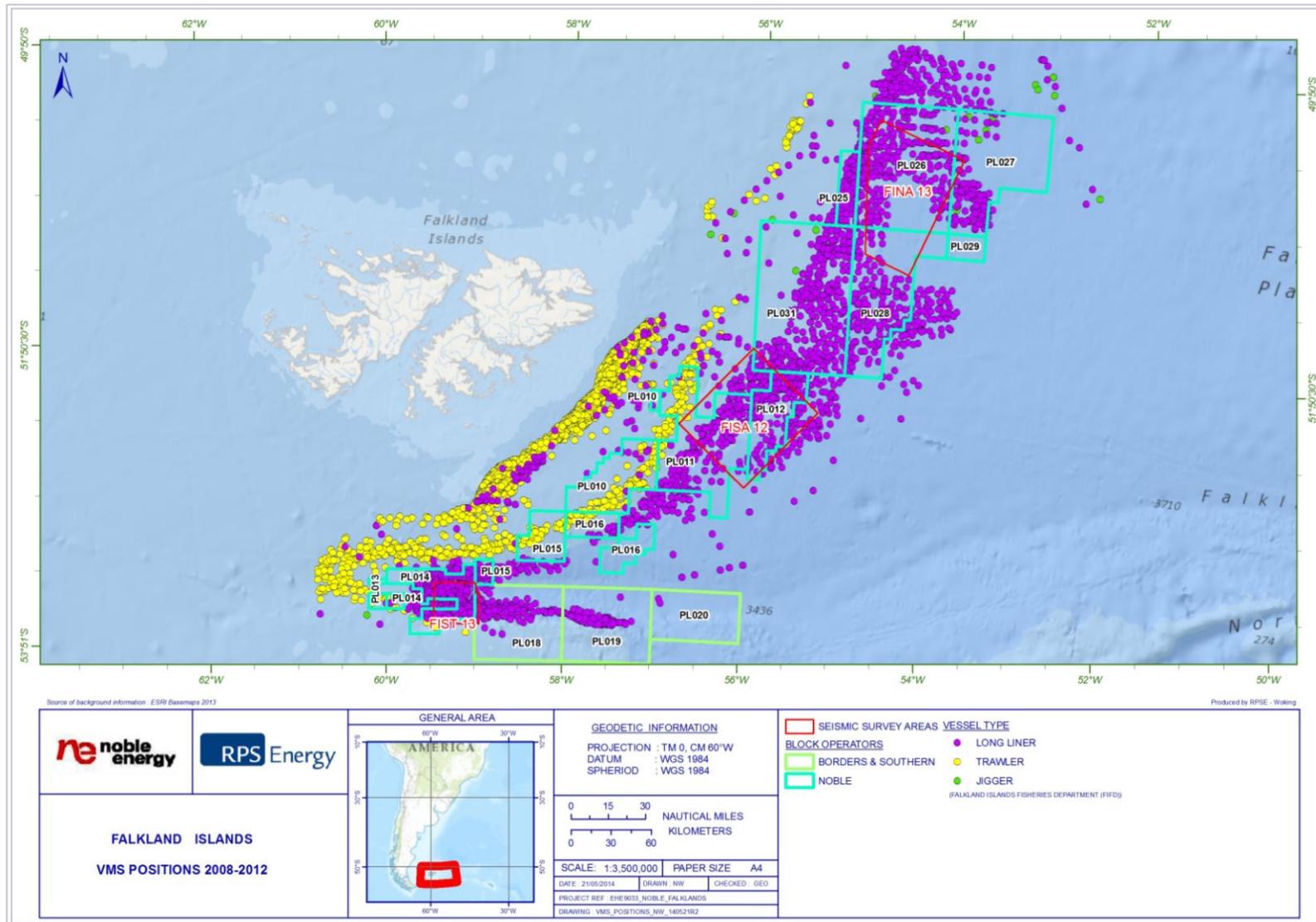


Figure E.19: Fishing effort for the year 2008 (FIG Department of Natural Resources – Fisheries Department, 2014)

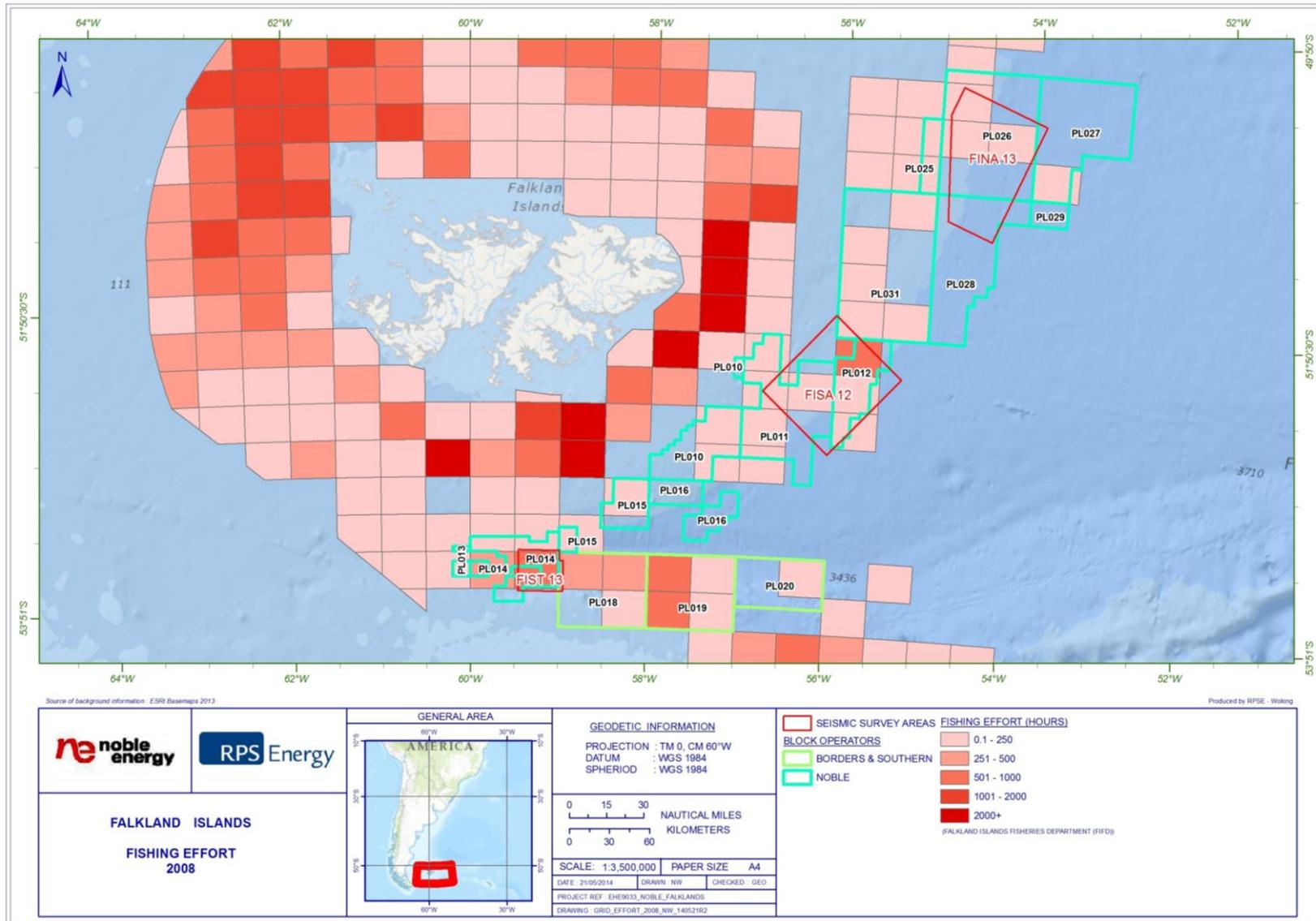


Figure E.20: Fishing effort for the year 2009 (FIG Department of Natural Resources – Fisheries Department, 2014)

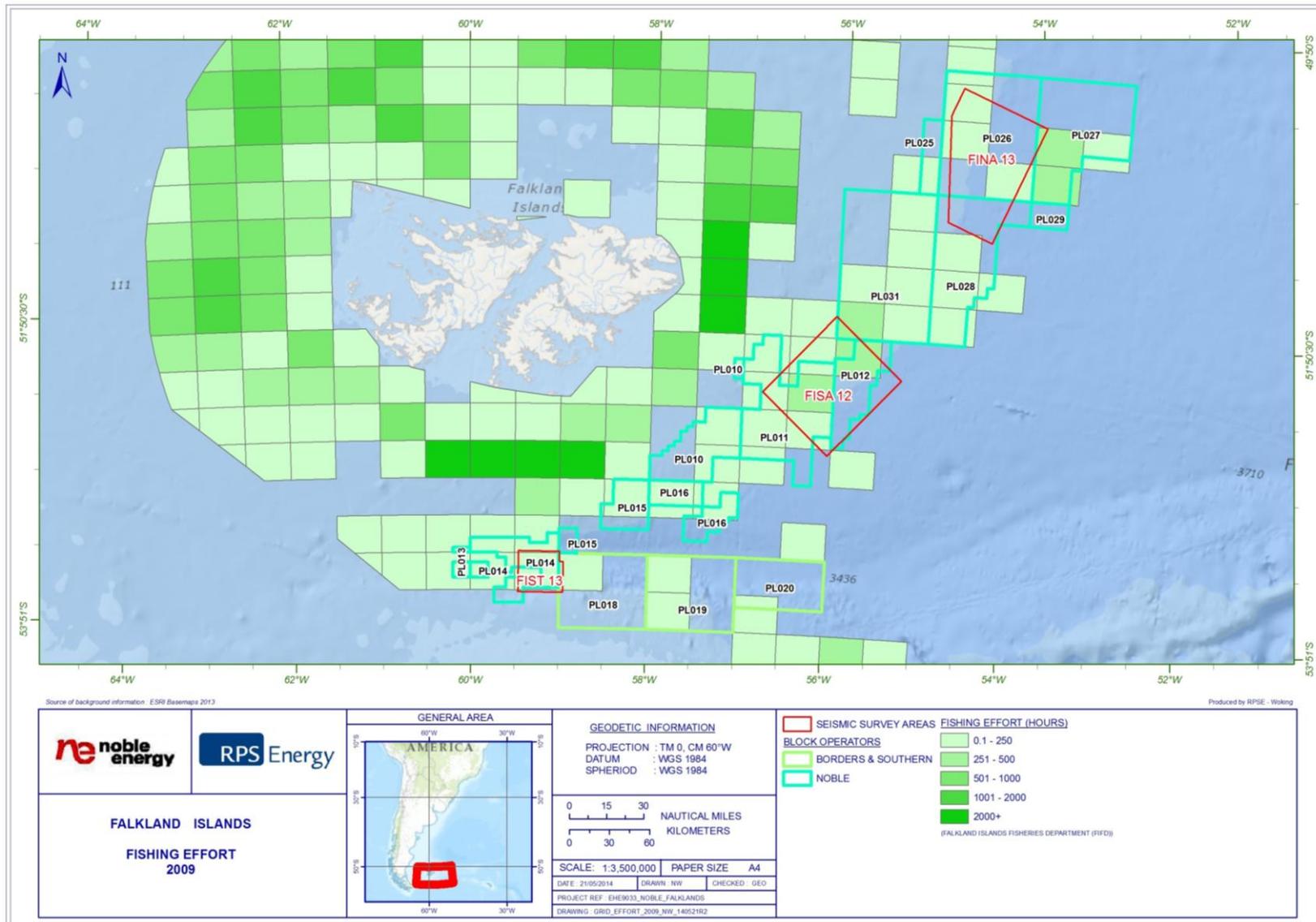


Figure E.21: Fishing effort for the year 2010 (FIG Department of Natural Resources – Fisheries Department, 2014)

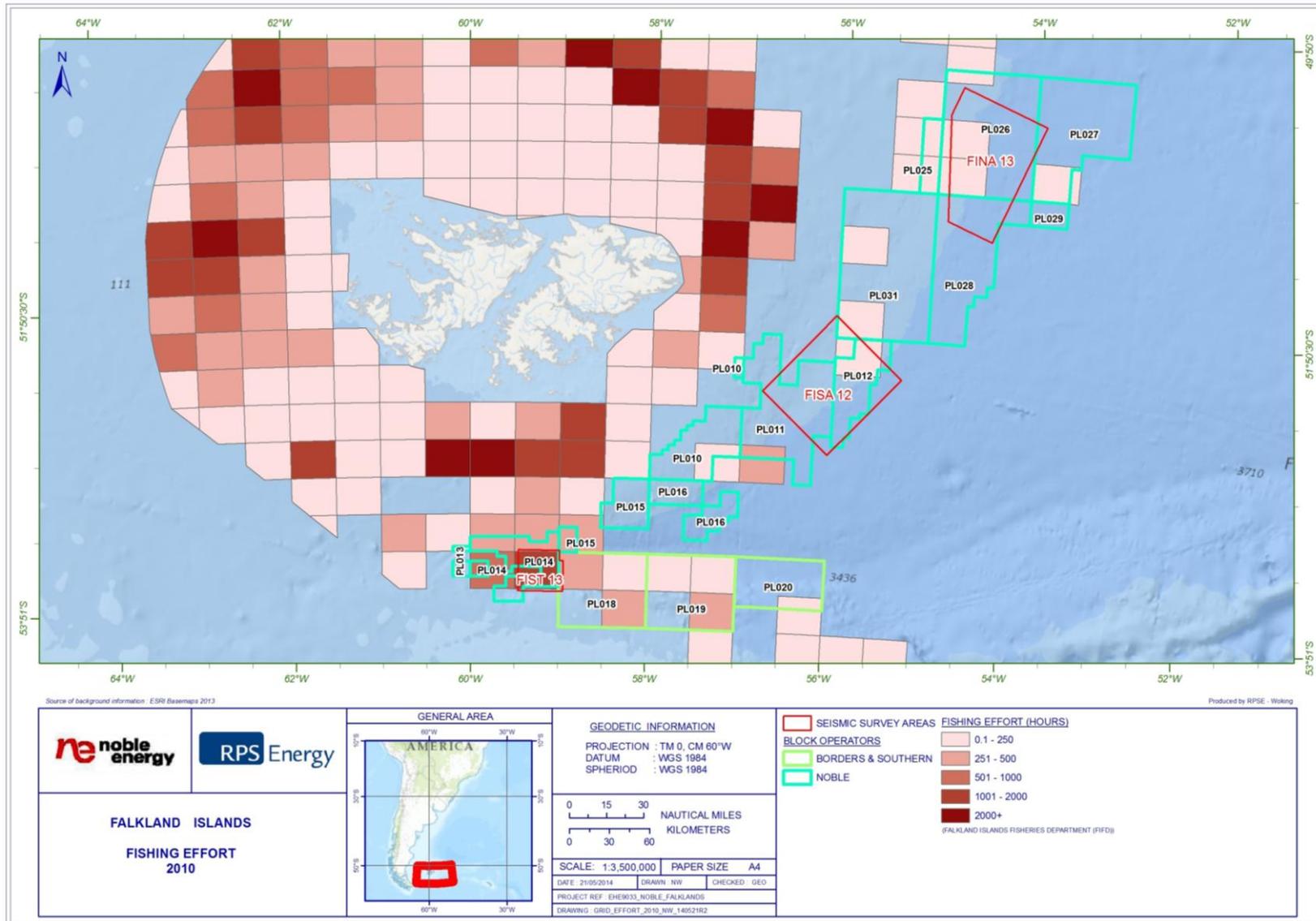


Figure E.22: Fishing effort for the year 2011 (FIG Department of Natural Resources – Fisheries Department, 2014)

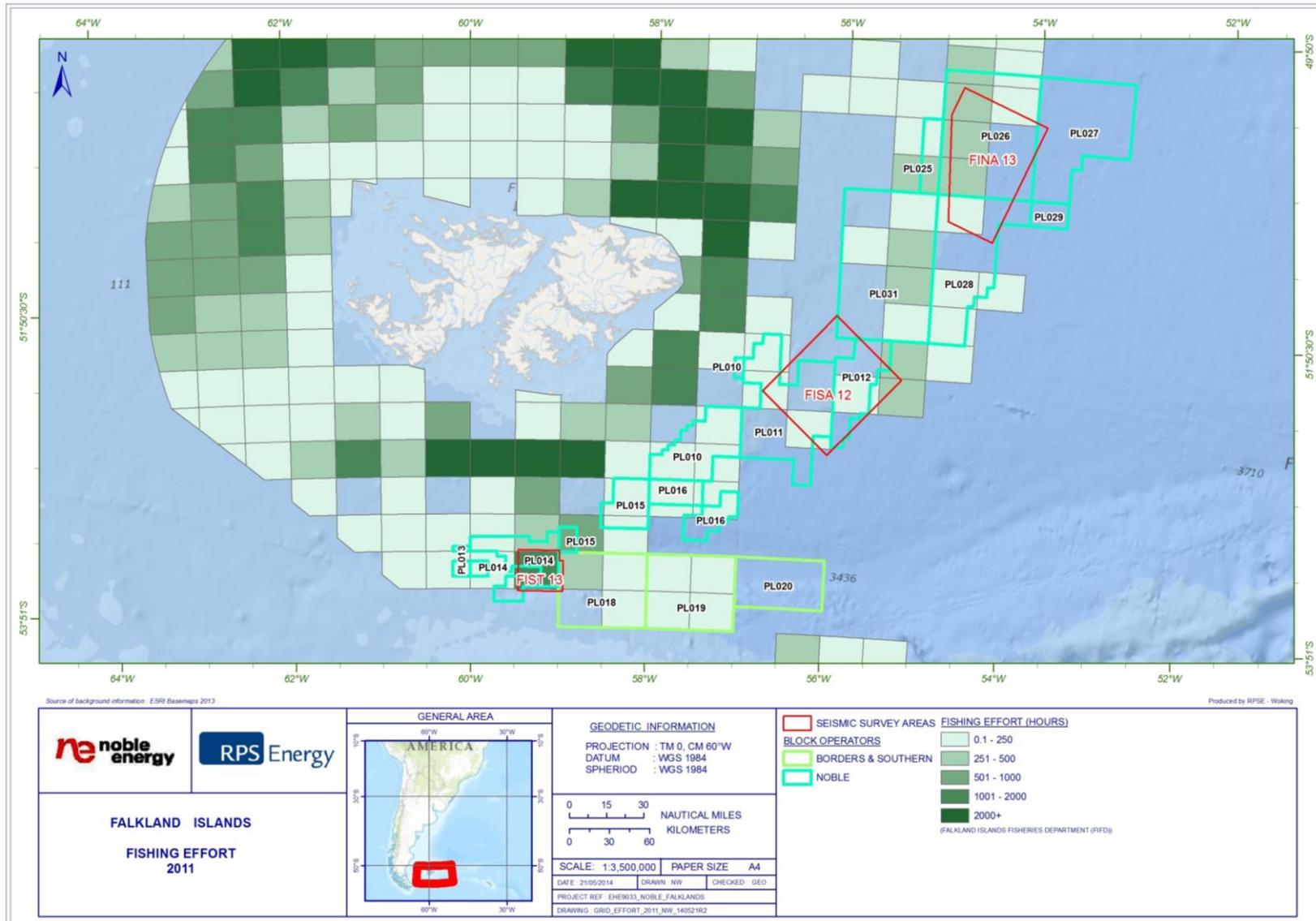
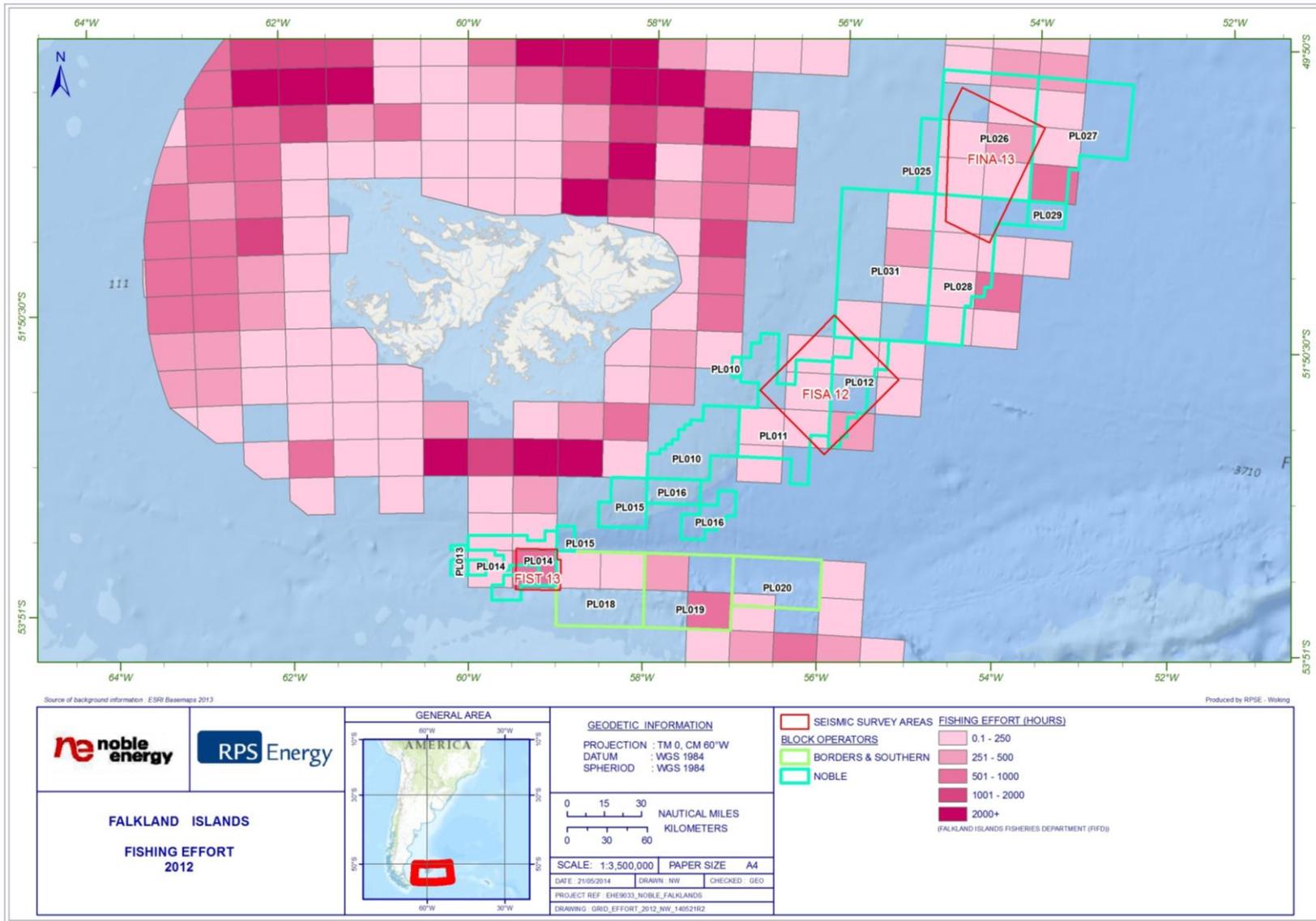


Figure E.23: Fishing effort for the year 2012 (FIG Department of Natural Resources – Fisheries Department, 2014)



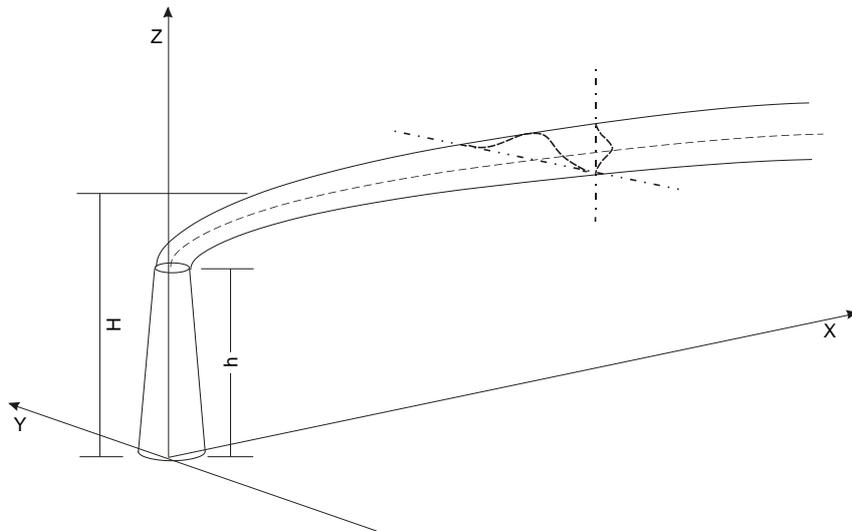
Appendix F: Atmospheric Modelling & Derivation of Global Warming Potential

F.1 Atmospheric Dispersion Modelling

The model used is spreadsheet based and derived from Davis & Cornwell (1991).

The model is an analytical model based on the Gaussian diffusion equation. The Gaussian element refers to the observation that the concentration of a gas released from a point follows an approximate normal distribution perpendicular to the centre line of the plume (Figure F.1).

Figure F.1: Diagram showing Gaussian diffusion



The concentration along the centre line is inversely proportionate to the distance from the source although very close to the source the concentration is decreased due to plume rise. Thus, a skewed concentration curve is characteristic of this sort of model. The governing equation is:

$$X(x, y, 0, H) = \left[\frac{Q}{\pi s_y s_z u} \right] \left[\exp \left[-\frac{1}{2} \left(\frac{y}{s_y} \right)^2 \right] \right] \left[\exp \left[-\frac{1}{2} \left(\frac{H}{s_z} \right)^2 \right] \right]$$

where $X(x, y, 0, H)$ = downwind concentration at ground level, g/m^3

Q = emission rate of pollution, g/s

s_y, s_z = plume standard deviations, m

u = wind speed, m/s

The basic Gaussian diffusion equation has the following assumptions:

- Atmospheric stability, that is the amount of mechanical mixing in the air, is uniform throughout the layer into which the gas stream is discharged (normally the boundary layer).
- Turbulent diffusion is random and therefore the dilution of the contaminated gas stream in both the vertical and horizontal direction can be described by the Gaussian or normal equation.

- The gas stream is released into the atmosphere at a distance above ground level that is equal to the stack height plus the plume rise (caused by convection if the released gas is hotter than the ambient temperature).
- The degree of dilution is inversely proportional to the wind speed (although wind speed data is not actually used within this model).
- Pollutant material that reaches the ground is totally reflected back into the atmosphere.

The calculation of H is obtained from adding ΔH and h via Holland's formula:

$$\Delta H = \frac{v_s d}{u} \left[1.5 + \left(2.68 \times 10^{-2} (P) \left(\frac{T_s - T_a}{T_s} \right) d \right) \right]$$

where v_s = stack velocity, m/s

d = stack diameter

P = Pressure, kPa

T_s = stack temperature, K

T_a = air temperature, K

Specific assumptions for the modelling of the gas emissions associated with the Noble drilling programme are given in the following sections.

F.1.1 Assumptions

Physical Parameters

- Height of discharge (h) 50 metres above lowest astronomical tide (LAT) (taken to represent ground level).
- Temperature of (T_s) 200 degrees Celsius (473 Kelvin).

Atmospheric Conditions

- Text Wind speed (u) of 10 metres per second.
- Temperature (T_a) 15 degrees Celsius (288 Kelvin).
- Pressure (P) 95.0 kPa (thousand Pascals).
- Overcast conditions (neutral stability).

Discharge Characteristics

Power Generation:

- Molecular weight of gas of 22.
- Emission factors from UKOOA, 2006.

F.2 Calculating Global Warming Potential (GWP)

Just as radiative forcing (the difference between radiant energy (sunlight) received by the Earth and energy radiated back to space) provides a simplified means of comparing the various factors that are believed to influence the climate system to one another, global-warming potentials (GWPs) are one type of simplified index based upon radiative properties that can be used to estimate the potential future impacts of emissions of different gasses upon the climate system in a relative sense. GWP is based on a number of factors, including the radiative efficiency

(infrared-absorbing ability) of each gas, relative to that of carbon dioxide, as well as the decay rate of each gas (the amount removed from the atmosphere over a given number of years) relative to that of carbon dioxide.

The **radiative forcing capacity** (RF) is the amount of energy per unit area, per unit time, absorbed by the greenhouse gas that would otherwise be lost to space. It can be expressed by the formula:

$$RF = \sum_{n=1}^{100} Abs_i * F_i / (\text{pathlength} * \text{density})$$

where the subscript *i* represents an interval of 10 inverse centimeters, Abs_i represents the integrated infrared absorbance of the sample in that interval, and F_i represents the RF for that interval.

The Intergovernmental Panel on Climate Change (IPCC) provides generally accepted values for GWP. An exact definition of how GWP is calculated is to be found in the IPCC's 2001 Third Assessment Report. The GWP is defined as the ratio of the time-integrated radiative forcing from the instantaneous release of 1 kg of a trace substance relative to that of 1 kg of a reference gas:

$$GWP(x) = \frac{\int_0^{TH} a_x \cdot [x(t)] dt}{\int_0^{TH} a_r \cdot [r(t)] dt}$$

where TH is the time horizon over which the calculation is considered; a_x is the radiative efficiency due to a unit increase in atmospheric abundance of the substance (i.e., $\text{Wm}^{-2} \text{kg}^{-1}$) and $[x(t)]$ is the time-dependent decay in abundance of the substance following an instantaneous release of it at time $t=0$.

The denominator values (a_r and r) contain the corresponding quantities for the reference gas (i.e. CO_2). The radiative efficiencies a_x and a_r are not necessarily constant over time. While the absorption of infrared radiation by many greenhouse gasses varies linearly with their abundance, a few important ones display non-linear behaviour for current and likely future abundances (e.g., CO_2 , CH_4 , and N_2O). For those gases, the relative radiative forcing will depend upon abundance and hence upon the future scenario adopted.

Since all GWP calculations are a comparison to CO_2 , which is non-linear, all GWP values are affected. Assuming otherwise, as is done above, will lead to lower GWPs for other gasses than a more detailed approach. Clarifying this, while increasing CO_2 has less and less effect on radiative absorption as concentrations rise, more powerful greenhouse gasses like methane and nitrous oxide have different thermal absorption frequencies to CO_2 that are not filled up (saturated) as much as CO_2 . Therefore, rising concentrations of these gasses are far more significant.

Appendix G: Air Quality Limits

G.1 World Health Organisation (WHO) Guidelines

Data on air quality offshore is limited. Emissions of carbon dioxide (CO₂), oxides of nitrogen (NO_x), and oxides of sulphur (SO_x) will result from power generation by the drilling rig and by any associated vessels which are required to support drilling operations.

The World Health Organisation (WHO) first published Air Quality Guidelines for Europe in 1987, and these were subsequently updated in 2000 and partially again in 2005 (Table G.1). The Guidelines aim to provide a basis for protecting public health from the adverse effects of environmental pollutants, and eliminating or minimising exposure to those pollutants that are known or likely to be hazardous to human health or wellbeing.

Although health effects were the major consideration behind the establishment of the Guidelines, ecologically-based Guidelines for preventing adverse effects on terrestrial vegetation were also considered, and guideline values for vegetation protection for nitrogen and sulphur oxides and ozone have been established.

Table G.1: WHO Air Quality Guidelines for Europe

| Gas | Guideline Value | Averaging Time |
|--------------------------------|---|-------------------|
| Carbon Monoxide | 100 mg/m ³ | 15 min |
| | 60 mg/m ³ | 30 min |
| | 30 mg/m ³ | 1 hour |
| | 10 mg/m ³ | 8 hour |
| Ozone | 120 µg/m ³ | 8 hour |
| Nitrogen Dioxide | 200 µg/m ³ | 1 hour |
| | 40 µg/m ³ | annual |
| Sulphur Dioxide | 500 µg/m ³ | 10 min |
| | 125 µg/m ³ | 24 hour |
| | 50 µg/m ³ | annual |
| VOCS | | |
| Benzene | 6 x 10 ⁻⁶ (µg/m ³)-1 | UR / lifetime |
| 1,3 Butadiene | no guideline | |
| Dichloromethane | 3 mg/m ³ | 24 hour |
| Formaldehyde | 0.1 mg/m ³ | 30 min |
| PAH (BaP) | 8.7 x 10 ⁻⁵ (ng/m ³)-1 | UR / lifetime |
| Styrene | 0.26 mg/m ³ | 1 week |
| Tetrachloroethylene | 0.25 mg/m ³ | annual |
| Toluene | 0.26 mg/m ³ | 1 week |
| Trichloroethylene | 4.3 x 10 ⁻⁷ (µg/m ³)-1 | UR / lifetime |
| Ecotoxic Effects | | |
| SO ₂ critical level | 10 - 30 µg/m ³ a | annual |
| NO _x critical level | 30 µg/m ³ | annual |
| Ozone critical level | 0.2 - 10 ppm.h a | 5 days - 6 months |

The European Union Framework Directive 2008/50/EC¹ on ambient air quality assessment and management has been derived from the recommendations of the WHO. This directive came into force in May 2008 and had to be implemented by Member States, including the UK, by June 2010. The Directive aims to protect human health and the environment by avoiding, reducing or preventing harmful concentrations of air pollutants. The limit values are legally binding and the Secretary of State, on behalf of the UK Government, is responsible for their implementation.

G.2 Air Quality Strategy

The UK Air Quality Strategy (AQS) describes the Government's strategy for improving air quality in the UK. The current UK AQS² was published in July 2007 and updates the original strategy to set out new objectives. The current AQS includes objectives for eight pollutants: benzene, 1,3-butadiene, ozone, carbon monoxide, lead, nitrogen dioxide, particulates and sulphur dioxide. AQS objectives are in some cases more onerous than the limit values set out within the relevant EU Directives and the Air Quality Standards Regulations 2010.

There is no legal requirement to meet objectives set within the UK AQS, except where equivalent limit values are set within the EU Directives.

The limit values and objectives relevant to this assessment are summarised in Table G.2.

Table G.2: Summary of Relevant Air Quality Limit Values and Objectives

| Pollutant | Averaging Period | Objectives/ Limit Values | Not to be Exceeded More Than |
|-------------------------------------|------------------|--------------------------|------------------------------|
| Nitrogen Dioxide (NO ₂) | 1 hour | 200 µg.m ⁻³ | 18 times per calendar year |
| | Annual | 40 µg.m ⁻³ | - |
| Sulphur Dioxide (SO ₂) | 15 minute** | 266 µg.m ⁻³ | 35 times per calendar year |
| | 1 hour | 350 µg.m ⁻³ | 24 times per calendar year |
| | 24 hour | 125 µg.m ⁻³ | 3 times per calendar year |

***This Air Quality Objective is within the AQS but not the Regulations, therefore there is no legal requirement to meet this objective.*

¹ European Commission (EC) Council Directive 2008/50/EC of 21 May 2008 on Ambient Air Quality and Cleaner Air for Europe.

² Defra (2007), The Air Quality Strategy for England, Scotland, Wales and Northern Ireland, Volume 2.

Appendix H: Cuttings Dispersion Modelling Study

Falkland Islands | Preliminary Results – Drilling Discharges Modeling at Caperea-1

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1. Project Background and Geographic Location

RPS Group, plc. has contracted Applied Science Associates, Inc. (dba RPS ASA) to evaluate seabed deposition associated with operational drilling discharges within the FISA12 and FIST13 license areas, south and east of the Falkland Islands. Four exploration wells have been selected for dispersion modeling representing drilling at depths between 1,273 m and 1,880 m (Table 1).

Table 1. Location of the discharge sites selected for modeling offshore the Falkland Islands (grey text represents wells that will be included in future modeling).

| Site Name | Block Name | Latitude (S) | Longitude (W) | Water Depth (m) |
|-------------------|--------------|--------------|---------------|-----------------|
| Caperea-1 | FISA 12 area | 52.40496 | 56.06709 | 1,333 |
| Humpback | FISA 12 area | 52.14670 | 55.73342 | 1,273 |
| Finback | FISA 12 area | 52.20798 | 55.85475 | 1,285 |
| Scharnhorst North | FIST 13 area | 53.58889 | 59.06030 | 1,880 |

Preliminary drilling plans have been developed for each site, however, as the schedule of discharges is expected to change with further refinement of the drilling program, both RPS and their client (Noble Energy) have requested initial modeling only at the Caperea-1 site. The location was selected because the site (a) falls within the shallower of the two licence blocks (FISA12), and (b) is the closest proposed well to a site of cultural heritage (the *SMS Scharnhorst* wreck). Caperea-1 is located 150 km east of the Falkland Islands coastline at an approximate water depth of 1,330 m (Figure 1). Additional modeling will be performed for the remaining sites as the drilling periods and discharge schedules become available.

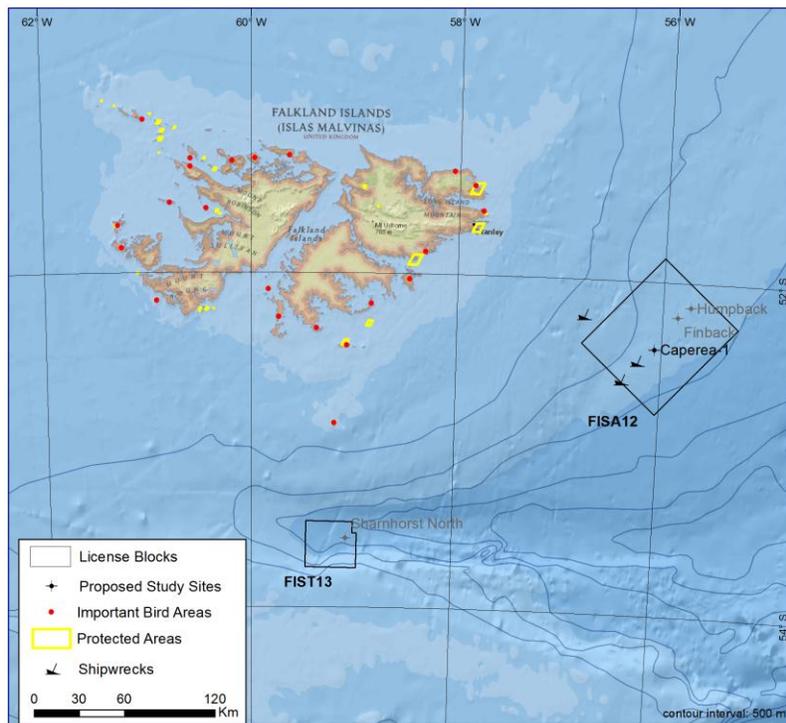


Figure 1. Drilling locations and nearby protected areas (future modeling sites shown in grey).

Discharge simulations at Caperea-1 were completed using ASA's MUDMAP modeling system (Spaulding et al., 1994). The MUDMAP model predicts the transport of solid releases in the marine environment and the resulting seabed deposition. The model requires information regarding the discharge characteristics (release location, rate of discharge, etc.), the properties of the sediment (particle sizes, density), and environmental characteristics (bathymetry and ocean currents), to predict the transport of solids through the water column. A technical description of the MUDMAP model is included in Appendix A.

2. Model Inputs

2.1. MetOcean Data

Hydrodynamic data from the HYCOM (HYbrid Coordinate Ocean Model) 1/12 degree global simulation was used as an environmental forcing for the discharge simulations. The HYCOM model is run daily by the U.S. Navy to provide a 5-day hydrodynamic forecast (+ 5 day of hindcast as best estimate) composed of 3-D daily mean temperature, salinity, sea surface height, zonal velocity and meridional velocity fields. Ocean dynamics including geostrophic and wind driven currents are reproduced by the model. The system uses the Navy Coupled Ocean Data Assimilation (NCODA) system for data assimilation (Cummings, 2005). The model domain has a spatial resolution defined by a 1/12 degree grid in the horizontal direction and a daily temporal resolution, which for this study was obtained for the period from January 2009 to December 2012.

Daily currents were obtained by interpolating the values from the nearest HYCOM model grid points. At the model cell closest to Caperea-1 release site, the water column is represented in 21 discrete vertical layers. Because the drilling period is currently unknown, vertically and time varied currents for two seasonal periods (beginning in January and June, 2012) were subset from the full HYCOM dataset and used as forcing for the MUDMAP dispersion model. The year 2012 was chosen as a representative period based on a qualitative assessment of the HYCOM record. The HYCOM record at Caperea-1 is presented in the following figures:

- Stick plot of HYCOM current speeds and directions with depth (2009-2012).
- Vertical profiles of current velocity based on the full HYCOM record.
- Current roses showing the distribution of current speed/direction at various depths and time periods.
- Monthly averaged current speeds derived from the HYCOM model at the sea surface and the seabed.

All figures display current data in the oceanographic convention (stick vectors/roses indicate the direction toward which currents are flowing).

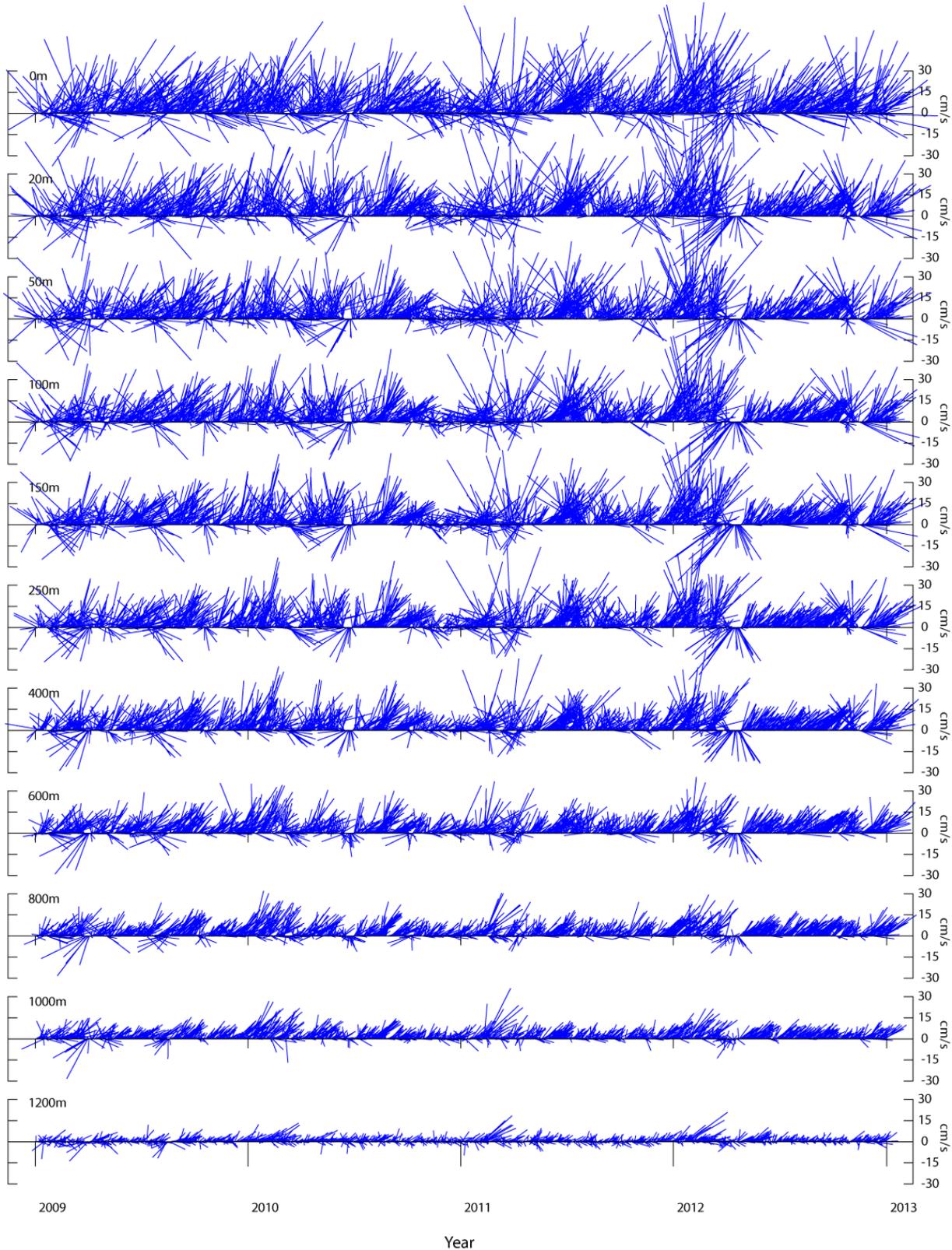


Figure 2. Time series of HYCOM model currents with depth at the Caperea-1 discharge site. Daily HYCOM currents for each depth interval are plotted as vectors representing current speed and direction. For clarity, every other depth interval is plotted.

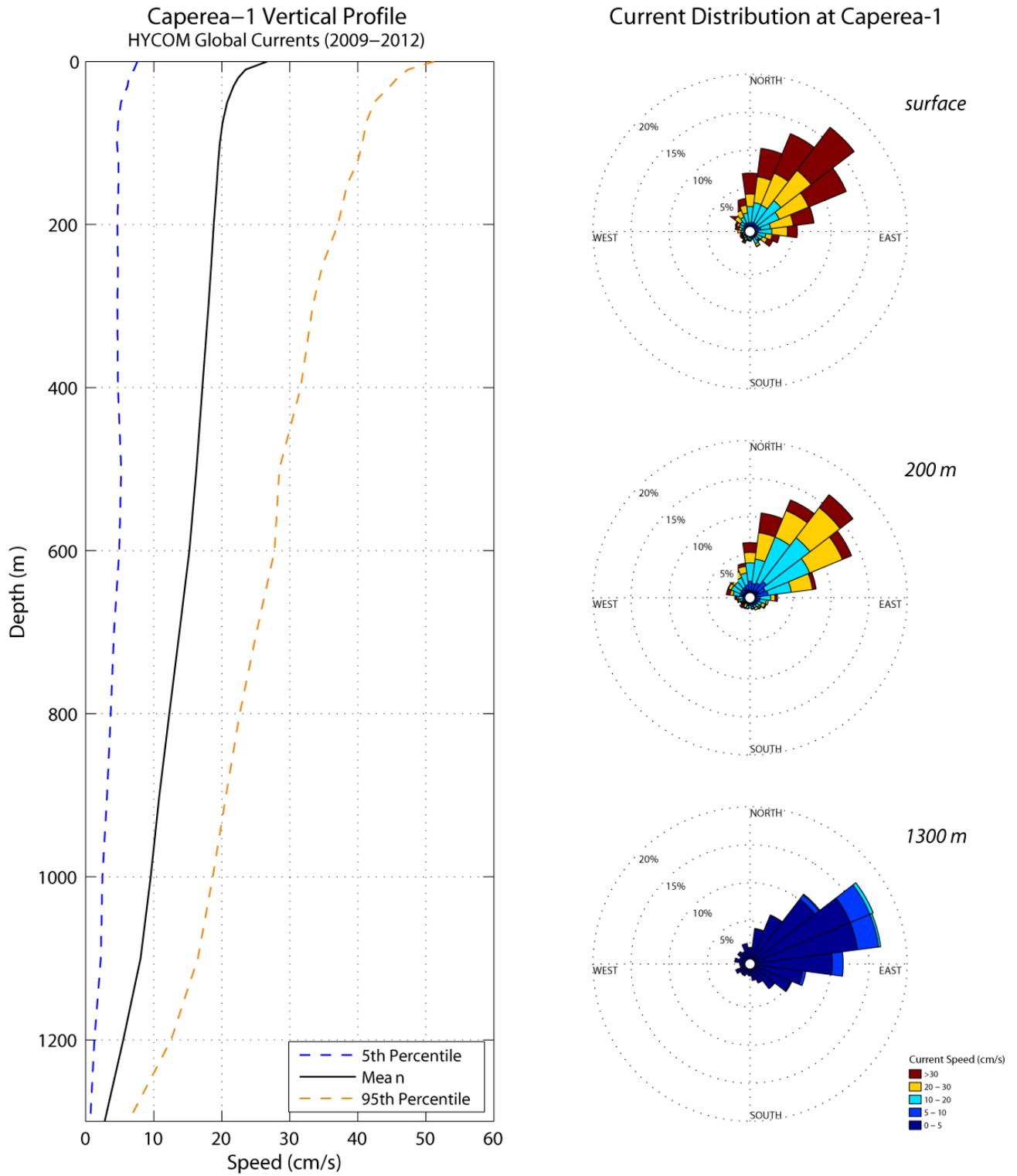


Figure 3. Vertical profile of averaged current speeds (left) and current roses showing the distribution of current speeds and directions (right) for Caperea-1, derived from HYCOM model currents between 2009 and 2012.

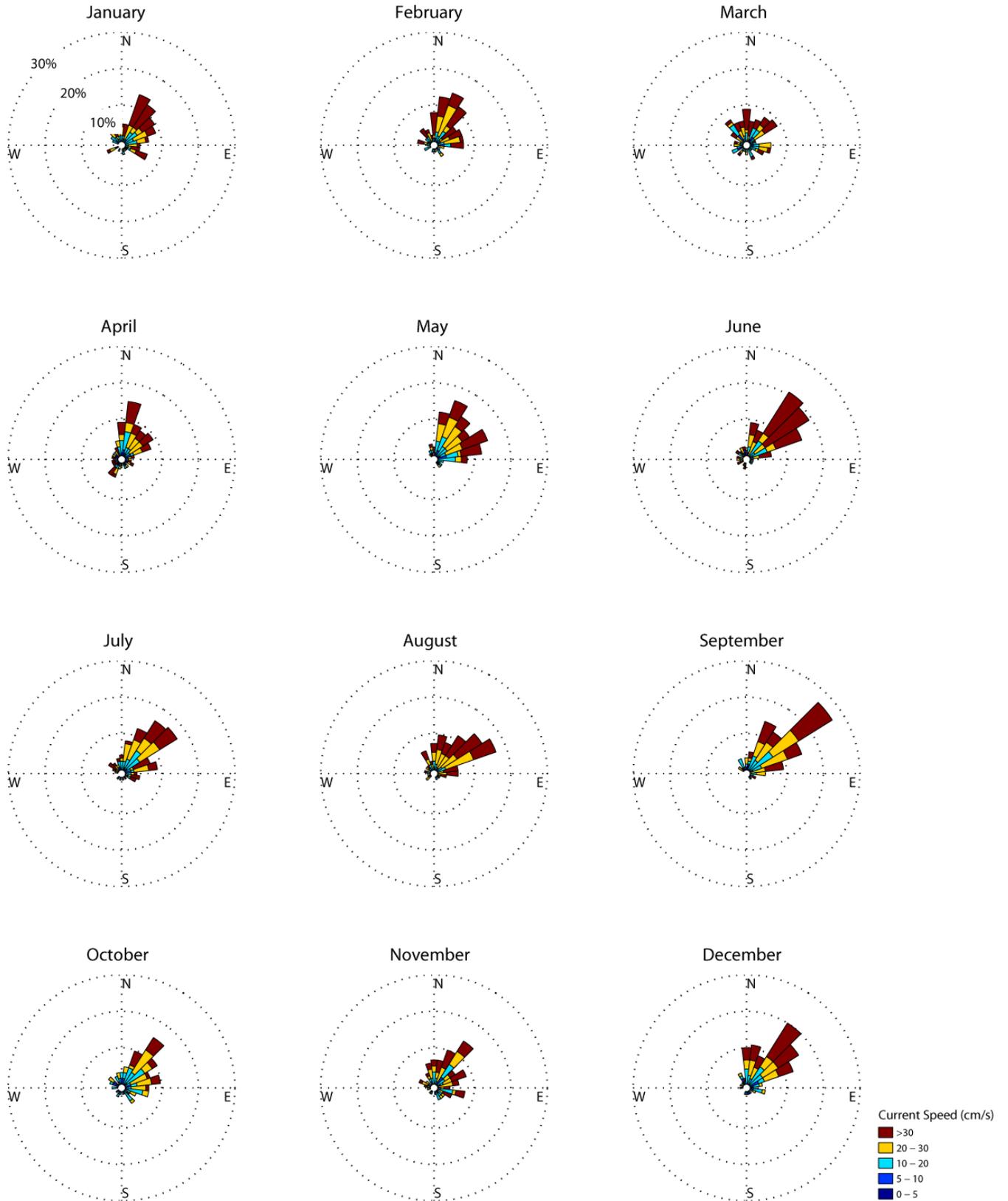


Figure 4. Current roses showing the distribution of HYCOM surface currents (speed and direction) by month at the Caperea-1 drill site (model period: 2009 and 2012).

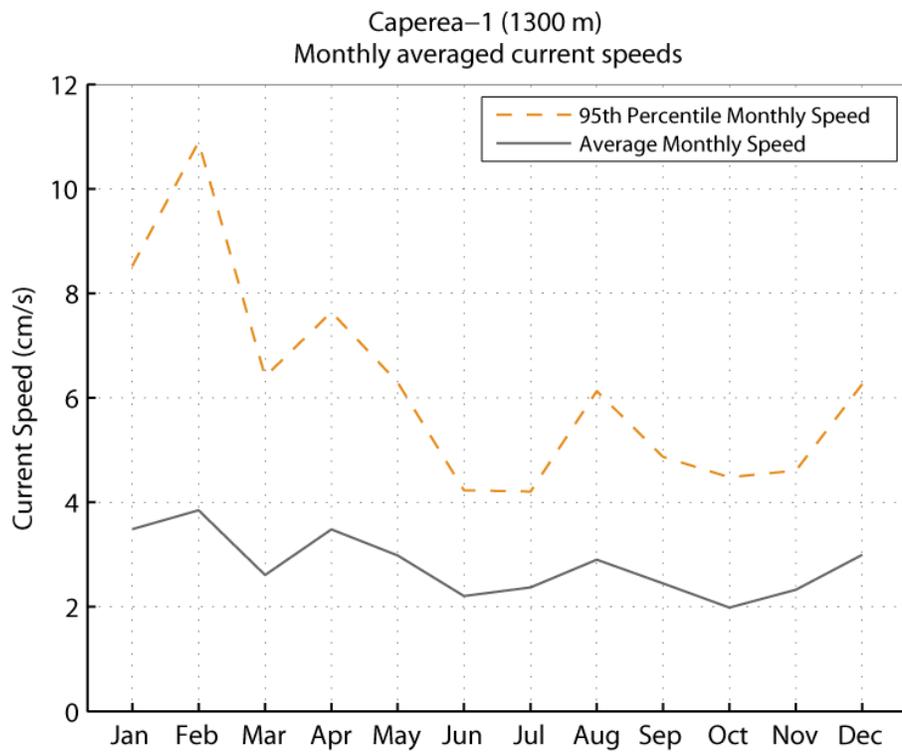
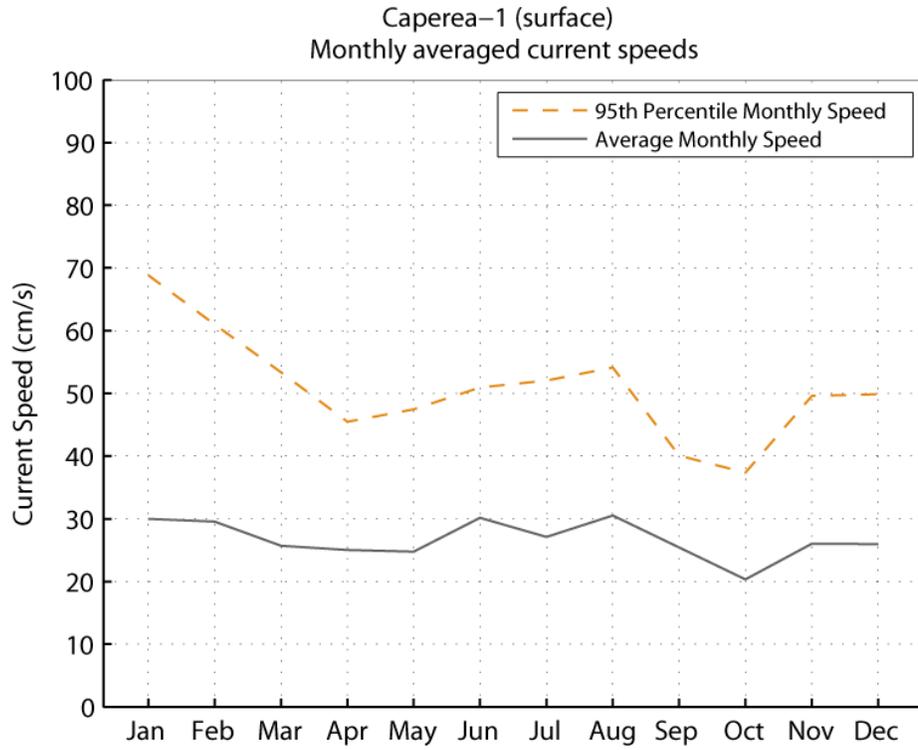


Figure 5. Monthly averaged current speeds at Caperea-1 derived from the HYCOM dataset. Average current speeds are shown for the surface (top figure) and 1,300 m (bottom figure) water depths.

2.2. Drilling Schedule

The preliminary schedule of discharges provided by RPS/Noble indicates that drilling at Caperea-1 will include (a minimum of) four well sections ranging from 42" to 12 ¼" (inches) in diameter. There is also the potential for drilling an additional by-pass section and a geological sidetrack hole for further reservoir evaluation (sections 5 and 6). The first two sections (riserless) are expected to utilize drilling fluid comprised of seawater and bentonite sweeps, which will be discharged directly to the seabed. All subsequent sections will require the use of a high performance water based mud and all cuttings and drilling fluids will be discharged from the drilling platform. The discharge of cuttings and muds is expected to occur continuously during drilling, over 75 days. For all intervals, barite is the primary weighting element of the drilling fluid.

Because the drilling programme may change, model simulations were performed for different periods (seasons) in order to evaluate the influence of potential variability in regional ocean currents. Based on a review of ocean circulation model data within the project area, operational releases were simulated for two (2) discharge periods to compare the impacts of drilling during the austral summer (Jan-Mar; Period 1), and winter (Jun-Aug; Period 2). Local currents are slightly weaker and more directionally variable during summer months as compared to the winter, which is characterized by relatively strong currents that are oriented toward the northeast at most depths.

Table 2. Drilling discharge program used for model simulations at Caperea-1.

| Section (in) | Release Depth ¹ | Release Duration (day) | Cuttings Discharges | | Drilling Fluids Discharges | |
|---------------------------------------|----------------------------|------------------------|---------------------|-------------|----------------------------|----------------|
| | | | m ³ | MT | m ³ | MT Barite |
| 42" | seabed | 2.5 | 67 | 174 | 67 | 58 |
| 26" | seabed | 7.5 | 493 | 1282 | 493 | 1293.75 |
| 17-1/2" | sea surface | 10 | 219 | 571 | 219 | 152.01 |
| 12-1/4" | sea surface | 19.5 | 108 | 281 | 108 | 89.53 |
| 8-1/2" BP | sea surface | 20.5 | 52 | 135 | 52 | 62.13 |
| 12-1/4" ST | sea surface | 15 | 118 | 307 | 118 | 97.83 |
| Total (tonnes) | | | | 2750 | | 1753.25 |
| Discharged at Seabed (tonnes) | | | | 1456 | | 1351.75 |
| Discharged at Surface (tonnes) | | | | 1294 | | 401.5 |

Notes: 1. Releases were simulated at 5 m above seabed and 2 m below sea surface.

2.3. Discharged Solids Characteristics

Table 3. Composition of drilling discharges used for modeling (Brandsma and Smith, 1999).

| Discharged material | Bulk density (pounds per gallon [ppg]) | Average SG of solids fraction |
|---------------------|--|-------------------------------|
| WBM cuttings | 2650 | 2.65 |
| WBM (all sections) | 1132 | 3.377 |

Table 4. WBM cuttings settling velocities used for simulations (Brandsma and Smith, 1999; Dames and Moore, 1978).

| Size Class | Percent Volume | Settling Velocity | |
|------------|----------------|-------------------|----------|
| | | (cm/s) | (m/day) |
| 1 | 8.00 | 1.350E-04 | 0.12 |
| 2 | 6.00 | 1.686E-03 | 1.46 |
| 3 | 7.00 | 2.182E-02 | 18.86 |
| 4 | 3.00 | 2.328E-01 | 201.14 |
| 5 | 2.00 | 1.447E+00 | 1250.37 |
| 6 | 18.00 | 4.011E+00 | 3465.65 |
| 7 | 16.00 | 9.796E+00 | 8463.98 |
| 8 | 15.00 | 1.352E+01 | 11679.45 |
| 9 | 25.00 | 2.598E+01 | 22442.45 |

Table 5. Drilling mud (WBM) settling velocities used for simulations (Brandsma and Smith, 1999; O'Reilly et al., 1988).

| Size Class | Percent Volume | Settling Velocity | |
|------------|----------------|-------------------|---------|
| | | (cm/s) | (m/day) |
| 1 | 7.01 | 2.74E-03 | 2.37 |
| 2 | 7.99 | 6.10E-03 | 5.27 |
| 3 | 5.00 | 1.48E-02 | 12.77 |
| 4 | 10.00 | 3.00E-02 | 25.94 |
| 5 | 13.26 | 4.36E-02 | 37.66 |
| 6 | 13.26 | 5.12E-02 | 44.24 |
| 7 | 19.24 | 6.40E-02 | 55.30 |
| 8 | 19.24 | 8.23E-02 | 71.10 |
| 9 | 4.00 | 4.27E-01 | 368.69 |
| 10 | 1.00 | 1.12E+00 | 969.12 |

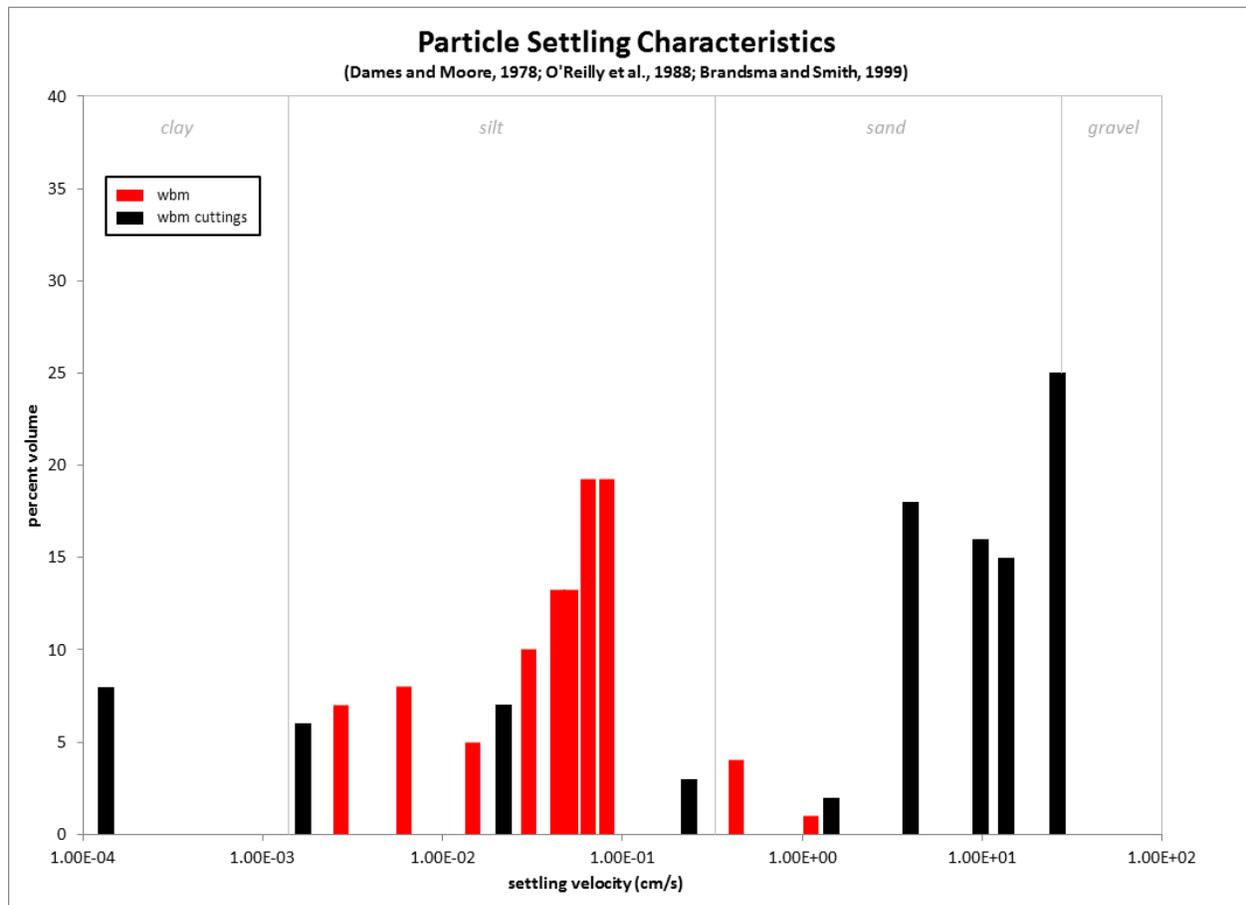


Figure 6. Comparison of settling velocities for solid discharges used in the modeling study. Size class divisions are from Gibbs et al. (1971).

3. Model Results

The fate of mud and cuttings released from operational drilling were assessed through two discharge model scenarios corresponding to the drilling schedule and release volumes shown in Table 6. For each scenario, the MUDMAP model was used to predict the resulting bottom deposition from individual drilling sections at Caperea-1, along with the pattern of cumulative deposits. Simulations were designed to continue tracking the far field dispersion for a minimum of 72 hours following the completion of each section, to account for settling of fine material from the water column. Figure 7 and Figure 8 show the plan view extents of the model-predicted seabed deposition during the austral summer (Period 1) and winter (Period 2), respectively. Table 7 through Table 8 summarize the areal extent of deposition for each scenario. Deposit thicknesses were calculated based on mass accumulation on the seabed and assume a sediment bulk density of $2,500 \text{ kg/m}^3$ and no void ratio (zero porosity).

Table 6. Summary of model parameters used for each scenario.

| Model Scenario | Discharge Period | Description | Discharged Cuttings (MT) | Discharged Mud (MT Barite) | Duration of Discharges (d) |
|----------------|------------------|------------------------------------|--------------------------|----------------------------|----------------------------|
| Scenario 1 | Jan-Mar 2012 | WBM and cuttings from sections 1-6 | 2,750 | 1,753 | 75 |
| Scenario 2 | Jun-Aug 2012 | WBM and cuttings from sections 1-6 | 2,750 | 1,753 | 75 |

For both scenarios, the thin, broad blanket of sediment that extends ~1-2 km from the discharge site results from the accumulation of very fine particles (which experience more variation in the current regime as they settle) and from discharges that originate at the sea surface (which disperse widely while settling in deep water). To that end, stronger and more uniform currents during Period 2 produce a cumulative deposit that is more elongated and extends nearly 2 km from the discharge site toward the northeast. The extent of deposition is considerably larger for thicknesses < 1 mm during Period 2. For Period 1, the overall deposit is more rounded and the footprint is confined to 945 m from the Caperea-1 site. Both scenarios impact a similar cumulative area for contours > 1mm, as shown Figure 9. Deposition at or above 10 mm is uniform and concentric around the well, which indicates that dispersion processes are nearly as influential as advection from currents due to the settling characteristics of material being released and the release depths.

Finally, Figure 10 shows the integrated footprint from both discharge scenarios with respect to the best known coordinates of the *SMS Scharnhorst* (as reported by Wrecksite.eu), a shipwrecked German cruiser (lost December 1914) that lies within the FISA12 license area. The results from all model runs were integrated to define the likely area of coverage for both current conditions modelled above the 0.1 mm minimum thickness threshold. A 10 km buffer has been applied to the wreck location due to the positional uncertainty of the shipwreck.

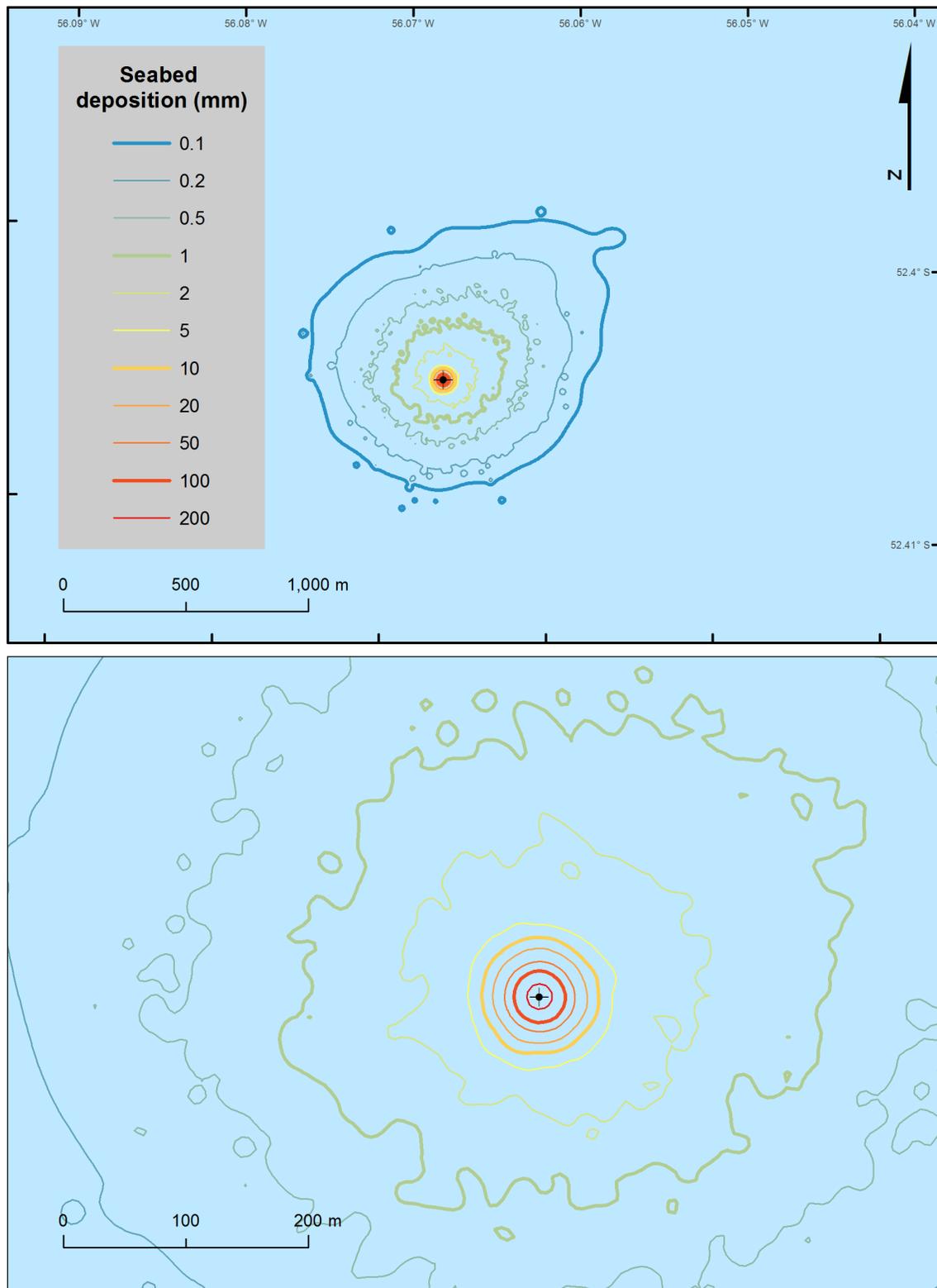


Figure 7. Predicted thickness of drilling discharges at Caperea-1 (Period 1; Jan-Mar). Top: composite deposition resulting from all drilling intervals. Bottom: contours above 1 mm (bold green) shown at an expanded scale.

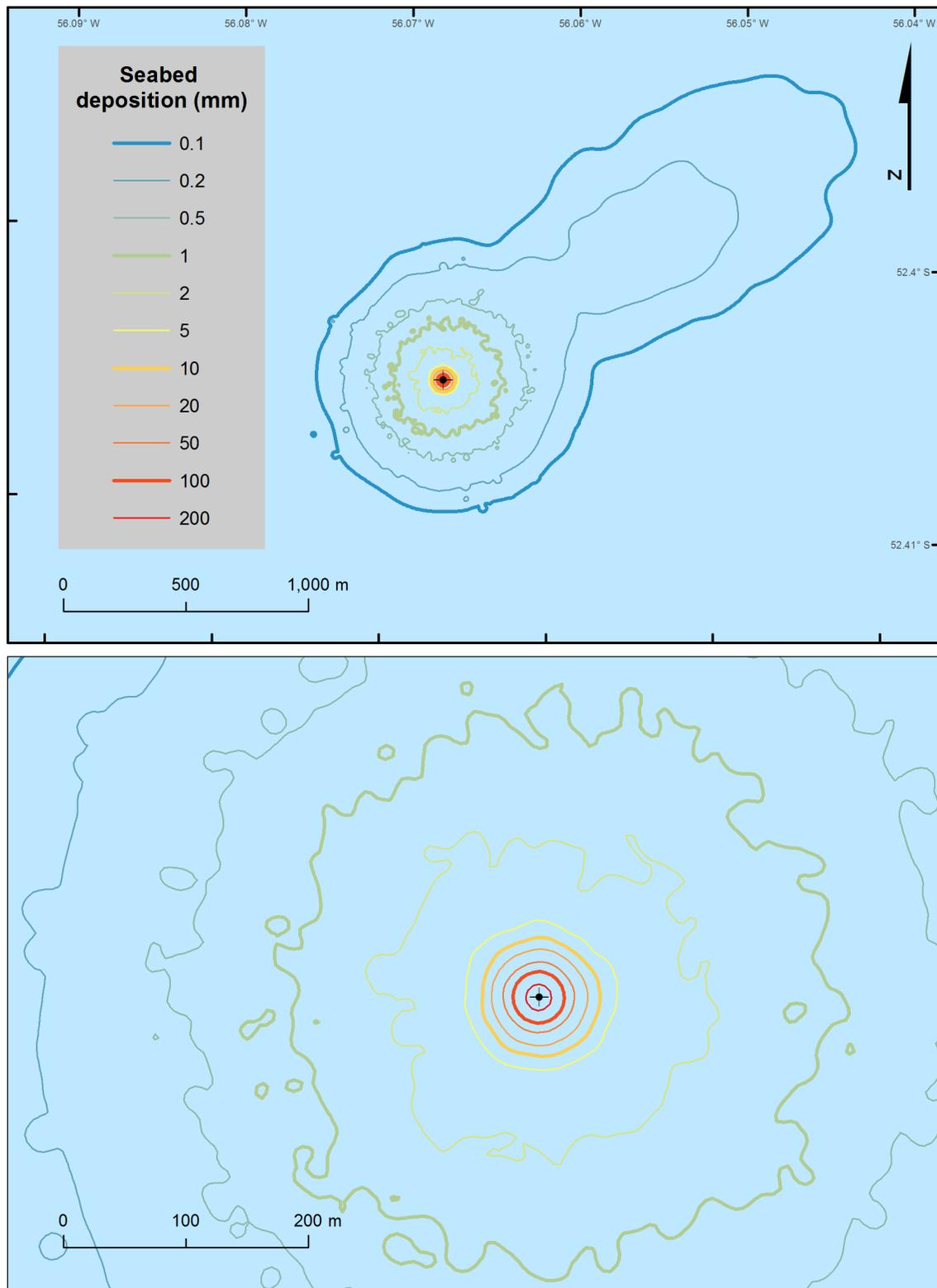


Figure 8. Predicted thickness of drilling discharges at Caperea-1 (Period 2; Jun-Aug). Top: composite deposition resulting from all drilling intervals. Bottom: contours above 1 mm (bold green) shown at an expanded scale.

Table 7. Areal extent of seabed deposition (by thickness interval) for each model scenario.

| Deposition Thickness (mm) | Cumulative Area Exceeding (ha) | |
|---------------------------|--------------------------------|----------------|
| | Period 1 | Period 2 |
| 0.1 | 103.164 | 193.141 |
| 0.2 | 63.706 | 97.999 |
| 0.5 | 29.838 | 31.589 |
| 1 | 13.747 | 15.713 |
| 2 | 4.316 | 5.200 |
| 5 | 1.124 | 1.149 |
| 10 | 0.693 | 0.718 |
| 20 | 0.464 | 0.466 |
| 50 | 0.257 | 0.247 |
| 100 | 0.135 | 0.142 |
| 200 | 0.035 | 0.032 |

Table 8. Maximum extent of thickness contours (distance from release site) for each model scenario.

| Deposition Thickness (mm) | Maximum extent from discharge point (m) | |
|---------------------------|---|----------|
| | Period 1 | Period 2 |
| 0.1 | 945 | 1980 |
| 1 | 308 | 265 |
| 10 | 50 | 50 |
| 100 | 22 | 22 |

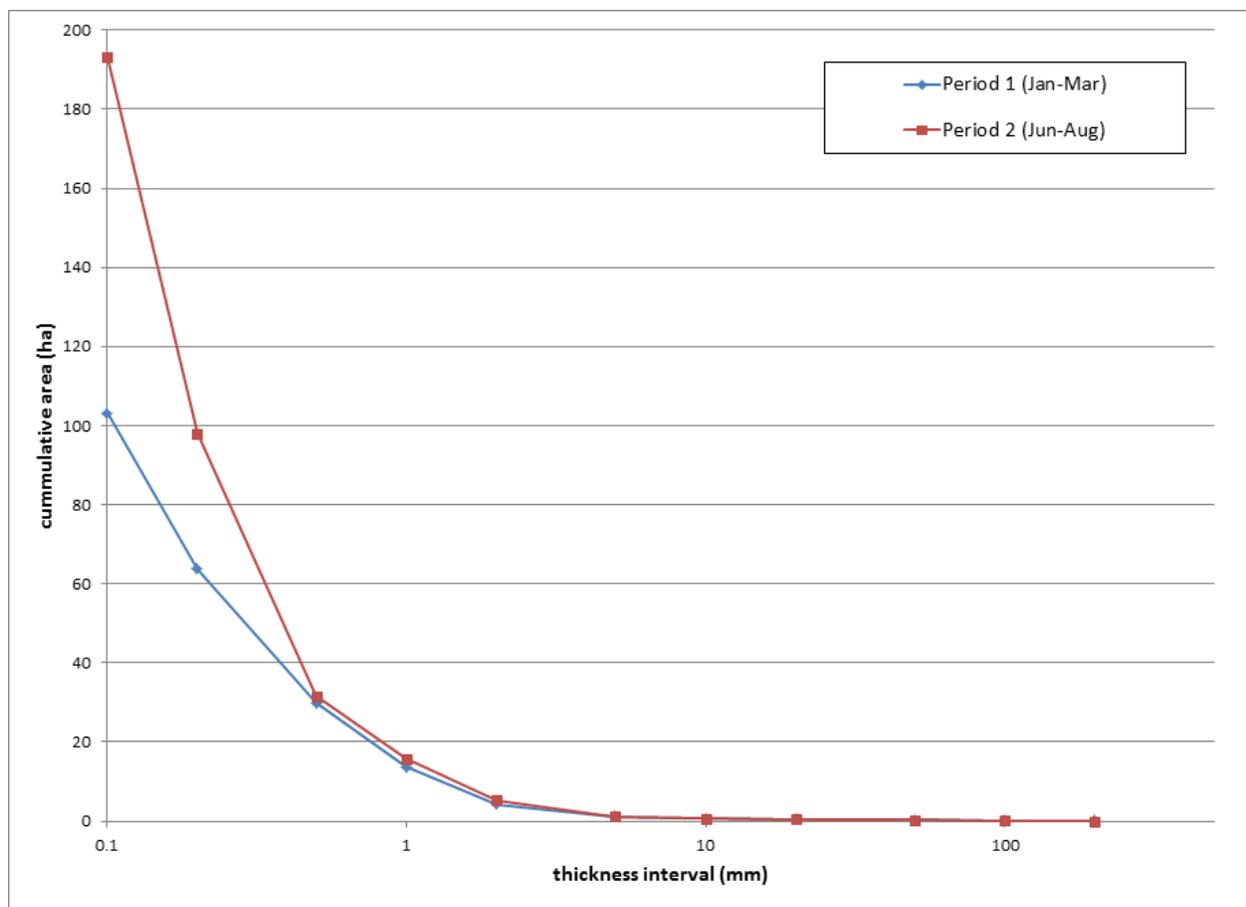


Figure 9. Comparison of seabed deposition (by thickness interval) for discharges originating from Caperea-1. Blue – discharges during the Jan-Mar period, Red – discharges during the Jun-Aug period.

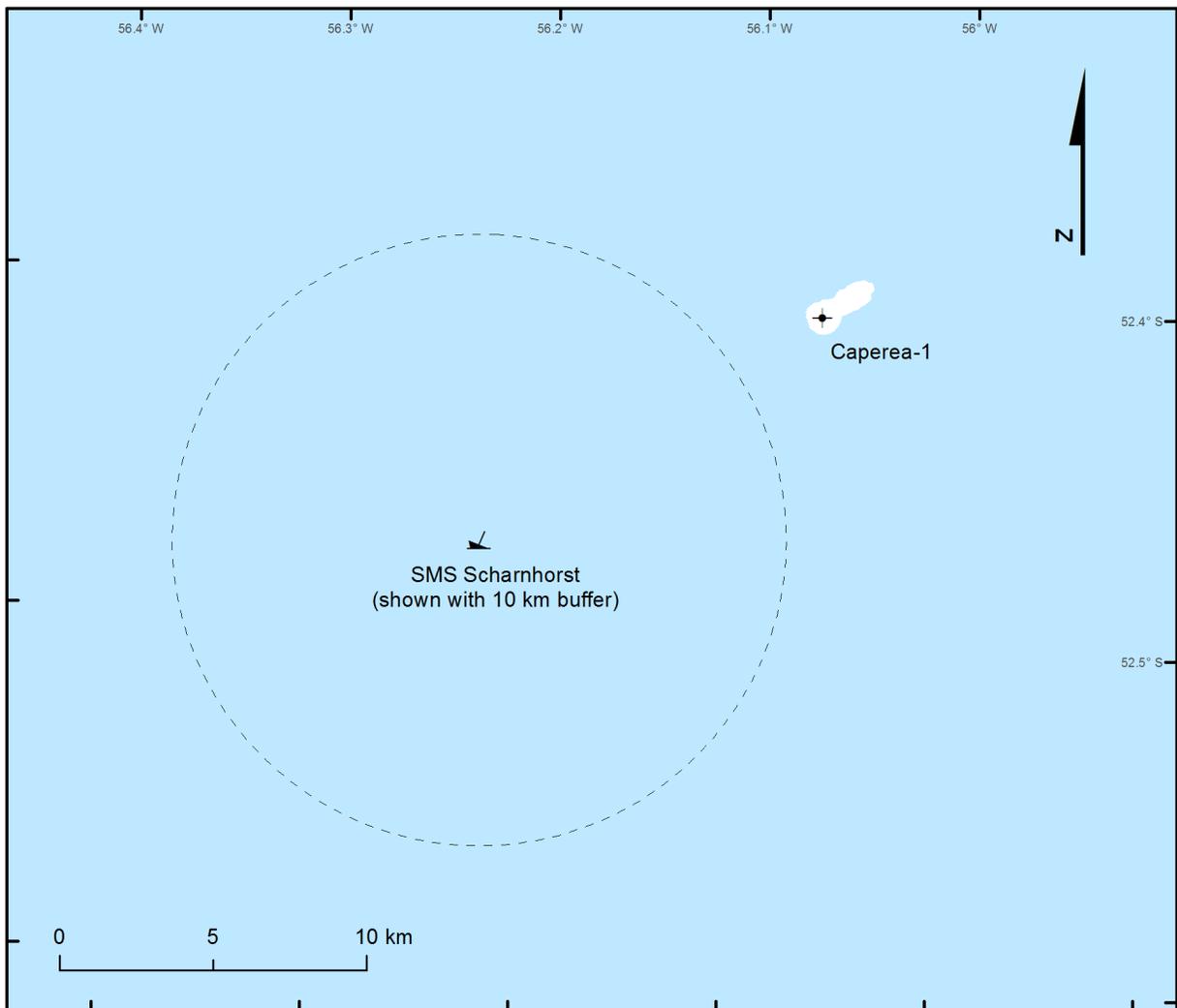


Figure 10. Integrated footprint of deposition at Caperea-1 for both model periods (white polygon) shown with shipwreck location (SMS Scharnhorst) (as reported by Wrecksite.eu). Dashed line shows the positional uncertainty (10 km) of the wreck.

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Appendix A: MUDMAP Model Description

MUDMAP is a personal computer-based model developed by ASA to predict the near and far-field transport, dispersion, and bottom deposition of drill muds and cuttings and produced water (Spaulding et al; 1994). In MUDMAP, the equations governing conservation of mass, momentum, buoyancy, and solid particle flux are formulated using integral plume theory and then solved using a Runge Kutta numerical integration technique. The model includes three stages:

Stage 1: Convective decent/jet stage – The first stage determines the initial dilution and spreading of the material in the immediate vicinity of the release location. This is calculated from the discharge velocity, momentum, entrainment and drag forces.

Stage 2: Dynamic collapse stage – The second stage determines the spread and dilution of the released material as it either hits the sea surface or sea bottom or becomes trapped by a strong density gradient in the water column. Advection, density differences and density gradients drive the transport of the plume.

Stage 3: Dispersion stage – In the final stage the model predicts the transport and dispersion of the discharged material by the local currents. Dispersion of the discharged material will be enhanced with increased current speeds and water depth and with greater variation in current direction over time and depth.

MUDMAP is based on the theoretical approach initially developed by Koh and Chang (1973) and refined and extended by Brandsma and Sauer (1983) and Khondaker (2000) for the convective descent/ascent and dynamic collapse stages. The far-field, passive diffusion stage is based on a particle based random walk model. This is the same random walk model used in ASA's OILMAP spill modeling system (ASA, 1999).

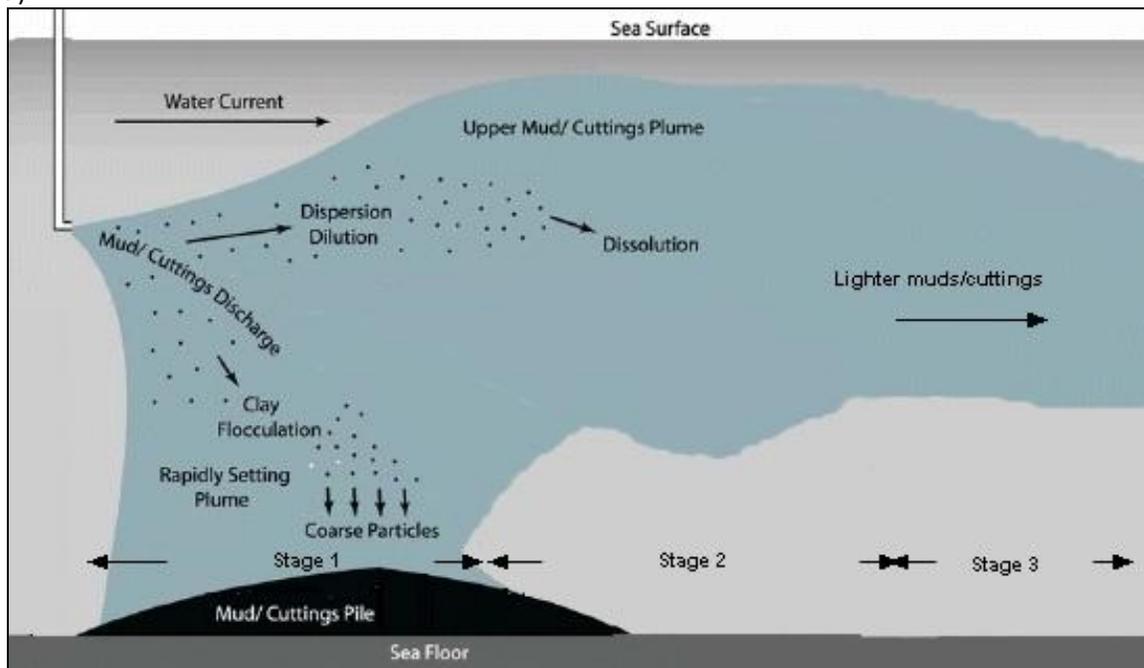


Figure A1. Conceptual diagram showing the general behavior of cuttings and muds following discharge to the ocean and the three distinct discharge phases (after Neff 2005).

The model's output consists of calculations of the movement and shape of the discharge plume, the concentrations of soluble (i.e. oil in produced water) and insoluble (i.e. cuttings and muds) discharge components in the water column, and the accumulation of discharged solids on the seabed. The model predicts the initial fate of discharged solids, from the time of discharge to initial settling on the seabed. As MUDMAP does not account for resuspension and transport of previously discharged solids, it provides a conservative estimate of the potential seafloor concentrations (Neff 2005).

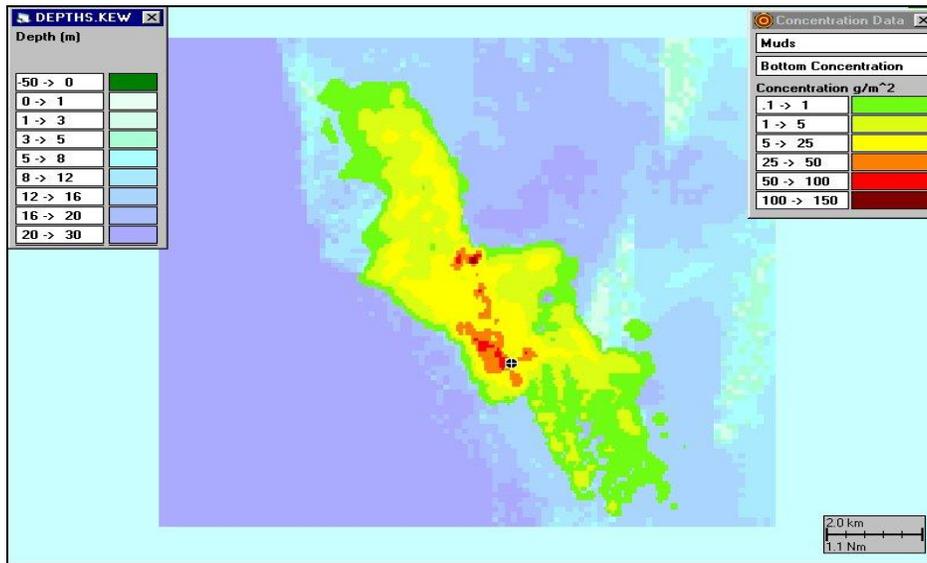


Figure A2. Example MUDMAP bottom concentration output for drilling fluid discharge.

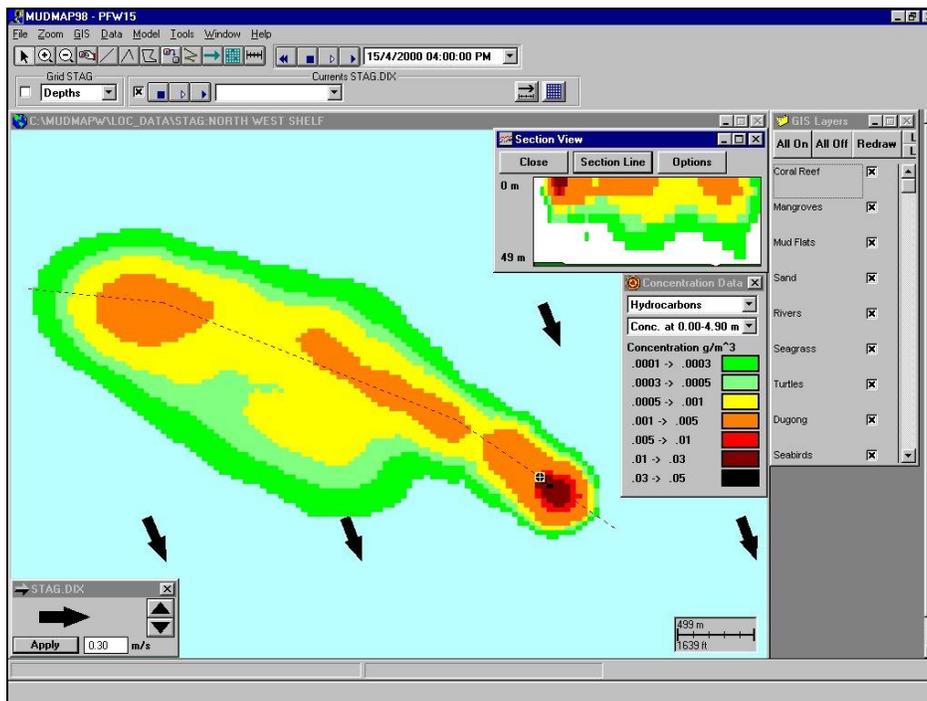


Figure A3. Example MUDMAP water column concentration output for drilling fluid discharge.

MUDMAP uses a color graphics-based user interface and provides an embedded geographic information system, environmental data management tools, and procedures to input data and to animate model output. The system can be readily applied to any location in the world. Application of MUDMAP to predict the transport and deposition of heavy and light drill fluids off Pt. Conception, California and the near-field plume dynamics of a laboratory experiment for a multi-component mud discharged into a uniform flowing, stratified water column are presented in Spaulding et al. (1994). King and McAllister (1997, 1998) present the application and extensive verification of the model for a produced water discharge on Australia's northwest shelf. GEMS (1998) applied the model to assess the dispersion and deposition of drilling cuttings released off the northwest coast of Australia.

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Appendix I: Oil Spill Modelling Study

RPS | Falkland Islands. Preliminary Oil Spill Modelling Results

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1. Geographic Location - Area of Interest

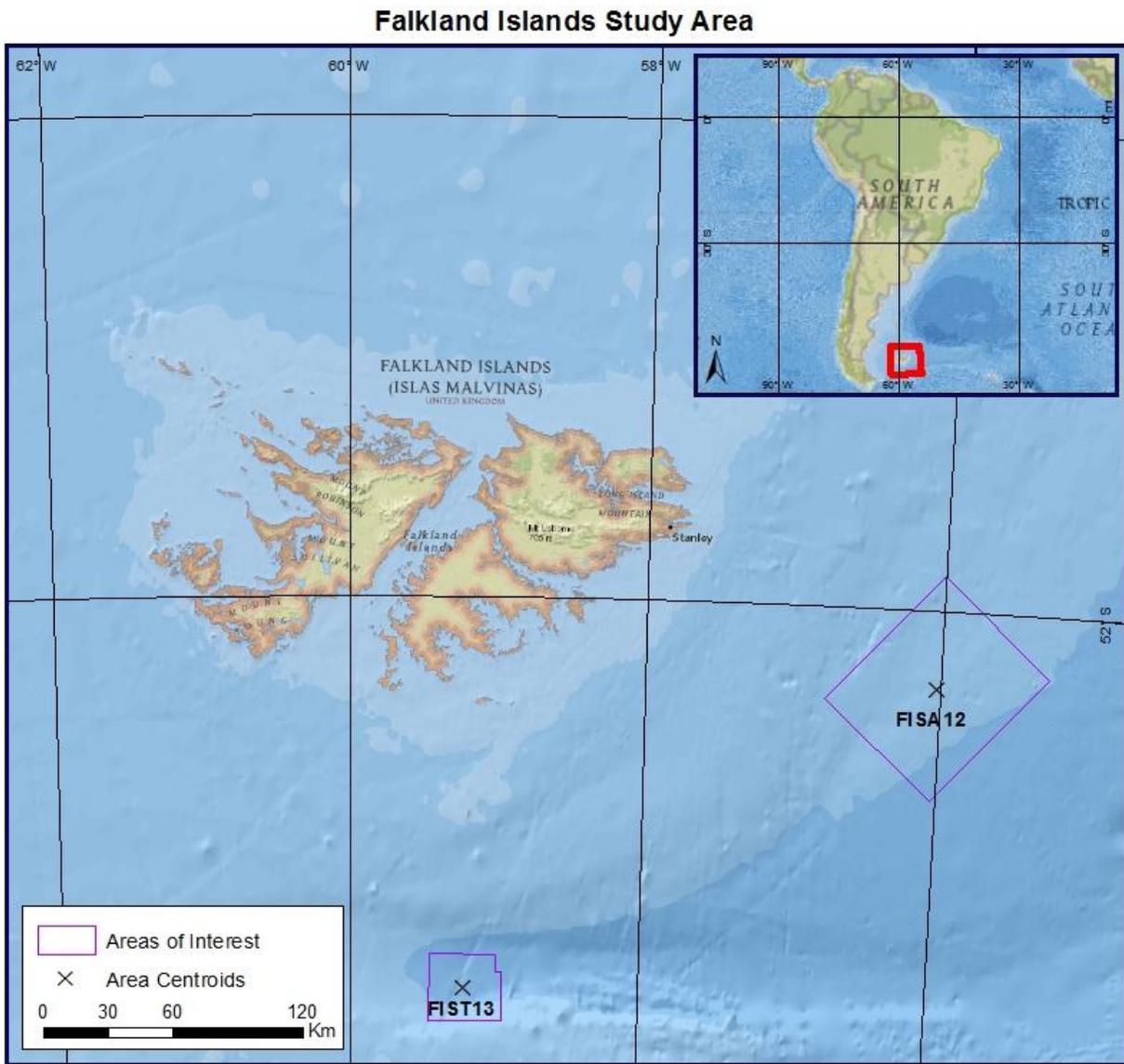


Figure 1. Geographic location of the study area.

Table 1. Coordinates of the sites of interest.

| Site Name | Comments | Latitude N | Longitude E | Distance from land (km) | Approximate Water Depth (m) |
|-----------|------------------|------------|-------------|-------------------------|-----------------------------|
| FISA12 | Center of FISA12 | -52.3232 | -56.0231 | 137 (Cape Pembroke) | 1,177 |
| FIST13 | Center of FIST13 | -53.6283 | -59.2109 | 76 (Beauchene Island) | 1,527 |

2. Environmental Data Analysis

The environmental datasets used for this study include the temperature and salinity dataset, WOA-13 (hosted by the National Oceanographic and Atmospheric Administration), the wind dataset, NOGAPS (a product of the United States Navy) and the current dataset, HYCOM (a product of the National Oceanographic Partnership Program). All wind and current figures presented in this report use subsets of these datasets that cover the period from January 1, 2009 to December 31, 2012. All wind roses follow standard meteorological convention (showing the direction from which winds are blowing) while all current roses follow standard oceanographic convention (showing the direction towards which currents are flowing).

2.1. Wind Dataset – NOGAPS

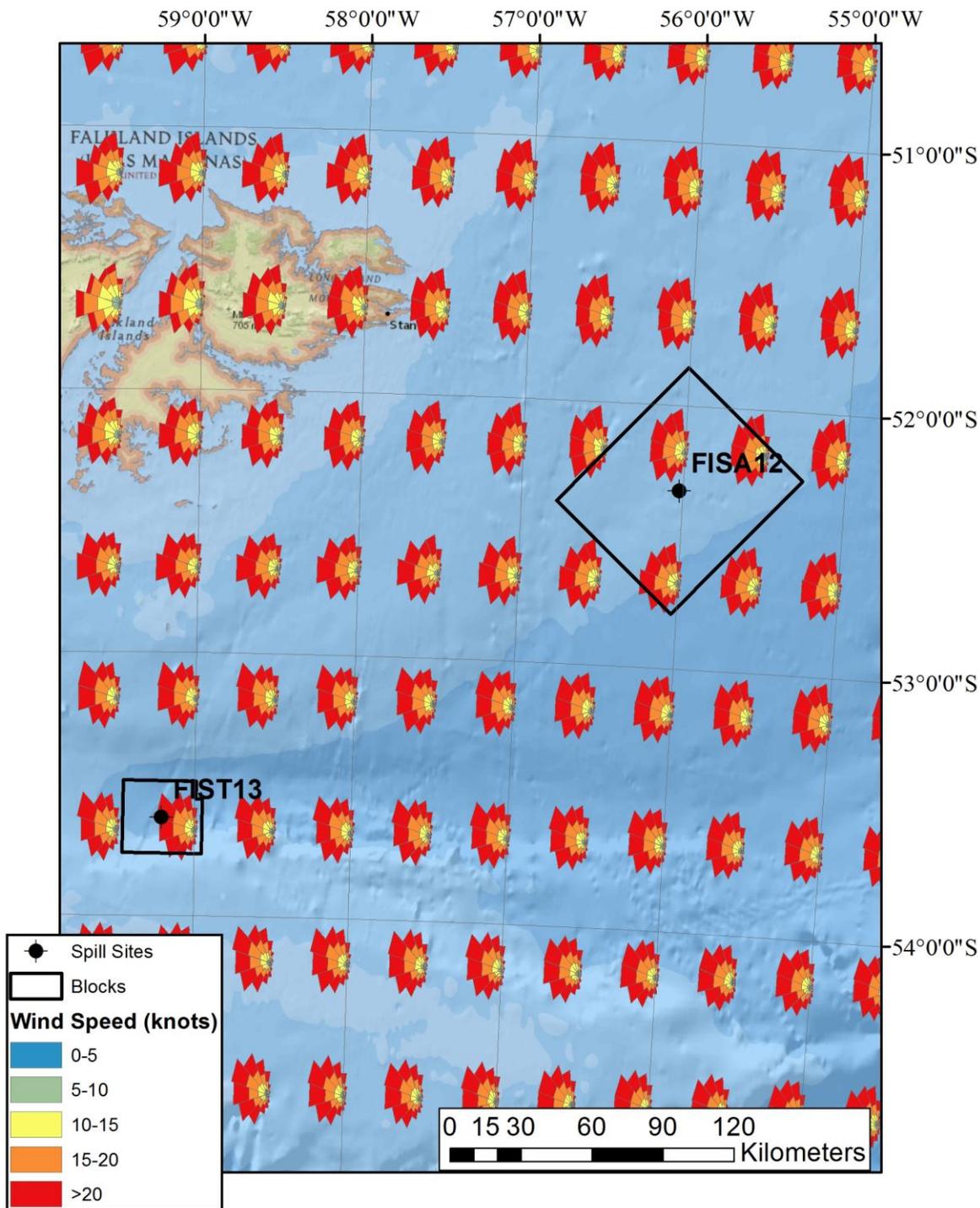


Figure 2. Annually-averaged NOGAPS wind roses representing the spatial variability of the wind field in the area of interest.

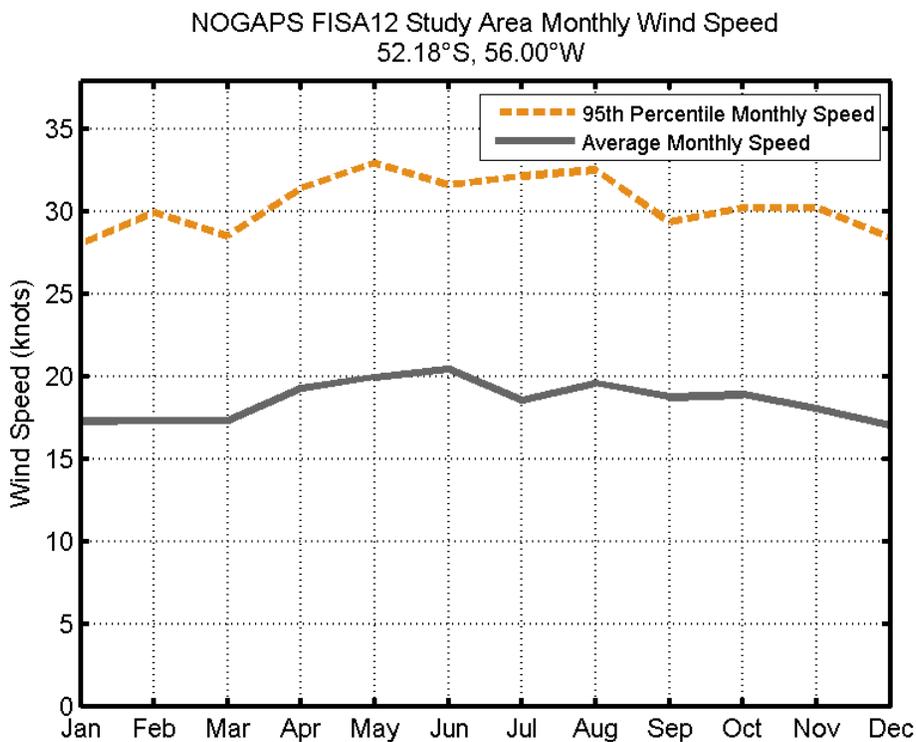


Figure 3. NOGAPS monthly wind speed statistics for FISA12.

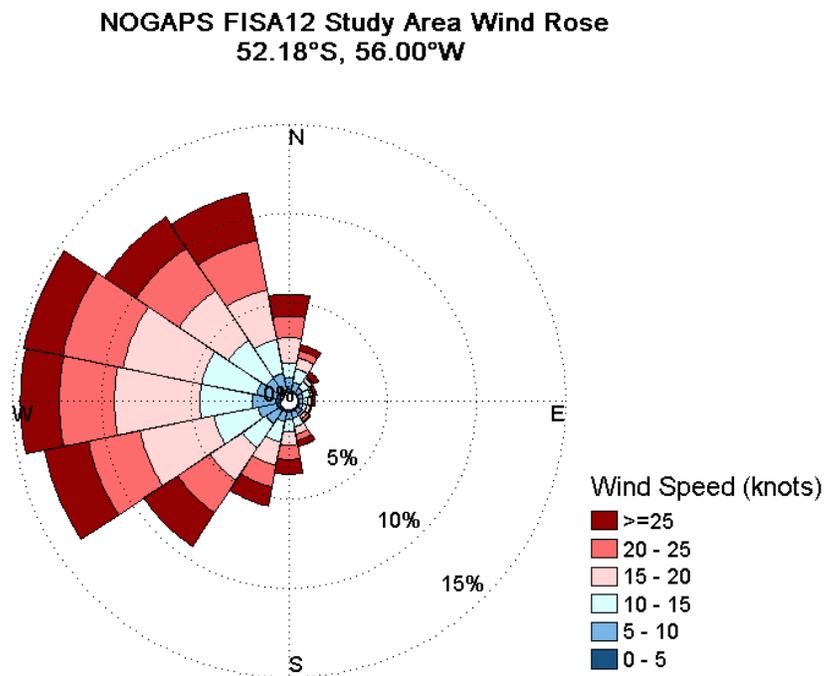


Figure 4. Annually-averaged NOGAPS wind rose for FISA12.

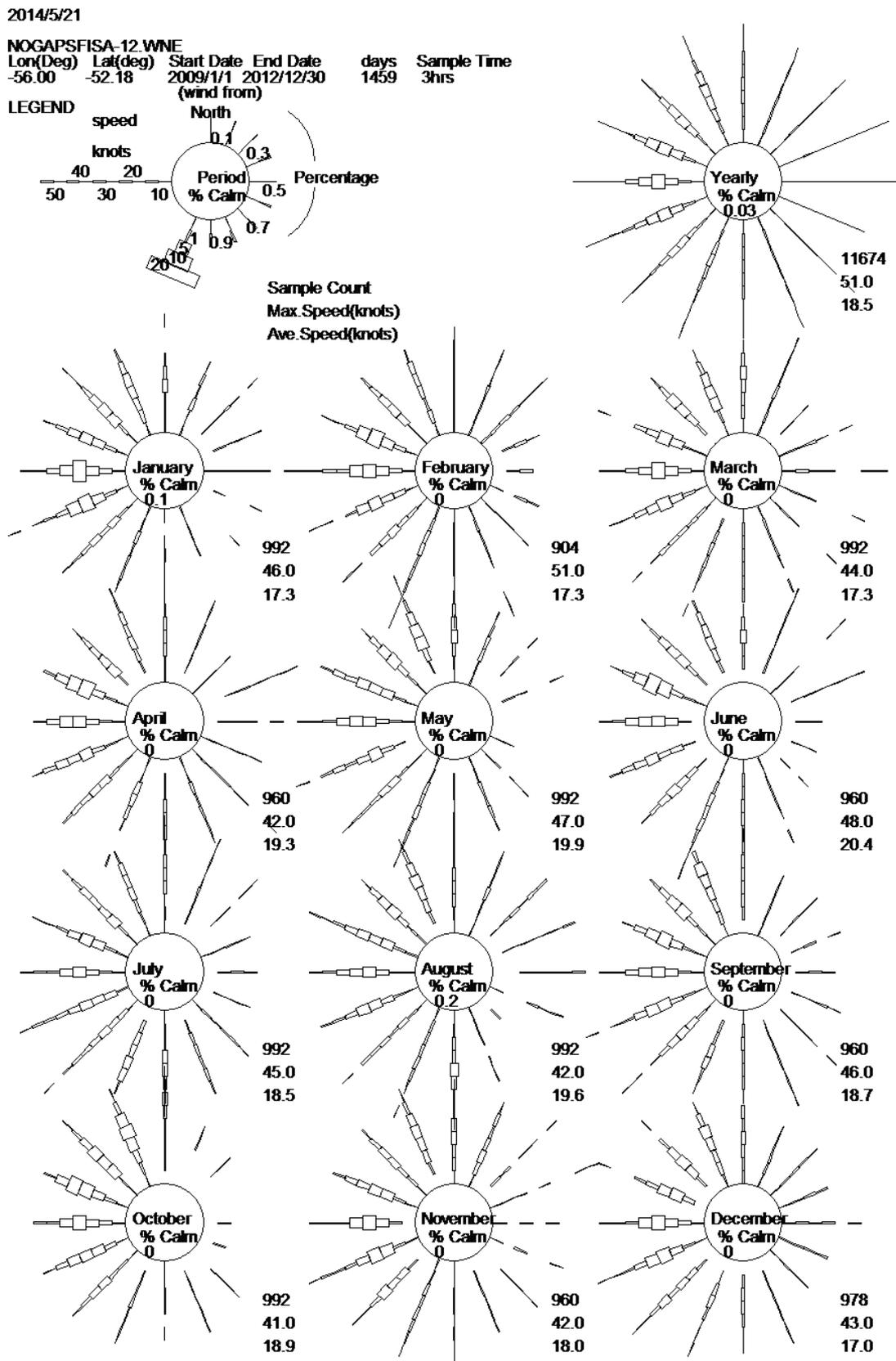


Figure 5. Monthl-y and annually-averaged NOGAPS wind roses for FISA12.

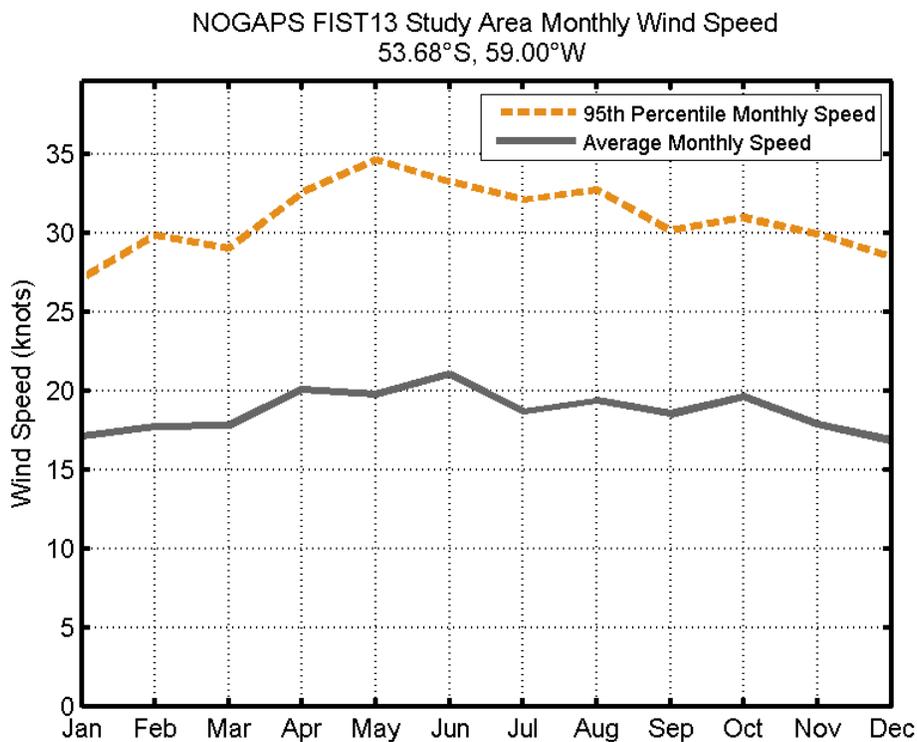


Figure 6. NOGAPS monthly wind speed statistics for FIST13.

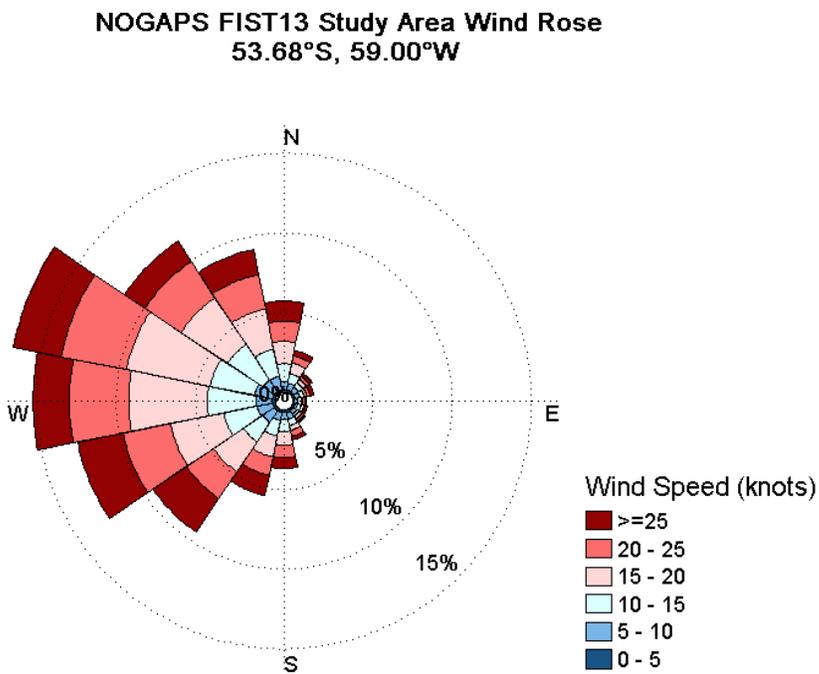


Figure 7. Annually-averaged NOGAPS wind rose for FIST13.

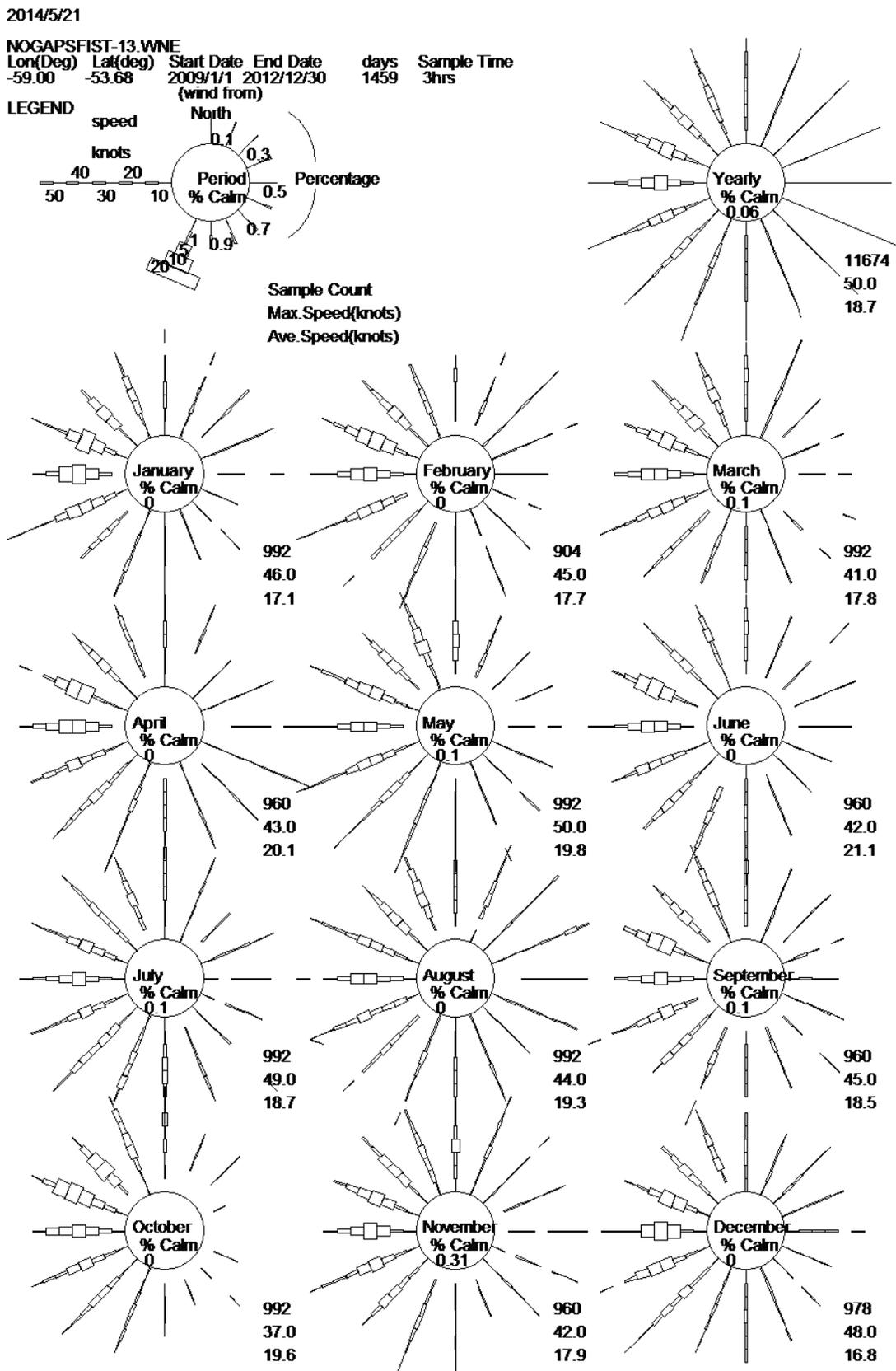


Figure 8. Monthly and annually-averaged NOGAPS wind roses for FIST13.

2.2. Current Dataset – HYCOM

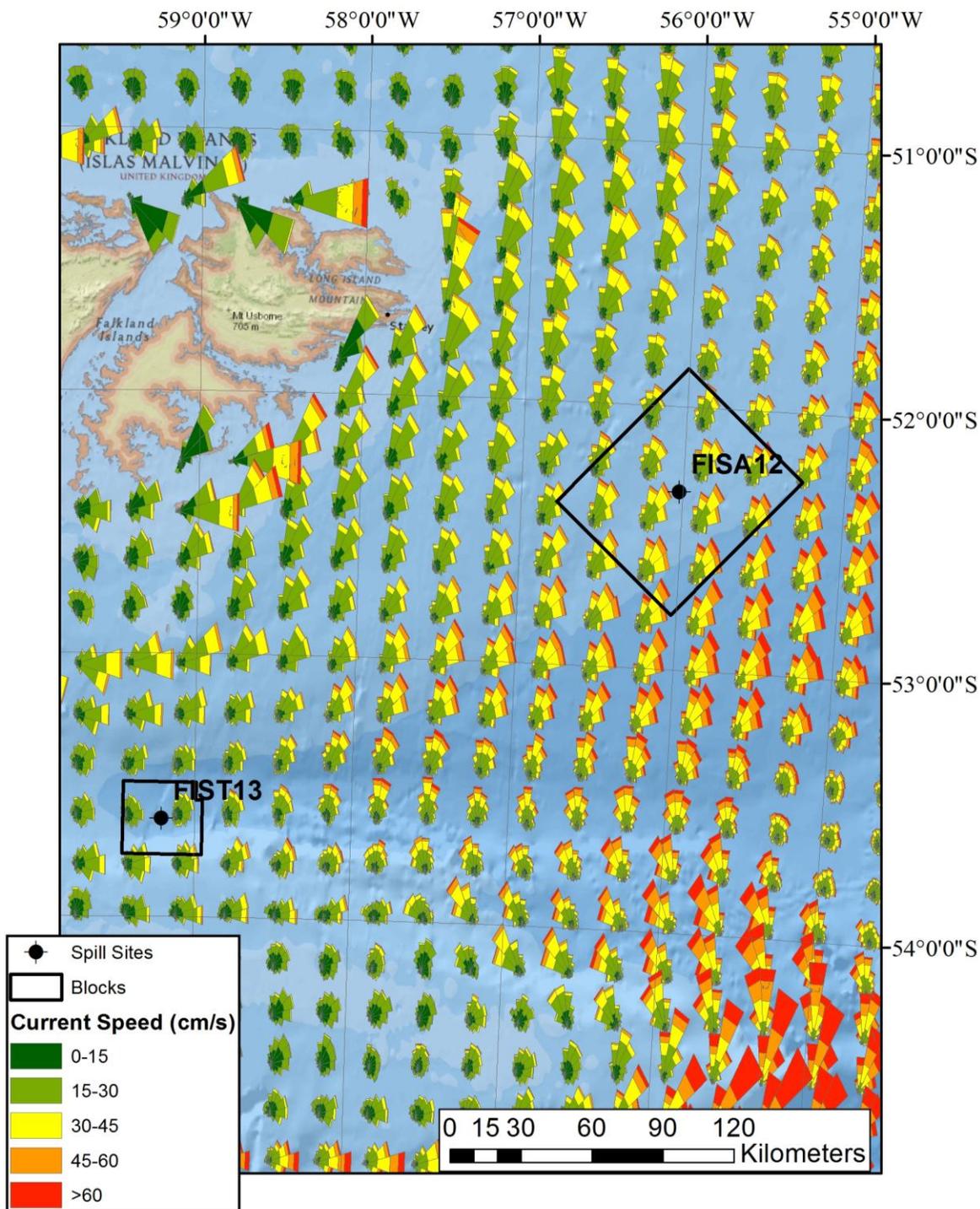


Figure 9. Annually-averaged HYCOM current roses representing the spatial variability of currents in the area of interest.

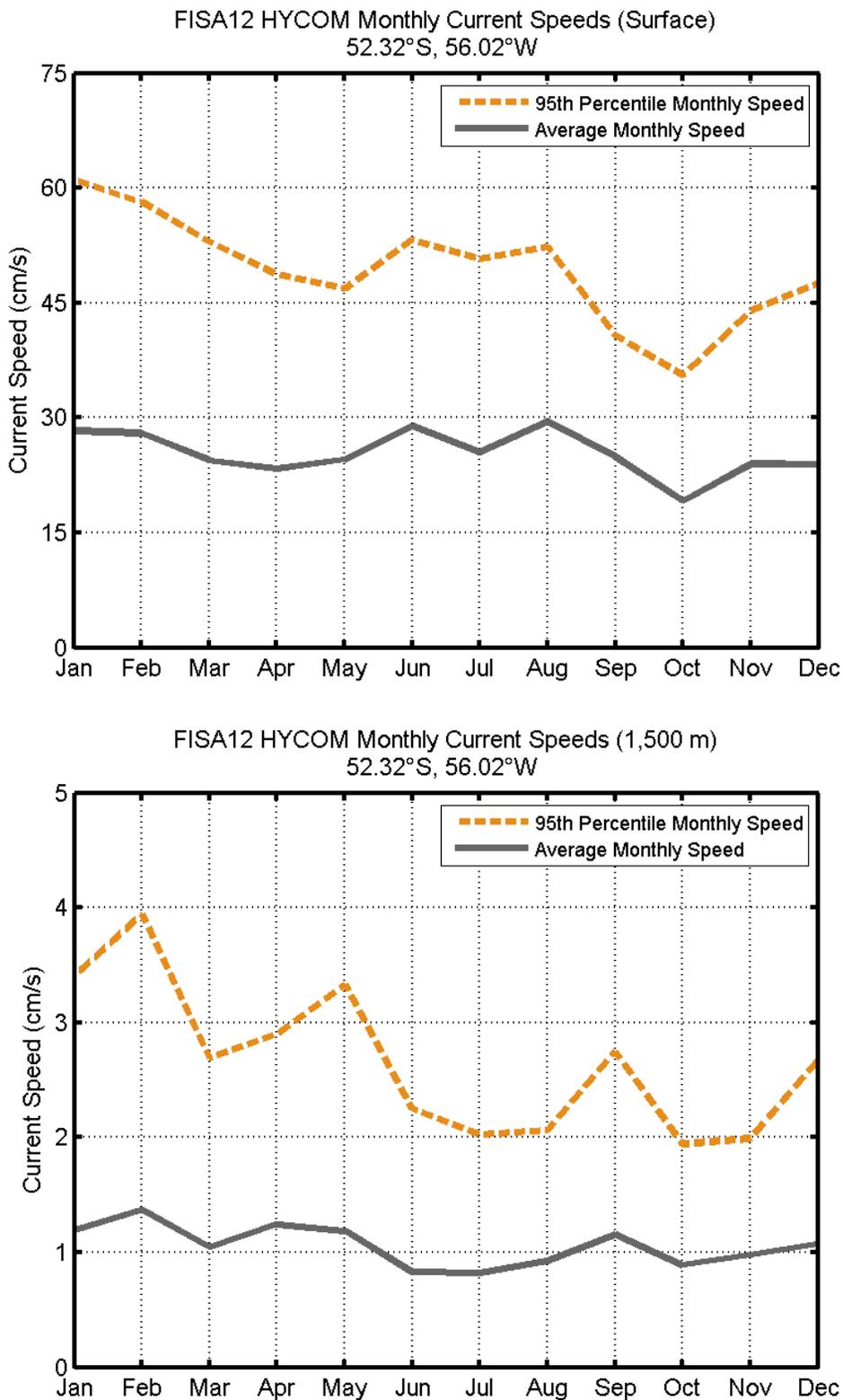


Figure 10. HYCOM monthly current speed statistics for FISA12.

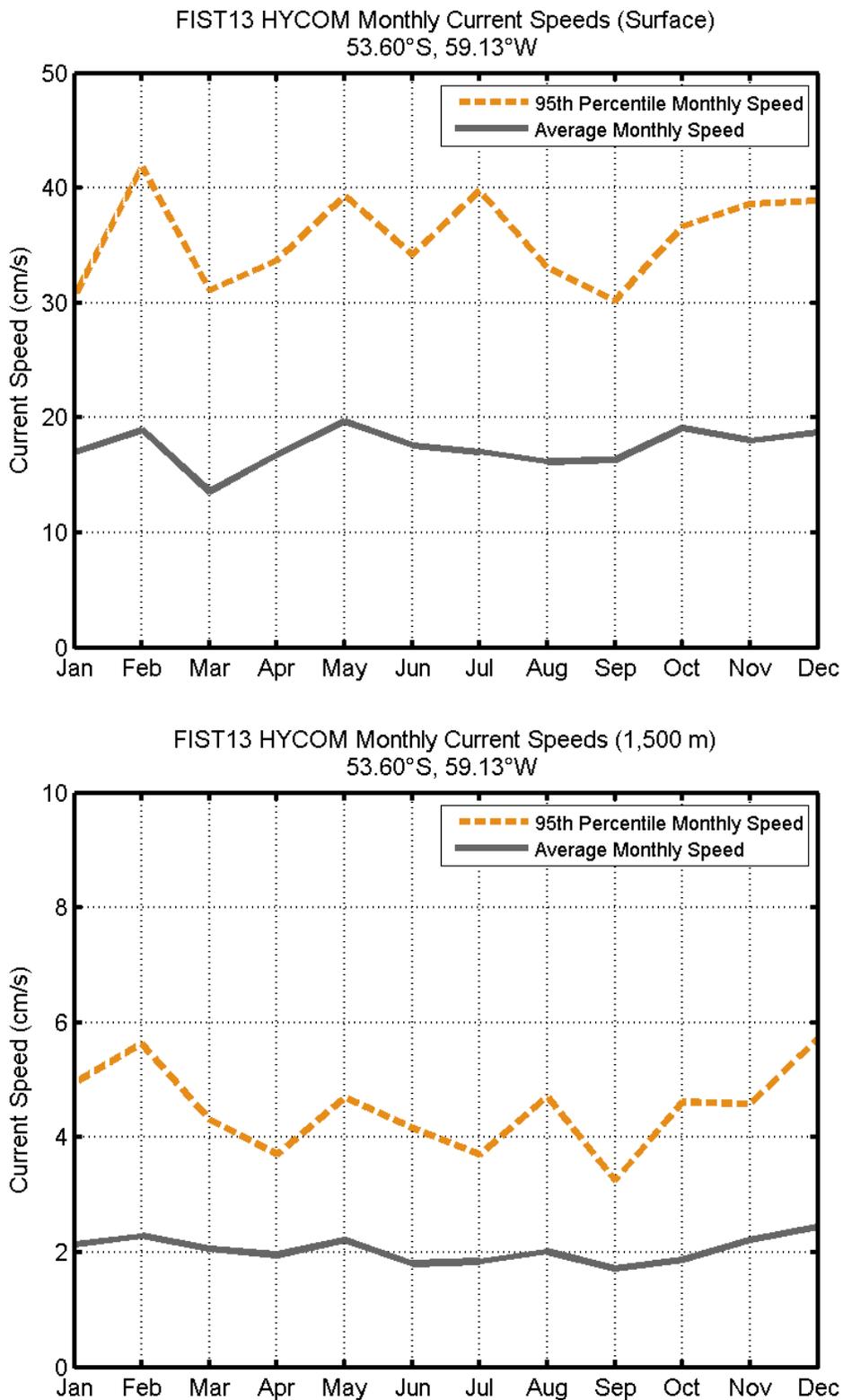
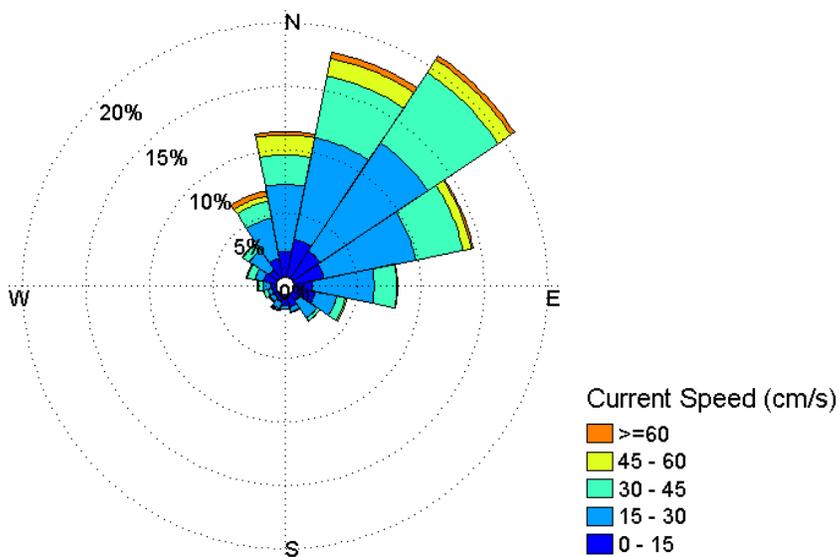


Figure 11. HYCOM monthly current speed statistics for FIST13.

HYCOM Current Rose near FISA12 (Surface)
52.32°S, 56.02°W



HYCOM Current Rose near FISA12 (1,500 m)
52.32°S, 56.02°W

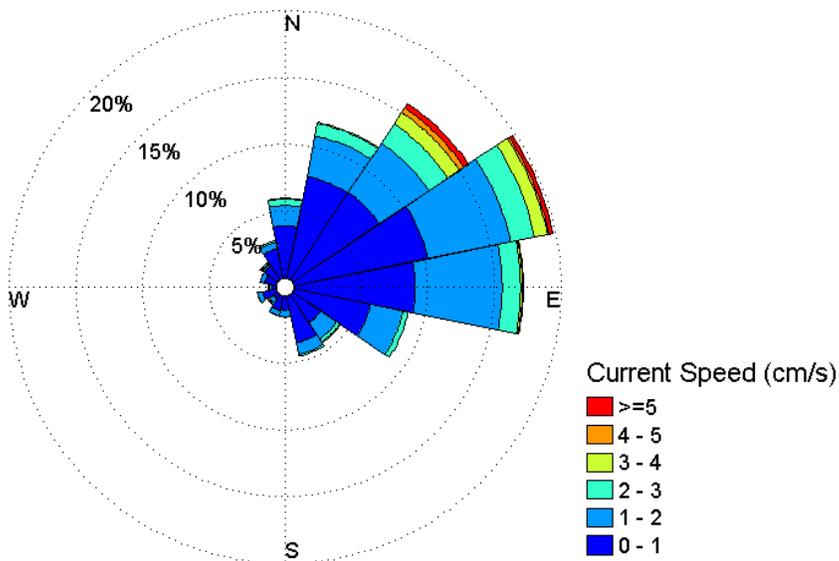


Figure 12. Annually-averaged HYCOM current roses for FISA12.

Monthly current roses at FISA12 (Surface)
 52.32°S, 56.02°W

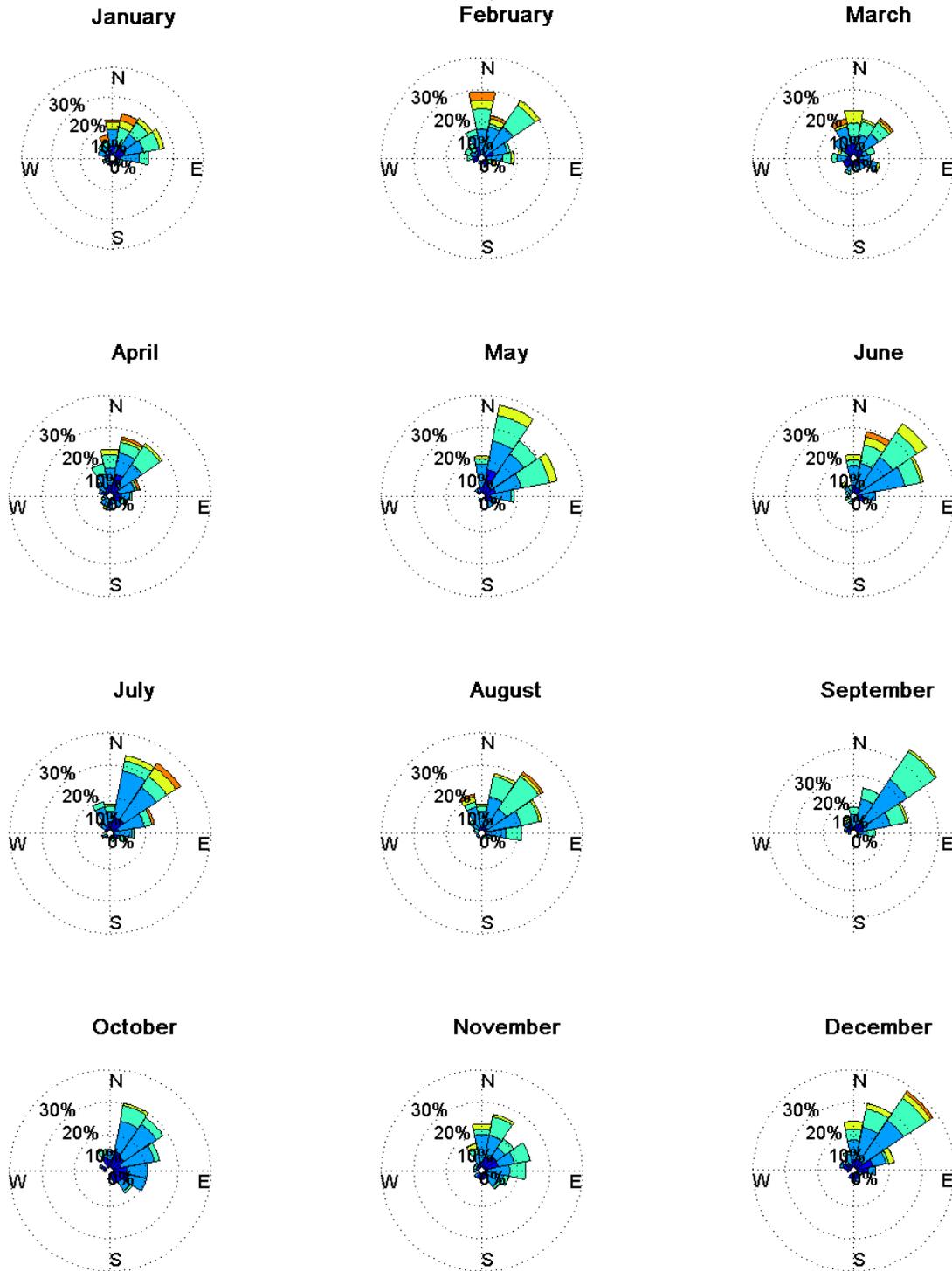
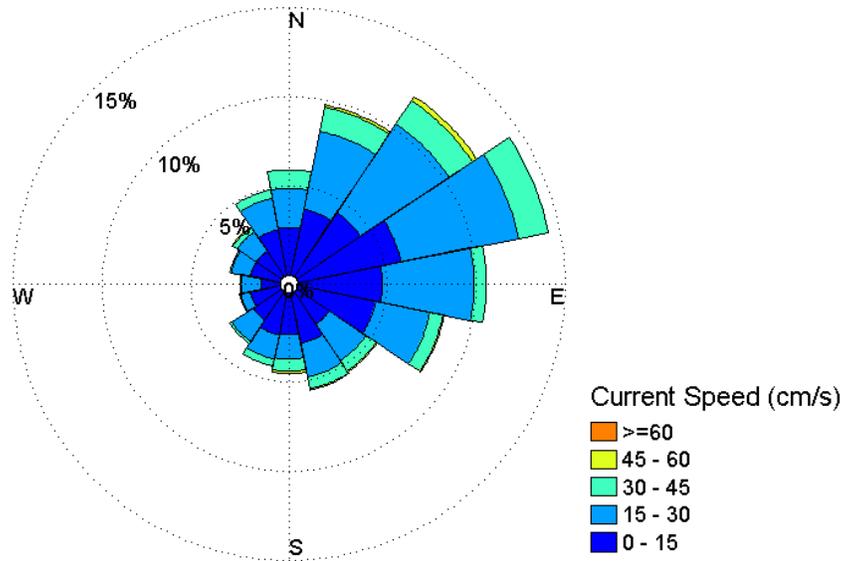


Figure 13. Monthly-averaged HYCOM current roses for FISA12.

HYCOM Current Rose near FIST13 (Surface)
53.60°S, 59.13°W



HYCOM Current Rose near FIST13 (1,500 m)
53.60°S, 59.13°W

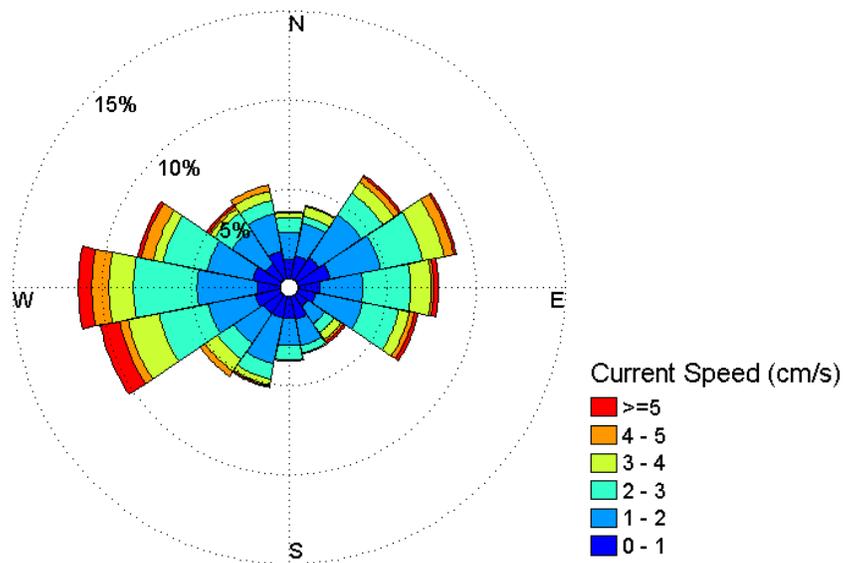


Figure 14. Annually-averaged HYCOM current roses for FIST13.

Monthly current roses at FIST13 (Surface)
53.60°S, 59.13°W

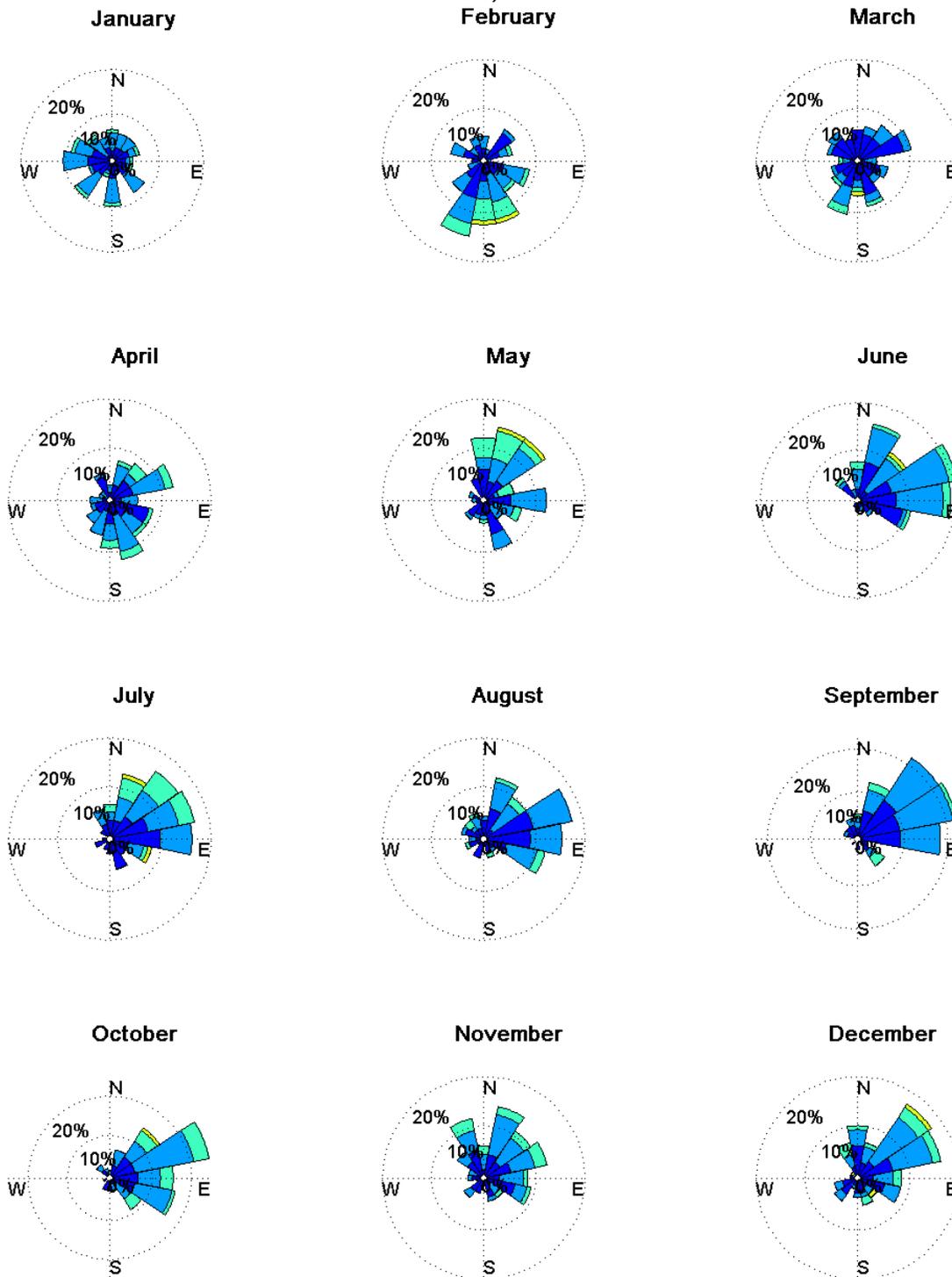


Figure 15. Monthly-averaged HYCOM current roses for FIST13.

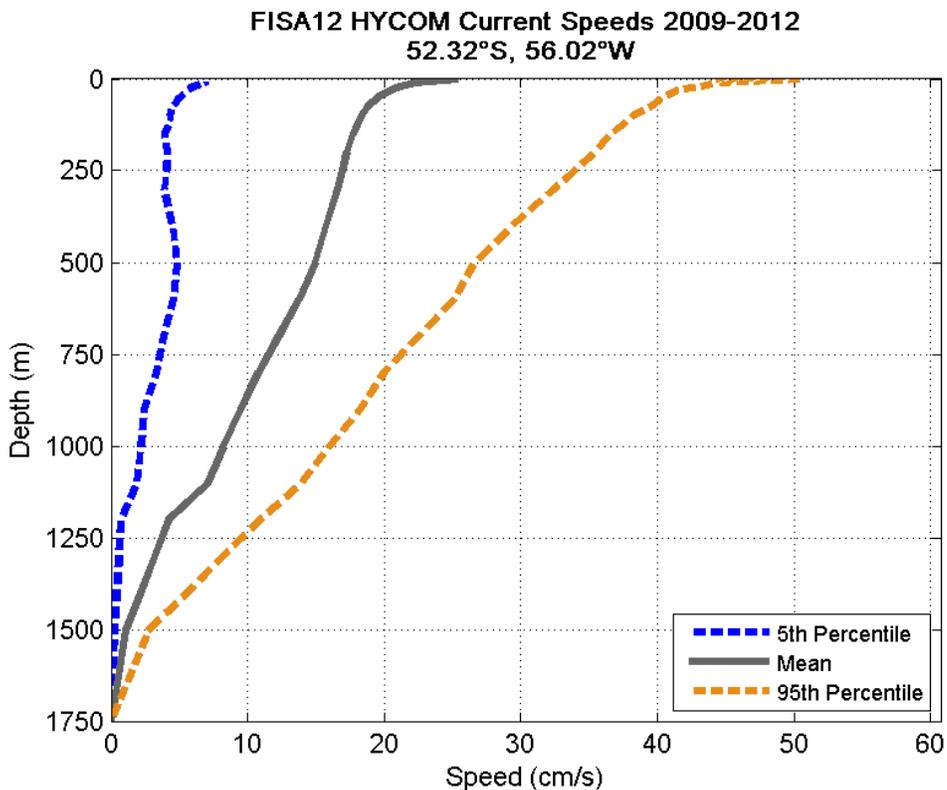


Figure 16. Annually-averaged HYCOM current speed statistics with depth for FISA12.

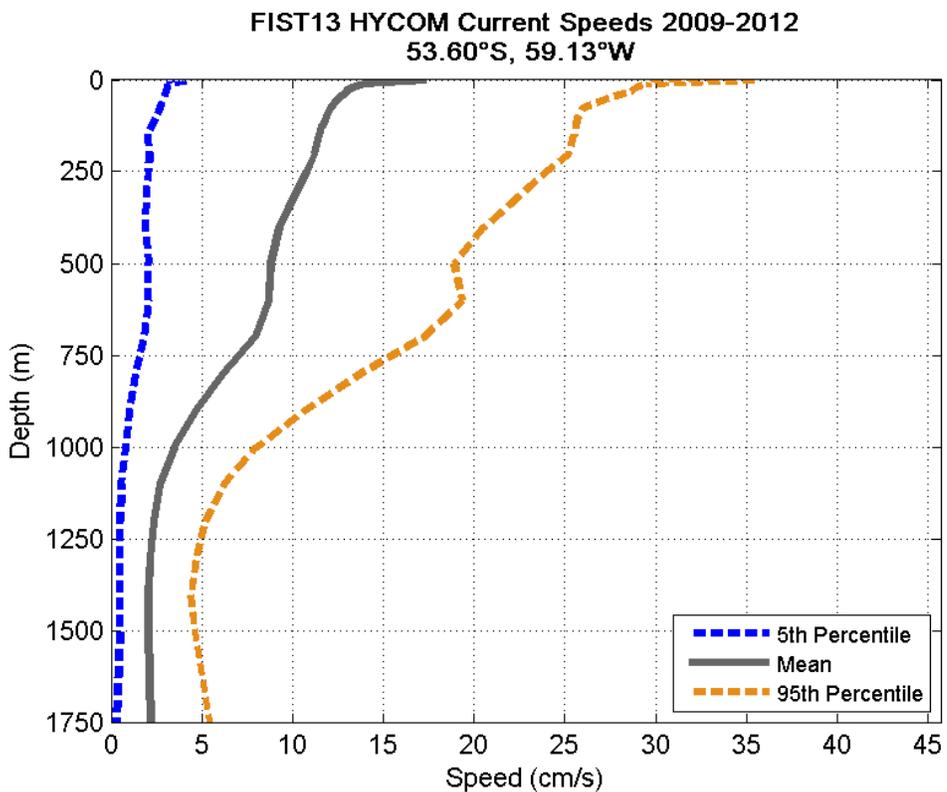


Figure 17. Annually-averaged HYCOM current speed statistics with depth for FIST13.

2.3. Water Column Vertical Structure

Temperature, Salinity and Density for FISA12
 52.13°S, 56.13°W

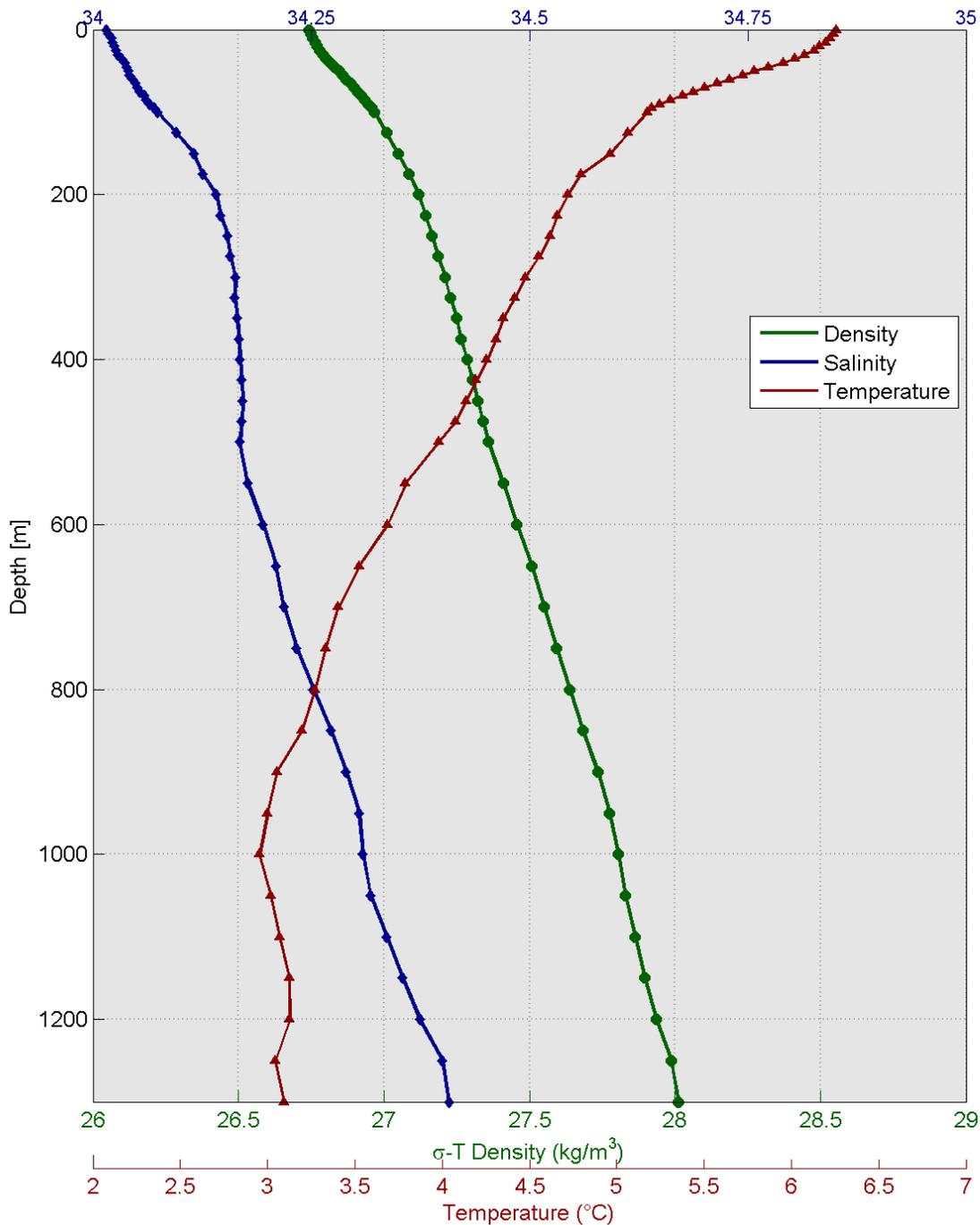


Figure 18. Annually-averaged temperature, salinity, and density vertical profiles for FISA12.

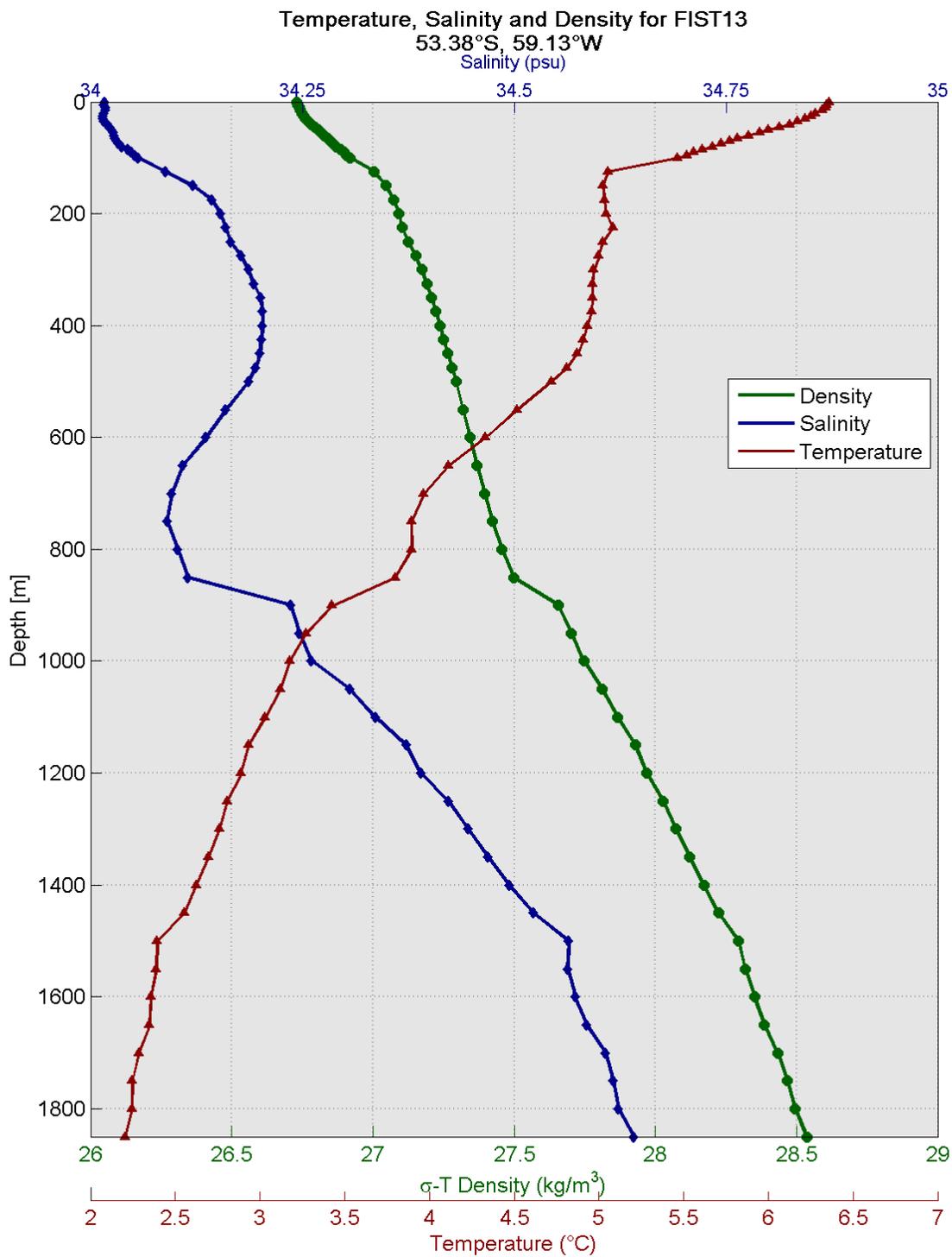


Figure 19. Annually-averaged temperature, salinity, and density vertical profiles for FIST13.

3. Oil Spill Simulations – Preliminary Results

3.1. Modelling Inputs

Table 2. Summary of oil properties used in the simulations.

| Oil Type | API Gravity (°) | Viscosity (cP at 25°C) | Interface Tension (dyne/cm) | Emulsion maximum Water Content (%) |
|-------------|-----------------|------------------------|-----------------------------|------------------------------------|
| Light Crude | 38.9 | 4.00 | 26.1 | 89.6 |
| Diesel | 38.8 | 2.76 | 27.5 | 0 |

Table 3. Blowout conditions used in the subsurface simulation.

| Site | Water Depth of the release (m) | Gas to Oil Ratio (e.g. scf/bbl) | Opening Diameter (e.g. inches) | Discharge Temperature (°C) |
|--------|--------------------------------|---------------------------------|--------------------------------|----------------------------|
| FISA12 | 1,177 | 600 scf/bbl | 13.375" | 104 |
| FIST13 | 1,527 | 600 scf/bbl | 13.375" | 104 |

Table 4. Simulation periods used.

| Period | Months to Use |
|--------------------------|--------------------|
| Period 1: Austral Summer | October - February |
| Period 2: Austral Winter | March - September |

Table 5. Parameters of the oil spill scenarios.

| ID | Spill Site | Spill Event | Oil Type | Period | Spill Rate | Spill Duration | Total Spilled Volume | Simulation Duration |
|---|------------|---|-----------|-----------------|--------------|----------------|----------------------|---------------------|
| Stochastic Scenarios | | | | | | | | |
| 1 | FISA12 | Drilling Rig Fuel Oil Inventory | Diesel | Period 1 (10-2) | Instant | Instant | 4,631 m ³ | 14 Days |
| 2 | FISA12 | Drilling Rig Fuel Oil Inventory | Diesel | Period 2 (3-9) | Instant | Instant | 4,631 m ³ | 14 Days |
| 3 | FIST13 | Drilling Rig Fuel Oil Inventory | Diesel | Period 1 (10-2) | Instant | Instant | 4,631 m ³ | 14 Days |
| 4 | FIST13 | Drilling Rig Fuel Oil Inventory | Diesel | Period 2 (3-9) | Instant | Instant | 4,631 m ³ | 14 Days |
| 5 | FISA12 | Surface Blowout | Crude Oil | Period 1 (10-2) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 6 | FISA12 | Surface Blowout | Crude Oil | Period 2 (3-9) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 7 | FIST13 | Surface Blowout | Crude Oil | Period 1 (10-2) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 8 | FIST13 | Surface Blowout | Crude Oil | Period 2 (3-9) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 9 | FISA12 | Subsurface Blowout | Crude Oil | Period 2 (3-9) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 10 | FIST13 | Subsurface Blowout | Crude Oil | Period 2 (3-9) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| Deterministic Scenarios (Worst Cases from Stochastic) | | | | | | | | |
| 5 | FISA12 | Surface Blowout (worst case: shortest time from stochastic) | Crude Oil | Period 1 (10-2) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 6 | FISA12 | Surface Blowout (worst case: shortest time from stochastic) | Crude Oil | Period 2 (3-9) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 7 | FIST13 | Surface Blowout (worst case: shortest time from stochastic) | Crude Oil | Period 1 (10-2) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |
| 8 | FIST13 | Surface Blowout (worst case: shortest time from stochastic) | Crude Oil | Period 2 (3-9) | 50,071 bbl/d | 10 days | 500,710 bbl | 30 Days |

3.2. Blowout Near-Field Modelling Results

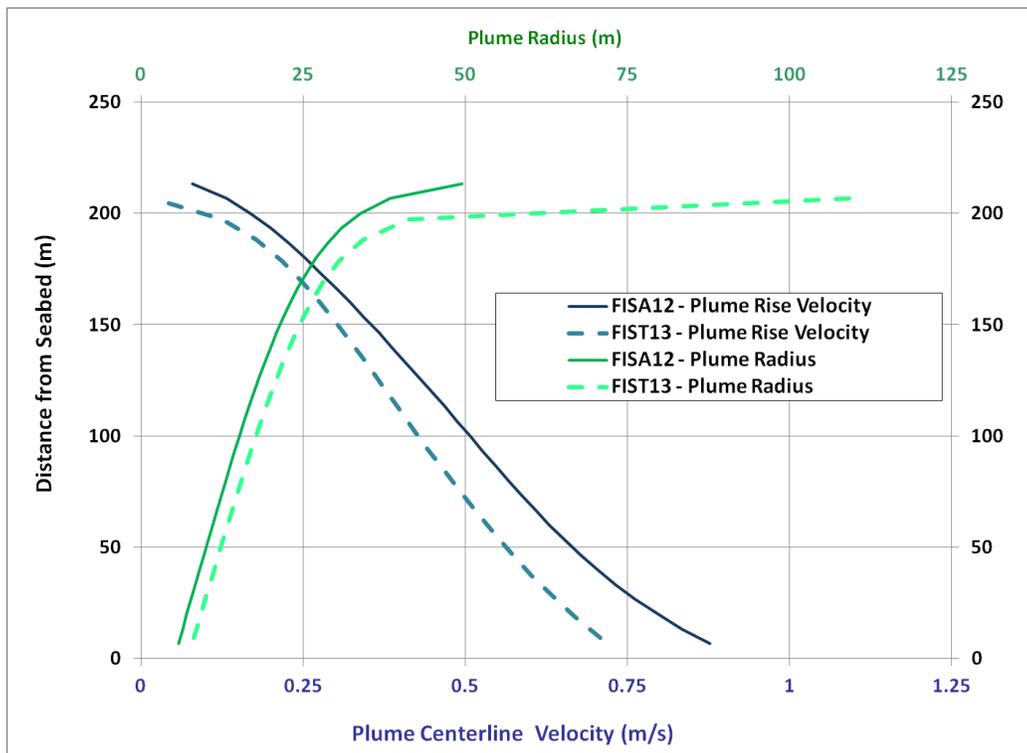


Figure 20. Predicted plume radius and plume centreline velocity for the subsurface blowout events.

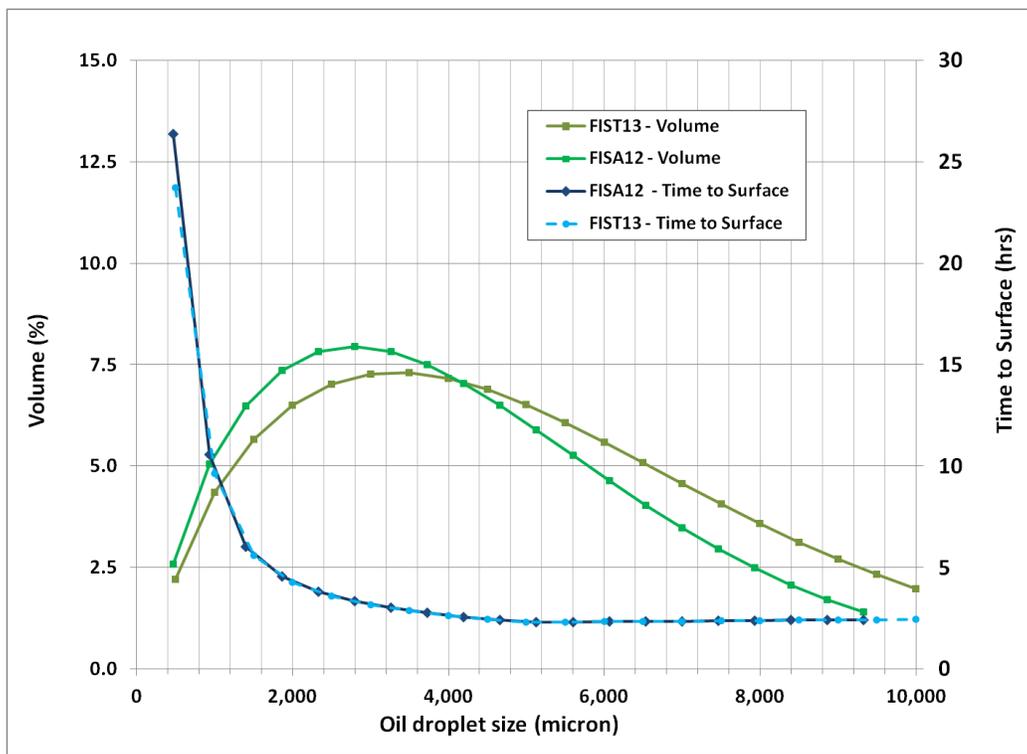


Figure 21. Predicted droplet size distribution and droplet rise time for the subsurface blowout events.

3.3. Stochastic Modelling Results

Table 6. Oil Spill Stochastic results – Predicted shoreline impacts.

| Spill Site | Oil Type | Spill Type | Simulation Period | Total Volume Released | Sims. Reaching Shore (%) | Time to Reach Shore (days) | |
|------------|-----------|---------------------------------|-------------------|-----------------------|--------------------------|----------------------------|------|
| | | | | | | Min. | Avg. |
| FISA12 | Diesel | Drilling Rig Fuel Oil Inventory | Period 1 (10-2) | 4,631 m ³ | 0 | - | - |
| FISA12 | Diesel | Drilling Rig Fuel Oil Inventory | Period 2 (3-9) | 4,631 m ³ | 0 | - | - |
| FIST13 | Diesel | Drilling Rig Fuel Oil Inventory | Period 1 (10-2) | 4,631 m ³ | 0 | - | - |
| FIST13 | Diesel | Drilling Rig Fuel Oil Inventory | Period 2 (3-9) | 4,631 m ³ | 0 | - | - |
| FISA12 | Crude Oil | Surface Blowout | Period 1 (10-2) | 500,710 bbl | 0 | - | - |
| FISA12 | Crude Oil | Surface Blowout | Period 2 (3-9) | 500,710 bbl | 2 | 10.0 | 10.5 |
| FIST13 | Crude Oil | Surface Blowout | Period 1 (10-2) | 500,710 bbl | 13 | 1.7 | 16.3 |
| FIST13 | Crude Oil | Surface Blowout | Period 2 (3-9) | 500,710 bbl | 32 | 1.7 | 10.2 |
| FISA12 | Crude Oil | Subsurface Blowout | Period 2 (3-9) | 500,710 bbl | 2 | 11.0 | 12.0 |
| FIST13 | Crude Oil | Subsurface Blowout | Period 2 (3-9) | 500,710 bbl | 18 | 4.0 | 10.9 |

The following figures illustrate the spatial extent of surface and shoreline oiling probabilities and associated minimum travel times for the spills; only oiling above a threshold of 0.04 μm is included.

For each scenario, two figures are presented. For scenarios where there is a model-predicted potential for shoreline oiling, two additional figures are presented.

1. **Probability of surface oil exceeding 0.04 μm :** The map defines the area in which sea surface oil has at least a 1% chance to exceed 0.04 μm and the associated probability of exceeding the threshold based on analysis of the resulting trajectories from the ensemble of individual simulations run for each spill scenario. The map does not imply that the entire contoured area would be covered with oil in the event of a spill. The map also does not provide any information on the amount of oil in a given area.
2. **Minimum time for surface oil to exceed 0.04 μm :** The footprint on this map corresponds to the surface probability map, and illustrates the shortest time required for oil to reach any point within the footprint. These results are also based on the ensemble of all individual simulations.
3. **Probability of shoreline oil exceeding 0.04 μm :** The map defines the area in which beached oil has at least a 1% chance to exceed 0.04 μm and the associated probability of exceeding the threshold based on analysis of the resulting trajectories from the ensemble of individual simulations run for each spill scenario. The map does not imply that the entire area would be covered with oil in the event of a spill. The map also does not provide any information on the amount of oil in a given area. In the absence of data, all shoreline segments are assumed to be 10-m-wide sandy beaches. Using actual shoreline data may alter the results, but the results provided are likely on the conservative side for shoreline impacts.
4. **Minimum time for shoreline oil to exceed 0.04 μm :** The footprint on this map corresponds to the shoreline probability map, and illustrates the shortest time required for oil to reach any point within the footprint. These results are also based on the ensemble of all individual simulations. In the absence of data, all shoreline segments are assumed to be 10-m-wide sandy beaches. Using actual shoreline data may alter the results, but the results provided are likely on the conservative side for shoreline impacts.

Stochastic Results: Instantaneous Release of 4,631 m³ of Diesel from FISA12 (Oct-Feb)

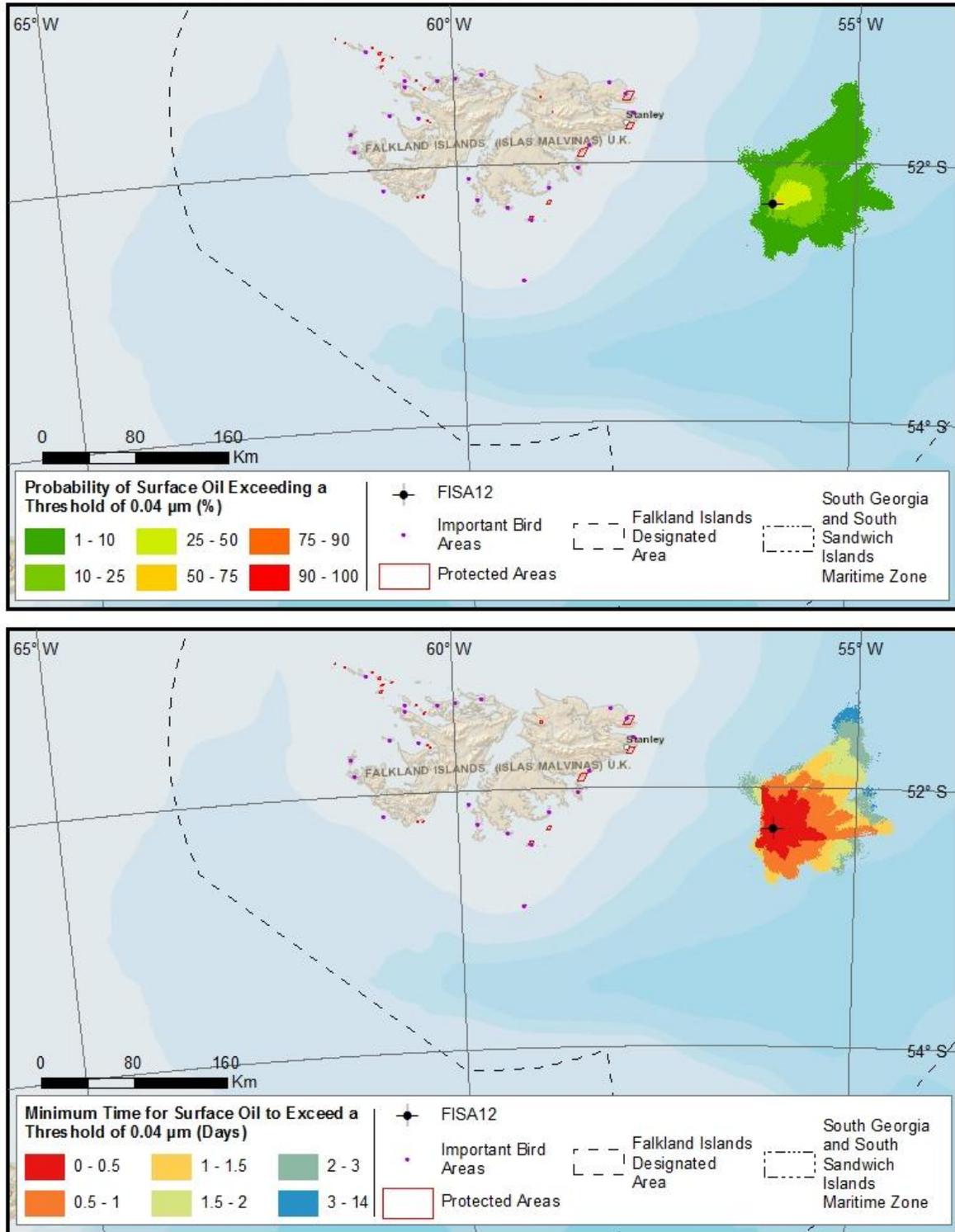


Figure 22. Scenario 1 stochastic maps for potential water surface contamination.

Stochastic Results: Instantaneous Release of 4,631 m³ of Diesel from FISA12 (Mar-Sep)

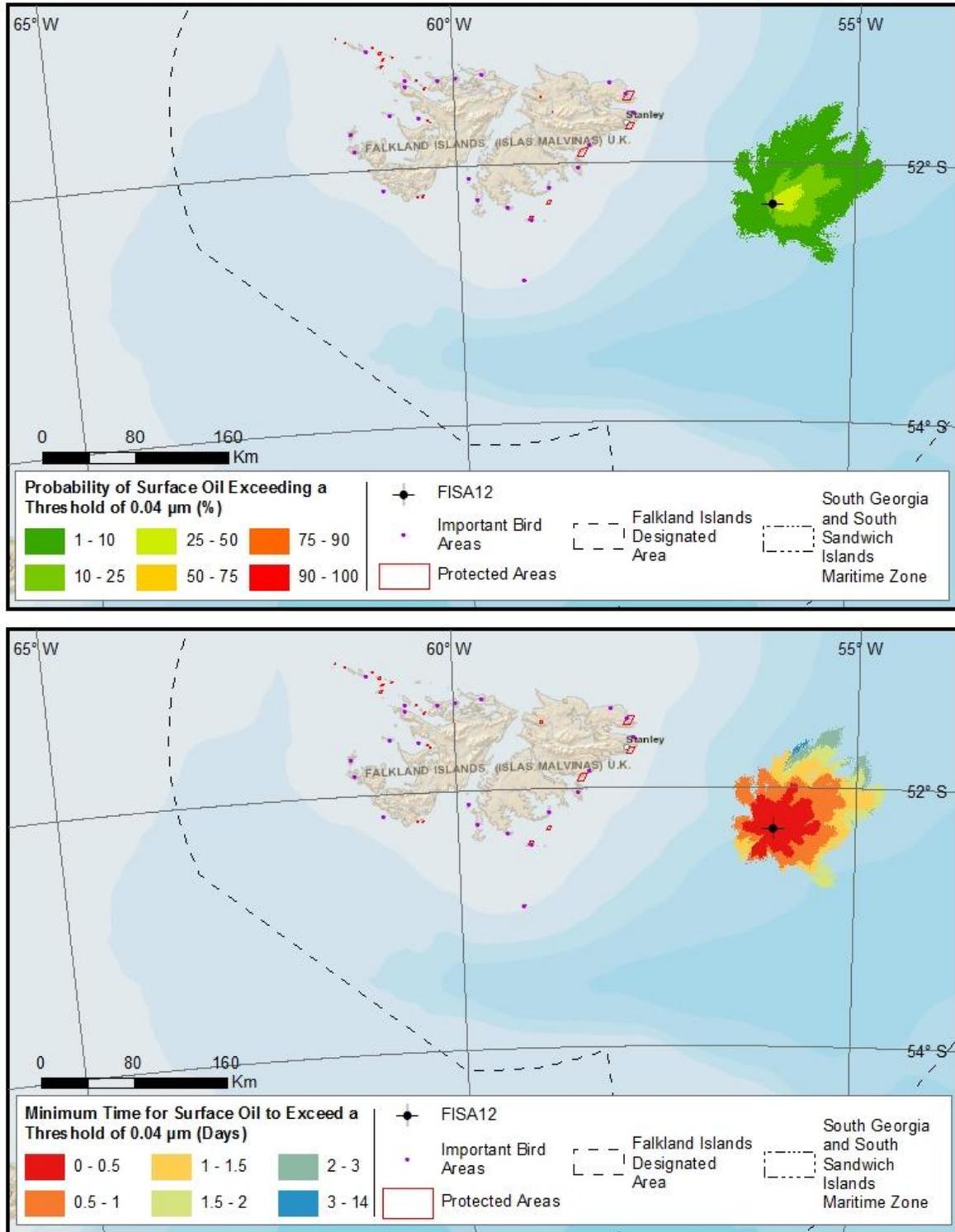


Figure 23. Scenario 2 stochastic maps for potential water surface contamination.

Stochastic Results: Instantaneous Release of 4,631 m³ of Diesel from FIST13 (Oct-Feb)

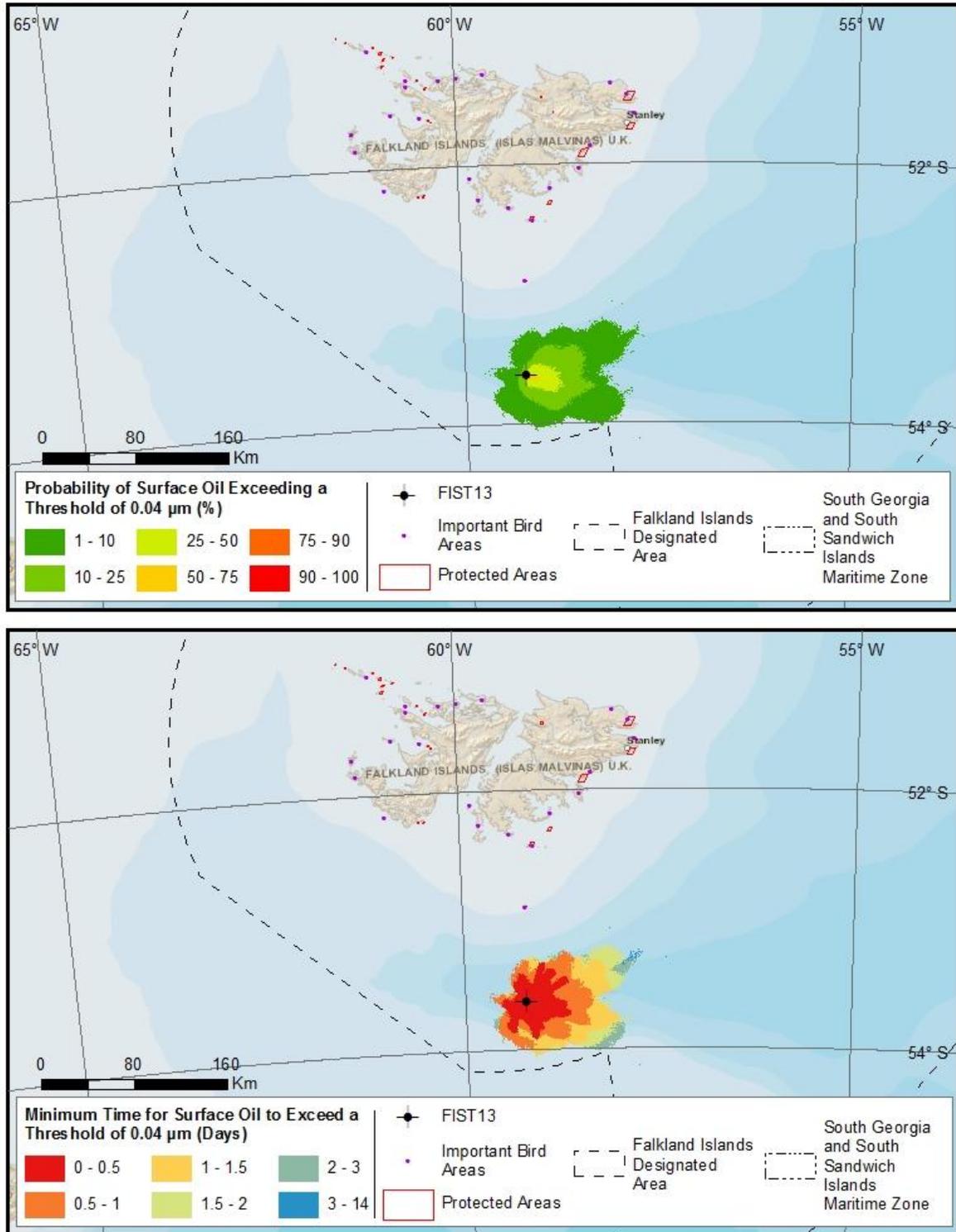


Figure 24. Scenario 3 stochastic maps for potential water surface contamination.

Stochastic Results: Instantaneous Release of 4,631 m³ of Diesel from FIST13 (Mar-Sep)

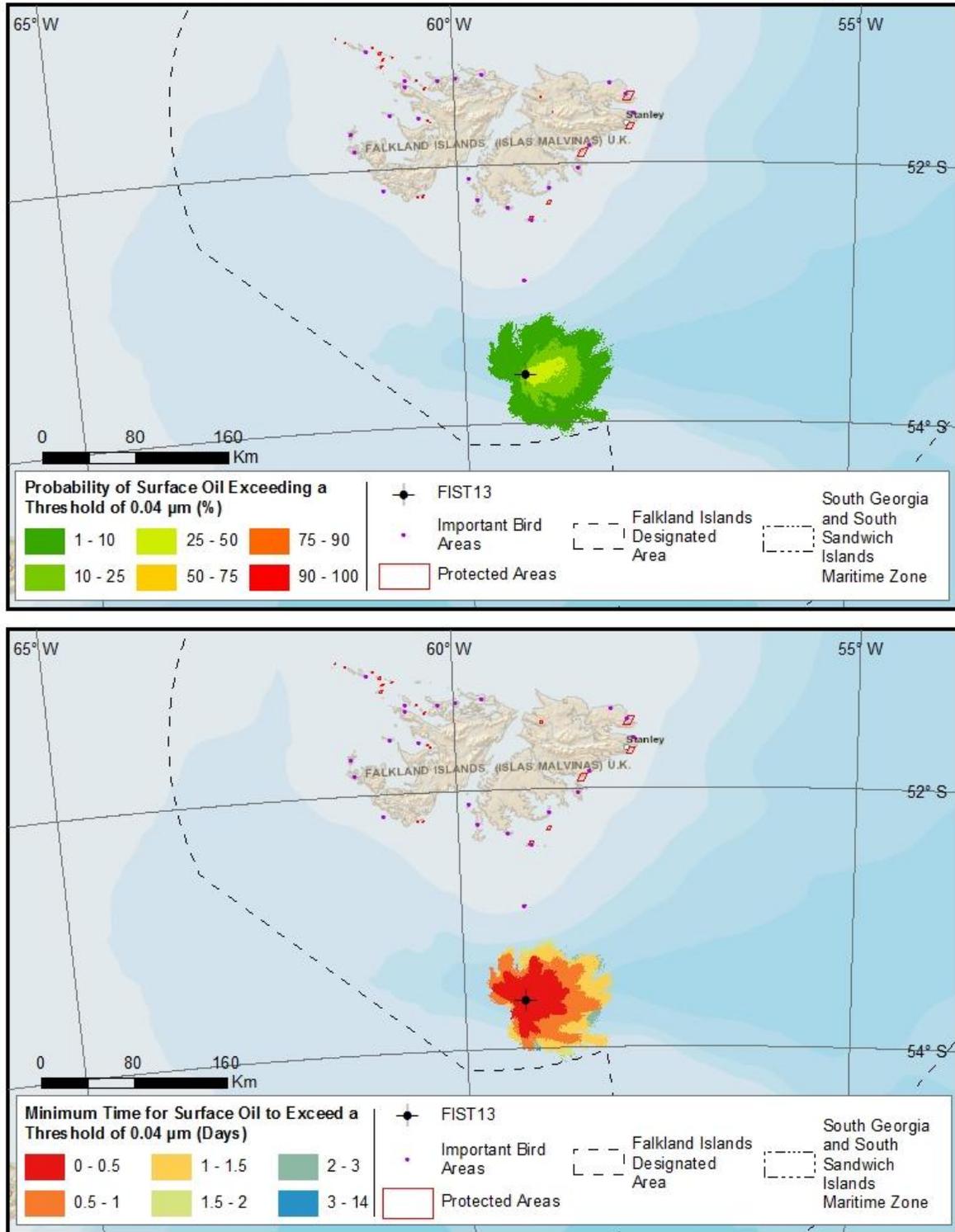


Figure 25. Scenario 4 stochastic maps for potential water surface contamination.

Stochastic Results: 10-Day Surface Blowout of 500,710 bbl of Crude Oil from FISA12 (Oct-Feb)

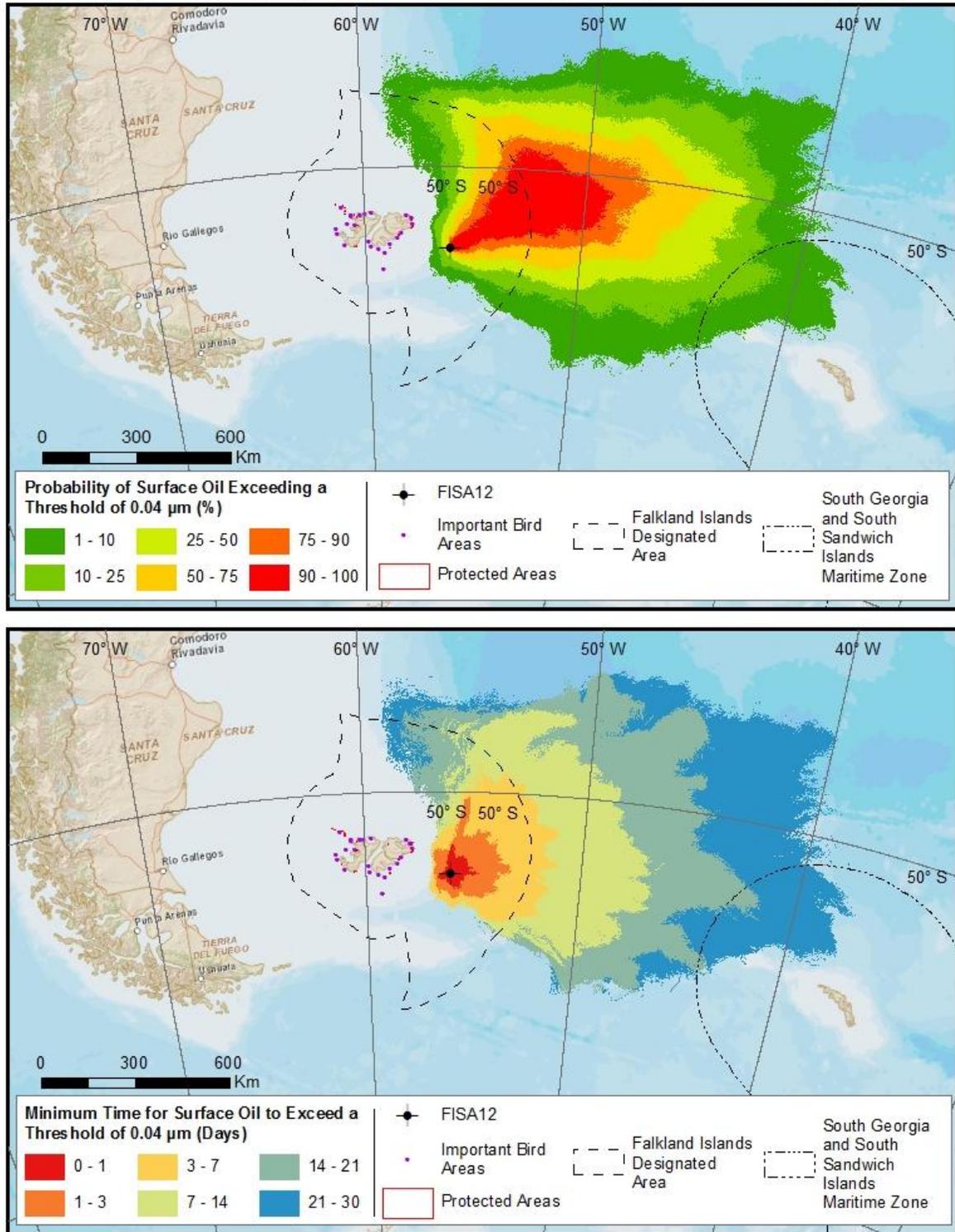


Figure 26. Scenario 5 stochastic maps for potential water surface contamination.

Stochastic Results: 10-Day Surface Blowout of 500,710 bbl of Crude Oil from FISA12 (Mar-Sep)

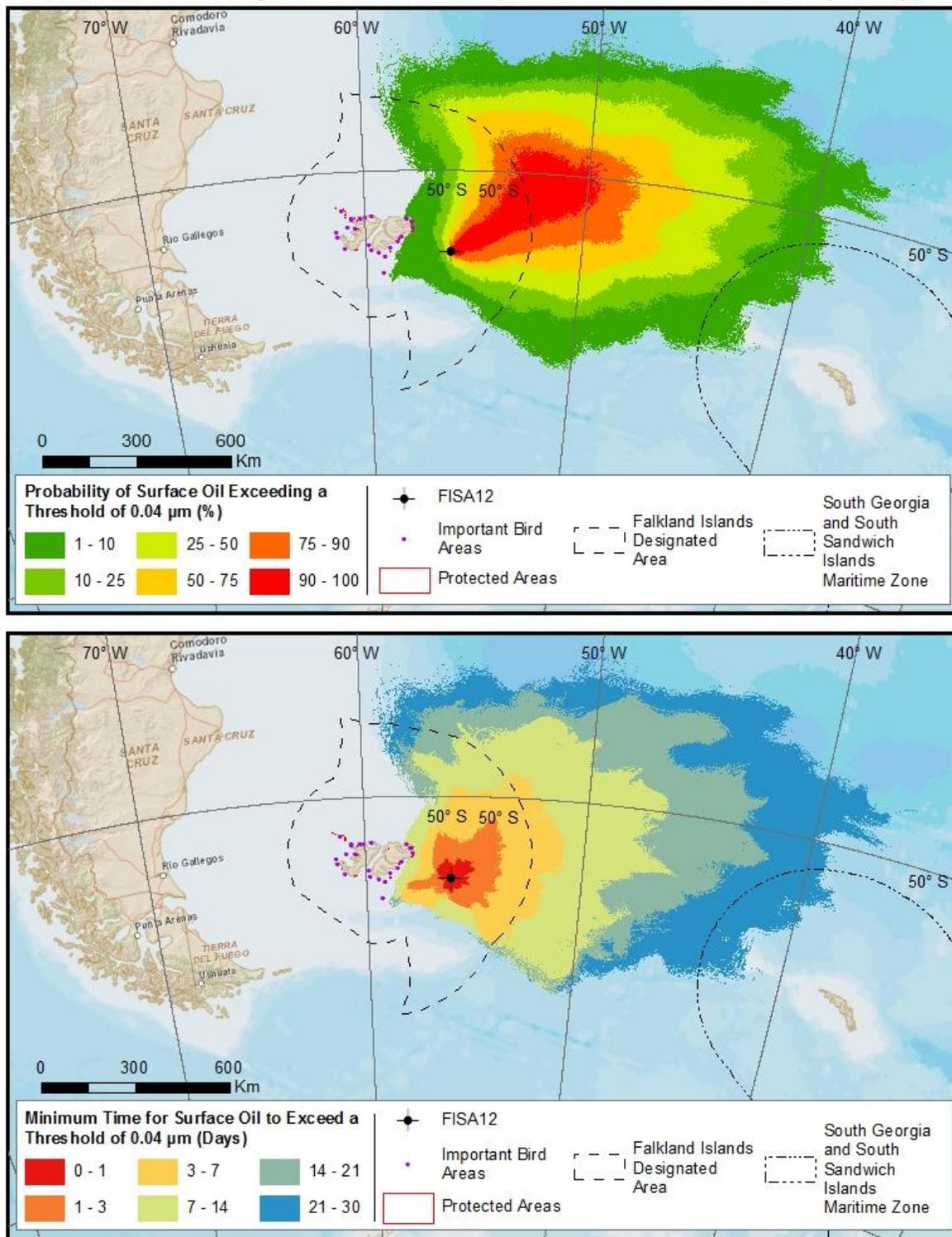


Figure 27. Scenario 6 stochastic maps for potential water surface contamination.

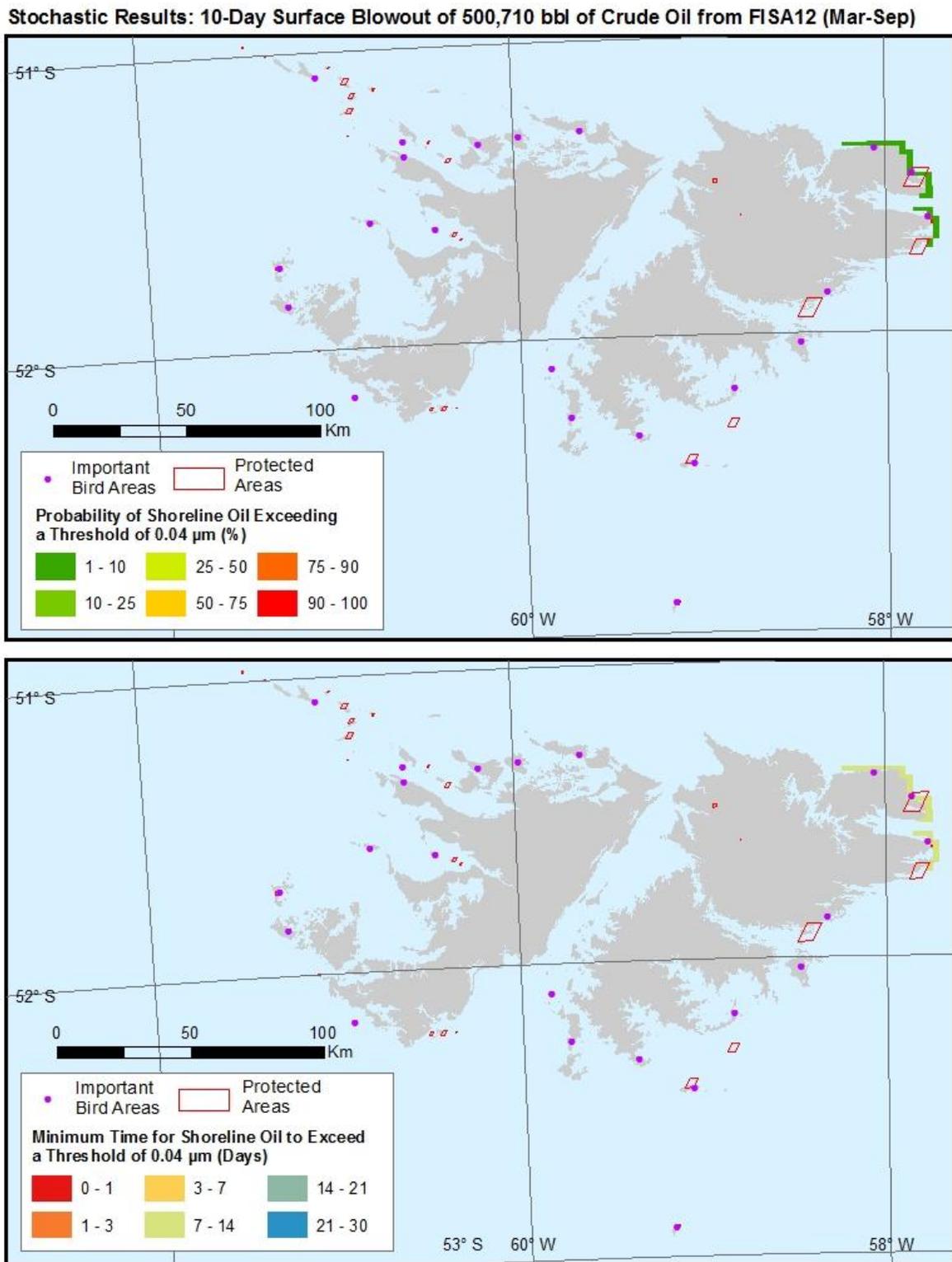


Figure 28. Scenario 6 stochastic maps for potential shoreline contamination.

Stochastic Results: 10-Day Surface Blowout of 500,710 bbl of Crude Oil from FIST13 (Oct-Feb)

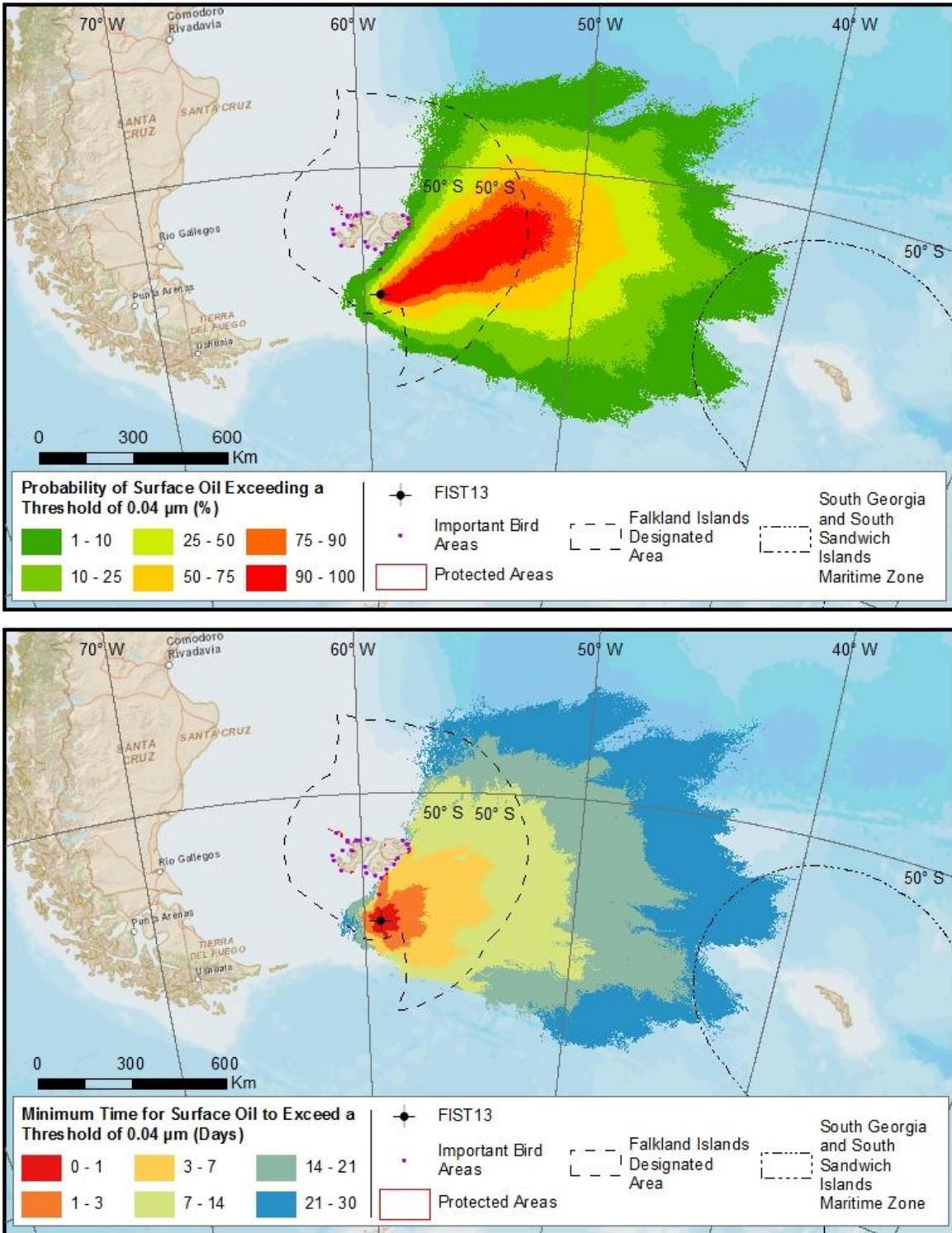


Figure 29. Scenario 7 stochastic maps for potential water surface contamination.

Stochastic Results: 10-Day Surface Blowout of 500,710 bbl of Crude Oil from FIST13 (Oct-Feb)

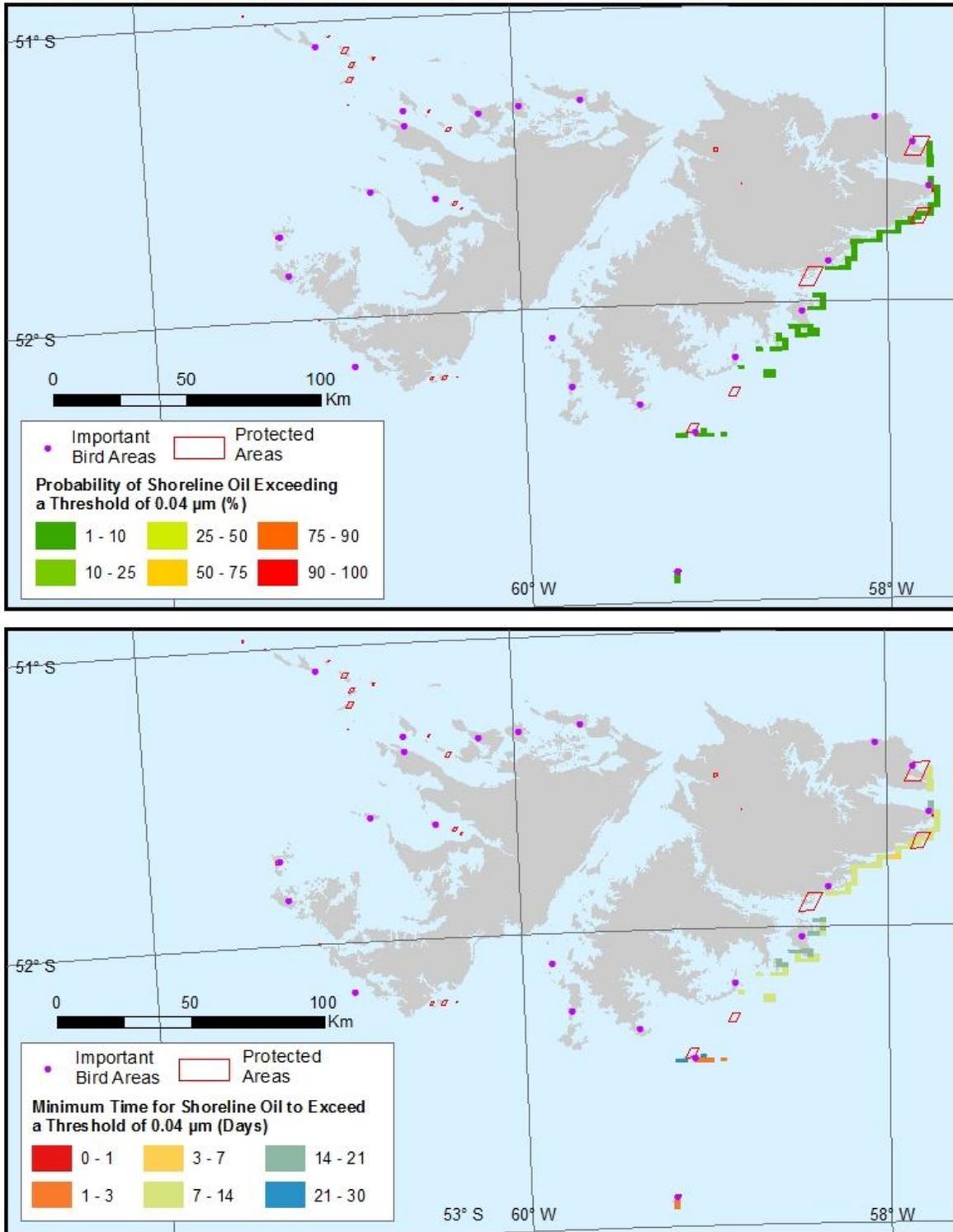


Figure 30. Scenario 7 stochastic maps for potential shoreline contamination.

Stochastic Results: 10-Day Surface Blowout of 500,710 bbl of Crude Oil from FIST13 (Mar-Sep)

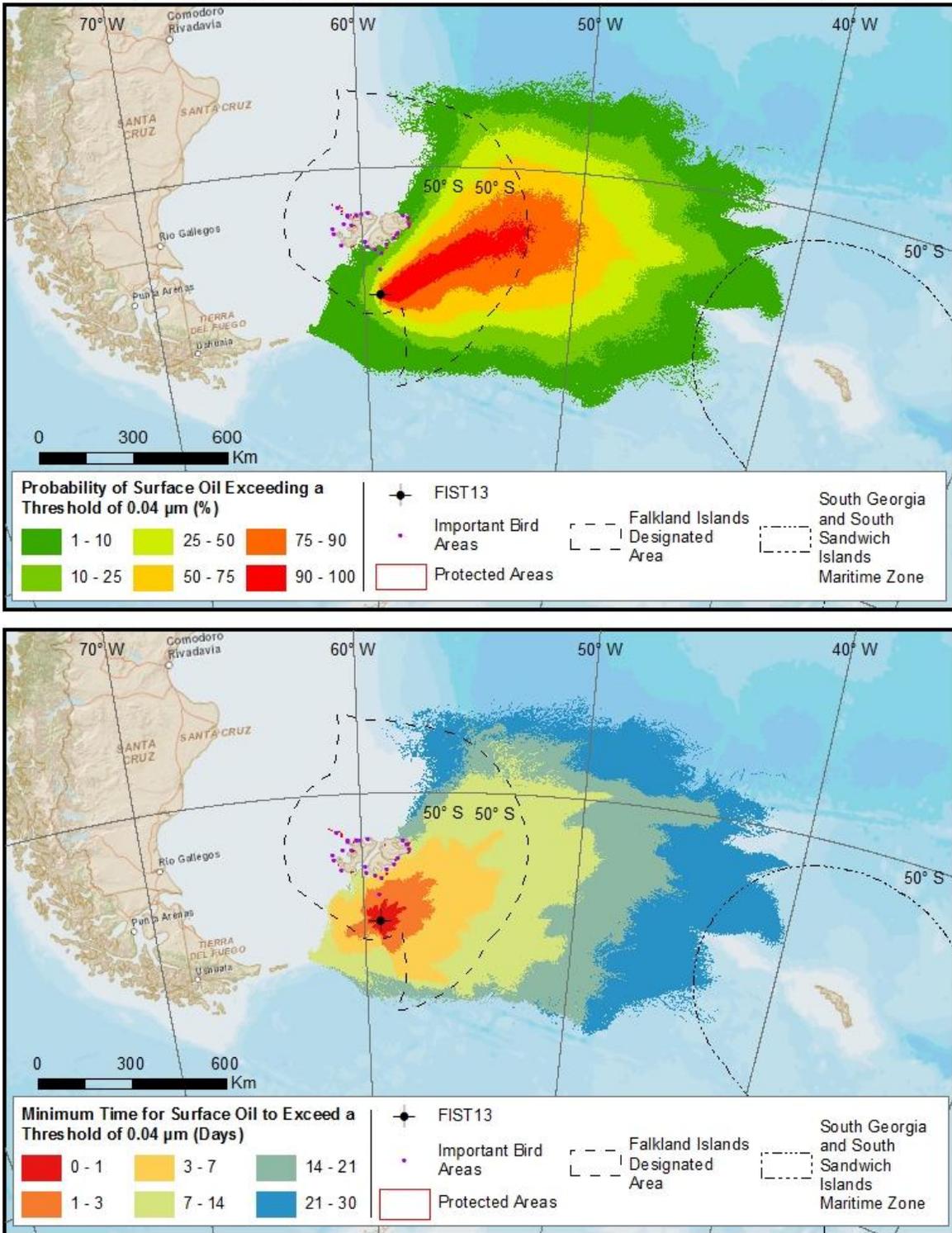


Figure 31. Scenario 8 stochastic maps for potential water surface contamination.

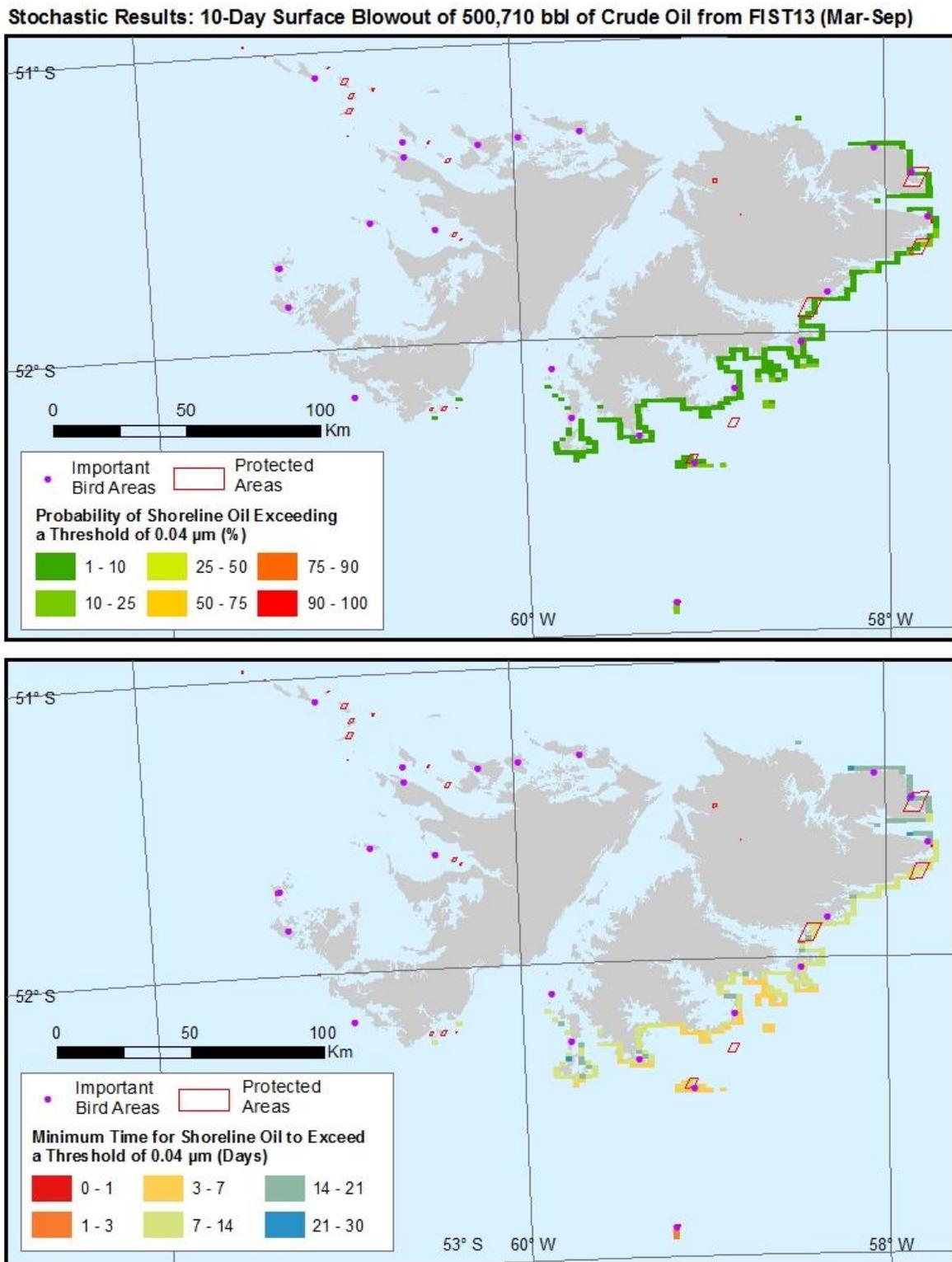


Figure 32. Scenario 8 stochastic maps for potential shoreline contamination.

Stochastic Results: 10-Day Subsurface Blowout of 500,710 bbl of Crude Oil from FISA12 (Mar-Sep)

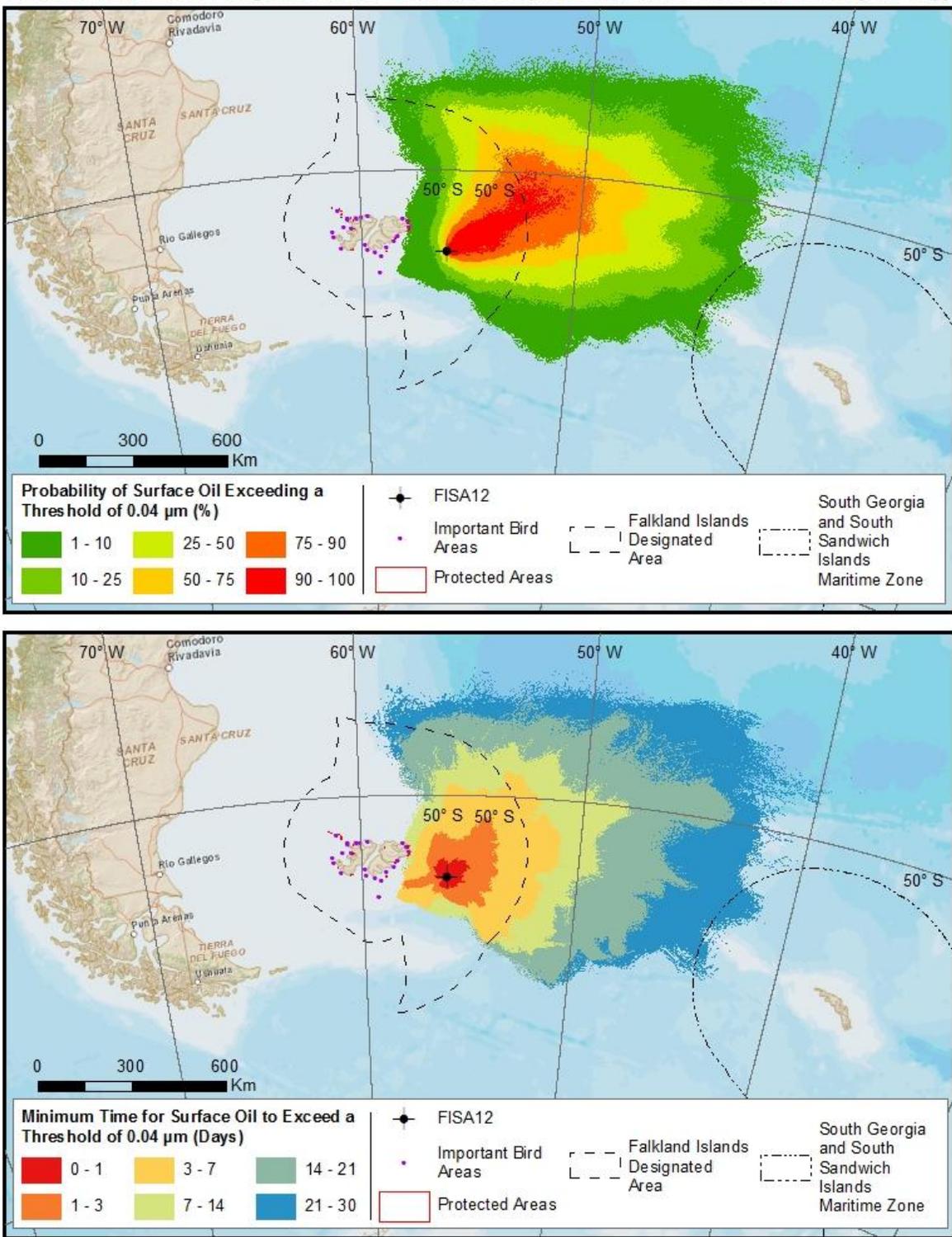


Figure 33. Scenario 9 stochastic maps for potential water surface contamination.

Stochastic Results: 10-Day Subsurface Blowout of 500,710 bbl of Crude Oil from FISA12 (Mar-Sep)

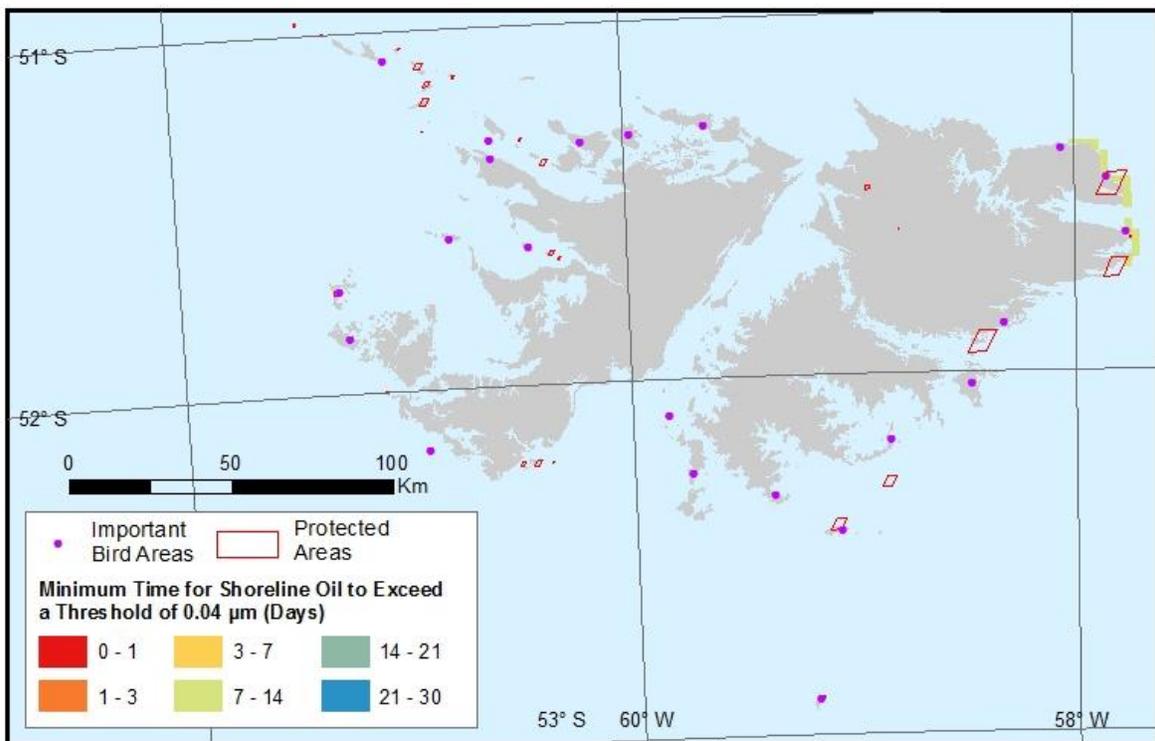
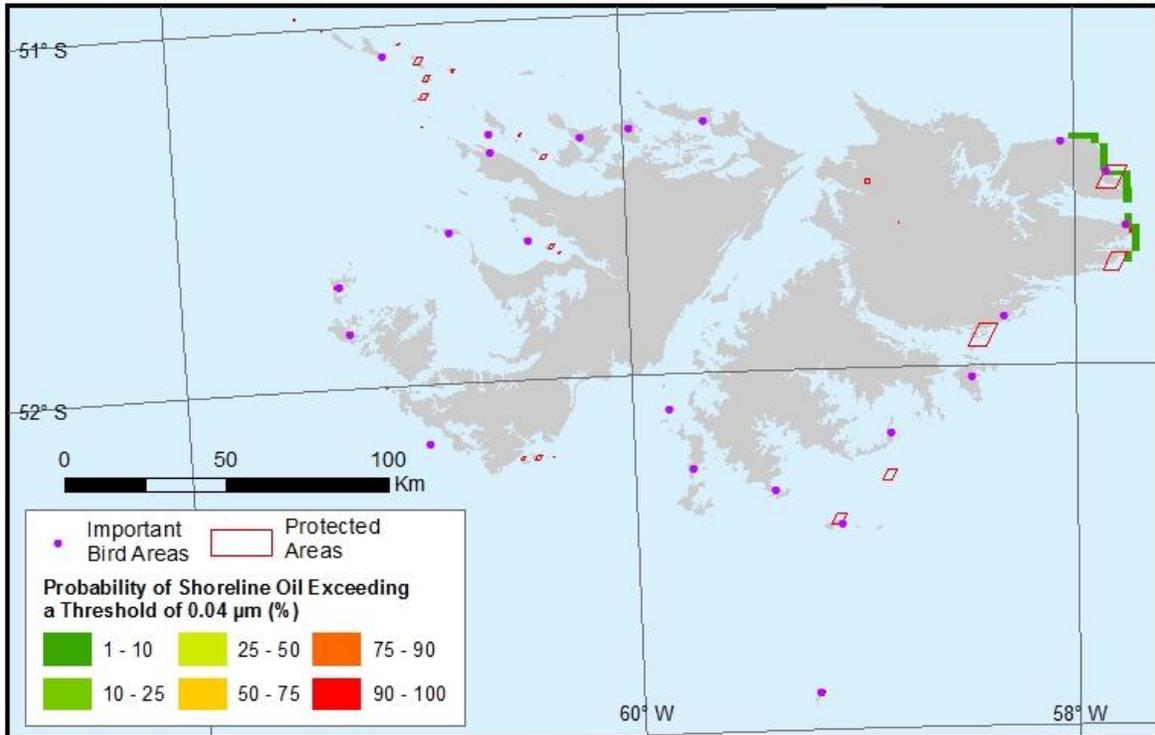


Figure 34. Scenario 9 stochastic maps for potential shoreline contamination.

Stochastic Results: 10-Day Subsurface Blowout of 500,710 bbl of Crude Oil from FIST13 (Mar-Sep)

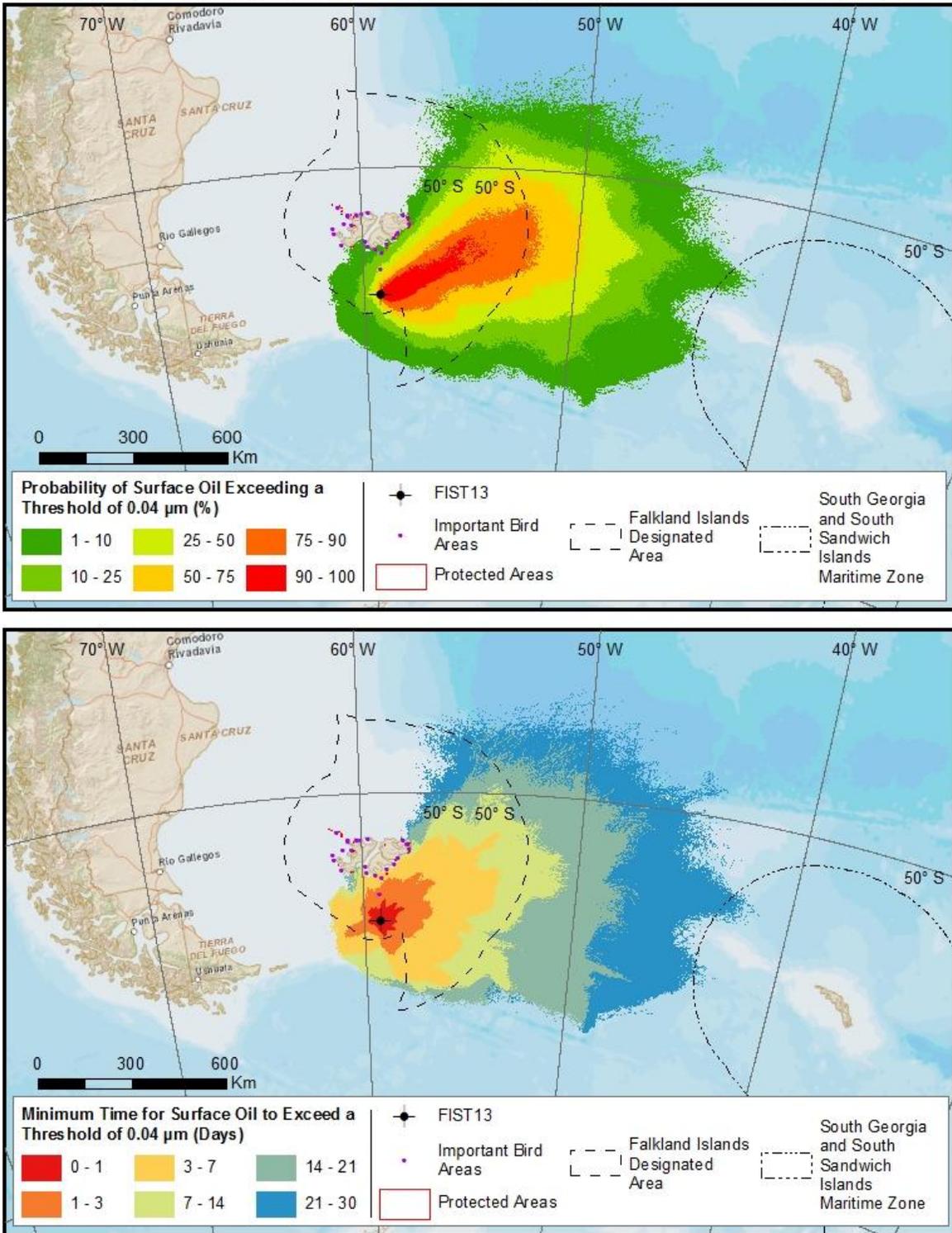


Figure 35. Scenario 10 stochastic maps for potential water surface contamination.

Stochastic Results: 10-Day Subsurface Blowout of 500,710 bbl of Crude Oil from FIST13 (Mar-Sep)

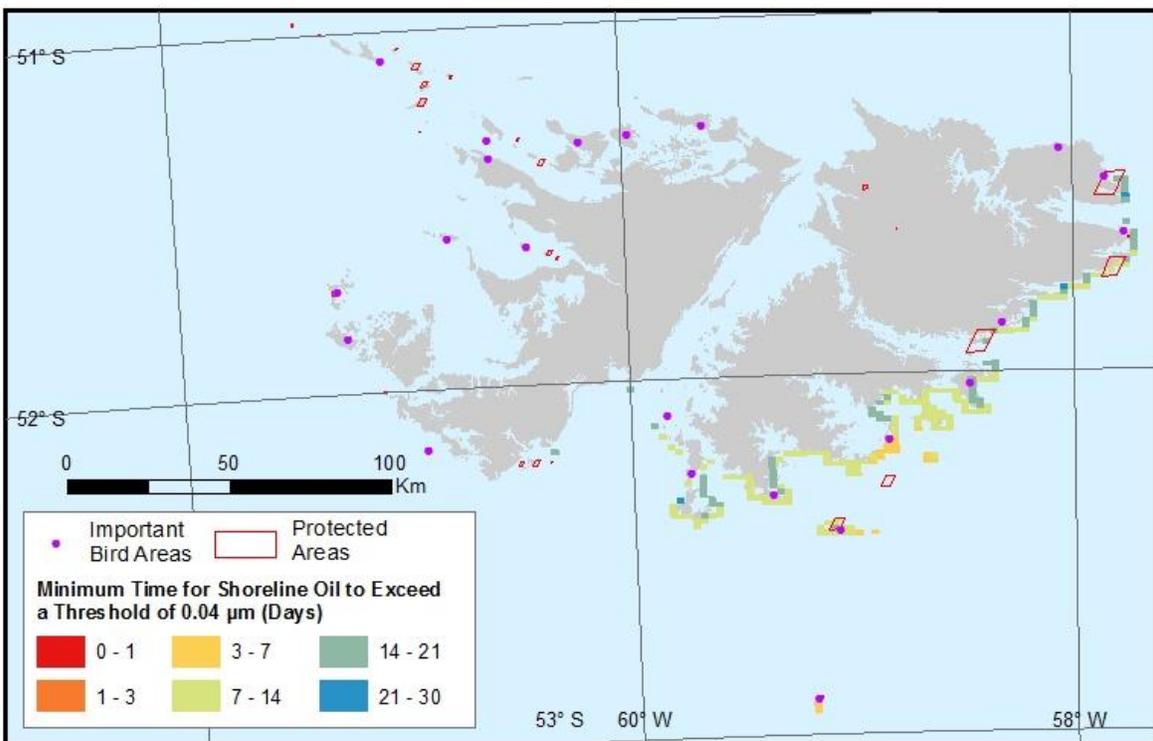
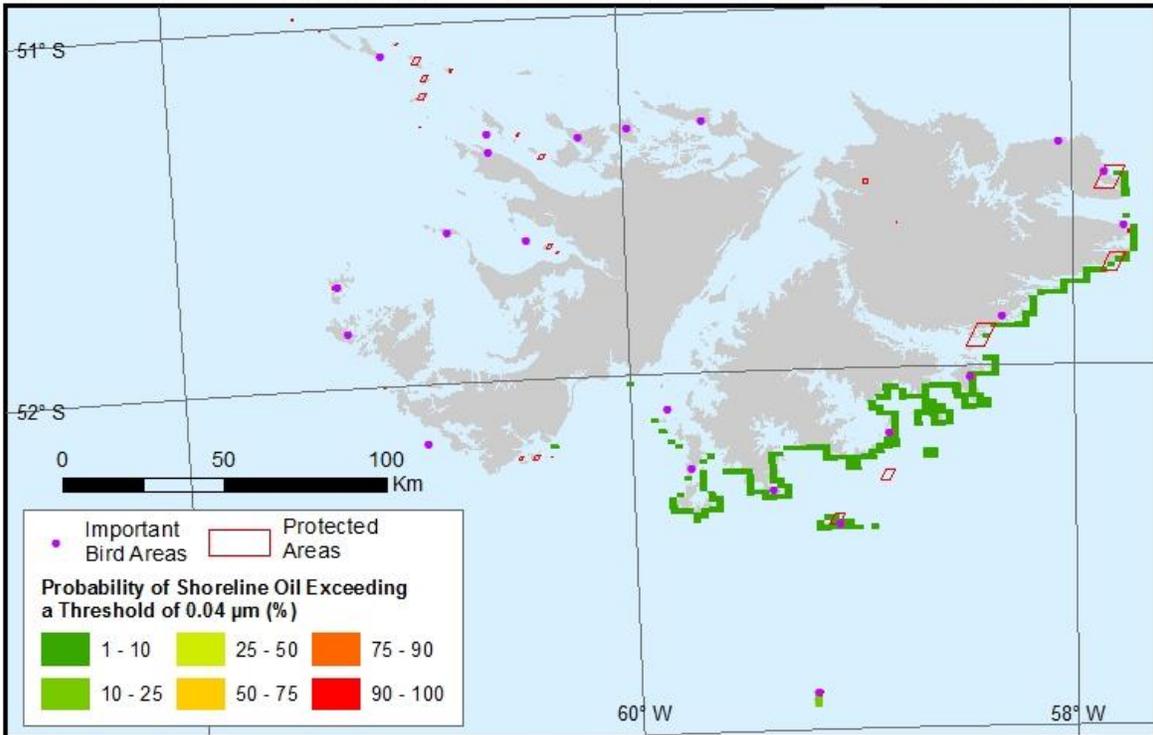


Figure 36. Scenario 10 stochastic maps for potential shoreline contamination.

3.4. Deterministic Modelling Results

For each stochastic surface blowout scenario where there was any shoreline impact, a worst case scenario was selected based on the time it took for oil to arrive at the shore. From each of these stochastic scenarios, the individual trajectory that impacted the coast in the shortest time was chosen for the deterministic modelling scenario. This criterion was chosen to represent the worst case because it would require the quickest response efforts. For the surface blowout scenario where no shoreline oiling was predicted, the trajectory that came closest to impacting the Falkland Islands was chosen as a representative case.

All trajectory/fate simulations were run using the same variable winds and current forcing used for the corresponding stochastic simulation from which it was identified.

Table 7. Selected worst case deterministic runs for each spill scenario.

| ID | Spill Site | Oil Type | Spill Type | Total Spilled Volume (bbl) | Selected Deterministic Case | Run Type |
|----|------------|-----------|-----------------|----------------------------|-----------------------------|----------------|
| 5 | FISA12 | Crude Oil | Surface Blowout | 500,710 | 2/21/2011 | Representative |
| 6 | FISA12 | Crude Oil | Surface Blowout | 500,710 | 8/12/2009 | Worst Case |
| 7 | FIST13 | Crude Oil | Surface Blowout | 500,710 | 2/13/2010 | Worst Case |
| 8 | FIST13 | Crude Oil | Surface Blowout | 500,710 | 3/13/2010 | Worst Case |

Table 8. Predicted shoreline contamination information for the selected deterministic runs.

| ID | Spill Site | Oil Type | Spill Type | Total Spilled Volume (bbl) | Time To Shore (days) | Amount of oil Ashore (bbl) | |
|----|------------|-----------|-----------------|----------------------------|----------------------|----------------------------|-------|
| | | | | | | Peak | End |
| 5 | FISA12 | Crude Oil | Surface Blowout | 500,710 | - | - | - |
| 6 | FISA12 | Crude Oil | Surface Blowout | 500,710 | 10 | 9,646 | 8,012 |
| 7 | FIST13 | Crude Oil | Surface Blowout | 500,710 | 1.6 | 2,309 | 1,803 |
| 8 | FIST13 | Crude Oil | Surface Blowout | 500,710 | 1.7 | 506 | 309 |

The following figures are presented for the deterministic modelling results:

1. **Maximum mass of floating oil per unit area (~ oil slick thickness):** The map depicts the maximum mass per unit area of oil on the water surface that passed by a given area at some point during the simulation.
2. **Maximum mass of shoreline oil per unit area (for trajectories that impacted the coast):** The map depicts the maximum mass per unit area of oil that beached on a given area of the shoreline at some point during the simulation. In the absence of data, all shoreline segments are assumed to be 10-m-wide sandy beaches. Using actual shoreline data may alter the results, but the results provided are likely on the conservative side for shoreline impacts.
3. **Predicted mass balance:** The graph shows the model-predicted mass balance for the spilled oil. The mass balance graph shows the degree of weathering that the oil undergoes during the period of the simulation.

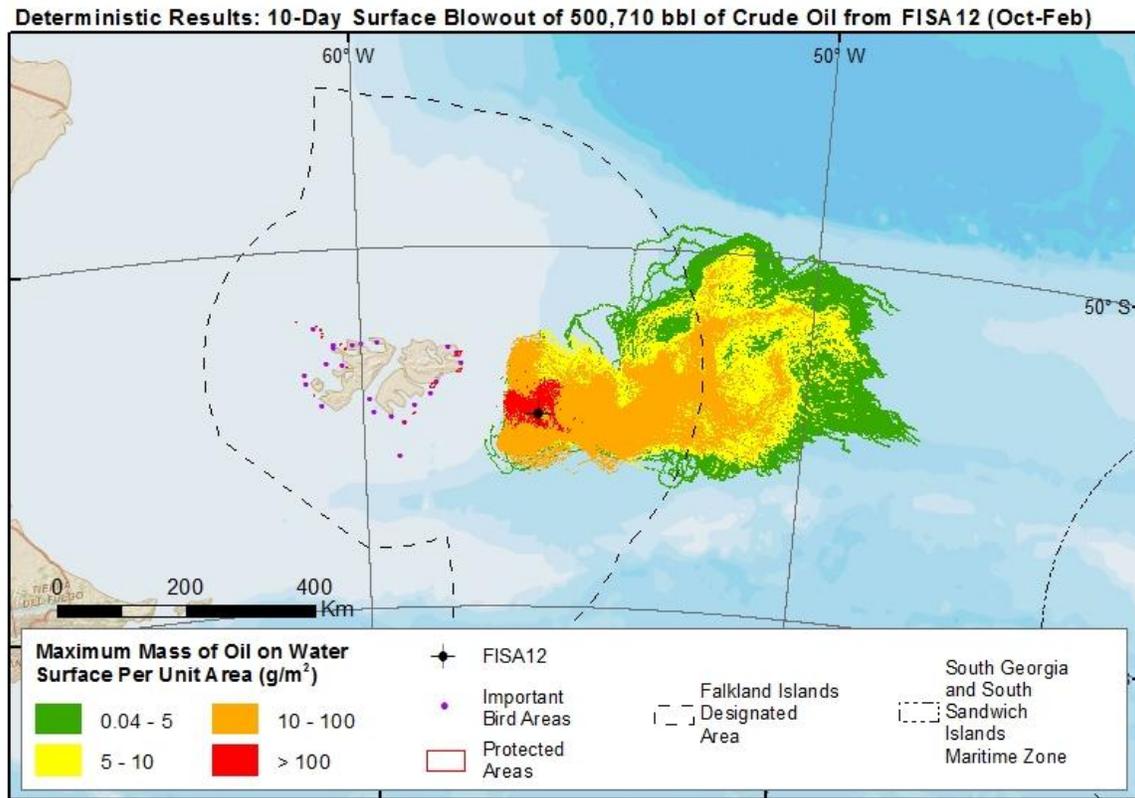


Figure 37. Scenario 5 deterministic map for potential water surface contamination.

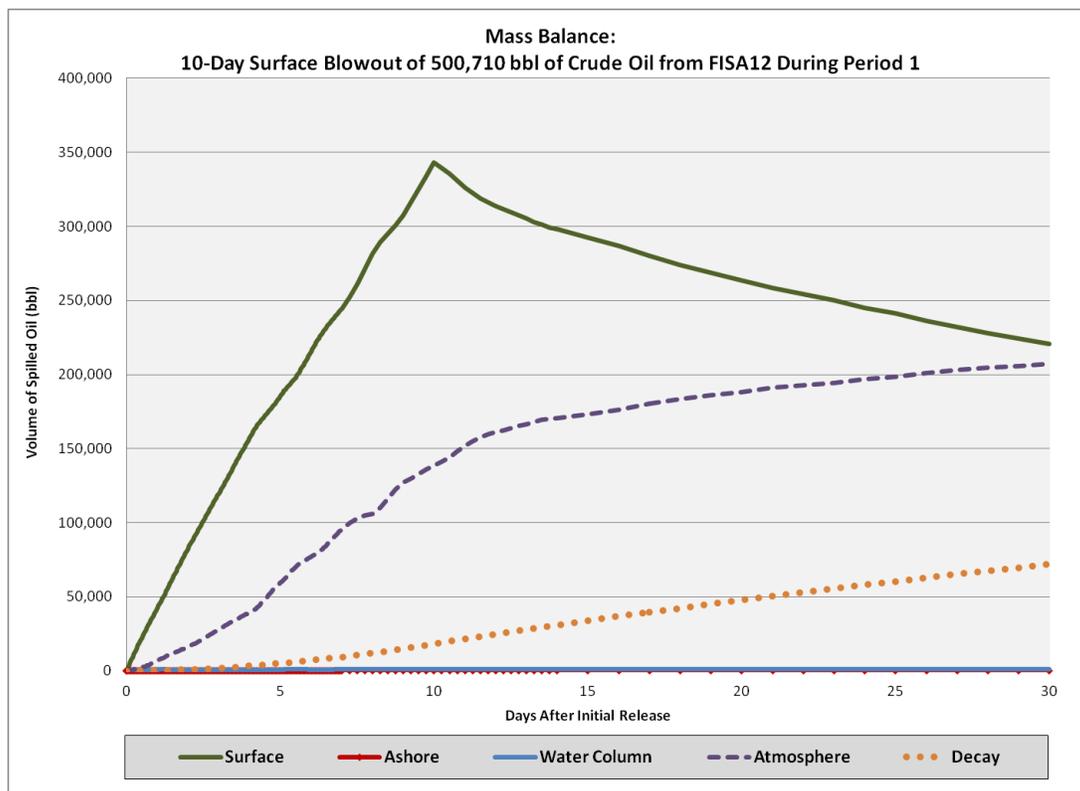


Figure 38. Scenario 5 mass balance graph.

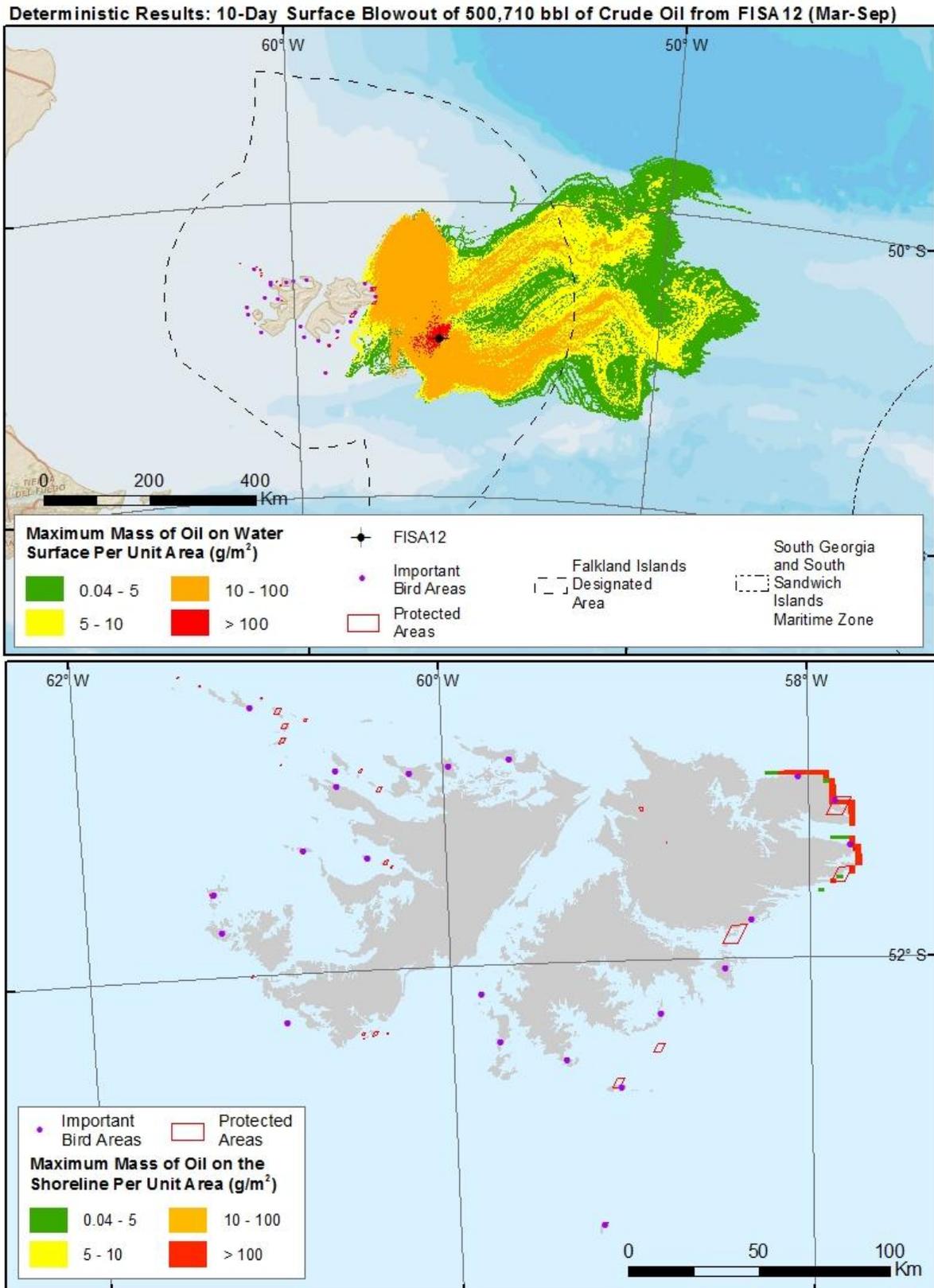


Figure 39. Scenario 6 deterministic maps for potential water surface (top) and shoreline (bottom) contamination.

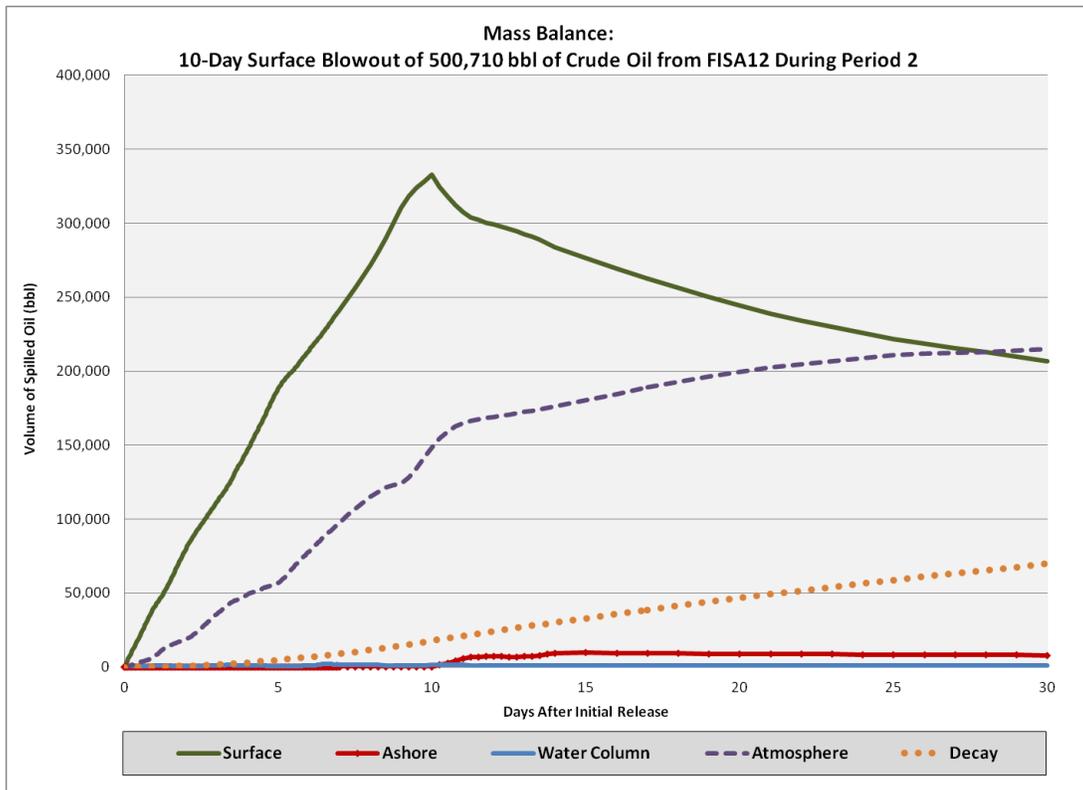


Figure 40. Scenario 6 mass balance graph.

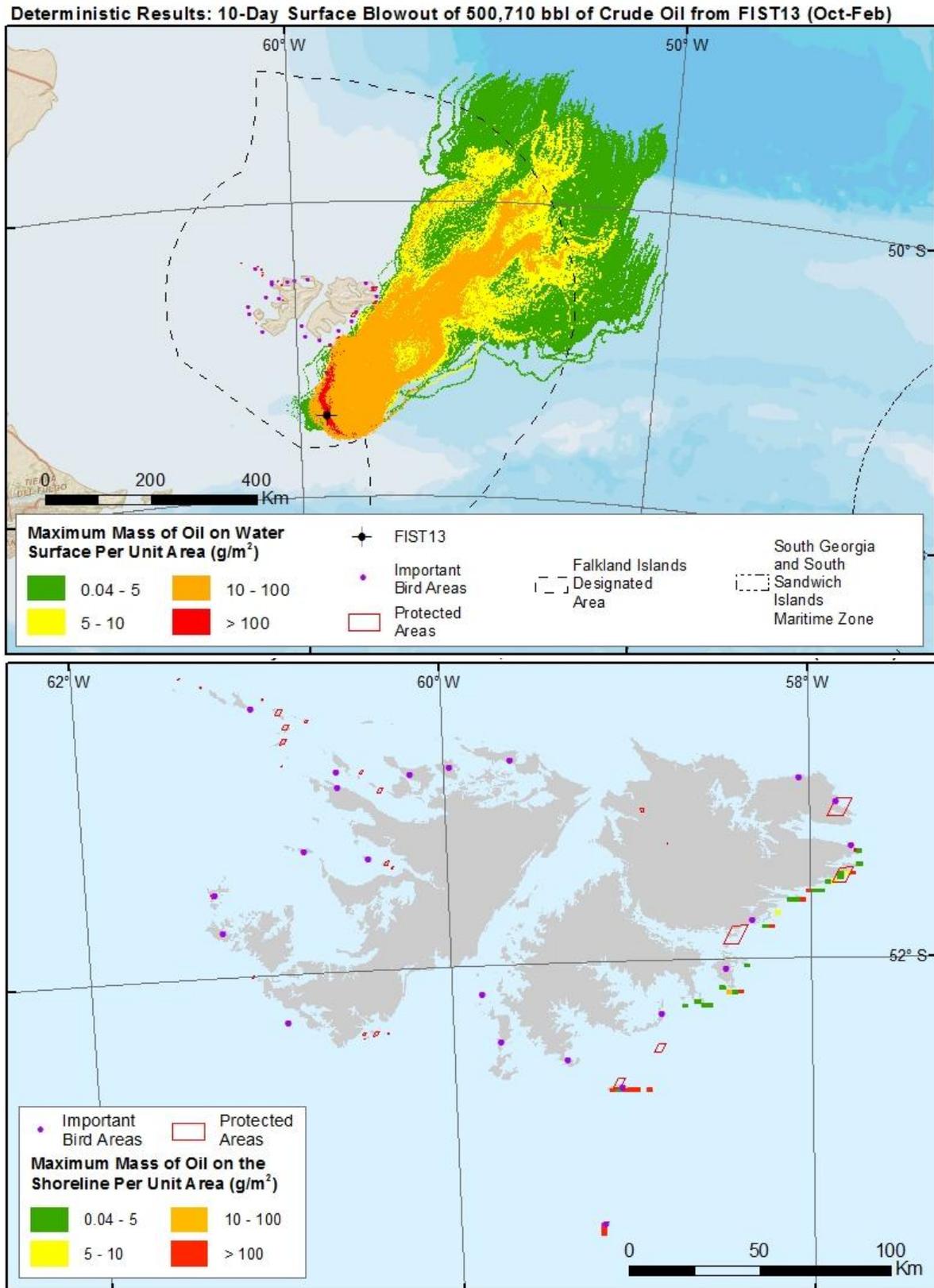


Figure 41. Scenario 7 deterministic maps for potential water surface (top) and shoreline (bottom) contamination.

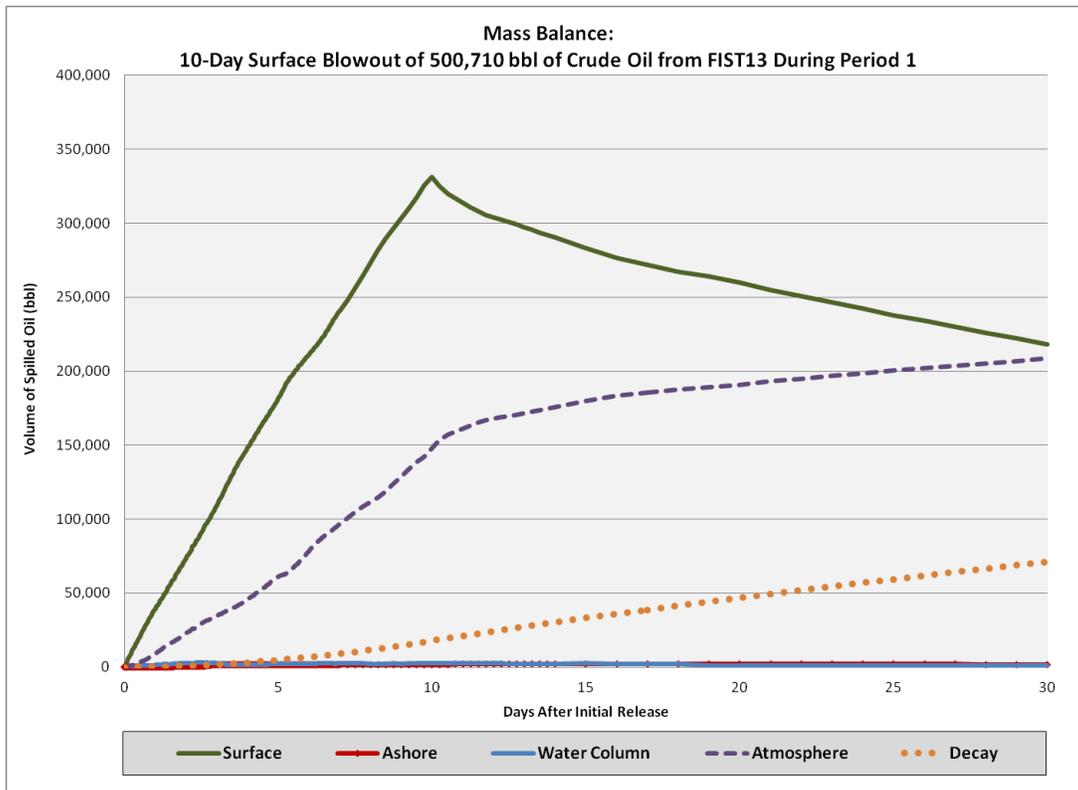


Figure 42. Scenario 7 mass balance graph.

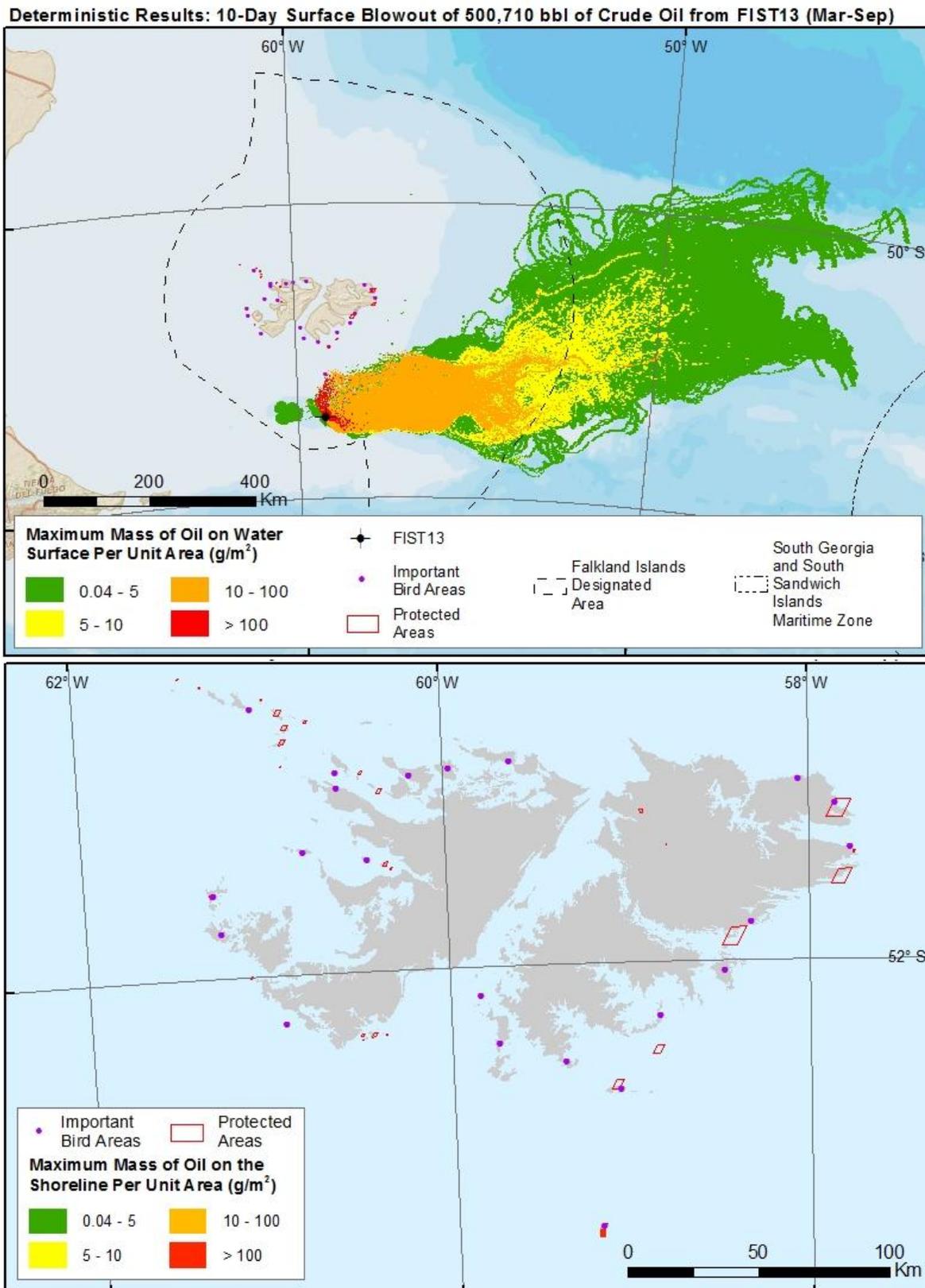


Figure 43. Scenario 8 deterministic maps for potential water surface (top) and shoreline (bottom) contamination.

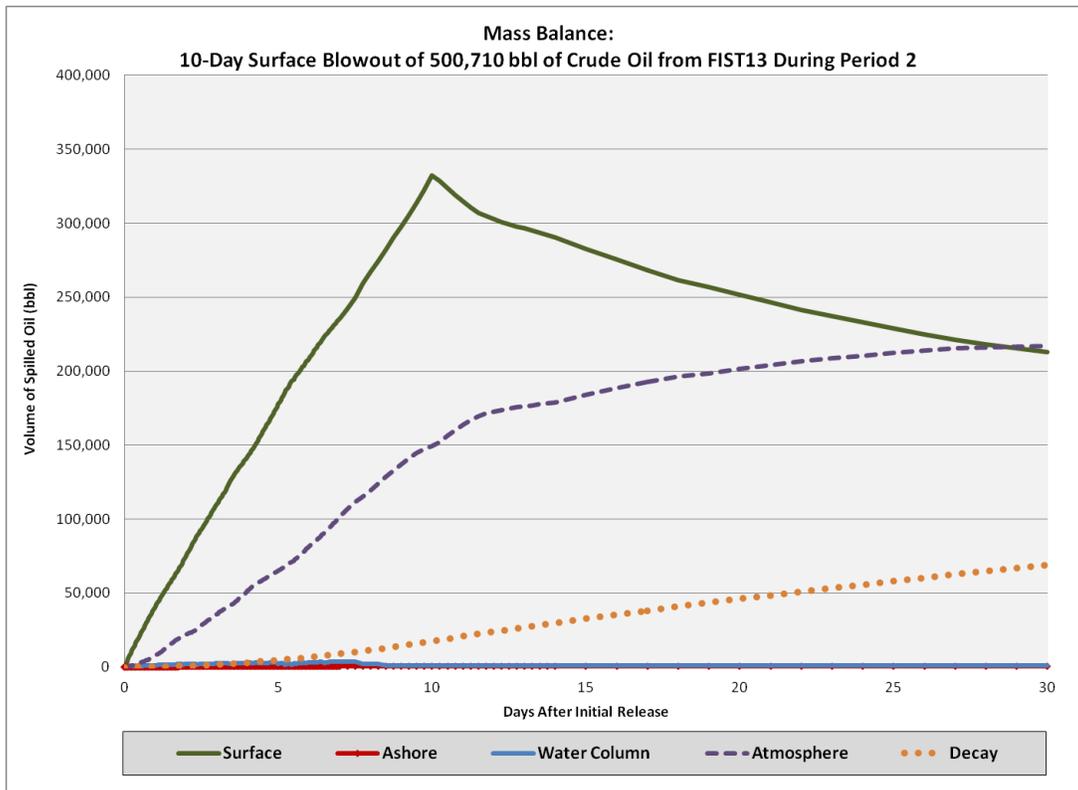


Figure 44. Scenario 8 mass balance graph.

RPS | Falkland Islands. Results of the Oil Spill Modelling DECC Scenarios

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1. Geographic Location - Area of Interest

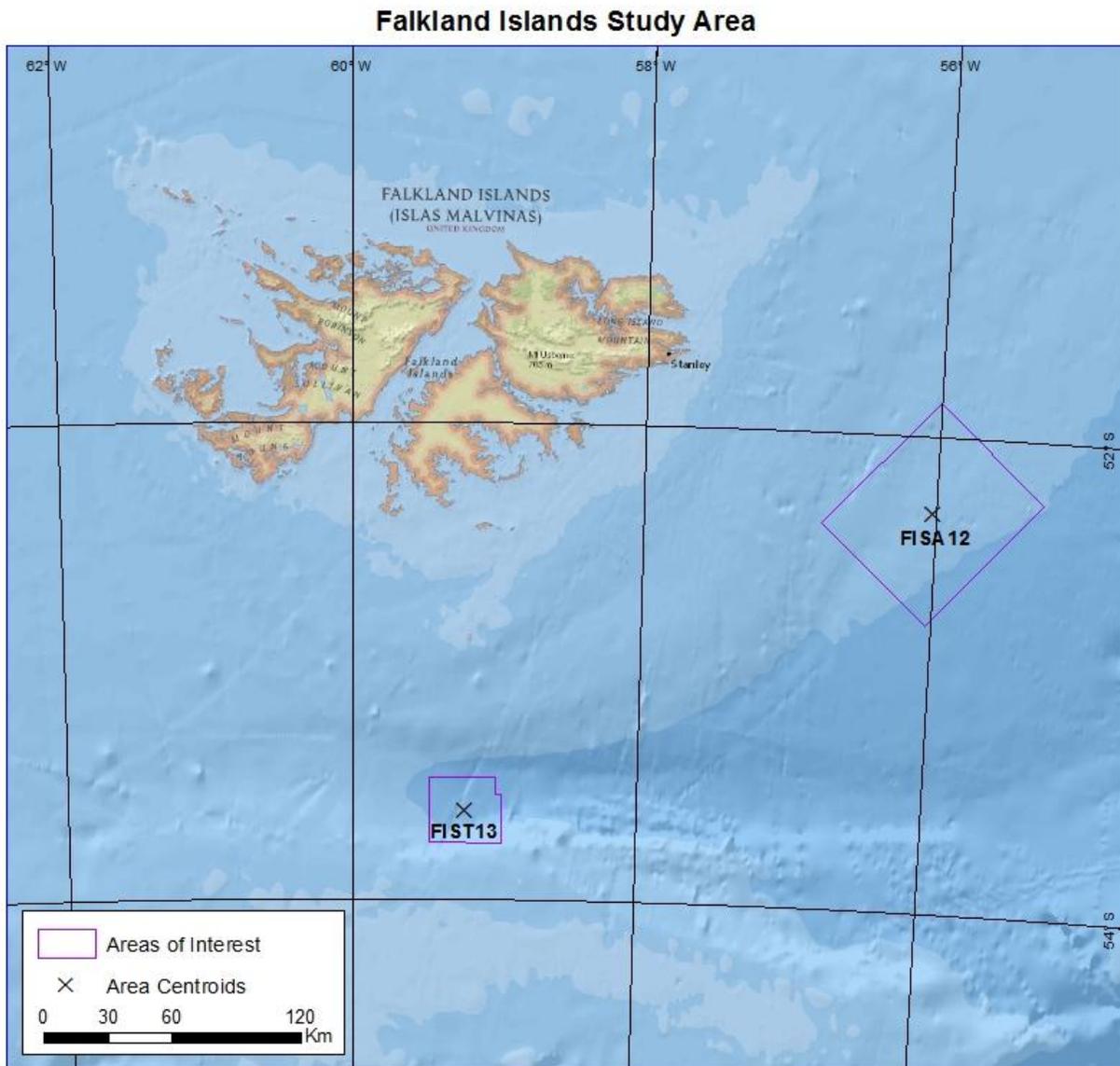


Figure 1. Geographic location of the study area.

Table 1. Coordinates of the sites of interest.

| Site Name | Comments | Latitude N | Longitude E | Distance from land (km) | Approximate Water Depth (m) |
|-----------|------------------|------------|-------------|-------------------------|-----------------------------|
| FISA12 | Center of FISA12 | -52.3232 | -56.0231 | 137 (Cape Pembroke) | 1,177 |
| FIST13 | Center of FIST13 | -53.6283 | -59.2109 | 76 (Beauchene Island) | 1,527 |

2. Oil Spill Simulations – Preliminary Results

2.1. Modelling Inputs

Each surface blowout scenario has been evaluated to fulfil what is currently referred to as “worst case” conditions under DECC guidelines. These worst case conditions require a constant 30-knot wind speed to be used in both the direction of the closest shoreline as well as the direction of the closest median line. These conditions are expected to provide information on necessary response measures considering the time to shore as well as the time to reach foreign waters.

Table 2. Summary of oil properties used in the simulations.

| Oil Type | API Gravity (°) | Viscosity (cP at 25°C) | Interface Tension (dyne/cm) | Emulsion maximum Water Content (%) |
|-------------|-----------------|------------------------|-----------------------------|------------------------------------|
| Light Crude | 38.9 | 4.00 | 26.1 | 89.6 |

Table 3. Parameters of the oil spill scenarios.

| ID | Spill Site | Spill Event | Wind Direction | Oil Type | Spill Rate (bbl/d) | Spill Duration (days) | Total Spilled Volume (bbl) | Simulation Duration (days) |
|--|------------|--|----------------|-----------|--------------------|-----------------------|----------------------------|----------------------------|
| Deterministic Scenarios (Current DECC Scenarios) | | | | | | | | |
| 1 | FISA12 | 30 knot onshore wind speed (surface) | From ESE | Crude Oil | 50,071 | 10 | 500,710 | 30 |
| 2 | FISA12 | 30 knot wind speed towards median line (surface) | From ENE | Crude Oil | 50,071 | 10 | 500,710 | 30 |
| 3 | FIST13 | 30 knot onshore wind speed (surface) | From S | Crude Oil | 50,071 | 10 | 500,710 | 30 |
| 4 | FIST13 | 30 knot wind speed towards median line (surface) | From ENE | Crude Oil | 50,071 | 10 | 500,710 | 30 |

2.2. Deterministic Modelling Results

For each scenario, 6 maps are provided with snapshots of the mass per unit area of oil on the water surface at specific times after the release. Due to the uniform wind direction, oil slowly beaches in the same location until the holding capacity of the shoreline is met. After that point, additional oil collects on the water surface against the shoreline. This excess oil is referred to as “shoreline overflow” in Table 5. Under normal environmental conditions, the wind and current variability may result in more extensive transport along the shore, thereby increasing the amount of shoreline oil and decreasing shoreline overflow. Note that volumes listed in Table 5 are specific to artificial parameters (30 knot onshore winds) and in reality, the natural variability of environmental conditions in the area would likely result in very different volumes.

Table 4. Predicted impact information for each deterministic run.

| ID | Spill Site | Wind Direction | Total Spilled Volume (bbl) | Time To Shore (days) | Time To Median Line (days) | Countries Impacted | |
|----|------------|------------------------|----------------------------|----------------------|----------------------------|-----------------------------|------------------|
| | | | | | | Water Surface | Shore |
| 1 | FISA12 | Onshore (from ESE) | 500,710 | 3.0 | - | Falkland Islands | Falkland Islands |
| 2 | FISA12 | Median Line (from ENE) | 500,710 | 16 | 8.3 | Falkland Islands; Argentina | Argentina |
| 3 | FIST13 | Onshore (from S) | 500,710 | 1.7 | - | Falkland Islands | Falkland Islands |
| 4 | FIST13 | Median Line (from ENE) | 500,710 | 7.8 | 2.0 | Falkland Islands; Argentina | Argentina |

Table 5. Predicted volumes ashore (including shoreline overflow) for 6 time steps.

| Scenario | Approximate Volume Ashore + Shoreline Overflow (bbl) | | | | | |
|----------|--|--------|---------|---------|---------|---------|
| | Day 1 | Day 3 | Day 7 | Day 10 | Day 20 | Day 30 |
| 1 | - | - | 110,257 | 190,604 | 235,070 | 206,992 |
| 2 | - | - | - | - | 102,500 | 205,700 |
| 3 | - | 14,670 | 115,586 | 188,413 | 221,630 | 196,356 |
| 4 | - | - | - | 30,719 | 231,400 | 207,000 |

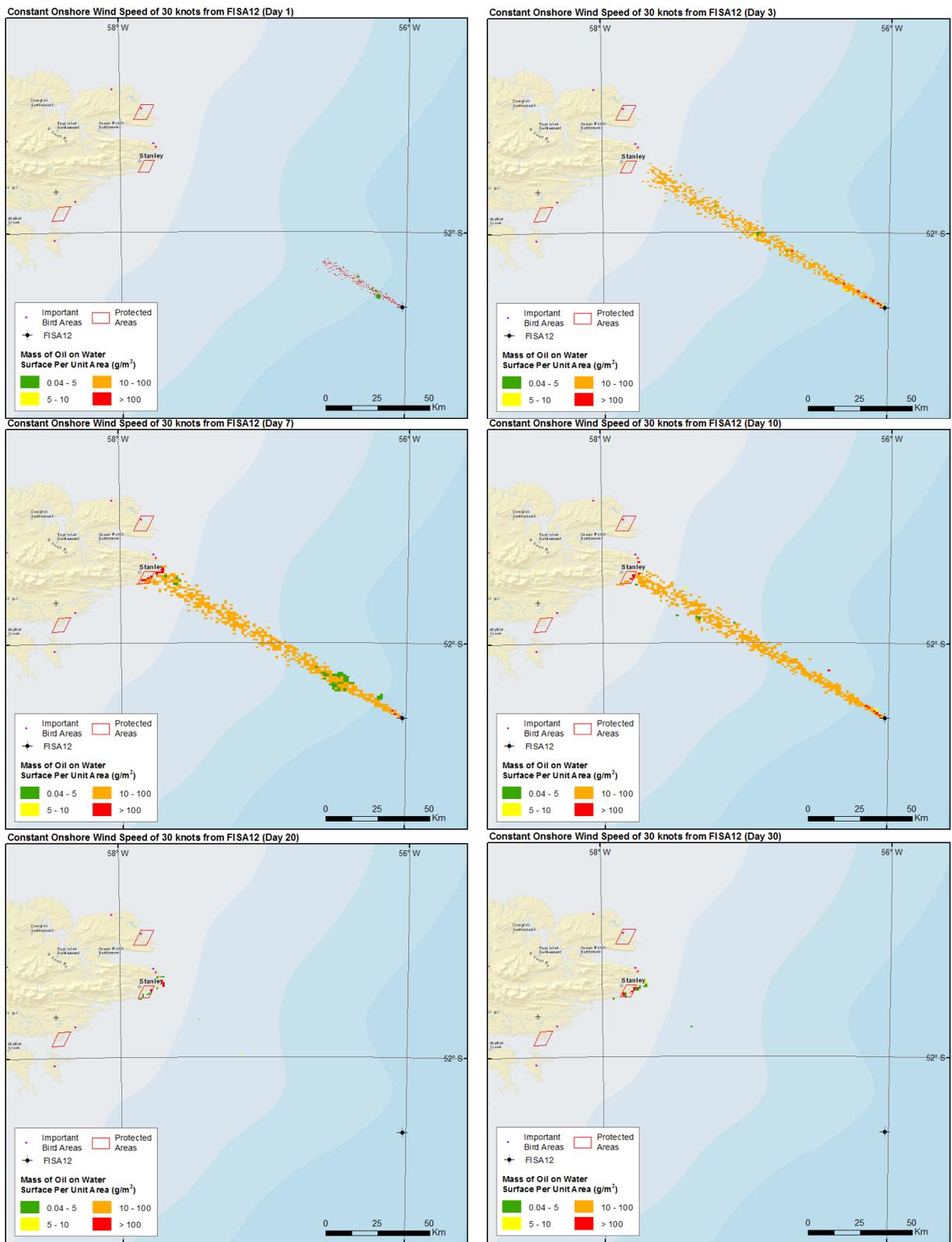


Figure 2. DECC1 deterministic snapshots.

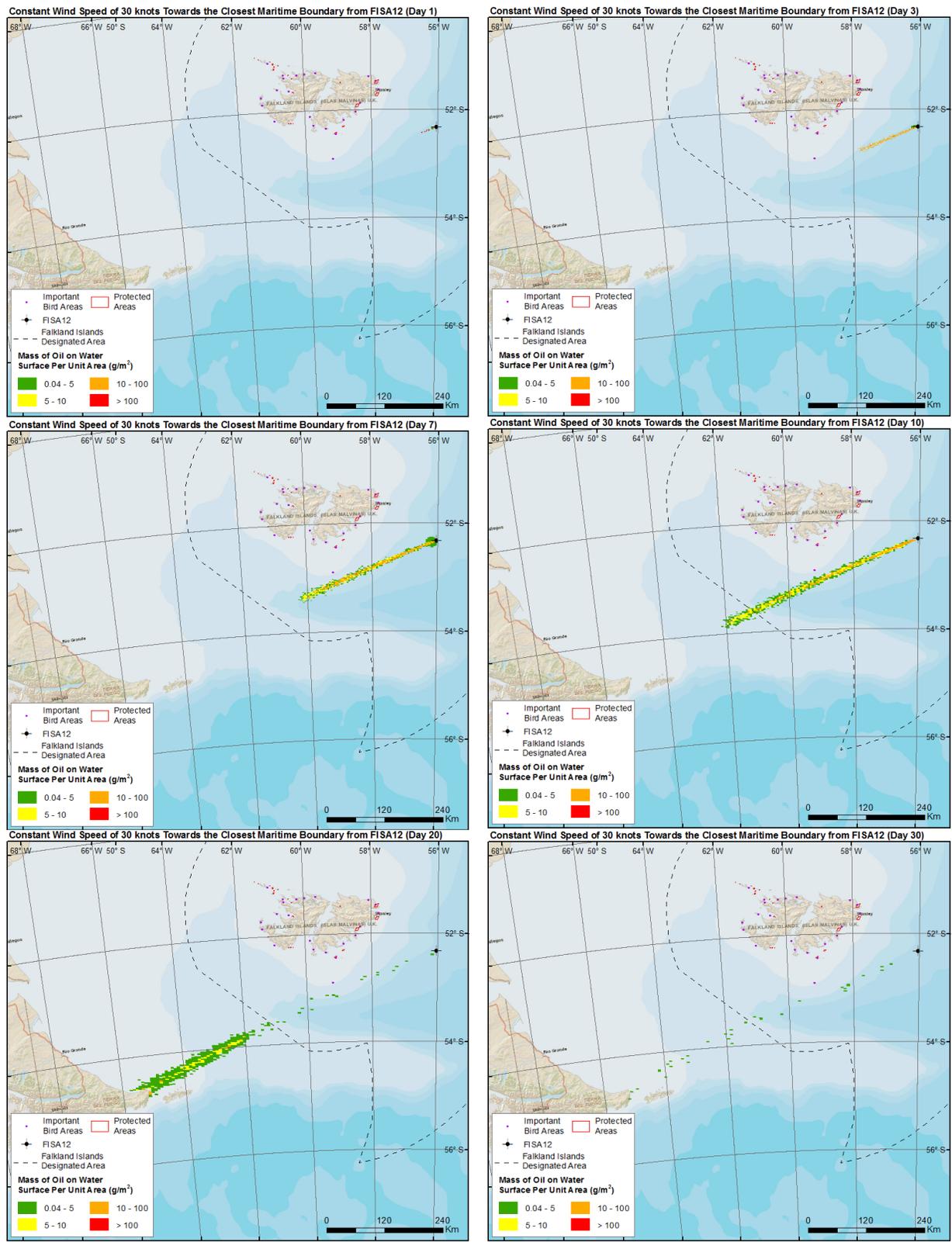


Figure 3. DECC2 deterministic snapshots.

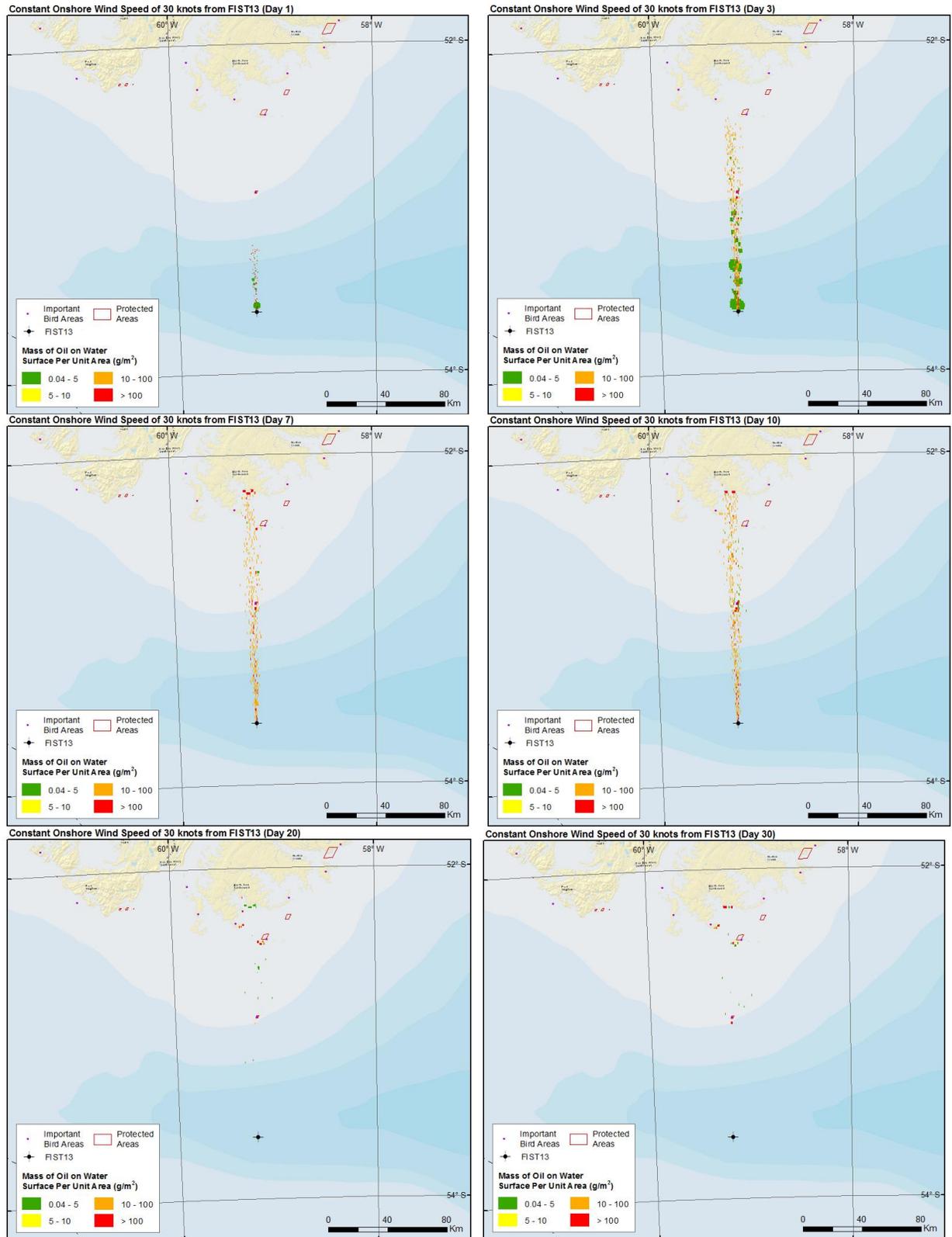


Figure 4. DECC3 deterministic snapshots.

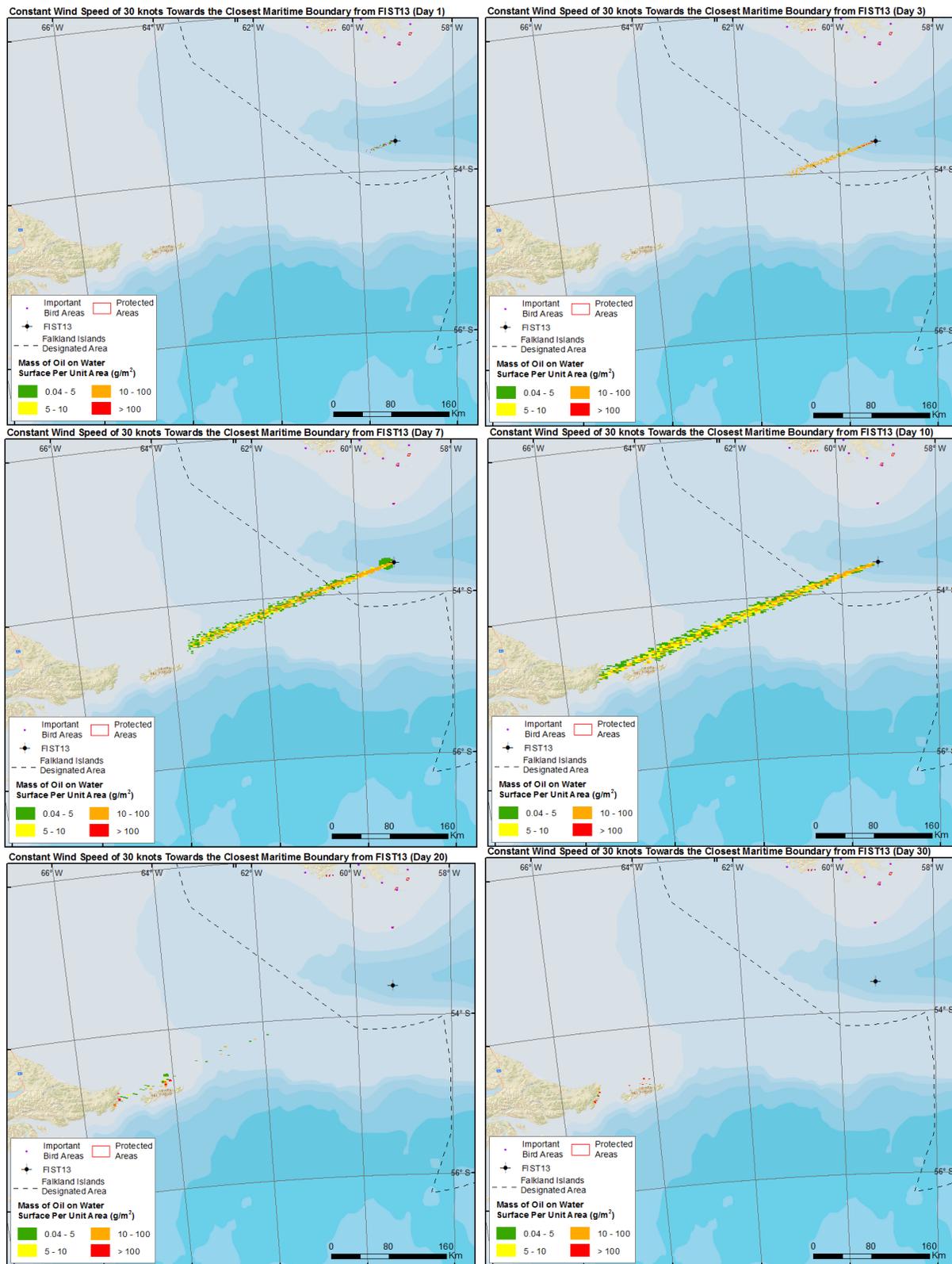


Figure 5. DECC4 deterministic snapshots.

Appendix J: Stakeholder Engagement Plan (SEP)

Noble Energy Falklands Limited

**Stakeholder Engagement Plan: Falkland Islands Exploration Drilling
Environmental, Social and Health Impact Assessment (ESHIA)**

Date: May 2014

Revision: 00L



Noble Energy Falklands Limited

Stakeholder Engagement Plan: Falkland Islands Exploration Drilling Environmental, Social and Health Impact Assessment (ESHIA)

| DATE | VERSION | DESCRIPTION | PREPARED | CHECKED | APPROVED |
|----------|---------|------------------------------------|----------|---------|----------|
| 06.09.13 | 00J | Update to Appendix B | SJS | AJP | |
| 14.01.14 | 00K | Update to Appendix B | SJS | AJP | |
| 08.05.14 | 00L | Update to Appendix B and timeframe | SJS | AJP | |

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|--|---------|--|--|
| NOBLE ENERGY DOCUMENT CONTROL | | | |
| DATE | VERSION | DOCUMENT OWNER | DOCUMENT APPROVER |
| 08.05.14 | 00L | EHSR Coordinator – International Frontier Ventures | EHSR Manager – International Frontier Ventures |
| | | | |
| Document Number: 178-13-EHSR-SEP-PA-T4 | | | |

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Abbreviations

| Abbreviation | Description |
|----------------|--|
| CBO | Community Based Organisation |
| DECC | Department of Energy and Climate Change |
| EHS Guidelines | Environmental, Health and Safety Guidelines |
| EIA | Environmental Impact Assessment |
| EIS | Environmental Impact Statement |
| EP | Equator Principles |
| ESHIA | Environmental, Social and Health Impact Assessment |
| ESMP | Environmental and Social Management Plan (ESMP) |
| Espoo | UNECE Convention on Environmental Impact Assessment in a Trans-boundary Context (Espoo Convention) |
| ExCo | Executive Council |
| FCZ | Falklands Conservation Zone |
| FGD | Focus Group Discussion |
| FI | Falkland Islands |
| FIG | Falkland Islands Government |
| FIFCA | Falkland Islands Fishing Companies Association |
| FIGAS | Falkland Islands Government Air Service |
| FOGL | Falkland Oil and Gas Company |
| ICP | Informed Consultation and Participation |
| IFC | International Finance Corporation |
| IFC PS | IFC Performance Standards |
| JNCC | Joint Nature Conservation Committee |
| NGO | Non-Governmental Organisation |
| NOSCP | National Oil Spill Contingency Plan |
| NTS | Non-Technical Summary |
| SAERI | South Atlantic Environmental Research Institute |
| SEP | Stakeholder Engagement Plan |
| SMSG | Shallow Marine Surveys Group |
| TOR | Terms of Reference |

Definitions

| Terms | Description |
|-------------------------------------|---|
| Comment Form | A standard 'hard copy' form in which stakeholders can submit written comments, views and opinions. Often distributed at public meetings and made available in locations accessible to stakeholders. |
| Culturally appropriate | An engagement process that identifies a practical and appropriate approach for sharing information and comments/views/opinions that is compatible with local cultural norms and behaviour. |
| Espoo Convention | UNECE Convention on Environmental Impact Assessment in a Transboundary Context (named after the town of Espoo in Finland where the meeting that produced the Convention was held). |
| Engagement | Generic term for activity including both two-way disclosure of information/consultations and feedback to stakeholders on the way in which their inputs to an ESHIA process were used. |
| Formal meeting | Face-to-face meeting with a stakeholder (or stakeholders) organized in advance, by letter or e-mail, often with an agreed agenda and held on the premises of a stakeholder. Refer to 'Informal' meeting' below. |
| Grievance | A complaint from an individual, or group, who raise a concern about any aspect of the project, or the way in which stakeholder engagement activities are being implemented. |
| Grievance Procedure | A description of the process used to record, track and respond to a grievance, after it has been raised, until the grievance is resolved. |
| Informal meeting | Face-to-face meeting with a stakeholder (or stakeholders) organized in advance, by letter, email or phone, to discuss project-related issues. Can occur in any mutually agreeable location(s). |
| Disadvantaged and vulnerable groups | Individuals and groups that may be directly and differentially or disproportionately affected by the project because of their disadvantaged or vulnerable status. This disadvantaged or vulnerable status may stem from an individual's or group's race, colour, gender, language, religion, political or other opinion, national or social origin, property, birth, or other status. In addition, other factors may be relevant, such as age, ethnicity, culture, literacy, sickness, physical or mental disability, poverty or economic disadvantage, and dependence on unique natural resources. |
| Mitigation | Specific measures developed through the ESHIA process to, prevent, avoid and reduce adverse impacts to a level considered to be as low as reasonably practical. Can also include measures to enhance positive outcomes. |
| Project leaflet | Brief publication containing information about a project which is made available to stakeholders. |
| Public disclosure | Disclosure of project or ESHIA-related information to stakeholders undertaken during a wider consultation process. |
| Public meeting | A meeting held in a suitable location which is open to any individual. |
| Stakeholders | Stakeholders are persons or groups who are directly or indirectly affected by a project, as well as those who may have interests in a project and/or the ability to influence its outcome, either positively or negatively. |
| Stakeholder Engagement | A process of sharing information, ideas and concerns in a two-way dialogue between project proponent and stakeholders, allowing stakeholders to express their views and opinions, and for these views and opinions to be considered in the 'project' design and the project 'approval' decision-making processes. |
| Stakeholder Identification | A process of identifying who is likely to be affected by the project both directly and indirectly, and who may have an interest in the project or influence over the project. |
| Stakeholder Mapping | Process of identifying stakeholders for the purpose of an ESHIA project. |

1 Introduction

This document is the Stakeholder Engagement Plan (SEP) that has been prepared for the Environmental, Social and Health Impact Assessment (ESHIA) of the proposed exploration drilling and seismic survey operations in the Diomedia and Tilted Fault Block areas of the southern licence region (the project), offshore the Falkland Islands. The project is being led by Noble Energy Falklands Limited (hereafter referred to as Noble). The SEP presents and describes the stakeholder engagement activities (primarily information acquisition, disclosure and consultations, and disclosure and dissemination of key issues/results) that will take place during key stages of the ESHIA.

This document is a live document and will be updated throughout the project as necessary.

1.1 Objectives of the Stakeholder Engagement Plan

The objectives of this Stakeholder Engagement Plan (SEP) are to:

- Describe the regulatory, lender, company (i.e. Noble) and/or other requirements for consultation and disclosure;
- Identify and prioritise key stakeholder groups, focusing on Affected Communities;
- Describe the strategy and present the timetable for sharing information and consulting with each of these groups;
- Describe the internal resources and individual responsibilities assigned to implement stakeholder engagement activities; and
- Describe how the effectiveness of the SEP will be monitored and how lessons learned will be recorded, with the aim of improving stakeholder engagement activities during the ESHIA and during future implementation of the project itself.

1.2 Project Description

1.2.1 Background

Noble Energy Inc. is a US-based company with a headquarters in Houston, Texas, which is engaged in global oil and gas exploration and production operations.

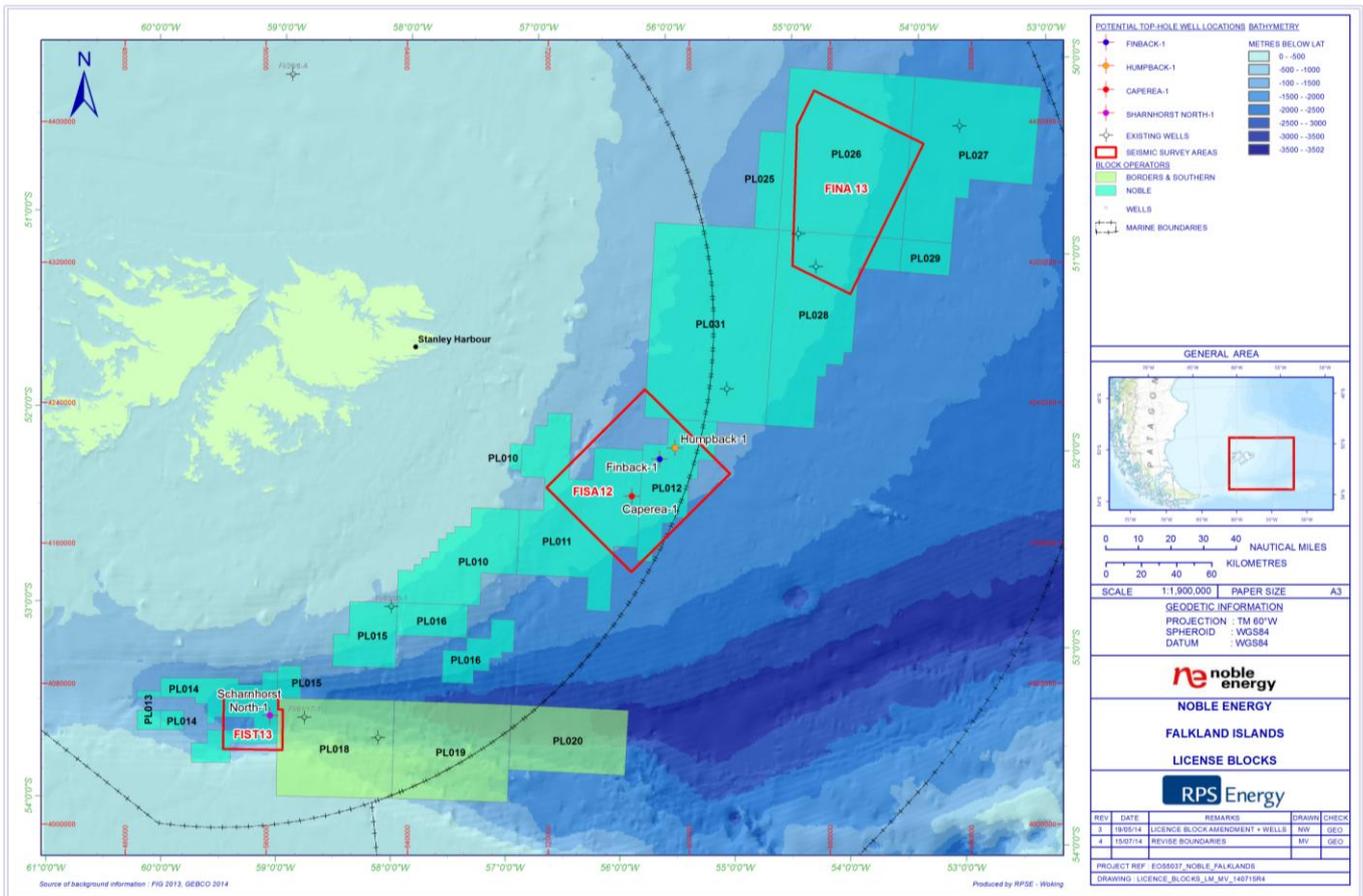
In 2012, Noble acquired licence interests in the southern and northern licence areas offshore the Falkland Islands. As of 2014, Noble currently holds operatorship of these licenses. Figure 1.1 shows the location of the Noble operated licenses, and the areas that have been subject to seismic survey.

1.2.2 Proposed Southern Licence Area Exploration

The planned drilling programme consists of two exploration wells and one optional exploration/appraisal well (a potential total of three wells), all to be located within the southern area licenses.

In accordance with the Falkland Islands regulatory regime, an Environmental Impact Assessment (EIA) process is required for the project leading to the preparation of an Environmental Impact Statement (EIS). Noble has decided to prepare an Environmental, Social and Health Impact Assessment (ESHIA), which meets the requirements outlined in the *Offshore Minerals Ordinance 1994*, Noble internal policies and guidelines, and the requirements of the Equator Principles (EQ) and International Finance Corporation (IFC) Sustainability Framework (2012) which includes the IFC Performance Standards (PS) and accompanying Environmental, Health, and Safety (EHS) Guidelines (2007).

Figure 1.1: Noble Falkland Islands licence areas



1.3 Document Structure

This SEP contains the following sections:

- Section 2:** Regulations and requirements;
- Section 3:** Summary of previous stakeholder engagement activities;
- Section 4:** Stakeholder identification;
- Section 5:** Stakeholder engagement programme;
- Section 6:** Engagement timescale;
- Section 7:** Internal and external roles and responsibilities;
- Section 8:** Noble Energy Community Feedback Mechanism; and
- Section 9:** Monitoring and reporting.

2 Regulations & Requirements

This section provides a summary of the applicable regulations and requirements for stakeholder engagement that have contributed to the development of this SEP.

2.1 National Regulatory Requirements

The *Offshore Minerals Ordinance 1994* PART VI 'Miscellaneous and General' Sections 64 to 67, provides the regulatory framework associated with EIAs and EISs in the Falkland Islands.

Section 64 of the *Offshore Minerals Ordinance 1994*, "*Environmental Impact Assessments and Environmental Impact Statements*", provides for whether or not an EIA or EIS will be required. An EIA or EIS may be required if it is considered by the Governor, that the environment may be substantially affected by the activity in question. Paragraph 2 states that:

"(2) The Governor may, if he considers that the environment might be substantially affected were he to grant an application, cause an environmental impact assessment to be prepared and submitted to him by such person or persons as the Governor directs and in relation to the likely adverse and beneficial effects upon the environment if the application were to be granted, and Schedule 4 shall have effect as to the matters to be dealt with by an environmental impact assessment".

Paragraph 1 defines EIA and EIS as follows:

"(1) (c) "environmental impact assessment" means an assessment commissioned by the Governor under subsection (2);

(1) (d) "environmental impact statement" means a statement prepared by or on behalf of the applicant pursuant to a requirement made by the Governor under subsection (3) and dealing with all, or such as the Governor may require of the matters mentioned in paragraph 2 of Schedule 4".

The basic scope and content of an EIA and EIS are specified within Schedule 4 of the *Offshore Minerals Ordinance 1994* and are essentially the same. An EIA commissioned by the Governor does not have to go through a public consultation period, whereas an EIS submitted by an applicant will be required to go through a 42 calendar day public consultation period.

Sections 65 and 66 of the *Offshore Minerals Ordinance 1994* contain general provisions for commissioning by the Governor of an EIA.

Section 67 of the *Offshore Minerals Ordinance 1994* contains general provisions for applicants who have prepared an EIS and the provision of further information:

"(1) The Governor, when dealing with an application in relation to which the applicant has furnished an environmental impact statement (and whether pursuant to a requirement under section 71(3) or otherwise), may require the applicant to provide such further information as the Governor may specify concerning any matter which is required to be, or may be, dealt with in the environmental impact statement.

(2) The Governor may in writing require an applicant to produce such evidence as he may reasonably call for to verify any information in the applicant's environmental impact statement.

(3) For the purposes of section 65(4) and 66(2), further information furnished pursuant to subsection (1) of this section and evidence furnished pursuant to subsection (2) of this section shall be treated as forming part of the applicant's environmental impact statement".

The *Offshore Minerals Ordinance 1994* was updated in 2011 by the *Offshore Minerals Amendment Ordinance 2011*. The update contains additional requirements within Sections 64 to 67 in the form of new Sections 64A to 64C being inserted after Section 64:

Section 64A “*Environmental impact assessment and environmental impact statements required for applications to drill regulated wells in controlled waters*” states that:

“(1) An environmental impact assessment and an environmental impact statement are required for each application for permission to drill a regulated well in controlled waters”.

“(2) An applicant for permission to drill a regulated well in controlled waters must comply with the requirements of section 64C(1) before making the application”.

Section 64B provides powers to require EIA or EIS for applications for activity other than exploratory drilling.

Section 64C contains general provisions for EIA and EIS contents and considerations of the Governor prior to granting consent:

“(1) If an environmental impact assessment and an environmental impact statement are required for an application:

(a) the applicant must conduct an environmental impact assessment of the likely adverse and beneficial effects upon the environment that there would be if the application were to be granted;

(b) the applicant must deliver to the Governor an environmental impact statement that contains (at least) the information required by schedule 4; and

(c) the applicant must publish that environmental impact statement and consult upon it in accordance with sections 65, 65A and 65B.

(2) The Governor must not determine an application for which an environmental impact assessment and an environmental impact statement are required until the applicant has complied with subsection (1).

(3) If an environmental impact assessment and an environmental impact statement are required for an application and the Governor grants that application:

(a) the Governor may impose conditions on the consent for one or more of the following purposes:

(i) to eliminate or reduce significant adverse effects on the environment of the project and the infrastructure associated with the project;

(ii) if possible, to remedy those effects; and

(iii) to offset them; and

(b) the Governor may impose those conditions even if there is no other power to do so.

(4) When considering an application for which an environmental impact assessment and an environmental impact statement are required, the Governor must take the following into account before deciding whether or not to grant it and whether or not to impose conditions:

(a) the environmental impact statement;

(b) if the Governor has sent the environmental impact statement to a technical expert for review, the representations made by that technical expert;

(c) representations from the public (and representations in reply from the applicant) submitted to the Governor in accordance with section 65B; and

(d) if the Governor has requested additional information or evidence under section 66, that additional information or evidence”.

Section 65 provides further information on the consultation and approval process, including new sections 65A and 65B:

“(1) Whenever an environmental impact statement is delivered by an applicant to the Governor in accordance with section 64C(1)(b), the applicant must seek to agree with the Governor the date on which the process of consultation on it will start.

(2) If no agreement is reached within a reasonable period, the Governor may give a direction as to the date on which the process will start.

(3) In sections 65A and 65B, the following definitions apply in relation to an environmental impact statement —

(a) “start date” means either —

(i) the date agreed between the applicant and the Governor under subsection (1); or

(ii) the date directed by the Governor under subsection (2);

(b) “closing date” means the date 42 days after the start date;

(c) “consultation period” (during which members of the public may make representations) means the period starting on the start date and ending on the closing date; and

(d) “follow-up period” (during which the applicant may make representations in reply) means the period —

(i) starting on the date on which confirmation is given to the applicant under section 65B(3) that copies of all of the representations made during the consultation period have been forwarded under section 65B(2); and

(ii) ending on the date 28 days after that date”.

65A. Publicity for environmental impact statement and consultation process

“(1) The Governor must arrange for a notice to be issued in the Gazette on the start date for each environmental impact statement.

(2) That notice must refer to the publication of the environmental impact statement and describe the consultation process.

(3) The applicant must make arrangements for each of the following things to happen:

(a) throughout the consultation period, a paper copy of the environmental impact statement must be available in Stanley (and, if the Governor directs, at one or more other places in the Falkland Islands) for the public to inspect (without charge) during at least normal government office hours;

(b) a paper copy of the non-technical summary of the environmental impact statement must be provided (without charge and as soon as possible) to each member of the public who requests one during the consultation period;

(c) an electronic copy of the environmental impact statement must be provided (without charge, as soon as possible and in a format that has been approved by or on behalf of the Governor) to each member of the public who requests one during the consultation period;

and;

(d) an electronic copy of the non-technical summary of the environmental impact statement must be provided (without charge, as soon as possible and in a format that has been approved by or on behalf of the Governor) to each member of the public who requests one during the consultation period.

(4) The applicant must also arrange for the publication of the environmental impact statement and the arrangements made under subsection (3) to be advertised in the following way:

(a) on the start date, there must be at least one announcement on the broadcast service provided by the Media Trust under section 5(1)(aa) of the Media Trust Ordinance (Title 59.1);

(b) throughout the rest of the consultation period, there must be further announcements on that radio service either —

(i) as agreed with the Governor; or

(ii) if an agreement cannot be reached, as directed by the Governor; and

(c) throughout the consultation period, there must be a notice in each edition of the newspaper published by the Media Trust under section 5(1)(a) of the Media Trust Ordinance.

(5) The announcements made and notices given under subsection (4) must also inform members of the public about —

(a) their right to make representations under section 65B;

(b) how to make those representations; and

(c) the closing date (by which those representations need to be made).

(6) The applicant may take other steps to publish the environmental impact statement and publicise the consultation process”.

65B. Representations

“(1) During the consultation period, anyone who wishes to do so may make written representations to the Governor about:

(a) the contents of the environmental impact statement; and

(b) in particular, the applicant’s proposals to protect the environment from adverse effects that there might be if the application were to be granted.

(2) The Governor must arrange for copies of the written representations made during the consultation period to be forwarded to the applicant as soon as possible after they are received.

(3) The Governor must arrange that, as soon as copies of all of the representations made during the consultation period have been forwarded under subsection (2), this is confirmed to the applicant.

(4) During the follow-up period, the applicant may make written representations to the Governor in reply to representations made under subsection (1)”.

Section 66 provides amendments to the text relating to the provision of further information in support of the EIS or evidence to verify the content of the EIS.

Section 67 provides powers for the applicant to request an exemption from the requirement of EIA on the basis that the environment would not be significantly affected. However, for drilling operations, it is generally accepted that an EIS is required.

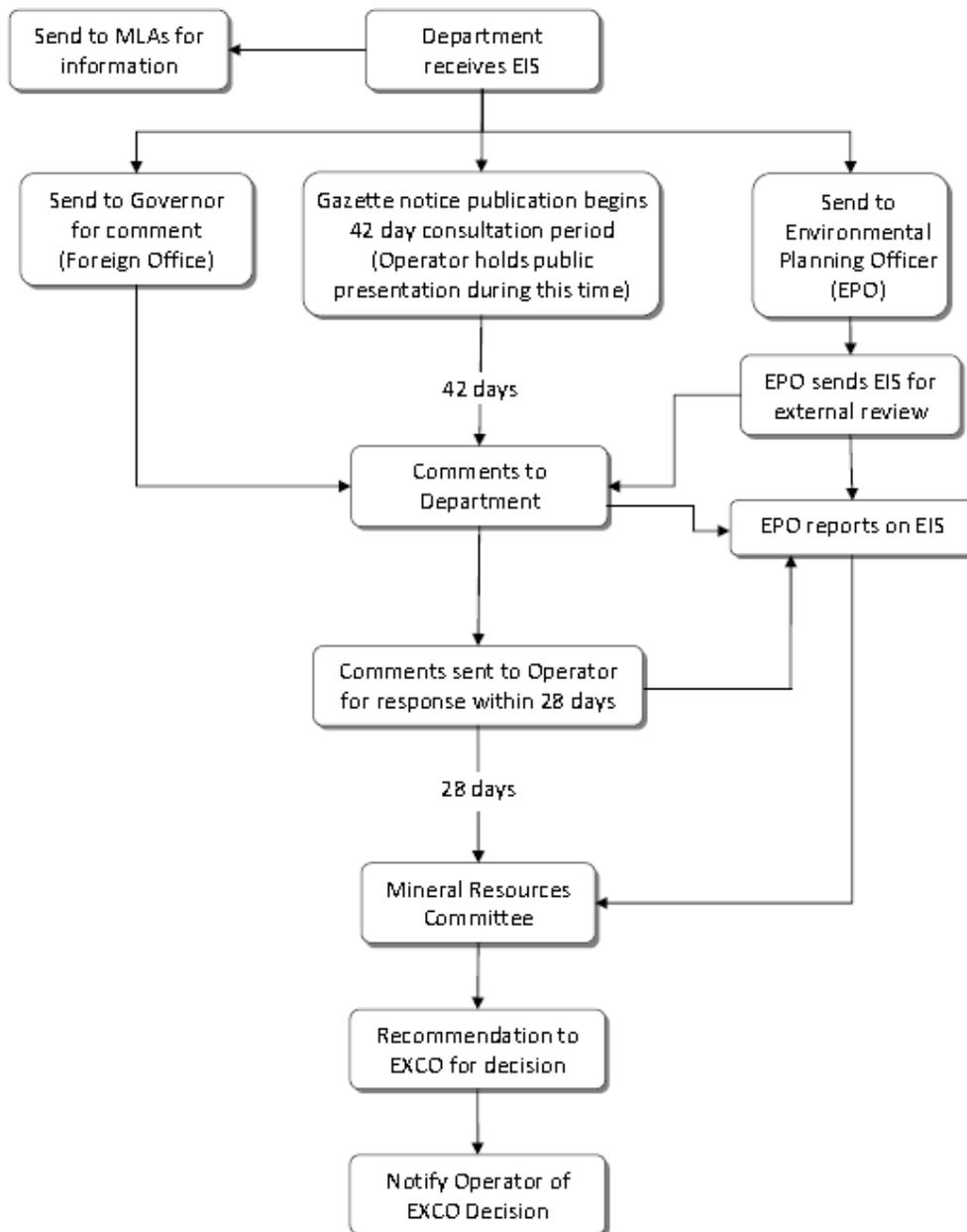
Amendments to Schedule 4 are also provided which contain further clarification on the requirements of the basic content of an EIS.

The *Offshore Minerals ordinance 1994* is currently the only piece of legislation in the Falkland Islands related to EIA for the oil and gas industry.

2.1.1 Guidance on the EIS Submission Process

The FIG Department of Mineral Resources has provided guidance to RPS on the way in which the ESHIA should be submitted and used for the basis of consultation during the approval process. A copy of a flow chart diagram received is provided in Figure 2.1.

Figure 2.1: Falkland Islands Government EIA / EIS consultation and approval process (Department of Mineral Resources, 2012)



2.1.2 RPS' Interpretation on guidance received from the Department of Mineral Resources

The FIG Department of Mineral Resources (DMR) and Environmental Planning Department (EPD) are responsible for assessment of applications under the *Offshore Minerals Ordinance 1994* (as

amended). Impact assessment documents are submitted to the DMR, however, their review is independently led by the EPD. Figure 3.1 outlines Nobles process for submission and approval of the exploration drilling ESHIA, and includes an outline of the Falkland Islands application and public consultation process, which is described below.

Preparation & Submission of the ESHIA

Following receipt of comments on the Scoping Report, the ESHIA document will be prepared. Once complete, the ESHIA will be submitted to the FIG DMR. At this point, an advertisement will be placed in the Falkland Islands Gazette, which will outline the project and state that a formal public consultation period is now open. An announcement will also be made to this end on the local radio station. The consultation period starts on the day that the advertisement is published in the Falkland Islands Gazette. The consultation period lasts for 42 calendar days.

The 42 calendar day period is the period from the date when the availability of the ESHIA is advertised in the FI Gazette, to the deadline for comments closing. During this time, the document is also presented to the Executive Council by FIG.

Accessibility of the ESHIA

During the consultation period, two copies of the ESHIA document will be made available in a public place for review, which will be the public library in Stanley. This will be made clear in the advertisement in the FI Gazette and during the announcement on the local radio station.

Fifteen full colour hard copies of the ESHIA document will be supplied to the DMR as part of the submission for internal FIG use and distribution. Hard copies should also be made available to interested local agencies from the Noble local office in Stanley during the consultation process to whoever requests such a copy. These hard copies will be printed locally in the Falkland Islands in the local print shop in Stanley. Demand is anticipated to be less than 50 copies.

FIG may also choose to host and publicise the ESHIA on its own website.

Public Disclosure Meetings & Stakeholder Comment on the ESHIA

During the 42 day consultation period, Noble will hold two public disclosure meetings in Stanley on consecutive days to present the ESHIA to the public. The meeting format will comprise a formal presentation given by Noble and/or RPS, outlining the project activities and the key findings of the ESHIA process, followed by an opportunity for members of the public to ask questions. This public session will be held in the second half of the 42 calendar day consultation period in order to give the public time to receive and review the report.

It is envisaged that the two public disclosure meetings will take place in the FIG Chamber of Commerce (subject to availability) and that the minutes from both days, will be recorded to comprise a formal consultation record. The meeting may also be used to provide general material relating to the project, in the form of public information leaflets and/or posters.

During the public consultation period, stakeholders will be able to comment on the ESHIA, using comment forms available on the day, or sending comments directly to the DMR and/or by attending and verbally commenting at one of the two public disclosure meetings.

ESHIA Review

At the end of the formal public consultation period, the FIG DMR, EPD and Mineral Resources Committee formal review of the ESHIA will take place. The review of the document will be led by the EPD. EPD often contracts an external UK scientific agency for review of the document, in addition to their own review. EPD also contact other UK government agencies for review of the document, which may include the UK Department of Energy and Climate Change (DECC) and/or the Joint Nature Conservation Committee (JNCC), although these agencies have no direct powers of intervention in any decision or recommendation made by FIG. This process can take up to an additional 12 weeks from the date of the end of the 42 day public consultation period.

Responses to Comments

After the formal review period, FIG will submit their comments to Noble on the ESHIA. There is a limited timeframe to reply to these comments (normally 28 calendar days). A direct formal reply to the comments in the form of a separate document is normally an adequate response and an updated ESHIA incorporating all comments is not normally required. However, the ESHIA report can be updated, if this is deemed necessary.

From previous experience on Falklands EIA projects it has been noted that the timing of comments received on the document can be sporadic; they can appear at various times throughout the formal FIG review period. RPS Energy will endeavour to respond to comments received as soon as possible upon their receipt. RPS Energy will draft responses to comments and submit them to Noble for review. Noble will have final approval of any response to comments and reserves the right to coordinate the response communication.

During the process, FIG will ensure that all responses received from the operator on stakeholder comments are satisfactory.

Notification of Decision

Notification of the Authorities decision on the ESHIA will be made formally by the FIG DMR, EPD and Mineral Resources Committee shortly after they are satisfied that all stakeholder comments have been addressed. The approval is in the form of a formal letter issued by the DMR. The decision is likely to be publicised by FIG on its own website. There may be certain conditions and requirements for the consented drilling activities and/or for further work or reports attached to any approval.

2.2 International Requirements from the IFC

The requirement for stakeholder engagement is outlined in IFC Performance Standard (PS) 1, paragraphs 26 through to 31 which, in general terms, require the following:

- Stakeholder analysis and planning;
- Disclosure and dissemination of information;
- Consultation and participation;
- The availability of a grievance mechanism; and
- On-going reporting to affected communities.

Specifically, IFC PS1 requires the following:

- The development and implementation of a Stakeholder Engagement Plan (this document) that is scaled to the project risks and impacts and development stage; and
- The disclosure of relevant project information to help Affected Communities and other stakeholders understand the risks, impacts and opportunities of the project. The client (Noble) should provide Affected Communities with access to relevant information on:
 - The purpose, nature, and scale of the project;
 - The duration of proposed project activities;
 - Any risks to and potential impacts on such communities and relevant mitigation measures;
 - The envisaged stakeholder engagement process; and
 - The grievance mechanism.

When Affected Communities are subject to identified risks and adverse impacts from a project, the client (Noble) will undertake a process of consultation in a manner that provides the Affected

Communities with opportunities to express their views on project risks, impacts and mitigation measures, and allows the client to consider and respond to them (this will take place during the 42 calendar public disclosure and consultation period using the ESHIA).

Furthermore, IFC PS1 emphasizes that effective consultation is a two-way process that should:

- Begin early in the process of identification of environmental and social risks and impacts (i.e. during ESHIA Scoping) and continue on an ongoing basis as risks and impacts arise;
- Be based on the prior disclosure and dissemination of relevant, transparent, objective, meaningful and easily accessible information which is in a culturally appropriate local language(s) and format and is understandable to Affected Communities;
- Focus inclusive engagement on those directly affected as opposed to those not directly affected;
- Be free of external manipulation, interference, coercion, or intimidation;
- Enable meaningful participation, where applicable; and
- Be documented.

IFC PS1 also requires, for projects with potentially significant adverse impacts on Affected Communities, an Informed Consultation and Participation (ICP) process be conducted that results in the Affected Communities' informed participation. ICP involves a more in-depth exchange of views and information, and an organized and iterative consultation. This process leads to the client's incorporating the views of the Affected Communities on matters that affect them directly, such as the proposed mitigation measures, the sharing of development benefits and opportunities, and implementation issues into the decision-making process.

3 Summary of Previous Stakeholder Engagement Activities

This section provides an overview of issues and concerns raised by specific stakeholders during previous exploration and drilling environmental impact assessments carried out by RPS Energy.

Table 3.1: Summary of previous stakeholder engagement and issues raised

| Stakeholders | Issues Raised | | | Summary |
|--|---|--|--|--|
| | Argos Resources Limited (April 2011) Exploration and Appraisal Drilling EIS (Licence PL001) | Borders & Southern Petroleum plc. (February 2010) Offshore Falkland Islands Exploration Drilling EIS (Licence PL018) | Falklands Oil and Gas Limited (November 2011) Exploration Drilling EIS (Licences PL011, 12, 26, 27, 28) | |
| Falklands Islands Department of Mineral Resources | Review of FIG Oil and Gas Legislation. Argos to provide technical updates. | Borders & Southern to provide technical updates. | FOGL to provide technical updates and mitigation measures in line with DECC Guidance (July 2011). | Operator to provide mitigation measures for negative impacts. |
| Civil Aviation and Stanley Airport Falkland Islands Government Air Service (FIGAS) | MoD is integral to all aviation issues. Charter flights to the Islands can be organised. | MoD is integral to all aviation issues. Charter flights to the Islands can be organised. | Adequate aerial surveillance can be provided by FIGAS, but no dispersant spraying capability exists in the event of an oil spill. | FIGAS can provide adequate aerial surveillance in the event of an oil spill, however they have no dispersant spraying capacity (such as that comparable to the UKCS dispersant aircraft). |

| Stakeholders | Issues Raised | | | Summary |
|--|--|---|---|---|
| | Argos Resources Limited (April 2011) Exploration and Appraisal Drilling EIS (Licence PL001) | Borders & Southern Petroleum plc. (February 2010) Offshore Falkland Islands Exploration Drilling EIS (Licence PL018) | Falklands Oil and Gas Limited (November 2011) Exploration Drilling EIS (Licences PL011, 12, 26, 27, 28) | |
| <p>Environmental Planning Department</p> <p>Falklands Conservation</p> | <p>Collection and incorporation of updated environmental data.</p> <p>Impact of drilling activities on benthic communities is currently a “hot topic” which should be considered in the EIA process.</p> <p>Seabirds are the main vulnerability due to the risk of oil spills.</p> | <p>Seabirds are the main vulnerability due to the risk of oil spills.</p> | <p>Limited waste disposal capacity on the islands; only non-hazardous waste can be accepted whereas hazardous waste must be shipped to UK.</p> <p>Highlighted international protection status of a number of vulnerable areas within the FI and lack of local protection measures.</p> <p>Seabirds and mammals are the main vulnerability to any offshore work (through spills).</p> <p>Emphasised limited capabilities for Wildlife response (Tier 1) and operator’s responsibility for arranging Tier 2/3 response.</p> <p>Need for survey programmes and research on marine wildlife to improve the knowledge on baseline environment.</p> | <p>Environmental Planning Department:</p> <p>Limited waste disposal capacity for waste, no hazardous waste can be received, it must be exported. International protection measures for a number of vulnerable areas in the Islands and current lack of local protection measures in the event of an oil spill.</p> <p>Falklands Conservation:</p> <p>Seabirds and marine mammals are the main vulnerability to any offshore oil and gas activities through possible oil spills, therefore oil spills are of concern. Concerns over limited capabilities of the Falkland Islands for wildlife response in the event of a pollution incident (Tier 1) and the operator’s responsibility for arranging Tier 2/3 response. The need for further survey programmes and research on marine wildlife to improve the knowledge on baseline environment has also often been mentioned.</p> |

| Stakeholders | Issues Raised | | | Summary |
|--|--|---|---|---|
| | Argos Resources Limited (April 2011) Exploration and Appraisal Drilling EIS (Licence PL001) | Borders & Southern Petroleum plc. (February 2010) Offshore Falkland Islands Exploration Drilling EIS (Licence PL018) | Falklands Oil and Gas Limited (November 2011) Exploration Drilling EIS (Licences PL011, 12, 26, 27, 28) | |
| Department of Fisheries and Marine Resources | Notice to Mariners via Fisheries and Marine Resources, and minimal response capability for oil spills. Dispersant Use Policy. | Notice to Mariners via Fisheries and Marine Resources, and minimal response capability for oil spills. | Notice to Mariners via Fisheries and Marine Resources. Minimal response capability for oil spills. Emphasised operator's responsibility for responding to small and large spill in cooperation with other Oil & Gas operators and FIG. | Disruption to fishing activities, concerns regarding oil spills, limited capacity for responding to oil spills in the Falklands. |
| Department of Public Works | Waste – very limited capacity for storage, management and/or onshore processing. Recycling is an option, but very limited. | Waste – very limited capacity for storage, management and/or onshore processing. Some recycling is available but is very limited. | Waste – very limited capacity for storage, management and/or onshore processing. Some recycling is available. Water resources are scarce during dry summer months and may impose restrictions on water demands for multiple drilling operations. | Waste storage, handling and onshore processing, as the Falkland Islands have very limited waste capacity. Water resources can be scarce during dry summer months and may impose restrictions on water demands. |
| Emergency Response: FI Defence Force, Fire/Rescue Service, KEMH, Police Chief | FIG Major Incident Plan. FIG National Oil Spill Contingency Plan. | National Emergency Response Plan. | National Emergency Response Plan. | Emergency plans developed by operators must take into account the Falkland Islands National Emergency Response Plan. Oil Spill Contingency Plans Developed must also take into account the National Oil Spill Contingency Plan (NOSCP). |

| Stakeholders | Issues Raised | | | Summary |
|--|--|---|--|--|
| | Argos Resources Limited (April 2011) Exploration and Appraisal Drilling EIS (Licence PL001) | Borders & Southern Petroleum plc. (February 2010) Offshore Falkland Islands Exploration Drilling EIS (Licence PL018) | Falklands Oil and Gas Limited (November 2011) Exploration Drilling EIS (Licences PL011, 12, 26, 27, 28) | |
| British Military Base at Mount Pleasant | Understanding military operations and resources, and communicating with the military on proposed activities. | Understanding military operations and resources, and communicating with the military on proposed activities. | Not engaged with. | Keep military informed of all proposed activities. |

4 Stakeholder Identification

4.1 Introduction

In accordance with the IFC Sustainability Framework (2012) Guidance Note 1 on IFC PS1 paragraph 95, stakeholders are defined as: *“persons, groups or communities external to the core operations of a project who may be affected by the project or have interest in it. This may include individuals, businesses, communities, local government authorities, local nongovernmental and other institutions, and other interested or affected parties”*.

The Falklands Islands are characterised by a relatively isolated, geographically constrained collection of communities (settlements) primarily based on East Falkland. This is reinforced by the relatively low population of the Falkland Islands which in the 2012 census (*Falkland Islands Government, 2013*) was recorded to be 3,135 people; this includes 295 persons who were classified as temporary visitors to the island. Almost all of the population lives on East Falkland (75% in Stanley) and elsewhere, with just 200 people living on West Falkland.

Due to the recent high level of oil and gas exploration activity, which was started by Desire Petroleum plc exploration drilling in February 2010, there is an established precedent of stakeholder groups who are regularly engaged for planned oil and gas activities.

4.2 Identification Process

For the purpose of the ESHIA, the process used to identify stakeholders comprised the following steps:

- Identifying individuals, groups, local communities and other stakeholders that may be affected by the project, positively or negatively, and directly or indirectly, particularly those directly and adversely affected by project activities, including those who are disadvantaged or vulnerable (*this includes representatives of the fishing, tourism industry and local residents [refer to note about Affected Communities below]*);
- Identifying broader stakeholders who may be able to influence the outcome of the project because of their knowledge about the Affected Communities or political influence over them (*this includes national regulators and public institutions who are able to influence the outcome of the ESHIA*);
- Identifying legitimate stakeholder representatives, including elected officials, non-elected community leaders, leaders of informal or traditional community institutions, and elders within the Affected Community (*for the purpose of the ESHIA this has been expanded to include representatives of civil society groups such as NGOs*); and
- Mapping the impact zones by placing the Affected Communities within a geographic area, which should help the client define or refine the project’s area of influence (*see note about Affected Communities below*).

The economic benefits derived from offshore hydrocarbon extraction have the potential to impact all the communities and residents of the Falklands Island through increased provision of social infrastructure services (such as improved road transport, and social resources such as health and educational establishments), as well as increased employment available within the oil and gas sector.

Any significant, environmental, socio-economic or health impact from the offshore oil and gas industry, (such as, for example, a large unintentional released of hydrocarbons (e.g. a blowout event) during drilling that results in an oil spill impacting the coastline and fishing industry) has the potential to directly or indirectly affect all communities and residents on the Falkland Islands. This type of impact is likely to be perceived by local communities and residents as an ‘island-wide’ event potentially impacting the onshore economy, the price of goods and services (such as food), the fishing and tourism industries and the farming industry. Due to the reasons outlined

above, and for the purpose of the ESHIA, the communities and residents of the Falklands Islands have been considered in the SEP to comprise a single 'Affected Community'.

4.2.1 Categories of Stakeholders to be consulted

Table 4.1 describes the categories of stakeholders who will be consulted during the ESHIA process.

Table 4.1: Categories of stakeholders to be contacted

| Stakeholder Group | Connection to the Project |
|--|---|
| Public institutions and regional authorities | Ministries, departments and agencies who implement legislation associated with planning and approval for oil and gas exploration activities, statutory agencies associated with environmental protection or that have a role in the project planning and approval process. The Governor of the Falkland Islands and functional departments within the administration with regulatory responsibilities delegated from the Falkland Islands Government (FIG) that are relevant to the project planning and approval process. |
| Local businesses | Private companies with interest in the Falkland Islands whose business may be impacted by the proposed exploration drilling activities. Service companies that are relevant to the project (including for example, logistics support). |
| Users of the sea | People and businesses reliant on the quality of the sea and seashore for fishing, recreational activities, and tourism. |
| Affected Communities | The communities and residents on the Falkland Islands who collectively, represent a single Affected Community that may potentially be impacted (either directly or indirectly) by various aspects of the proposed exploration drilling activities. This includes disadvantaged people and/or vulnerable groups. |
| Local, national and international environmental NGOs and research institutions | Organisations with interests in sustainability and the environment, who aim to represent the views and interests of their members and/or the general public with regards to exploration drilling activities. |

4.2.2 Stakeholder Identification

The identities of stakeholders who will be engaged with during the ESHIA are presented in Table 4.2.

Table 4.2: Stakeholders to be engaged with during the ESHIA process

| Stakeholder Identity | Stakeholder profile and justification for inclusion into the SEP |
|--|--|
| Public institutions and regional authorities | |
| Falkland Islands Department of Mineral Resources | Government department responsible for providing exploration and production licences for oil and gas activities within the Falkland Islands Designated Exploration Area. Expected to have updated recent socio-economic data associated with fishing activity. |
| Falklands Islands Environmental Planning Department | Government department responsible for town planning, building and the environment, including waste management. The department also administers the Mineral Resources Committee. |
| British Military Base at Mount Pleasant | Centre of military defence in the Falkland Islands. There is a potential for the project to interfere with planned, or ongoing, military operations within the offshore blocks. The military must also be kept informed of any proposed oil and gas activity. |
| Civil Aviation and Stanley Airport and Falkland Islands Government Air Service (FIGAS) | Government-operated airline that provides regular military and passenger services between the Falkland Islands and the UK. Can provide aerial surveillance in the event of an offshore oil spill response that may be implemented in the event of a major unintentional hydrocarbon release during the project. |
| Falkland Islands Fisheries Department | Government department responsible for the sustainable development of the fisheries occurring within the waters of the Falkland Conservation Zones (FCZs). Details about how a marine notice to mariners can be issued are required, as this comprises a key mitigation measure associated with avoiding marine accidents and vessel collisions. |
| Falkland Islands Public Works Department | Responsible for the provision of services in the Falkland Islands which include, but are not limited to, waste management infrastructure. The project will generate quantities of non-hazardous and hazardous waste. |
| The Royal Falkland Islands Police | Responsible for the development and implementation of the Falkland Island National Oil Spill Response Plan that may be implemented in the event of a major incident during the project. |
| Falkland Islands Tourist Board | Government department responsible for the management and promotion of tourism activities on the Falkland Islands. |
| The UK Joint Nature Conservation Committee (JNCC) | Public body that advises the UK Government and devolved administrations on UK-wide and international nature conservation. Originally established under the <i>Environmental Protection Act 1990</i> , JNCC was reconstituted by the <i>Natural Environment and Rural Communities (NERC) Act 2006</i> . |
| Users of the sea | |
| Falkland Islands Fishing Companies Association (FIFCA) | An organisation that aims to jointly represent the business interests of companies involved in fishing activities. Information on the location, type and intensity of fishing activities within the offshore blocks is required. |
| Falkland Islands Yacht Club | Formed in 2010 to promote boating activities in the Islands and aims to represent the interests of the recreational boating community. Information on the location, type and intensity of recreational sailing within the offshore blocks is required. |
| Affected Communities | |
| Affected Community of the Falkland Islands | The communities and residents of the Falkland Islands. Taking consideration of the views and opinions of Affected Communities in the decision making process is an IFC requirement. |
| Local, national and international environmental NGOs and research institutions | |
| Falklands Conservation | An NGO working to protect all the wildlife in the Falkland Islands for future generations. Undertakes practical conservation projects, surveys and scientific studies, conducts annual monitoring of seabird populations, rescues wildlife in trouble, publishes guides and information on many aspects of the Falkland Islands environment, and involves islanders of all ages in its activities. May have recent environmental data that is of relevance to the ESHIA. |
| Shallow Marine Surveys Group (SMSG) | Local organisation that exists to coordinate and conduct assessments of the status of inshore resources around the Falkland Islands. Headed by a core group of experienced biologists, divers and assisted by volunteers from the local community, they have been collecting, identifying and photographing marine animals since June 2006. May have recent environmental data that is of relevance to the ESHIA. |
| South Atlantic Environmental Research Institute (SAERI) | Locally active academic organisation conducting scientific research in the South Atlantic from the tropics down to the ice in Antarctica. SAERI's remit includes the natural and physical sciences. Aims to conduct world class research, teach students, and build capacity within and between the South Atlantic Overseas Territories. May have recent environmental data that is of relevance to the ESHIA. |
| New Island Conservation Trust | Locally active NGO that aims to promote the study and appreciation of ecology and wildlife conservation throughout the Falkland Islands, and to assist in developing plans for the management and conservation of its exceptional natural environment for the future. May have recent environmental data that is of relevance to the ESHIA. |
| Birdlife International | Global partnership of conservation organisations that strives to conserve birds, their habitats and global biodiversity, working with people towards sustainability in the use of natural resources. May have recent environmental data that is of relevance to the ESHIA. |

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5 Stakeholder Engagement Programme

Stakeholder engagement activities will take place at the following key stages of the ESHIA process: During ESHIA Scoping, content development of the ESHIA Scoping Report and content development of the ESHIA.

5.1 ESHIA Scoping

The way in which stakeholders will be engaged with during ESHIA Scoping is summarised in Table 5.1.

Table 5.1: Stakeholders to be engaged with during ESHIA Scoping

| Stakeholder Name | Format to be used for Engagement | Materials to be Disclosed | Method used to Record Results |
|--|--|--|---|
| Falklands Islands Department of Mineral Resources | Initial contact made by e-mail. A formal meeting held individually at their office. | A 4-page summary of the project will be sent to the stakeholder as an attachment to the initial e-mails. During the meetings, a large-scale, annotated, UK Hydrographic Office Admiralty Chart will be used to identify the project area. | ESHIA Scoping Consultation Record. A copy of the template to be used is provided in Appendix A. |
| Falkland Islands Environmental Planning Department | | | |
| British Military Base at Mount Pleasant | | | |
| Civil Aviation and Stanley Airport and Falkland Islands Government Air Service (FIGAS) | | | |
| Falkland Islands Fisheries Department | | | |
| Falkland Islands Public Works Department | | | |
| The Royal Falkland Islands Police | | | |
| Falkland Island Tourist Board | | | |
| Falkland Islands Fishing Companies Association | Initial contact made by e-mail. | A 4-page summary of the project will be sent to the stakeholder as an attachment to the letter. | The stakeholder's written response will be filed. |
| Falkland Island Yacht Club | A combined meeting. | | |
| The UK Joint Nature Conservation Committee (JNCC) | Formal letter. | A 4-page summary of the project will be sent to the stakeholder as an attachment to the e-mails. During the meetings, a large-scale, annotated, UK Hydrographic Office Admiralty Chart will be used to identify the project area. | ESHIA Scoping Consultation Record. A copy of the template to be used is provided in Appendix A. |
| BirdLife International | | | |
| Falklands Conservation | Initial contact made by e-mail. A round-table meeting will be held with all stakeholders on the same day. | A 4-page summary of the project will be sent to the stakeholder as an attachment to the e-mails. During the meetings, a large-scale, annotated, UK Hydrographic Office Admiralty Chart will be used to identify the project area. | ESHIA Scoping Consultation Record. A copy of the template to be used is provided in Appendix A. |
| Birdlife International | | | |
| Shallow Marine Surveys Group (MSG) | | | |
| South Atlantic Environmental Research Institute (SAERI) | | | |
| New Island Conservation Trust | A single, 3-hour focus group discussion (refer to Section 5.1.1). | PowerPoint slides to provide a summary of the project. | Formal minutes of meeting. |
| Affected Community of Falkland Island | | | |

5.1.1 Focus Group Discussion

A single, 3-hour focus group discussion (FGD) will be held with 12-15 people during ESHIA Scoping. The aim of the FGD is to gather a collection of local residents who can provide their views and opinions on the project in a transparent and semi-formal environment, at an early stage in the impact identification and evaluation process. In order to achieve a representative group of the Falkland Island Affected Community, the following selection criteria and targets will be used for the individuals attending:

- Gender - with the aim of achieving a male/female ratio of at least 60:40 so that the views and interests of women are adequately represented;
- Age - so that both young people and the elderly are adequately represented;
- Vulnerability - so that any individuals or groups who may be proportionally disadvantaged by the project are adequately represented;
- Representatives of educational establishments such as teachers or educational professionals; and
- Location - individuals from West Falkland will be specifically invited to attend, particularly those involved in farming activities.

In order to facilitate the FGD, RPS will provide suitable hotel facilities, refreshments and a positive, relaxed atmosphere. The FGD will be facilitated by an experienced RPS professional.

5.2 Content of the ESHIA Scoping Report

The way in which stakeholders will be engaged with using the content of the ESHIA Scoping Report is summarised in Table 5.2.

Table 5.2: Stakeholders to be engaged with, using the ESHIA Scoping Report

| Stakeholder Name | Format to be used for Engagement | Materials to be Disclosed | Method used to Record Results |
|--|----------------------------------|--|---|
| Falklands Islands Department of Mineral Resources | Electronic exchange of report. | Electronic copy of the ESHIA Scoping Report. | The stakeholder's written response will be filed. |
| Falklands Islands Environmental Planning Department | | | |
| British Military Base at Mount Pleasant | | | |
| Civil Aviation and Stanley Airport and Falkland Islands Government Air Service (FIGAS) | | | |
| Falkland Islands Fisheries Department | | | |
| Falkland Islands Public Works Department | | | |
| The Royal Falkland Islands Police | | | |
| Falkland Island Tourist Board | | | |
| Falkland Islands Fishing Companies Association | | | |
| Falkland Island Yacht Club | | | |
| The UK Joint Nature Conservation Committee (JNCC) | | | |
| Birdlife International | | | |
| Falklands Conservation | | | |
| Shallow Marine Surveys Group (SMSG) | | | |
| South Atlantic Environmental Research Institute | | | |
| New Island Conservation Trust | | | |
| Affected Community of Falkland Island | (none) | | |

5.3 Content of the ESHIA

The way in which stakeholders will be engaged with using the content of the ESHIA is prepared is summarised in Table 5.3.

Table 5.3: Stakeholders to be engaged with, using the ESHIA

| Stakeholder Name | Format to be used for Engagement | Materials to be Disclosed | Method used to Record Results |
|--|--|--|---|
| Falklands Islands Department of Mineral Resources | Electronic and/or hardcopy exchange of report. | Electronic and/or colour hard copy of the ESHIA. | The stakeholder's written response will be filed. |
| Falklands Islands Environmental Planning Department | | | |
| British Military Base at Mount Pleasant | | | |
| Civil Aviation and Stanley Airport and Falkland Islands Government Air Service (FIGAS) | | | |
| Falkland Islands Fisheries Department | | | |
| Falkland Islands Public Works Department | | | |
| The Royal Falkland Islands Police | | | |
| Falkland Island Tourist Board | | | |
| Falkland Islands Fishing Companies Association | | | |
| Falkland Island Yacht Club | | | |
| The UK Joint Nature Conservation Committee (JNCC) | | | |
| Birdlife International | | | |
| Falklands Conservation | | | |
| Shallow Marine Surveys Group (MSG) | | | |
| South Atlantic Environmental Research Institute | | | |
| New Island Conservation Trust | | | |

| Stakeholder Name | Format to be used for Engagement | Materials to be Disclosed | Method used to Record Results |
|---|--|---|--|
| <p>Affected Community of Falkland Islands (including the public).</p> | <p>42 calendar day public consultation process. 2 separate day walk-in sessions.</p> | <p>Notice placed in the Falkland Islands Gazette indicating the availability of the ESHIA.</p> <p>A radio announcement advertising the availability of the ESHIA will be made on the start date of the public consultation period via the local radio service.</p> <p>All announcements made in the Falkland Island Gazette and radio will inform the public about their right to make a representation, how this should be done and the applicable closing date.</p> <p>An electronic copy of the ESHIA and non-technical summary may (at the discretion of the Governor) be available on the FIG website/other websites during the consultation period.</p> <p>Colour hard copy of the ESHIA and non-technical summary will be available for inspection at the public library in Stanley (and other locations if the Governor directs) for review throughout the consultation period during normal government office hours</p> <p>Additional hard/electronic copies of the ESHIA and non-technical summary will be available to any member of the public who requests it, without charge or undue delay during the consultation period.</p> | <p>Comment forms.</p> <p>Written submission to the Governor or other public institution.</p> |

6 Engagement Timescale

The time schedule for the engagement that will take place during the ESHIA is provided in Table 6.1.

Table 6.1. Time schedule for stakeholder engagement

| <i>ESHIA Stage</i> | <i>Indicative Schedule</i> |
|-----------------------------------|--|
| <i>ESHIA Scoping Consultation</i> | <i>29th July 2013 to 13th September 2013</i> |
| <i>ESHIA Scoping Report</i> | <i>6th May to 20th May 2014</i> |
| <i>ESHIA</i> | <i>Public consultation will take place for 42 days following submission.</i> |

7 Internal & External Roles & Responsibilities

In relation to implementation of the SEP, RPS will be responsible for the following:

- The initial preparation of the SEP and any subsequent updates;
- Day-to-day coordination of all stakeholder engagement activities;
- The practical and logistical organisation of all stakeholder engagement activities including contacting stakeholders to arrange meeting times/locations;
- The preparation and issuing of all stakeholder engagement invitation letters by e-mail;
- The preparation of all stakeholder engagement disclosure materials which includes e-mails, PowerPoint slides, public advertisements, and the ESHIA Scoping Report and the ESHIA;
- Actions associated with the use of the Community Feedback Mechanism in close liaison with representatives from Noble;
- Attendance/facilitation of all stakeholder engagement meetings/discussions, including hotel conference room meetings, and the recording of all results and issues raised;
- Placement of the advertisement in the Falklands Island Gazette and The Penguin News at the start of the 42 days public consultation process; and
- The printing of all stakeholder engagement disclosure materials, including hard-copies of the ESHIA Scoping Report and hard-copies of the ESHIA for public consultation.

Noble is responsible for the following:

- Review and approval of the SEP and any subsequent updates;
- The approval of all stakeholder engagement disclosure materials, prior to their use; and
- Actions associated with the use of the Community Feedback Mechanism in close liaison with RPS.

8 Noble Energy Community Feedback Mechanism

A dedicated local telephone number and e-mail address have been set up for the Falkland Islands community to provide feedback directly to Noble:

- Phone line: +500 22986
- Email address: FalklandsSocialResponsibility@nobleenergyinc.com

The feedback mechanism is to be used by any party to raise any concerns, questions or complaints associated with the project at any stage of the project lifecycle.

9 Monitoring & Reporting

During the implementation of the SEP, stakeholder engagement activities and results will be closely recorded with the aim of improving the overall process of engagement over time.

For example, after each formal meeting at the start of the ESHIA Scoping Phase, an internal discussion will be held to review the success of the meeting and the way in which information was disclosed to try and improve on the overall performance of engagement, during subsequent meetings.

In the event that a grievance is received on the way in which stakeholder engagement activities are being conducted, all activities may temporarily cease until the grievance is resolved.

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APPENDIX A: ESHIA Scoping Template Consultation Record

| | | | |
|---|--|---|--|
| Stakeholder Name: | | Attendance / Representatives name: | |
| Date and Time (start/end): | | Location: | |
| Specific objectives of consultation: | | | |
| Record of Information Disclosed | | | |
| | | | |
| Scoping results – key areas of stakeholder concern | | | |
| | | | |
| Baseline data | | | |
| | | | |
| Influence on content of ESHIA | | | |
| | | | |

APPENDIX B: Stakeholder Contact Information

| Stakeholder | Website | Main contact and email address: | Tel/ Fax | Postal address: |
|---|---|---|------------------------------------|---|
| Falkland Islands Government Department of Mineral Resources: | http://www.bgs.ac.uk/falklands-oil/ | Steve Luxton; sluxton@mineralresources.gov.fk Roberto Cordiero; RCordeiro@mineralresources.gov.fk | T: +500 27322 F: +500 27321 | Department of Mineral Resources, Falkland Islands Government, Government House, Ross Road, Stanley, Falkland Islands FIQQ 1ZZ. |
| Falkland Islands Government Environmental Planning Department | http://www.epd.gov.fk/ | Nick Rendell (Environmental Officer) NREndell.planning@taxation.gov.fk epdfig@mail.com | T: +500 28480 F: +500 27391 | Environmental Planning Department, Falkland Islands Government, Government House, Ross Road, Stanley, Falkland Islands FIQQ 1ZZ. |
| British Military Base at Mount Pleasant | http://www.raf.mod.uk/currentoperations/opsfalklands.cfm | Roy Smith (Environmental Officer) hqtepo@MOUNTPLEASANT.mod.uk | +500 76845 (Mobile: +500 76049) | Ministry of Defence, Mount Pleasant Airbase, Darwin Road, Falkland Islands FIQQ 1ZZ |
| Civil Aviation and Stanley Airport and Falkland Islands Government Air Service | <i>No website</i> | Jan Ross jross@figas.gov.fk | +500 27219 | FIGAS, Port Stanley Airport, Airport Road, Port Stanley, Stanley, Falkland Islands FIQQ 1ZZ. |
| Falkland Islands Government Department of Natural Resources - Fisheries Department | http://fis.com/falklandfish/ | John Barton (Director of Fisheries) director@fisheries.gov.fk Dr. Paul Brewin (Scientific Officer) PBrewin@fisheries.gov.fk | T: +500 27260 F: +500 27265 | Falkland Islands Fisheries Department, PO BOX 598, Stanley, Falkland Islands FIQQ 1ZZ. |
| Falkland Islands Public Works Department | http://www.falklands.gov.fk/ | Manfred Keenleyside (Director of Public Works) admin@pwd.gov.fk | T: +500 27193 F: +500 27191 | Falkland Islands Public Works Department, Falkland Islands Government, Government House, Ross Road, Stanley, Falkland Islands FIQQ 1ZZ. |
| Attorney Generals Chambers | http://www.crownofficechambers.com/ | Alison Carter CrownCounsel@sec.gov.fk Mark Lewis MLewis@sec.gov.fk | T: +500 28461 | - |

| Stakeholder | Website | Main contact and email address: | Tel/ Fax | Postal address: |
|--|---|--|--------------------------------|--|
| Royal Falkland Islands Police | http://www.falklands.gov.fk/ | Barry Marsden (Director of Emergency Services, Chief Police Officer and Principal Immigration Officer). Lisa Martin (Secretary) LMartin@police.gov.fk | T: +500 28100 F: +500 28110 | Royal Falkland Islands Police Headquarters, Ross Road, Stanley, Falkland Islands FIQQ 1ZZ. |
| Falkland Islands Tourist Board | http://www.falklandislands.com/ | Tony@falklandislands.com | T: +500 22215 | Falkland Islands Tourist Board, Jetty Visitor Centre, PO BOX 618, Stanley, Falkland Islands FIQQ 1ZZ. |
| Stanley Services | http://www.stanley-services.co.fk/ | Robert Rowlands rrowlands@stanley-services.co.fk | T: +500 22622 F: +500 22623 | Stanley Services Limited, Airport Road, P.O Box 117, Stanley, Falkland Islands FIQQ 1ZZ |
| Falkland Islands Company | http://www.the-falkland-islands-co.com/ | Roger Spink rks@fic.co.fk | T: +500 27600 F: +500 27603 | - |
| UK Joint Nature Conservation Committee (JNCC) | http://jncc.defra.gov.uk/page-4402 | Anne Saunders (UK South Atlantic Overseas Territories Officer). Anne.Saunders@jncc.gov.uk | - | Joint Nature Conservation Committee, P.O. Box 585, Stanley, Falkland Islands, FIQQ 1ZZ. |
| Falkland Islands Harbour Master / Port Authority | - | Malcolm Jamieson MJamieson@fisheries.gov.fk | - | - |
| Falkland Islands Fishing Companies Association (FIFCA) | <i>No website</i> | Andy Pollard (Secretary). fifca@horizon.co.fk | T: +500 22317 | PO Box 664, Room 11, Stanley Services, By-Pass Road, Stanley, Falkland Islands FIQQ 1ZZ. |
| Falkland Islands Yacht Club (FIYC) | http://www.falklandsailing.com/index.html | Tony Blake (Chair – Falkland Islands Yacht Club Committee). sailing@horizon.co.fk Ken Passfield kenpassfield@yahoo.co.uk | - | - |
| Falklands Conservation | http://www.falklandsconservation.com/ | Dr. David Doxford (Chief Executive Officer) ceo@conservation.org.fk Andrew Stanworth CO@conservation.org.fk | T: +500 22247 F: +500 22288 | Falklands Conservation, Jubilee Villas, Ross Road, Stanley, Falkland Islands FIQQ 1ZZ. (UK contact: Sarah Brennan, Tel: 01767 650 639). |

| Stakeholder | Website | Main contact and email address: | Tel/ Fax | Postal address: |
|---|---|---|---------------|--|
| Shallow Marine Surveys Group | http://smsg-falklands.org/ | Paul Brickle (Fisheries Biologist / Marine Ecologist). pbrickle@smsg-falklands.org | T: +500 27260 | Shallow Marine Surveys Group, PO Box 598, Stanley, East Falkland, Falkland Islands FIQQ 1ZZ. |
| South Atlantic Environmental Research Institute (SAERI) | http://www.south-atlantic-research.org/ | Paul Brickle (Fisheries Biologist / Marine Ecologist). pbrickle@env.institute.ac.fk | T: +500 27374 | PO Box 609, Stanley Cottage, Stanley, Falkland Islands FIQQ 1ZZ. |
| New Island Conservation Trust | http://www.falklandswildlife.com/ | Phyl Rendell phylrendell@horizon.co.fk | - | New Island Conservation Trust, The Dolphins, Snake Hill, Stanley, Falkland Islands FIQQ 1ZZ. |
| Beaver Island Land Care | - | Sally Poncet sallyponcet@horizon.co.fk | - | - |
| Birdlife International | http://www.birdlife.org/worldwide/national/falkland_islands/ | Professor John Croxall (Chair BirdLife International Global Seabird Programme). John.Croxall@birdlife.org | - | <i>As per Falklands Conservation.</i> |
| Rural Business Association | - | Nyree Heathman rba@horizon.co.fk | - | - |

Appendix K: FISA12 Environmental Baseline Survey Report



Client

Contractor

Project: **Noble Energy Falklands Limited**

FINAL

**NOBLE ENERGY; SOUTH FALKLANDS BASIN ENVIRONMENTAL
AND GEOCHEMICAL PROGRAM
December 2013 – March 2014**

**FISA ENVIRONMENTAL BASELINE AND HABITAT
SURVEY REPORT**

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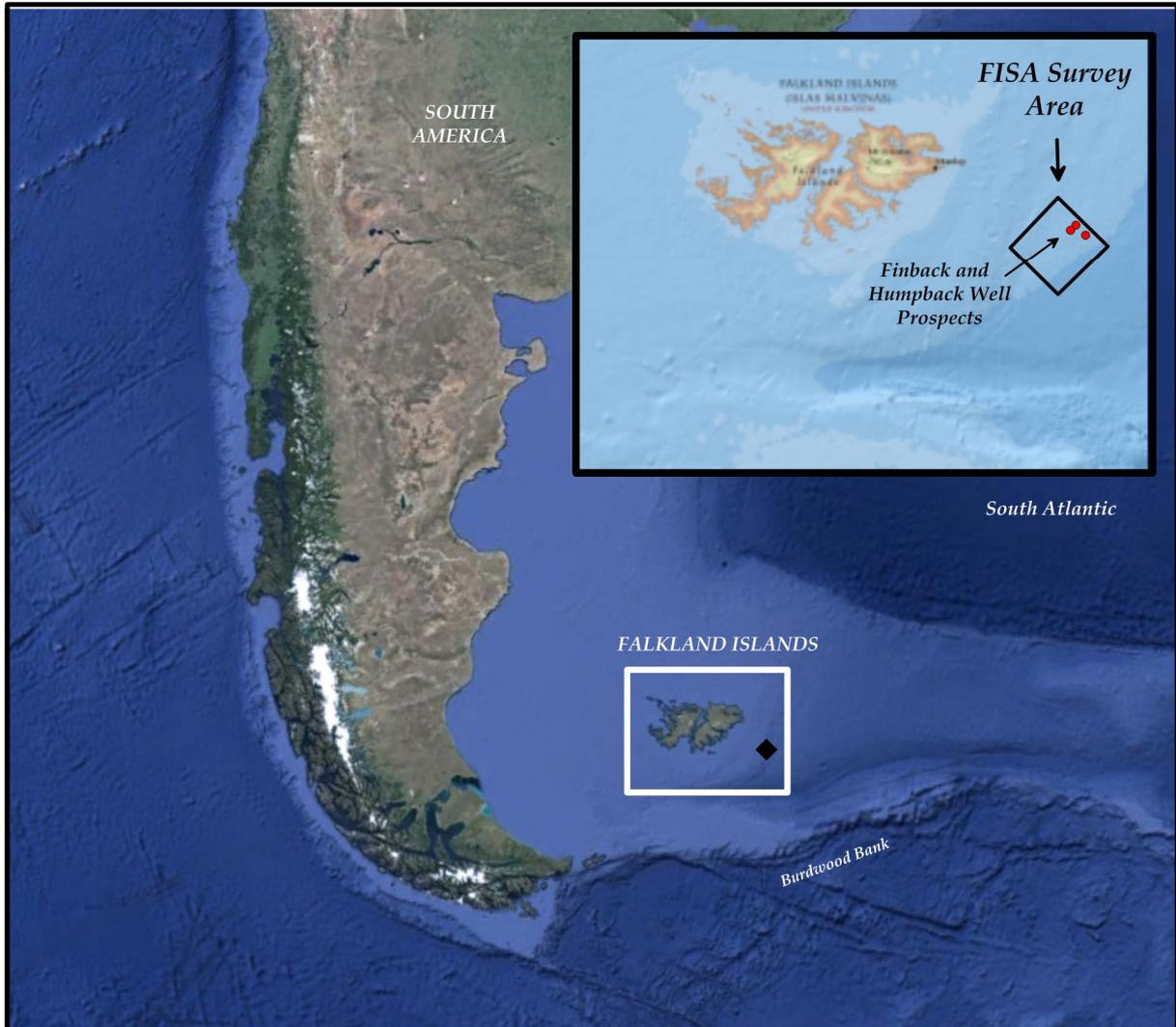
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| Authors | Hung Liu | Date : 08/01/2015 | | |
| Checked | Ian Wilson | Date : 18/09/2014 | | |
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GLOSSARY

| | |
|---------------------|---|
| BDC | Biodiversity Committee |
| Bedform | Morphology of the seabed created by currents (i.e. ripples) |
| Bioherm | Mound or rock composed mainly of sedentary organisms (i.e. corals, algae, or molluscs) and surrounded by rock of different origin |
| Bioturbation | Reworking of surface sediments by marine benthos |
| Box Corer (BC) | Type of seabed sampler (0.25m ² surface area) |
| BSL | Benthic Solutions Limited |
| Burdwood Bank | A large Bank approximately 200m in depth, part of the Scotia Arc projecting approximately 600km from Cape Horn 200km south of the Falkland Islands |
| CBD | Convention on Biological Diversity |
| CCAMLR | Convention on the Conservation of Antarctic Marine Living Resources |
| CITES | Convention on International Trade in Endangered Species |
| Clast | Large fragment of rock, typically pebble through to boulder in size |
| Drop stones | Drop stones are gravel sized rock found in sedimentary material deposited on the sea floor by glacial activity and ice rafting |
| EBS | Environmental Baseline Survey report |
| EHD | European Habitats Directive (92/43/EEC) |
| EOL | Camera end of line |
| Finback-1 | Proposed exploration well site |
| FISA | Falkland Island South Area, encompassing proposed Humpback-1 and Humpback-2 well location |
| FCO | Falkland Islands Constitution |
| HC | Hydrocarbons (saturates and polycyclic aromatics) |
| HCl | Hydrogen Chloride |
| HM | Heavy and trace metals |
| HSG | Headspace gas |
| Humpback-1 & 2 | Proposed exploration well site |
| Vinson | Historical well location and survey in the south of FISA |
| IMS | Industrial Methylated Spirit |
| IUCN Red List | International Union for the Conservation of Nature Red List of Threatened Species |
| <i>lebensspuren</i> | Evidence of life in the sediment (i.e. animal tracks, burrows etc.) |
| LPH | Liquid phase hydrocarbons |
| MDAC | Methane derived authigenic carbonate |
| MEA | Multi-lateral environmental agreements |
| MG3 | MG3 (Survey) Limited |
| MPOG | Microbial prospecting for oil and gas |
| MSL | Mean Sea Level |
| Natura 2000 | Habitats designated for protection under the European Habitats Directive |
| OSPAR | Oslo-Paris Commission |
| PSA | Particle size analysis |
| Red List Codes | Extinct (EX), extinct in the wild (EW), regionally extinct (RE), critically endangered (CR), endangered (EN), vulnerable (VU), near threatened (NT) or data deficient (DD). |
| SAC | Special Areas of Conservation |
| SOL | Camera start of line |
| SPA | Special Protection Areas |
| SS.SSa.OSa | Offshore circalittoral sand (UK JNCC Habitat Classification) |
| CR.LCR | Low energy circalittoral rock (UK JNCC Habitat Classification) |
| TOC | Total organic carbon |

EXECUTIVE SUMMARY

A regional environmental survey was carried out within the FISA prospect area of the South Falklands Basin for Noble Energy Falklands by MG3 (Survey) Limited and their subcontractor Benthic Solutions Limited (BSL) in December, 2013 to March, 2014. The FISA site is located to the east of the Falklands Islands, covers an area of approximately 80km x 70km and surrounds two proposed well locations (Humpback-1 and Humpback-2) located within the northeast of the survey area.

The water depth within the FISA survey area varied from approximately 950m to 1650m. A regional habitat assessment was undertaken using a combination of acoustic and environmental ground truthing techniques. This was reviewed onboard in order to interpret the potential for environmentally sensitive habitats within the survey area. Field ground truthing was undertaken using a combination of high resolution seabed imagery and seabed sampling using different sampling devices.

The survey data revealed four general seabed types within the FISA survey area. The dominant sediment type in the north and west was relatively homogeneous, slightly gravelly, silty sand associated with some Holocene sedimentary material over a glacial sediment. In the deeper water to the south and east, the proportion of gravels increased at the surface becoming a non-depositional, gravelly, silty sand. Separating these two areas in the central part of the survey area is a complex bathymetry indicative of an irregular escarpment running from the southwest to the northeast. The sediments here have been interpreted as steep-sloped exposures of low energy bedrock or a hard underlying formation. The remaining sediment type is represented by an intermediate habitat type, marking the transition from homogeneous sands to the rock outcrops. This is also the dominant sediment type along the top of the escarpment. This sediment lies at a depth of 1150m above another sub-cropping geological feature which runs in a northeast-southwest orientation across the length of the survey area.

Environmental baseline sampling undertaken within the FISA regional survey area involved the collection of seabed photography from 34 stations, with 26 stations further analysed for physico-chemical and biological macro-invertebrate properties. In line with the general habitat variations recorded across the survey area, the particle size analysis showed some variation in the different proportions of sands, fines (below 63µm) and gravels (>2mm) with sediments ranging from sandy mud and muddy sand through to a gravelly sand, but with the dominant sediment type comprising slightly gravelly muddy sand as denominated by the Folk characterisation (Folk, 1954). The mean particle size ranged from 45µm to 354µm, demonstrating the slightly elevated proportions of silts and clays at two sites and gravels at four sites. The mean proportion of gravels was typically around 7%, but elevated levels between 16% and 27.3% were recorded at the four stations.

Only two locations recorded a proportion of fines greater than 50%, with a peak of 66.7% recorded at station A/26/ENV, which is the same location as an earlier proposed Finback well prospect. The mean proportion of sediment fines was 32%. In FISA, the majority of softer sediments were recorded above the escarpment feature north and west of the proposed well locations.

The physico-chemistry results for total organic carbon (TOC), total hydrocarbon content (THC) and saturates all showed relatively low background concentrations. THC concentrations gave a mean of

3.1µg.g⁻¹, whilst alkanes recorded a mean of 313ng.g⁻¹, approximately 10% of the total THC recovered. Inspection of the individual gas chromatograms at all stations revealed no background fingerprints relating to anthropogenic or petrogenic sources. Analysis of alkanes revealed a predominance of marine related saturates indicative of an offshore site influenced by Antarctic water masses, whilst polycyclic aromatic hydrocarbons (PAHs) recorded only low concentrations of pyrolytic origin. A review, however, of the lighter 2-3 ring PAHs represented by naphthalene, phenanthrene and dibenzothiophene (NPD) and a higher proportion of alkylated PAHs throughout the full range of PAHs, suggests that the site is influenced by very small chromic input from petrogenic origin, albeit at a trace level. This could be the result of migrated thermogenic material relating to a natural hydrocarbon seep in the general vicinity. A similar phenomenon was recorded at neighbouring sites to the south and west as previously recorded within the region (Toroa and Burdwood Bank). Overall total PAH concentrations (2-6 compounds) recorded a mean of 82.8ng.g⁻¹ whilst the mean of the NPD fraction is 43.8ng.g⁻¹.

The concentrations of total heavy and trace metals were consistent with historical surveys recorded within the area. The distribution of the metals often correlated with themselves (with other metals) and occasionally other sediment factors. The most obvious of these was that of aluminium (Al) which exhibited a strong correlation with % gravels and sorting coefficient. Iron recorded a huge variability across the survey area ranging from 10.7mg.g⁻¹ to 135mg.g⁻¹, interpreted as a function of residual glacial materials of volcanic origin reworked by the currents in many areas. Arsenic (As) showed a close correlation with iron (Fe), albeit at a modest concentration ranging from 3.2µg.g⁻¹ to 12.3µg.g⁻¹. For cadmium (Ca) and mercury (Hg) only trace levels were recorded, whilst the means for lead (Pb), barium (Ba), chromium (Cr), nickel (Ni), copper (Cu), vanadium (V) and zinc (Zn) all gave background concentrations.

Multiple water column profiles and discrete water samples confirmed a consistent water column with limited vertical structure. Temperature varied from around 6.70°C at the surface to around 2.60°C at 1432m. A broad array of analytes relating to water chemistry (metals, nutrients, hydrocarbons etc.) at discrete depths throughout the water column generally recorded trace or undetectable concentrations at all sites and depths sampled.

Macrofaunal analysis was carried out on all 59 replicates obtained from the 26 baseline sediment sites. Macrofaunal taxonomy identified a total of 2,874 individuals and 254 species, with 188 comprising infauna, dominated by small polychaetes and crustaceans. In rank order, the top five key dominant species consisted of the polychaete *Rhamphobranchium (Spinigerium) ehlersi*, the crustacean *Ampelisca*, the polychaete *Gymnonereis fauveli*, followed by a Copepoda and the polychaete and *Apistobranchus* sp I. Whilst the deep water coral *Lophelia pertusa* was recorded in the samples, all incidents were of relic debris with no live animals recovered. Camera operations, however, did identify small live examples of this species, but of morphology insufficient to be considered as an Annex I habitat under the European Habitats Directive, or the OSPAR list of threatened habitats. Also of note from the macrofaunal samples were a group of solitary corals (*Flabellum* sp.) and a very rare mollusc of the group Monoplacophora (*Neopilina*). Only about 150 specimens have ever been found worldwide, belonging to about 25 species. Univariate analysis of the data showed that the species richness and abundance were similar to previous surveys in the area by surface area, whilst multivariate analysis indicated no clear separation of faunal populations, based on geographical location, sediment type or other environmental factors.

In addition to the infaunal community, the qualitative presence of the epifaunal community was reviewed from the seabed samples and photographed, and provided a rich faunal assemblage, especially in areas where drop-stones were common. Key faunal groups included sponges (classes Hexactinellida, Calcarea and Demospongiae) and Cnidaria which were represented by nine genera of thecate and athecate Hydrozoa and two genera of Stylasteridae. Furthermore, solitary corals belonging to the species *Flabellum curvatum* (a potential CITES Appendix II coral species) and several Octocorallia including the sea pens Pennatulidae and two species of Gorgonacea were identified. Bryozoa were also very common constituents on pebbles and stones, with many species endemic.

The remainder of the survey area did not yield any evidence of particularly sensitive habitats, especially within the immediate vicinity of the proposed exploration wells. The habitats in these areas were considered to be homogenous, gravelly or slightly gravelly silty sands with limited sensitivity to the proposed operations.

A review process was completed at the Natural History Museum (NHM) to assess nomenclature and species identification prior to the finalisation of the taxonomic species list included in this report. This is discussed in further detail in Appendix II, Section II.5.2.

Incidental to the environmental baseline datasets, the MG3 were contracted to conduct an additional wreck investigation within the FISA survey area using acoustic survey equipment already installed on the hull of the survey vessel. This was to locate the two First World War wrecks, *SMS Scharnhorst* and *SMS Gneisenau*, which were believed to lie within or in close proximity to this study area. However, after an extensive survey of approximately 30km x 45km no conclusive evidence was found to verify the locations of the wrecks. This is thought to be due to the limitations of the available survey equipment for these water depths and the uncertainty of the actual positions.

1 INTRODUCTION AND SCOPE OF WORK

1.1 GENERAL

In December 2013 Noble Energy Falklands Limited commissioned MG3 to carry out a field environmental survey over a regional area of the South Falklands Basin, encapsulating several prospect areas. This included analysis and interpretation of benthic sample data acquired during the field acquisition program. Field operations were undertaken from the MG3 managed Icelandic survey vessel *MV Poseidon*, using a combination of a deep water camera system and seabed sampling devices. Sampling was undertaken at a total of 34 stations for the FISA survey area relating to sites selected for ground truthing specific bathymetric features and to maintain a good geographical coverage over the prospect area.

1.2 SCOPE OF WORK

Sampling aimed to maximise the knowledge of the region as a whole, so as to provide a context for individual sites and habitats. This includes the proposed well locations Finback-1, Humpback-1 and Humpback-2 (sampled by station A/27/Env; Table 1.1 and Figure 1.1).

The scope of work was as follows:

- To review the bathymetry processed from the seismic 3D surface return over the 80km x 70km area of interest.
- To delineate and map potentially environmentally sensitive seabed features within the survey area. In particular, the presence of potential Annex I Habitat communities (i.e. biogenic reefs such as *Lophelia* and *Madrepora* corals, or cold water seep communities) as specified under the European Habitats Directive.
- To obtain baseline data for all habitat types recorded within close proximity to the proposed drilling operations (via benthic sampling and seabed imagery) in line with international requirements.
- To gain an understanding of the regional environmental conditions and habitats present at the proposed wells.

Environmental survey operations were undertaken following a review of the acoustic data, in order to provide an overall picture of the existing marine habitats. Seabed photography was used to ground truth all key seabed habitats identified by the acoustic datasets. Of particular interest were clear bathymetric features and, if recorded, potential Annex I habitats (European Union Habitats Directive and key sensitive habitats (OSPAR) along with a delineation of their boundaries, and the presence of possible IUCN red list or CITES Appendix II species. A minimum of five high resolution digital photographs were to be acquired at each location, accompanied by a video overview covering a larger seabed area.

Baseline sampling was undertaken based on seabed sampling using either box corer or grab sampler (subject to surface geology), providing detailed physico-chemical information and multiple macro-invertebrate replicates of 0.1m² over a 500µm mesh sieve.

1.2.1 Geophysical Survey

Shallow geophysics and bathymetric survey operations were carried out at selected target areas within the proposed survey polygon measuring approximately 80km x 70km. This additional information was overlaid onto the regional bathymetric chart and used to refine the location of ground truthing, or to resolve minor artefacts recorded in the 3D seismic data. The following acoustic equipment packages were used during the survey and reviewed as part of the habitat analysis.

- 3D surface return seismic data that covers the complete survey area. The grid resolution of this dataset was 25m x 25m. This gave excellent coverage of the proposed survey area, but some minor textures and patterns recorded within the data were the result of artefacts within the processing.
- Multibeam echo sounder data were reduced and processed offshore to provide a digital terrain model enabling major bathymetric features and minor bathymetric changes to be identified and highlighted. Although the resolution of the grid produced by the system in this moderate water depth was slightly lower than that of the seismic 3D data (i.e. 40m x 40m), the data was without the same artefacts as previously recorded. Typically, only a single short line was run over each prospective site thereby allowing only a limited resolution of the seabed close to the centre of the line. A larger search area (approximately 35km x 47km) was additionally surveyed to try and locate the position of the Sharnhorst World War I wreck (this was not found, MG3, 2014).
- Sub-bottom profiler of shallow soils was used to clarify changes that might be seen in the seismic and surface bathymetry. The survey was carried out using a hull-mounted 9-element pinger array, but the effectiveness of this system at these water depths was extremely limited and offered only marginal information to the interpretation in the field.

Considering the water depth, the multibeam data were of sufficient quality to adequately map regional seabed types and identify the presence of any anomalous features on the seabed, where present.

1.2.2 Environmental Survey Strategy

Priority targets were selected close to the proposed well centres to investigate key bathymetric features and potential habitat changes as well as to achieve geographical coverage of the area. Additional sample locations were distributed throughout the survey area in order to gain a regional understanding, and to investigate potentially environmentally significant features and habitat changes.

In summary the sampling locations were selected based on the following criteria:

- Proposed well locations.
- Bathymetric features of potential environmental importance (e.g. discrete reefs).
- Bathymetric features expected to represent a change in habitat.
- Slope features expected to show a gradient of habitat change.
- Bathymetric highs and lows within the survey area.
- Geographical coverage of the survey area.

Table 1.1 - Proposed Finback-1, Humpback-1 and Humpback-2 Well Locations

| Well Location | Easting | Northing |
|---------------|-----------|-------------|
| Finback-1 | 783,725mE | 4,207,331mN |
| Humpback-1 | 792,814mE | 4,213,086mN |
| Humpback-2 | 799,122mE | 4,207,171mN |

Projection: TM 60W; Spheroid/datum: WGS84

Environmental sampling was attempted at all 34 locations between the 31st December and 16th March 2014. Surveyed sites were generally based on combined seabed sampling and camera operations as summarised in Table 1.2 and shown in Figure 1.1. Table 1.2 also indicates the number of photographs acquired, the video duration and level of sampling for subsequent physico-chemical and biological determination at each site. These samples were processed and are reported in this Environmental Baseline Survey report.

The camera system used in the survey was based on a “Sea Bug” developed by Subsea Technology and Rentals (STR) deployed from a seabed frame. The system is based on a housed G10 Canon Camera which can output up to a 14.7 megapixel quality (5 megapixel used for the current project to reduce operational upload time). Further information is presented in Appendix I. Generally, the images were of good quality and simple to review. Some images were taken from a position above the seabed and therefore of reduced resolution (but showing a greater surface coverage), whilst most were taken from the grounded sled with a fixed oblique view of approximately 1m². All images are scaled by a laser. Whilst high resolution seabed photographs were acquired at regular intervals, occasional lower resolutions images were also obtained directly via screen grabs from the video footage between stills images.

In addition to the seabed photography, a total of 19 box core, 30 double Van Veen grabs and 11 Hamon grab deployments were completed, resulting in 59 successful samples and 18 ‘no sample’ attempts (mostly due to the sampler failing to trigger, see Appendix VI). The penetration of the successful box cores varied from 13cm to 35cm. This is indicative of a relatively compacted seabed which has a significant proportion of sands and coarser granular sediments. Retained material was processed onboard for both macro-

invertebrate communities and surface physico-chemistry with an additional small core retained from the centre of the box as a complete archive sample.

Vertical water quality profiles (temperature, depth, salinity) were obtained as well as water samples collected for set of water chemistry analysis (nutrients, heavy metals, TOC, etc.). Further details are shown in Appendix I/I.2.5 with results presented in section 2.8.

Sample positions, including those sampled for baseline purposes, are listed in Table 1.2, plotted in Figure 1.1, and detailed in Appendix VI. Detailed photographic positions are presented in Appendix VII.

1.2.2.1 *Sample Analyses*

The recovered benthic samples were correctly stored prior to demobilisation and transportation of the material to the correct laboratory. This involved the freezing of all physico-chemical samples on recovery and hand-carried back to the UK, to be forwarded to a laboratory remaining frozen at all times. This material was analysed at the following laboratories:

- BSL: Particle size Analysis
- BSL: Macro-invertebrate Analysis
- Environmental Scientifics Group (ESG/Chemtech): Sediment Chemistry

A second set of residual (backup) material is currently being held by BSL for future analysis or delivery to the authorities following completion of the survey submission. A summary of the analytical methodologies applied and accreditation is given in Appendix II and VIII.

1.2.2.2 *Environmental Data Presentation using Contouring Software*

To aid in the interpretation and presentation of the environmental information acquired for this report, both hydrographic and environmental variables were processed using contouring and 3D surface mapping software (Surfer v11). This software allows a digital terrain model (DTM), or grid, to be interpolated from irregularly spaced geographical information (XYZ data) using a kriging interpolation algorithm. When large quantities of data are used (such as in swathe bathymetry), the level of interpolation is limited only to small spaces in between the data points. However, when processing environmental variables a diagrammatic circle has been used to colour illustrate the parameter level at each relevant site (the size of this circle is diagrammatically determined to be colour coded based on the scale of the Figures presented in Section 2). It should be remembered that this is done for presentation purposes only and that these data values are “not representative” for the whole of the geographical area covered by the circle. No interpolation is required in this instance except where these circles overlap due to the scaling of the Figure.

Table 1.2 - Summary of Environmental Ground Truthing Locations and Acquisition

| Station | Summary Rationale | Type | Easting | Northing | Biological Samples* | Chemistry** | Depth (m) | No. of Photos | Video (mins) |
|------------|---|-------|---------|----------|---------------------|---|-----------|---------------|--------------|
| A/3/ENV | Regional sample, very slightly steeper slope from original stations 1 and 2 | BC | 736830 | 4188434 | Fa, Fb | Environmental chemistry (+ water chemistry) | 1134 | 27 | 14 |
| | | SOL | 736983 | 4188568 | | | | | |
| | | EOL | 737018 | 4189208 | | | | | |
| A/106/ENV | Move upslope to slightly raised linear feature (NE-SW trending) | SOL | 743566 | 4180145 | No samples | No samples | 1302 | 24 | 37 |
| | | EOL | 743518 | 4180147 | | | | | |
| A/1008/ENV | (Moved from A/8/Env) 200m transect N-S flank of escarpment | DG/HG | 784301 | 4202040 | Fa, Fb | Environmental chemistry | 1280 | 41 | 40 |
| | | SOL | 784327 | 4202063 | | | | | |
| | | EOL | 784323 | 4201775 | | | | | |
| A/9/ENV | Sequence through medium amplitude flat seabed feature | DG | 774052 | 4201351 | Fa, Fb, Fc | Environmental chemistry | 1260 | 27 | 31 |
| | | SOL | 774025 | 4201278 | | | | | |
| | | EOL | 774051 | 4201411 | | | | | |
| A/10/ENV | Sequence through medium amplitude flat seabed feature | DG | 774368 | 4205141 | Fa, Fb, Fc | Environmental chemistry | 1261 | 15 | 19 |
| | | SOL | 774451 | 4205142 | | | | | |
| | | EOL | 774412 | 4205133 | | | | | |
| A/1011/ENV | At deepest point of hollow feature | DG | 784810 | 4213180 | Fa, Fb | Environmental chemistry | 1325 | 17 | 26 |
| | | SOL | 784868 | 4213089 | | | | | |
| | | EOL | 784691 | 4213134 | | | | | |
| A/12/ENV | Regional sample, relatively flat seabed | DG/HG | 783794 | 4203737 | Fa, Fc | Environmental chemistry | 1280 | 22 | 25 |
| | | SOL | 783903 | 4203852 | | | | | |
| | | EOL | 783796 | 4203863 | | | | | |
| A/1013/ENV | Move into 'eroded' hole location, at deepest point | HG | 790680 | 4204135 | Fa, Fb | Environmental chemistry | 1350 | 27 | 31 |
| | | SOL | 790773 | 4204080 | | | | | |
| | | EOL | 790629 | 4204226 | | | | | |
| A/14/ENV | Edge of regional flat area | BC | 782519 | 4194777 | Fa, Fb | Environmental chemistry | 1316 | 15 | 23 |
| | | SOL | 782529 | 4194695 | | | | | |
| | | EOL | 782604 | 4194854 | | | | | |
| A/1015/ENV | Move to deepest point of hollow feature | BC | 786510 | 4198430 | Fa, Fb | Environmental chemistry | 1358 | 41 | 41 |
| | | SOL | 786596 | 4198429 | | | | | |
| | | EOL | 786309 | 4198453 | | | | | |
| A/18/ENV | High amplitude in low relief area | BC | 768352 | 4173956 | Fa, Fb | Environmental chemistry (+ water profile) | 1415 | 24 | 32 |
| | | SOL | 768169 | 4173962 | | | | | |
| | | EOL | 767871 | 4173444 | | | | | |
| A/20/ENV | General area coverage | DG | 791101 | 4195297 | Fa, Fb, Fc | Environmental chemistry (+ water profile) | 1350 | 18 | 33 |
| | | SOL | 791005 | 4195281 | | | | | |
| | | EOL | 791066 | 4195328 | | | | | |
| A/21/ENV | General area coverage | BC | 798155 | 4195368 | Fa, Fb | Environmental chemistry | 1430 | 31 | 28 |
| | | SOL | 798271 | 4195359 | | | | | |
| | | EOL | 798150 | 4195466 | | | | | |
| A/22/ENV | High amplitude base of slope | BC | 767851 | 4163126 | Fa, Fb | Environmental chemistry | 1522 | 27 | 26 |
| | | SOL | 768009 | 4163221 | | | | | |
| | | EOL | 768082 | 4163559 | | | | | |
| A/26/ENV | Close to proposed well location, (Finback-1) | DG | 783724 | 4207331 | Fa, Fb, Fc | Environmental chemistry | 1260 | 19 | 21 |
| | | SOL | 783858 | 4207372 | | | | | |
| | | EOL | 783861 | 4207366 | | | | | |
| A/27/ENV | Close to proposed well location, (Humpback-1) | DG | 792420 | 4213837 | Fa, Fb, Fc | Environmental chemistry | 1260 | 25 | 23 |
| | | SOL | 792430 | 4213799 | | | | | |
| | | EOL | 792415 | 4213974 | | | | | |

| | | | | | | | | | |
|------------|---|-------|--------|---------|------------|---|------|----|----|
| A/201/ENV | 200m transect perpendicular to scarp in feature 'hole' | SOL | 754240 | 4216058 | - | - | 1120 | 41 | 40 |
| | | EOL | 734621 | 4206717 | | | | | |
| A/202/ENV | Linked to previous station 5 to pick out base of undulation | BC | 754100 | 4216220 | Fa, Fb | Environmental chemistry | 1153 | 33 | 43 |
| | | SOL | 754227 | 4216098 | | | | | |
| | | EOL | 754403 | 4216019 | | | | | |
| A/203/ENV | 200m (S-N) camera transect on isolated feature | SOL | 787583 | 4199232 | - | - | 1321 | 37 | 26 |
| | | EOL | 787537 | 4199102 | | | | | |
| A/204/ENV | Linked to 17, possible slump | BC | 758950 | 4166435 | Fa, Fb | Environmental chemistry | 1302 | 50 | 27 |
| | | SOL | 758970 | 4166483 | | | | | |
| | | EOL | 758020 | 4166114 | | | | | |
| A/205/ENV | 500m transect SE-NW, camera only, possible discrete slope failure | SOL | 792365 | 4178244 | - | - | 1540 | 12 | 40 |
| | | EOL | 792218 | 4178377 | | | | | |
| A/206/ENV | Transect on discrete feature | SOL | 776822 | 4163141 | - | - | 1530 | 29 | 32 |
| | | EOL | 776881 | 4163576 | | | | | |
| A/207/ENV | Camera transect over discrete 'reef' area | SOL | 786707 | 4209650 | - | - | 1247 | 35 | 27 |
| | | EOL | 786584 | 4209771 | | | | | |
| A/208/ENV | Geographical coverage | SOL | 773650 | 4228268 | - | - | 1200 | 25 | 40 |
| | | EOL | 773740 | 4228376 | | | | | |
| A/301/ENV | 200m transect NW-SE slope feature, sample on slope | DG/HG | 790757 | 4214853 | Fa, Fc | Environmental chemistry | 1250 | 35 | 29 |
| | | SOL | 790827 | 4215011 | | | | | |
| | | EOL | 790806 | 4214965 | | | | | |
| A/302/ENV | Regional sample to be redefined by bathy pinger | DG | 804634 | 4210621 | Fa, Fb, Fc | Environmental chemistry | 1330 | 24 | 22 |
| | | SOL | 804638 | 4210663 | | | | | |
| | | EOL | 804617 | 4210571 | | | | | |
| A/303/ENV | 200m transect NW/SE to look at slope feature | DG | 796137 | 4211766 | Fa, Fb, Fc | Environmental chemistry | 1285 | 31 | 34 |
| | | SOL | 796077 | 4211873 | | | | | |
| | | EOL | 796114 | 4211842 | | | | | |
| A/304/ENV | Regional sample | DG | 783354 | 4221219 | Fa, Fb, Fc | Environmental chemistry | 1250 | 18 | 21 |
| | | SOL | 783322 | 4221128 | | | | | |
| | | EOL | 783265 | 4221188 | | | | | |
| A/305/ENV | Regional sample | BC | 778056 | 4210814 | Fa, Fb | Environmental chemistry (+ water profile) | 1267 | 19 | 32 |
| | | SOL | 778143 | 4210780 | | | | | |
| | | EOL | 777923 | 4211004 | | | | | |
| A/306/ENV | 200m transect across trench feature | HG | 796843 | 4206384 | Fa, Fb | Environmental chemistry | 1350 | 32 | 33 |
| | | SOL | 796895 | 4206287 | | | | | |
| | | EOL | 796885 | 4206287 | | | | | |
| A/307/ENV | Regional sample | BC | 793230 | 4187957 | Fa, Fb | Environmental chemistry | 1400 | 30 | 30 |
| | | SOL | 793335 | 4187995 | | | | | |
| | | EOL | 793204 | 4188035 | | | | | |
| A/308/ENV | 200m transect N-S slump at base of escarpment | BC | 784209 | 4192313 | No samples | No samples | 1380 | 26 | 24 |
| | | SOL | 784228 | 4192273 | | | | | |
| | | EOL | 784282 | 4192427 | | | | | |
| A/309/ENV | Transect NW-SE investigate base of trench | BC | 790971 | 4193442 | Fa, Fb | Environmental chemistry | 1395 | 20 | 27 |
| | | SOL | 791043 | 4193418 | | | | | |
| | | EOL | 790951 | 4193618 | | | | | |
| A/3010/ENV | Repeat sample of 2011 (8666.1 FOGL Vinson West EBS) | BC | 773473 | 4162976 | Fa, Fb | Environmental chemistry | 1540 | 20 | 27 |
| | | SOL | 773456 | 4162973 | | | | | |
| | | EOL | 773586 | 4163385 | | | | | |

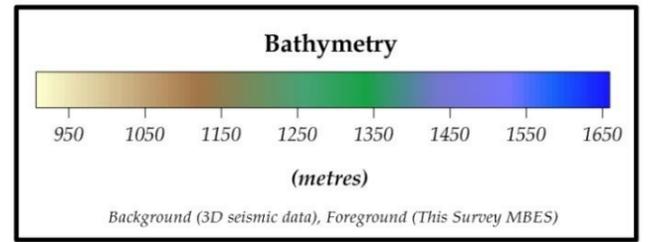
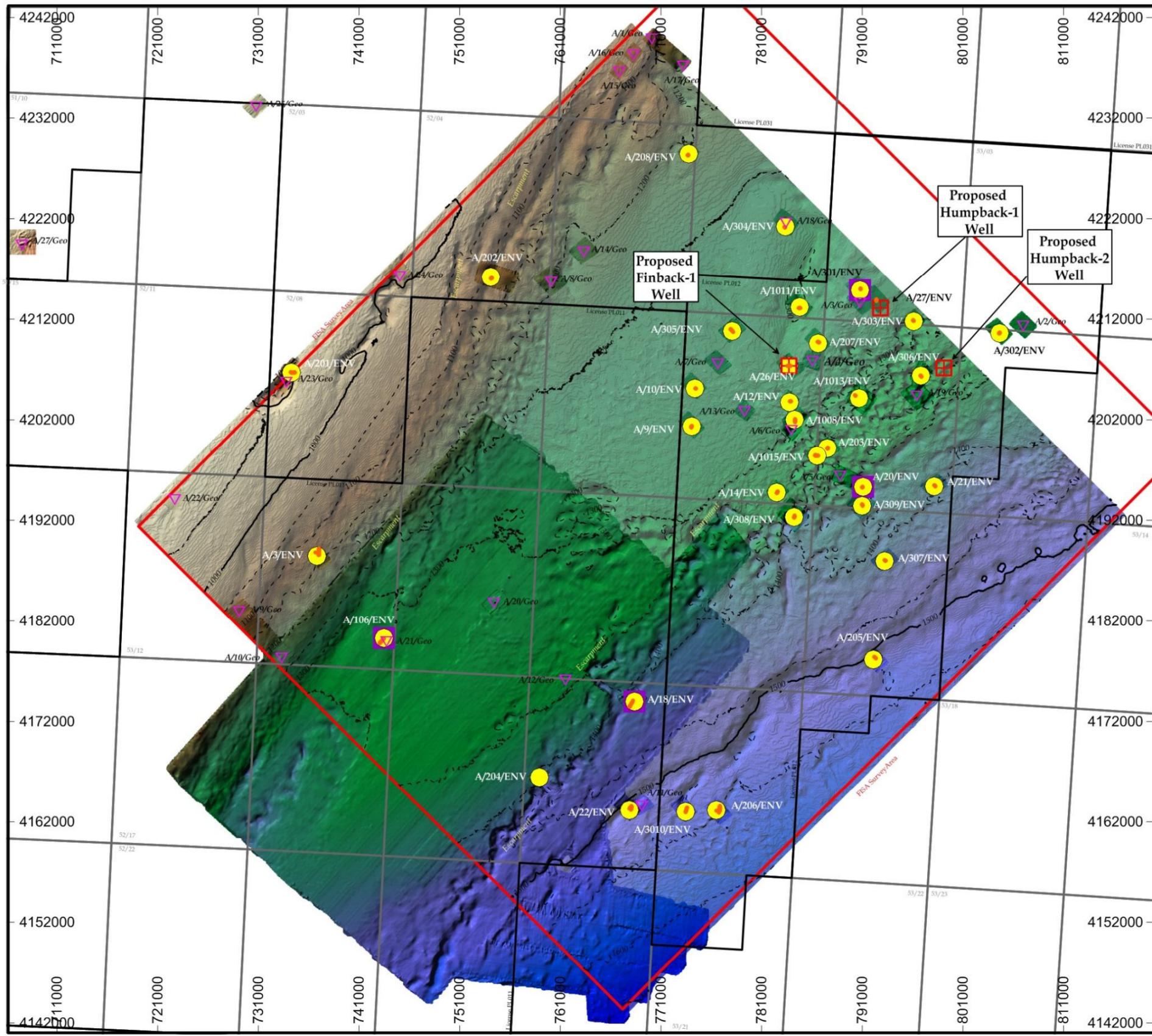
* Biological replicates Fa - Fc (0.1m² macro-invertebrate samples processed over 500µm)

** Environmental chemistry - PSA: particle size analysis; TOC: total organic carbon; HM: heavy and trace metals; HC: hydrocarbons (saturates and polycyclic aromatics),

BC = Box corer sampler; DG = Double grab sampler; HG = Hamon grab sampler; n.a. = not available due to beacon malfunctioning

SOL = camera start of line

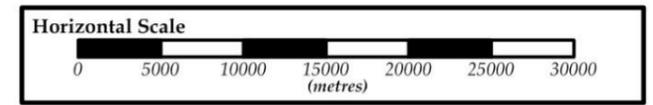
EOL = camera end of line



Legend

- Proposed Well Locations
- Environmental Sampling Location (Box Corer & Grab)
- Geophysical Sampling Location (Gravity Corer)
- Seabed Photographic Positions
- Water Quality Profiles

Surface Illumination
Horizontal 315°
Vertical 45°



Geodesy

Datum WGS84 (EPSG Code 6326), Spheroid WGS84 (EPSG Code 7030)
Projection Transverse Mercator, Central Meridian 60° West

CONTRACTOR

Benthic Solutions Limited, Elanco Works, Marsh Road, Hoveton, Norfolk NR12 8UH Tel: +44 (0) 1603 784726

CLIENT

Figure 1.1 Summary FISA Bathymetry and Sample Locations

| Rev | Date | Description | Drawn | Interp | Approved |
|-----|----------|--|-------|--------|----------|
| 0 | 12.08.14 | Bathymetry & Sample Locations for FISA | PRS | HL | IW |
| 1 | 20.08.14 | Bathymetry & Sample Locations for FISA | PRS | HL | IW |
| 2 | 18.09.14 | Bathymetry & Sample Locations for FISA | PRS | HL | IW |

1.3 GEODETIC PARAMETERS

| Project Geodesy | |
|------------------|----------------------------|
| Projection | Transverse Mercator CM 60W |
| Spheroid | WGS84 |
| Datum | WGS84 |
| Central Meridian | 60° West of Greenwich |
| False Easting | 500 000m |
| False Northing | 10,000,000m |

1.4 DATA COMPARISONS AND HISTORICAL DATASETS

1.4.1 Recent Surveys of Nearby Areas

Previous similar datasets have been recorded in earlier baseline surveys around the Falklands, in the North (Gardline, 1998), Eastern (Gardline, 2011a-f; Fugro 2009a-d) and South Falklands Basins (BSL, 2008). For the interpretation at FISA, data comparisons have been made from the previous surveys within the East Falklands Basin (Gardline, 2011b; Fugro 2009a and b). The FISA survey area has been partially surveyed previously and relates to the Vinson West site-survey location (Gardline, 2011b). All historical data acquired during these three earlier studies are displayed for comparison in parameter data tables (Section 2), as applicable (in blue). The station ENV 5 of the Vinson West study was coincidental to station A/3010/ENV of the present study and provides a direct comparison.

Sources used in this comparison are outlined below in Table 1.3 and plotted in Figure 1.2:

Table 1.3 - Historical Datasets Used for Comparison

| Source and Year | Reference of the report | Region and Comment |
|-----------------|---|---|
| Gardline 2011b | Block 60. Vinson West, Environmental Baseline Reference 8666.1. | Falklands: Vinson West site Inside the survey area |
| Fugro 2009a | Survey FIDA 61/05 Toroa. Environmental Baseline Reference 9763V3 | Falklands: Toroa site South-west of the survey area |
| Fugro 2009b | Survey FIDA 41/29 Nimrod. Environmental Baseline Reference 9763V5 | Falklands: Nimrod site North-east of the survey area |

Figure 1.2 FISA Sampling Locations and Historical Surveys in the Vicinity

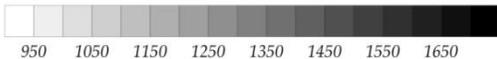
Legend

-  Proposed Well Locations
-  FISA Environmental Sampling Location
-  FISA Seabed Photography Only
-  Historical Toroa Location (2009)
-  Historical Nimrod Location (2009)
-  Historical Vinson West Location (2011)

Horizontal Scale (metres)

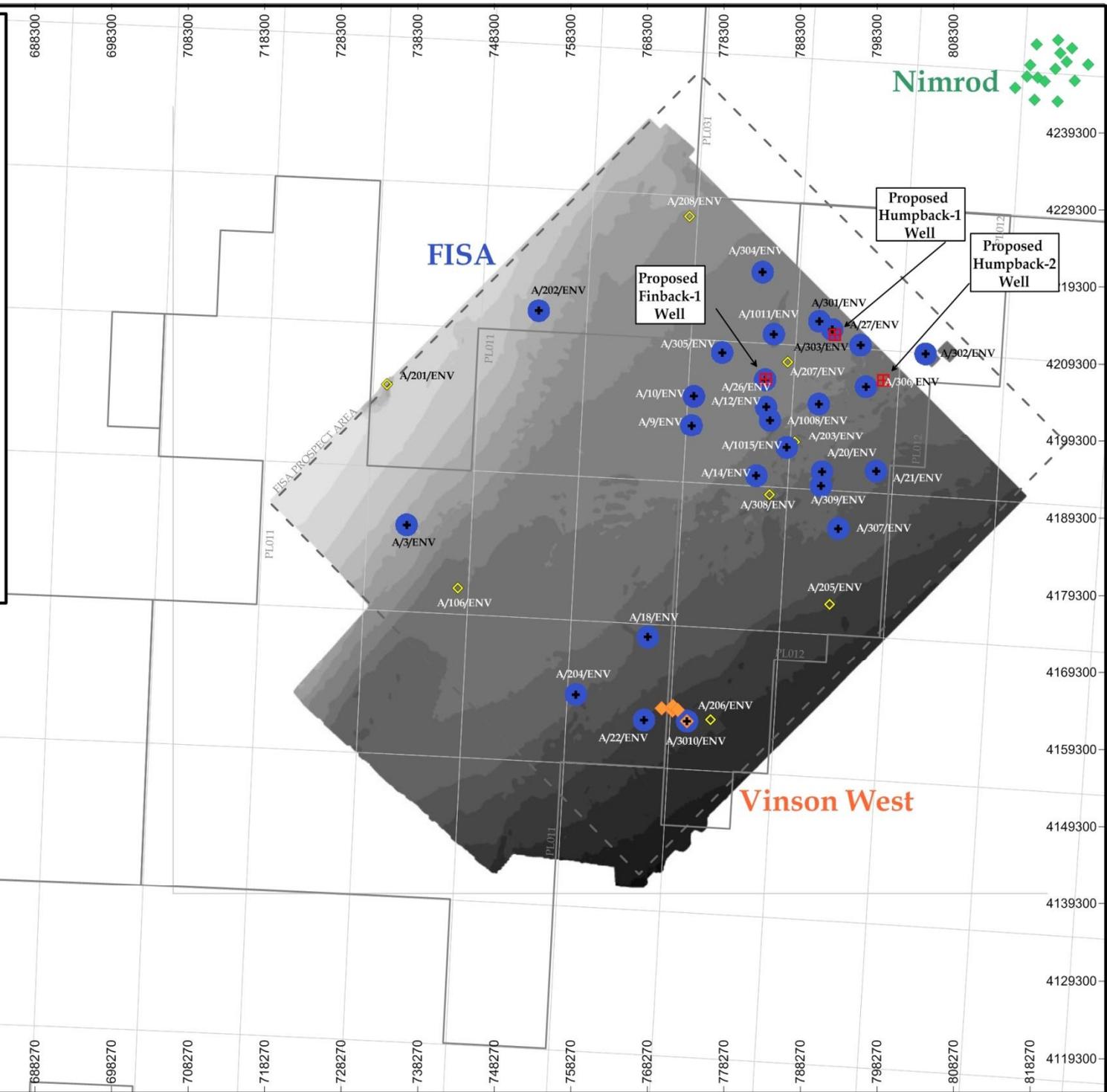


Background Bathymetry (metres)



Geodesy

Datum WGS84 (EPSG Code 6326), Spheroid WGS84 (EPSG Code 7030)
 Projection Transverse Mercator, Central Meridian 60° West



1.4.2 Regional Standards and Background Information

Details of the legislative background that apply to the Falkland Islands are provided in Volume 2 of this report (MG3, 2014b). This also includes details of the European environmental frameworks that are being applied to the habitats and species found around the islands based on the Oslo Paris (OSPAR) commission for threatened habitats, and the European Habitats Directive (92/43/EEC). Relevant IUCN Red list species (IUCN, 2013), the regulation of endangered species listed under the Convention on International Trade in Endangered Species (CITES), along with potential sensitive habitats, in particular relating to corals and sponge communities, are also discussed. All identification of Taxonomic Species has followed a review process with Natural History Museum (NHM) to ensure nomenclature and species identification follow a common standard for the region. This is discussed in further detail in Appendix II, Section II.5.2.

1.5 REPORTING STRUCTURE

Final reporting in the FISA area is separated into three volumes as follows:

- Volume 1 FISA Field Report.
- Volume 2 FISA Habitat Assessment Report.
- Volume 3 FISA Environmental Baseline Report (This Report).

2 RESULTS AND DISCUSSIONS

2.1 REGIONAL GEOLOGY AND SEABED FEATURES

The Falkland Islands are situated on an area known as the Falklands Plateau, separated to the north from the Argentine Basin by the Falklands Escarpment. This Falklands Plateau is characterised by a layer of fine to medium sand at the seabed, which may be up to 2m thick (Bastida *et al.*, 1992). Some areas are known to have a high percentage of gravel comprising either small pebbles or bioclasts with both gravels and sedimentary concretions recorded in earlier baseline surveys in the North (Gardline, 1998), Eastern (Gardline, 2011a-f; Fugro 2009a-d) and South Falklands Basins (BSL, 2008). The current survey area is located in an area east of the Falkland Islands, and north of the Burdwood Bank in the Falkland Plateau Basin. This Basin has a northeast to southwest trending western margin which is faulted and ends eastwards at the Maurice Ewing Bank which is a bathymetric high ~250km east of the islands (www.fig.gov.fk/minerals). The survey area has been partially surveyed previously, relating to the Vinson West well location (Gardline, 2011b). All of the previous work in the Falklands is available in the public domain from the FIG web site.

The Falkland Plateau Basin is essentially a margin basin that comprises Jurassic and Cretaceous passive margin-type rocks similar to those found in the South Falkland Basin. To the southwest, along the deep water trough known as the Falkland Chasm, the basin merges with the South Falkland Basin. This basin lies beneath 200m to 2500m of water and exhibits numerous major normal faults probably of Jurassic through Early Cretaceous age, contemporaneous with the deposition of sediments in the actively rifting trough. Some of the thrust faults, however, display reactivation into recent times (www.fig.gov.fk/minerals).

The prevalence of surface hard-bottom areas is not accurately known. Features recorded along the edge of several escarpments recorded in the bathymetry and the evidence provided from sampling and photography suggests that an irregular sedimentary material exists over the majority of the site along with some exposures of hard surfaces. Seabed sampling recorded a relatively inconsistent recovery of material and required three types of sampler, sometimes with multiple deployments, to acquire sufficient material for analysis. Whilst the gradient of the seabed was expected to be generally gradual within the survey area, localised bathymetric features such as the escarpments, hummocks, and rock outcrops may be encountered which will exhibit associated minor relief.

2.2 SURVEY BATHYMETRY

Bathymetry for the FISA survey area is summarised in Figure 1.1. This was produced from a combination of data collected during an earlier 3D seismic acquisition campaign (background data in Figure 1.1) and localised survey footprints acquired using multibeam echo sounder during the current survey (foreground data in Figure 1.1). For the most part, the latter dataset provides a more accurate representation of the bathymetry in the area due to removal of processing artefacts, despite its slightly lower resolution.

Water depths have been reduced to Mean Sea Level (MSL). More detailed representations of seabed sample photographs, as well as camera tracks and photographs are presented in Appendix VII.

The water depth for the proposed Finback-1, Humpback-1 and 2 well locations was approximately 1280m, 1270m and 1330m MSL. Depths across the survey area ranged from ~950m in the northwest, decreasing sharply to a maximum depth of ~1650m found in the southeast. The gradient within the well area was generally 2°, but this increased to around

2.3 SURVEY SURFACE GEOLOGY AND SEABED FEATURES

The seabed was variable throughout the FISA survey area, with the sediment typically comprising of silty SAND with varying levels of gravel, cobbles and boulders. To the northwest and southeast of the survey area, sediments were fine with varying levels of black basalt sands. Running through the centre of the survey area from northeast to southwest was a significant escarpment with associated slumps. Here, coarse material was abundant with exposed cobbles and boulders of glacial origin recorded, often at the base and shoulders of a steep slope. Biological debris was common throughout the escarpment area, especially that of the stony coral *Lophelia* sp. (order Scleractinia) which was found in small, but quite dense mats in discrete areas of the seabed (as evident at stations A/203/ENV, A/308/ENV, A/1008/ENV, A/1013/ENV and A/204/ENV). Station A/204/ENV indicated a *Lophelia* debris-field at the base of the escarpment in the southern part of the survey area. Incidents of live aggregations of hard corals were generally limited to only very small colonies 0.1m in area, with the largest 'thicket' of coral limited to 0.5m in size.

All of the fines observed were light grey to black in colour, whilst the sands were generally black, volcanic (i.e. basalt) in origin. This is generally associated with ice rafted material derived from islands in the Southern Ocean area and has been regularly recorded in sediments on previous surveys in the East Falklands Basin (Gardline, 2011b & d; Fugro 2009d).

Evidence from seabed photography, in particular the video data, shows that the escarpment features are the result of exposure by discordant formations displaying varying rates of erosion. The edge of these escarpments was shown to be capped with harder bedrock which are undercut by currents weathering older and softer substrates, eventually resulting in a collapse. This results in a well-defined escarpment edge in places, and the presence of a boulder field at the base of the slope. These features are discussed in further detail later in this report (Section 2.10.2).

2.4 PARTICLE SIZE DISTRIBUTION

The particle size interpretation of sediments across the survey area were based upon observations made from the acoustic data, seabed photography and the analytical results acquired from the surface sediment at 26 locations (Figure 2.1). Material for PSA analysis was recovered from the surface 5cm.

Sample stations were established at selected sites around the FISA survey area based on bathymetric, geological and spatial preferences as outline in Section 1.2.2. At a general level, the nature of the sediment was predominantly silty SAND with varying levels of gravel, cobbles and boulders.

On average, the sediment composed of these three main fractions:

- Sand: mean 60.31% ± 12.42 SD (standard deviation)
- Fine: mean 32.69% ± 10.74 SD
- Gravel: mean 7.01% ± 6.77 SD

The % proportions of these different sediment types for each station are listed in Table 2.1. These ranged from sandy mud and muddy sand through to gravelly sand, but with the dominant sediment type represented by slightly gravelly muddy sand as denominated by the Folk characterisation (Folk, 1954; Appendix III).

Analytical results were variable around a predominantly sandy seabed. Most distributions were either uni-modal, peaking around the medium SAND to fine SAND with a consistent tail of fines varying through silts and clays, or bi-modal with a slightly elevated proportion of coarse silts, and/or gravels. The mean particle size is outlined in Table 2.1, which ranged from 45µm to 354µm (Figure 2.1), demonstrating the varying proportions of silts, clays and gravels recorded around a consistent sand profile. A comparison of the full particle size distribution is presented in Figure 2.3. The majority of samples indicated similar distributions with representative size classes dominated by medium sands at around 15-20%, gravels below 5% and silts and clays between 4-6%. Outliers to this were stations A/1015/ENV, A/26/ENV, A/12/ENV and A/22/ENV which contained elevated levels of gravel, whilst stations A/3/ENV and A/26/ENV both recorded elevated fines. This particle size variation has also resulted in some variability in the sorting coefficient for the size distributions (Table 2.1), which varied from a poorly sorted 1.97 at A/309/ENV to an extremely poorly sorted 6.08 at A/26/ENV. The majority of stations, however, were generally very poorly sorted ranging from 2.3 to 2.9 based on a uni-modal distribution (albeit with an extended tail of fines).

Table 2.1 - Summary of Surface Particle Size Distribution

| Station | Mean Sediment Size | | Sorting | Skewness | Kurtosis | % Fines | % Sands | % Gravel |
|--------------------------|--------------------|-------------|-------------|-------------|-------------|--------------|--------------|-------------|
| | mm | Phi | | | | | | |
| A/304/ENV | 0.080 | 3.65 | 2.27 | 0.47 | 0.86 | 37.79 | 60.95 | 1.26 |
| A/303/ENV | 0.125 | 3.00 | 2.52 | 0.57 | 0.97 | 29.26 | 67.01 | 3.74 |
| A/09/ENV | 0.143 | 2.81 | 2.80 | 0.29 | 1.32 | 28.73 | 62.78 | 8.49 |
| A/204/ENV | 0.082 | 3.61 | 3.12 | 0.29 | 1.04 | 40.03 | 52.61 | 7.36 |
| A/202/ENV | 0.088 | 3.51 | 2.60 | 0.47 | 0.78 | 40.47 | 59.38 | 0.15 |
| A/22/ENV | 0.245 | 2.03 | 4.06 | -0.08 | 1.78 | 26.49 | 55.66 | 17.85 |
| A/20/ENV | 0.131 | 2.93 | 2.31 | 0.59 | 1.04 | 26.34 | 71.91 | 1.75 |
| A/12/ENV | 0.224 | 2.16 | 3.61 | 0.12 | 1.43 | 26.47 | 57.55 | 15.98 |
| A/21/ENV | 0.214 | 2.23 | 2.33 | 0.23 | 2.30 | 17.27 | 73.70 | 9.03 |
| A/1008/ENV | 0.141 | 2.82 | 2.56 | 0.57 | 0.96 | 27.87 | 71.18 | 0.95 |
| A/1011/ENV | 0.117 | 3.10 | 2.48 | 0.46 | 0.97 | 30.57 | 68.27 | 1.16 |
| A/3/ENV | 0.045 | 4.48 | 2.68 | 0.06 | 0.78 | 56.11 | 42.69 | 1.20 |
| A/307/ENV | 0.160 | 2.65 | 2.89 | 0.30 | 1.44 | 25.32 | 62.95 | 11.73 |
| A/18/ENV | 0.149 | 2.75 | 2.58 | 0.45 | 1.06 | 27.25 | 68.93 | 3.82 |
| A/1013/ENV | 0.094 | 3.41 | 3.18 | 0.22 | 1.12 | 37.93 | 49.45 | 12.62 |
| A/3010/ENV | 0.148 | 2.76 | 2.39 | 0.52 | 1.05 | 25.87 | 70.42 | 3.71 |
| A/301/ENV | 0.110 | 3.18 | 2.44 | 0.51 | 0.96 | 31.14 | 66.02 | 2.84 |
| A/14/ENV | 0.144 | 2.80 | 2.66 | 0.43 | 1.13 | 27.64 | 66.69 | 5.67 |
| A/27/ENV | 0.086 | 3.54 | 2.47 | 0.45 | 0.82 | 38.52 | 60.40 | 1.09 |
| A/306/ENV | 0.098 | 3.36 | 2.87 | 0.31 | 1.19 | 33.65 | 59.99 | 6.36 |
| A/305/ENV | 0.073 | 3.77 | 2.60 | 0.31 | 0.86 | 42.79 | 55.44 | 1.78 |
| A/10/ENV | 0.104 | 3.27 | 2.88 | 0.29 | 1.09 | 35.77 | 57.44 | 6.80 |
| A/309/ENV | 0.167 | 2.58 | 1.97 | 0.56 | 1.65 | 20.07 | 78.89 | 1.05 |
| A/1015/ENV | 0.354 | 1.50 | 4.13 | -0.03 | 0.80 | 27.63 | 45.06 | 27.30 |
| A/302/ENV | 0.177 | 2.50 | 2.73 | 0.27 | 1.85 | 22.18 | 65.88 | 11.94 |
| A/26/ENV | 0.084 | 3.58 | 6.08 | -0.58 | 2.62 | 66.71 | 16.76 | 16.53 |
| Mean | 0.14 | 3.00 | 2.89 | 0.31 | 1.23 | 32.69 | 60.31 | 7.01 |
| StDev | 0.07 | 0.64 | 0.83 | 0.26 | 0.47 | 10.74 | 12.42 | 6.77 |
| %Variance | 47.7 | 21.3 | 28.6 | 83.3 | 38.3 | 32.7 | 20.6 | 96.7 |
| Vinson West ENV 5 | 0.19 | 2.36 | - | - | - | 17.3 | 80.3 | 2.4 |
| Vinson West Mean | 0.19 | 2.35 | - | - | - | 18.4 | 76.9 | 4.7 |
| Vinson West SD | 0.03 | 0.22 | - | - | - | 2.9 | 4.5 | 4.3 |
| Nimrod Mean | 0.18 | 2.51 | 2.31 | - | - | 22.6 | 71.3 | 6.2 |
| Nimrod SD | 0.04 | 0.30 | 0.32 | - | - | 3.9 | 5.4 | 4.0 |
| Toroa Mean | 0.31 | 5.00 | 1.54 | - | - | 77.8 | 22.2 | 0.0 |
| Toroa SD | 0.02 | 0.08 | 0.01 | - | - | 1.9 | 1.9 | 0.0 |

Historical data are reported in blue

The distribution of coarser gravel sediment sizes (granules above 2mm in size or Phi -1) is presented geographically in Figure 2.3. Coarse sediments were recorded in all samples analysed ranging from 0.15% to 27.3%, with a mean of 7% (Table 2.1 and Figure 2.3). Four of the stations, A/12/ENV, A/22/ENV, A/1015/ENV and A/26/ENV revealed a significantly higher proportion of gravels ranging from 16% to 27.3%, indicative of mixed sediments routinely recorded within the survey area. Most of this material relates to ice-rafted glacial material in the form of drop-stones, although some weathered formations were also present, as described previously.

Figure 2.1 - Particle Size Comparison

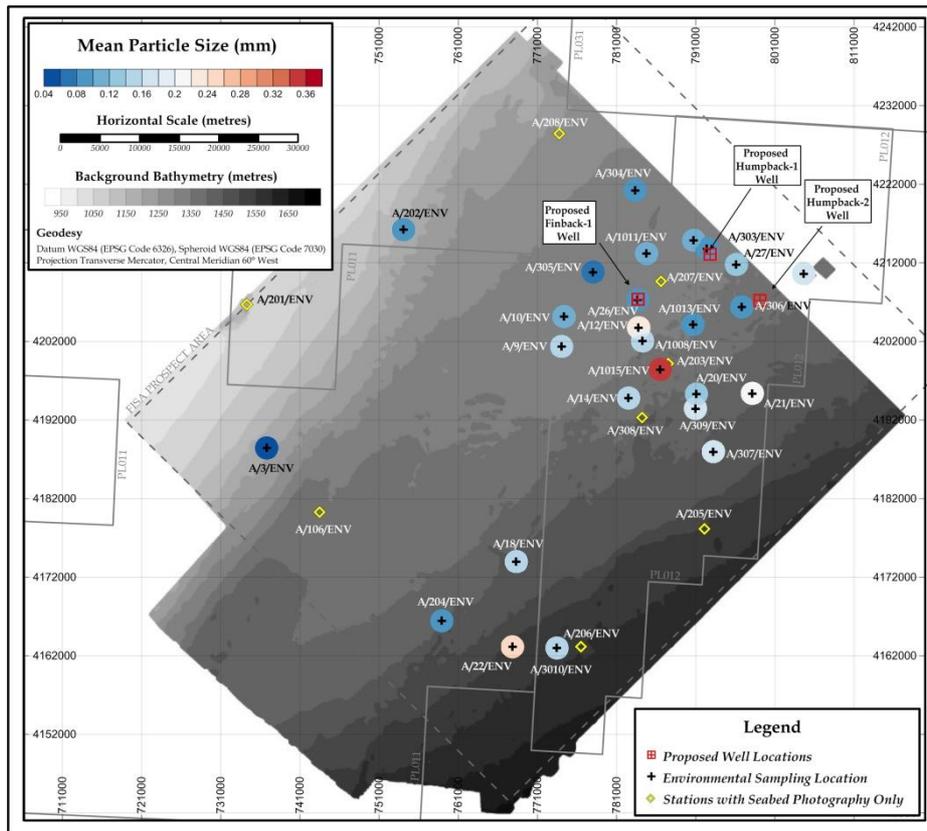
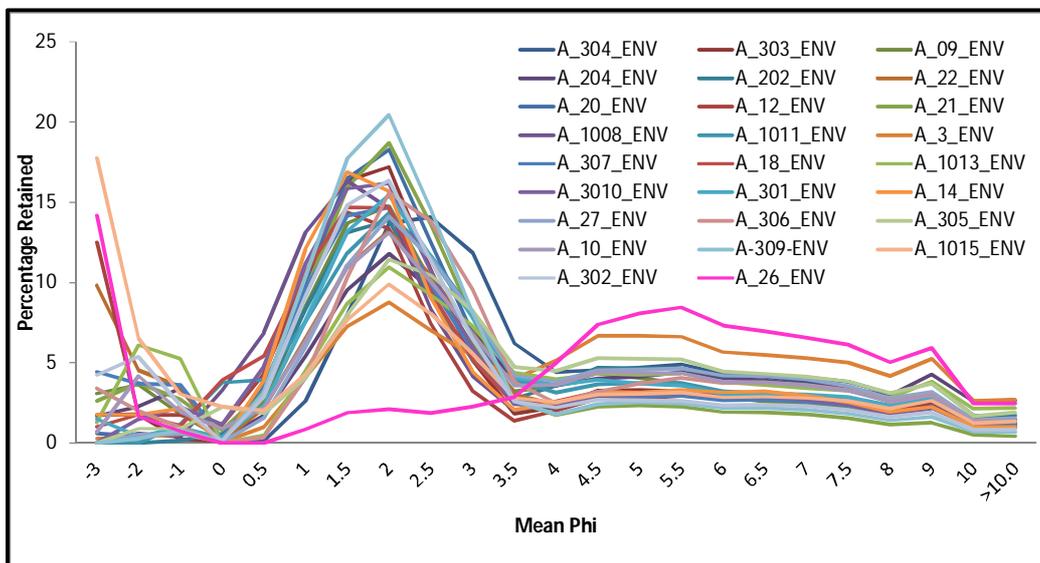


Figure 2.2 - Particle Size Distribution Comparison



Where mixed sediment occurs, the sampling process itself can be biased against coarser material, where only a relatively small sub-sample (approx. 250g) is recovered for analysis or where the efficiency of the sampler may be compromised by coarser sediments. The presence of larger clasts (cobbles and boulders) is well documented for many of the sites from the seabed photography, as is the requirement to use different samplers in order to recover suitable material for subsequent analysis. This means that the data obtained from

the particle size analysis should be treated with some caution and should only be used as an indication of the softer granular material populated by the macro-invertebrate communities in these areas, or used in combination with seabed photography where larger gravels are observed. The variability of gravels is readily identified in the residues from biological processing over a 500µm mesh (coarse sand) size and summarised in Appendix VII.

The geographical distribution of sediment fines (i.e. silts and clays <63µm), is shown in Figure 2.4. This shows a broad pattern of distribution with decreasing fines with increasing water depth ($P < 0.01$, Appendix XI), with the exception of station A/26/ENV, which indicated a peak 66.7% fines compared to a mean of 32% recorded elsewhere throughout the survey area. This pattern is considered unusual for a deep-water environment where, typically, the deeper waters indicate higher rates of pelagic sedimentation due to decreased water movement. This is clearly not the case at FISA with the majority of softer sedimentary material recorded above the escarpment feature north and west of the proposed well locations. Photographic evidence from sites around the escarpment and to the south and east, recorded rippled bedforms and sediment scour, both indicators of strong seabed currents. Station A/26/ENV indicated a slight aberration to other sediment distributions (Figure 2.2) with a very high proportion of fines, but almost equal proportion of fines and gravels (approximately 16.5%). The reason for this is not known, but must relate to a localised feature of the sediment that was not observed in the seabed photography.

A review of sediment types from the photographic evidence from both baseline and geological sample sites confirmed that the majority of the shallower sediments in the northwest were dominated by a relatively homogeneous and featureless slightly gravelly silty sand, associated with a weak sedimentary environment (i.e. slow sedimentation rate) throughout. The deeper sediments to the southeast were associated with a similar sediment type, but with greater erosion (reduced fines) and a significant gravel component, indicative of residual ice modification from drop stones. Separating these two areas in the central part of the survey area is a complex bathymetry indicative of an irregular escarpment running from the southwest to the northeast. The sediments here have been interpreted as steep sloped exposures of low energy bedrock or a hard underlying formation. The remaining sediment type was represented by an intermediate habitat type, marking the transition from homogeneous sands to the rock outcrops. This is essentially where the homogeneous sand thins to a veneer and is punctuated by occasional to numerous isolated rock outcrops, cobbles and boulders. This is the dominant sediment type along the top of the escarpment and at a depth of 1150m and runs in a northeast-southwest orientation across the length of the survey area. Further information on coarse sediment clasts has been provided from the seabed camera system and presented in Section 2.10.1, with the results of habitat variability presented in Section 2.10.2.

Figure 2.3 - Percentage of Gravel (%)

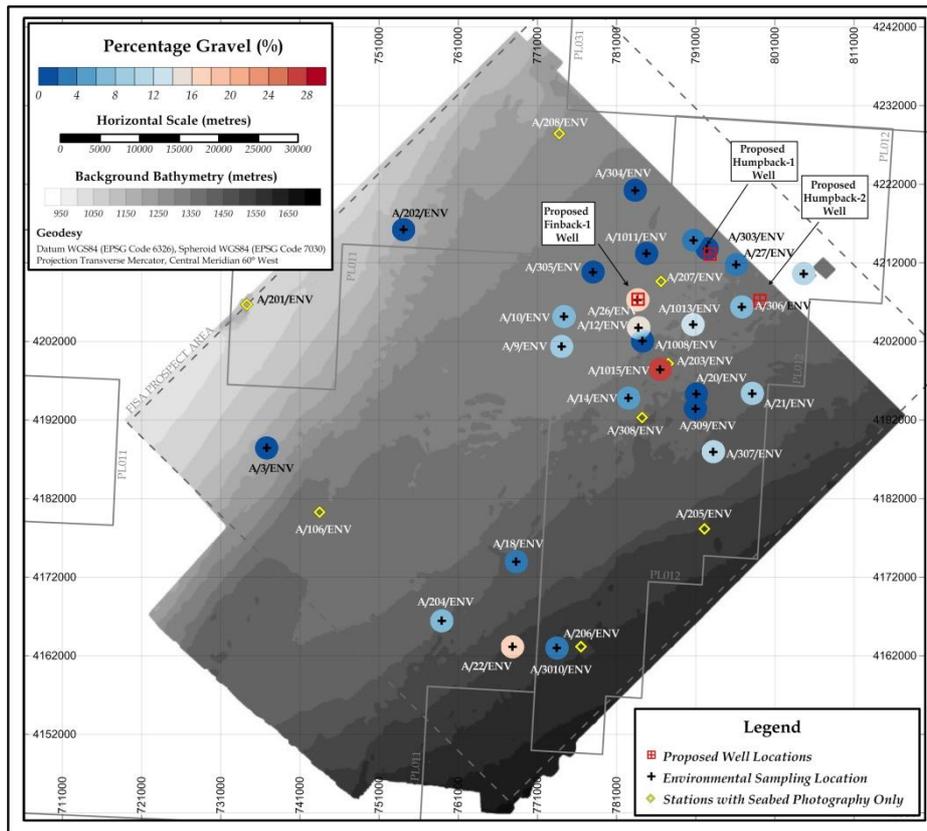
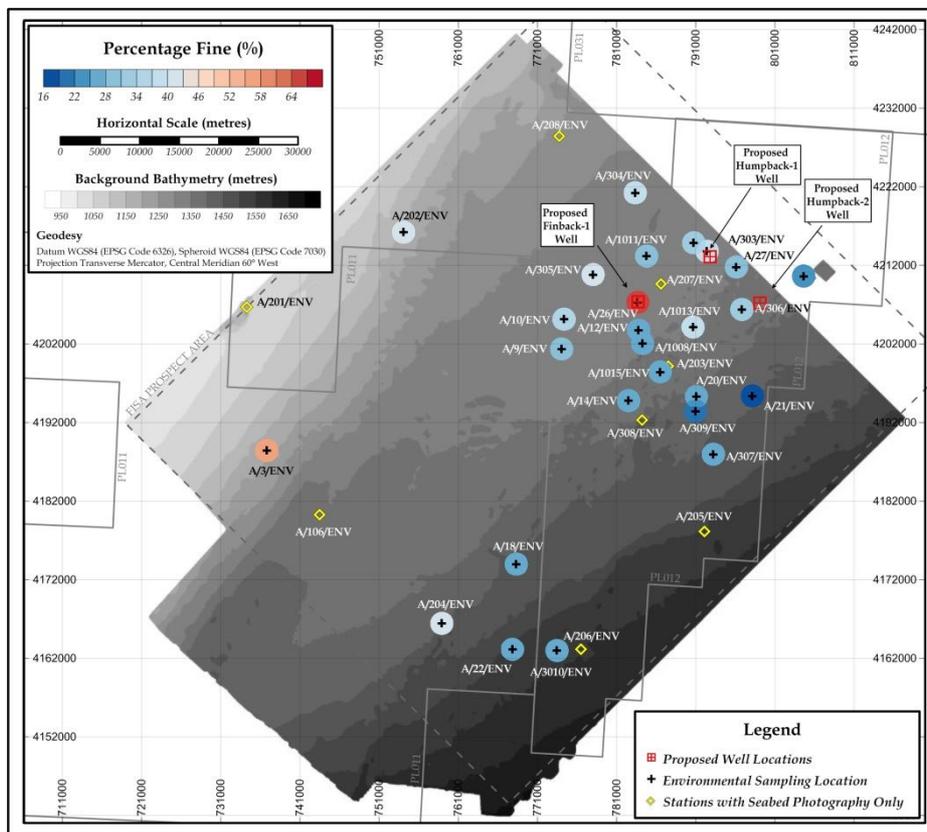


Figure 2.4 - Percentage of Fine (%)

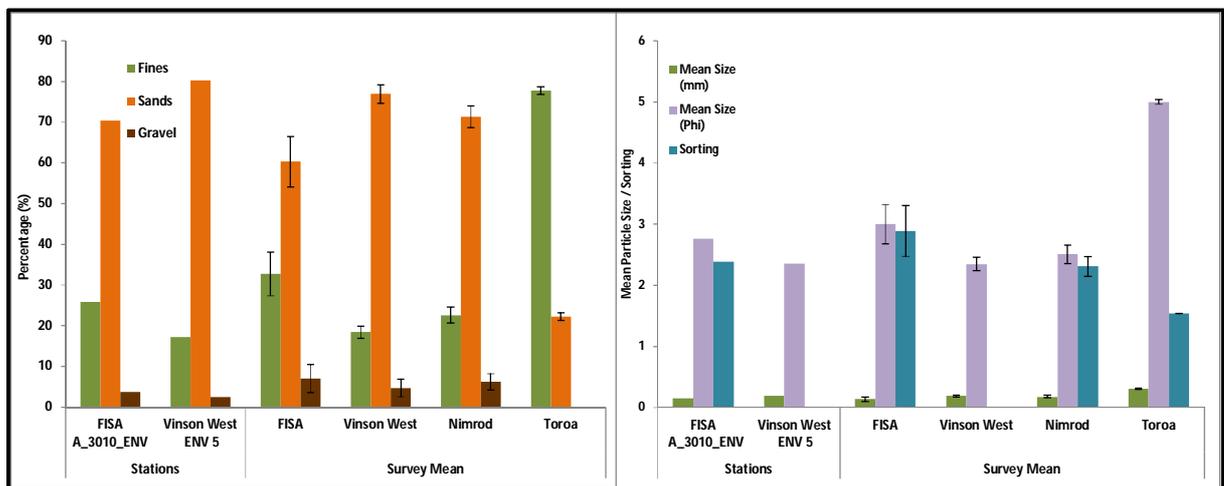


2.4.1 Historical Comparison with Particle Size Distribution

Whilst generally, relatively little is known about the sediments surrounding the Falklands, a number of historical environmental surveys have been undertaken in the South Falklands Basin, indicating similar results to those recorded at FISA. The FISA sample station A/3010/ENV is coincidental to the Vinson West survey (station ENV 5) providing a direct comparison between the two survey years (2009 and 2014). Figure 2.5 showed that both stations recorded approximately the same percentage of fine, sand, gravel and mean particles size (mm and Phi). The same figure also compares the average data with the additional surveys for Nimrod, Toroa as well as Vinson West. From this it is apparent that FISA and Vinson West recorded similar values and gave no significant indication for major changes in regards to particle size between 2009 and 2014.

Previous surface sediments analyses at the Toroa sampling locations, approximately 125km southwest of the present location, showed significantly finer sediments with a higher proportion of silts and clays of around 77.8%, compared to 32.7% recorded in the present study. This variation between sites was underpinned by a regional variation in bathymetry and possibly oceanography. Toroa was located in much shallower waters (approximately 650m) on the upper part of the continental slope and well above the complex bathymetry recorded around the escarpments in the present study (>1350m). The proportion of sands, at 22.2% and absence of coarser admixtures (such as gravels) at Toroa, were both much lower than found at FISA, reflecting the general trend of reduced sedimentation and greater erosion with increased water depth.

Figure 2.5 - Particle Size Comparison with Historical Data



Note sorting coefficient not available for Vinson West.

2.5 TOTAL ORGANIC CARBON

Sediments were analysed for TOC and moisture content and presented in Table 2.2. TOC is less susceptible to interference from other combustible components and represents the proportion of biological material and organic detritus within the substrates. Sometimes it can be recorded using crude combustion techniques, such as total organic matter by loss on ignition. TOC in surface sediments is an important source of food for benthic fauna (Snelgrove and Butman, 1994), although too much can lead to reductions in species richness and abundance due to oxygen depletion. Increases in TOC may also reflect increases in both physical factors (i.e. fines) and common co-varying environmental factor (through elevated sorption on increased sediment surface areas; Thompson and Lowe, 2004).

Table 2.2 - Summary of Moisture and Organic Carbon

| Station | Total Organic Carbon (% w/w) | Moisture Content (% w/w) |
|--------------------------|------------------------------|--------------------------|
| A/09/ENV | 0.11 | 28.1 |
| A/12/ENV | 0.21 | 21.8 |
| A/14/ENV | 0.17 | 26.9 |
| A/18/ENV | 0.23 | 26.6 |
| A/20/ENV | 0.15 | 23.2 |
| A/202/ENV | 0.22 | 30.0 |
| A/204/ENV | 0.15 | 33.1 |
| A/21/ENV | 0.19 | 21.3 |
| A/22/ENV | 0.24 | 24.2 |
| A/26/ENV | 0.40 | 37.5 |
| A/27/ENV | 0.21 | 25.0 |
| A/3/ENV | 0.32 | 33.5 |
| A/301/ENV | 0.24 | 26.2 |
| A/302/ENV | 0.15 | 22.2 |
| A/304/ENV | 0.17 | 30.8 |
| A/305/ENV | 0.17 | 28.5 |
| A/306/ENV | 0.24 | 23.3 |
| A/307/ENV | 0.23 | 23.4 |
| A/10/ENV | 0.32 | 28.4 |
| A/1008/ENV | 0.21 | 27.0 |
| A/1011/ENV | 0.28 | 29.1 |
| A/1013/ENV | 0.27 | 36.1 |
| A/1015/ENV | 0.31 | 28.6 |
| A/3010/ENV | 0.11 | 27.4 |
| A/303/ENV | 0.20 | 24.5 |
| A/309/ENV | <0.1 | 20.0 |
| Mean | 0.21 | 27.18 |
| StDev | 0.07 | 4.47 |
| %Variance | 34.2 | 16.5 |
| Vinson West ENV 5 | 0.37 | - |
| Vinson West Mean | 0.41 | - |
| Vinson West SD | 0.04 | - |
| Nimrod Mean | 0.27 | - |
| Nimrod SD | 0.02 | - |
| Toroa Mean | 0.73 | - |
| Toroa SD | 0.05 | - |

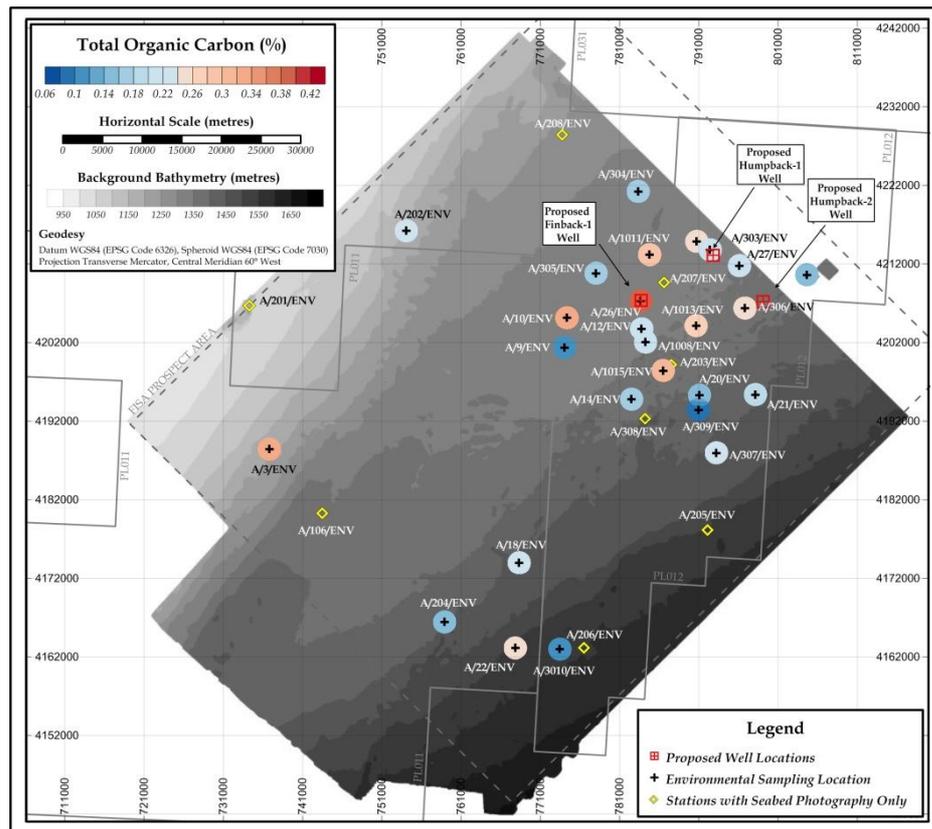
Historical data are reported in blue

The TOC results were low and reasonably consistent (Figure 2.6), ranging from <0.1% to 0.4% (mean $0.21 \pm 0.07SD$). Overall, these sediments can be considered to be organically poor. The data revealed no spatial pattern of distribution. Station A/26/ENV showed the highest percentage of TOC (0.4%), which also recorded the highest level of %fines (66.7%). This underlined a strong correlation with the proportion of sediment fines ($P < 0.01$, Appendix XI) recorded across the FISA area.

Overall, this level could be considered to be a limiting factor that may influence the distribution and abundance of some infaunal species within the substrates, in particular the deposit feeders within the sediments. TOC is expected to reflect autochthonous material and may give a broad measurement of a low carbon flux in this area due to relatively low nutrient enrichment of the water mass and limited terrestrial influences.

In addition to total organic carbon, the sediments were also analysed for moisture content. The moisture content remained consistent (mean $27.18 \pm 4.47SD$), indicative of similar texture and consolidation throughout; although slightly higher values were observed at stations A/26/ENV (37.5%) and A/1013/ENV (36.1%).

Figure 2.6 - Total Organic Carbon (TOC)



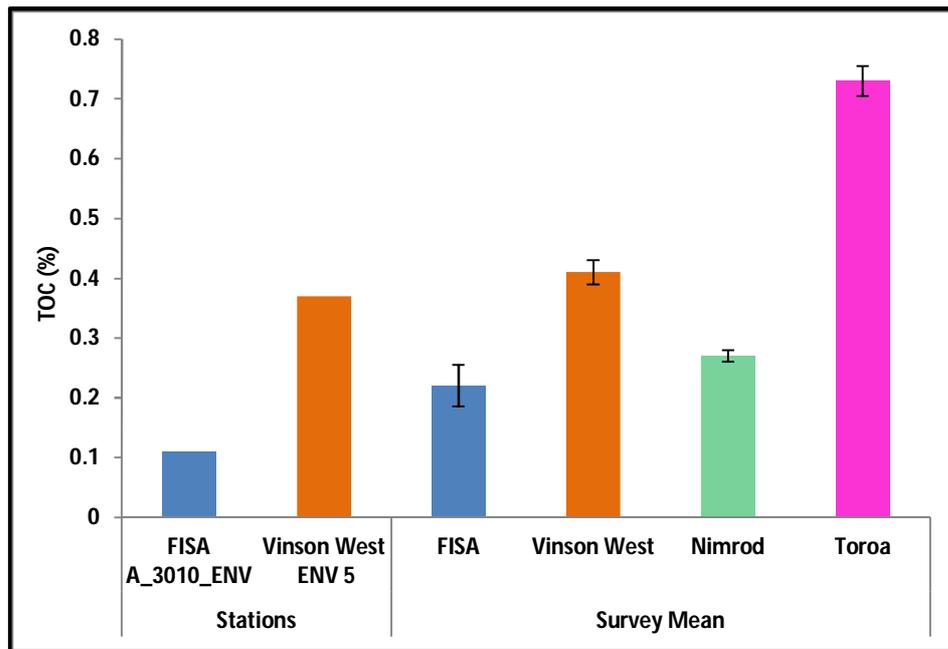
2.5.1 Historical Comparison with Total Organic Carbon

Overall, mean TOC levels in the FISA area can be compared to the data recorded previously in the region during similar baselines surveys at Vinson West ($0.41 \pm 0.04SD$) and Nimrod

($0.27 \pm 0.02SD$; Figure 2.7). This shows that TOC was slightly variable in this area between 2009 and 2014. The Toroa area revealed a higher mean value of $0.73 \pm 0.05SD$, corresponding to the finer sediment type previously recorded in this area. This was consistent with the particle size analysis (section above), as the mean percentage of fines (77.8%) at Toroa was about two to three times higher than measured at the other comparison sites.

The trend recorded in mean TOC was further reflected in the direct comparison between station A/3010/ENV on FISA and Vinson West ENV 5 which were also noticeable different (0.11% and 0.37%, respectively). This would suggest that the benthic population and deposition of organic material into the sediments, typically through detrital deposition from the water column (usually from autochthonous sources), varied between survey years. This was slightly lower in the current survey than previously recorded in 2011. As both surveys were undertaken in similar months of the year, the reason for this inconsistency is unclear.

Figure 2.7 - Total Organic Carbon (TOC) with Historical Data



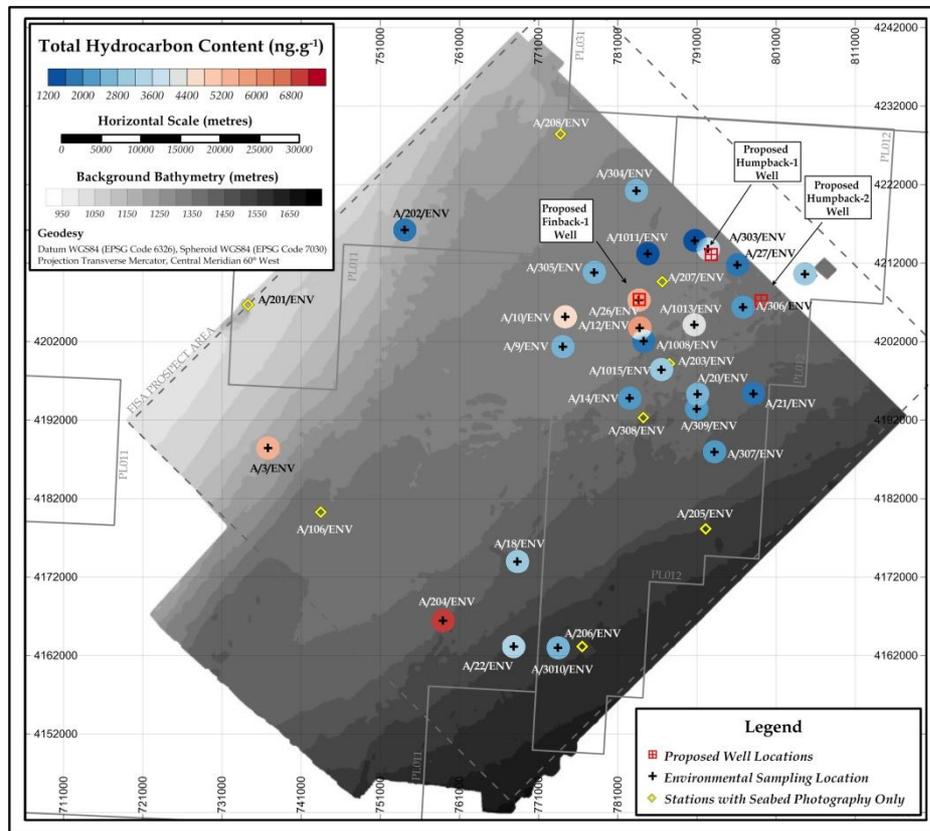
2.6 SEDIMENT HYDROCARBONS

2.6.1 Total Hydrocarbons Concentrations

Results for hydrocarbon analysis are summarised and tabulated as total hydrocarbon concentrations, total n-alkane and homologue ratios in Table 2.3, with individual alkanes (nC_{10} - nC_{37}) listed in Table 2.4. An example of a gas chromatogram showing the aliphatic hydrocarbon traces can be seen in Figure 2.11 (with the remaining gas chromatograms

presented in Appendix IV for each station), labelled with every fourth n-alkane, the isoprenoid hydrocarbon, pristane and phytane, along with the internal standards hepta-methylnonane (A), deuterated hexadecane (B) and 1-chlorooctadecane, (C).

Figure 2.8 - FISA Sediment Hydrocarbon Content



The THC concentrations of the sediments, measured by integration of all non-polarised components within the GC trace, showed generally low levels but slightly higher concentrations recorded at some stations. The data ranged from 1.23 $\mu\text{g.g}^{-1}$ to 7.12 $\mu\text{g.g}^{-1}$ (Table 2.3 and Figure 2.8). The mean for the whole of the survey area was 3.10 $\mu\text{g.g}^{-1}$ ($\pm 1.50\text{SD}$) but the variability between sites was quite noticeable, with a percentage variance (standard deviation over the mean) of 48%. The highest value of 7.1 $\mu\text{g.g}^{-1}$ was recorded at station A/204/ENV which was in close proximity to the escarpment feature. This variability cannot be explained by a geographical pattern of distribution (such as water depth) although THC negatively correlated with the % fines ($P < 0.05$, Appendix XI). These can be compared to a mean of 2.9 $\mu\text{g.g}^{-1}$ recorded in similar water depth west of the British Isles (AFEN, 1996).

Table 2.3 - Summary Hydrocarbon Concentrations

| Station | THC (µg.g ⁻¹) | Total n-alkanes (ng.g ⁻¹) | Carbon Preference Index | Pristane/phytane Ratio | Proportion of Alkanes (%) | Total PAHs (ng.g ⁻¹) | NPD (ng.g ⁻¹) |
|--------------------------|---------------------------|---------------------------------------|-------------------------|------------------------|---------------------------|----------------------------------|---------------------------|
| A/09/ENV | 2.5 | 189 | 1.37 | 10.3 | 7.50 | 53.53 | 28.96 |
| A/12/ENV | 5.8 | 435 | 1.24 | 3.5 | 7.51 | 102.85 | 52.88 |
| A/14/ENV | 2.2 | 208 | 1.25 | 11.3 | 9.47 | 56.73 | 29.77 |
| A/18/ENV | 3.1 | 324 | 1.23 | 18.7 | 10.46 | 88.86 | 46.42 |
| A/20/ENV | 2.6 | 246 | 1.25 | 3.4 | 9.36 | 68.09 | 37.41 |
| A/202/ENV | 1.6 | 155 | 1.30 | 5.5 | 9.46 | 37.73 | 22.11 |
| A/204/ENV | 7.1 | 731 | 1.22 | 5.7 | 10.27 | 207.65 | 108.08 |
| A/21/ENV | 1.8 | 273 | 1.48 | 6.0 | 15.13 | 74.94 | 37.36 |
| A/22/ENV | 3.3 | 306 | 1.21 | 5.3 | 9.32 | 82.32 | 40.34 |
| A/26/ENV | 5.3 | 355 | 1.25 | 2.3 | 6.76 | 105.83 | 55.93 |
| A/27/ENV | 4.0 | 401 | 1.34 | 5.1 | 10.04 | 103.85 | 53.35 |
| A/3/ENV | 5.4 | 510 | 1.30 | 3.4 | 9.44 | 147.35 | 80.28 |
| A/301/ENV | 1.2 | 155 | 1.29 | 6.3 | 12.50 | 35.18 | 21.02 |
| A/302/ENV | 2.9 | 253 | 1.31 | 16.7 | 8.71 | 71.06 | 37.49 |
| A/304/ENV | 2.7 | 230 | 1.34 | 16.8 | 8.47 | 60.21 | 31.72 |
| A/305/ENV | 2.5 | 352 | 1.46 | 4.4 | 14.04 | 91.01 | 46.03 |
| A/306/ENV | 2.1 | 282 | 1.42 | 5.6 | 14.65 | 57.21 | 31.21 |
| A/307/ENV | 2.2 | 241 | 1.03 | 18.1 | 10.97 | 59.49 | 32.52 |
| A/10/ENV | 4.7 | 461 | 1.38 | 11.2 | 9.84 | 138.09 | 72.24 |
| A/1008/ENV | 1.9 | 234 | 1.29 | 8.9 | 12.27 | 63.84 | 33.35 |
| A/1011/ENV | 1.4 | 297 | 1.45 | 6.7 | 12.02 | 39.88 | 23.08 |
| A/1013/ENV | 4.4 | 423 | 1.36 | 12.7 | 9.70 | 118.88 | 65.76 |
| A/1015/ENV | 3.1 | 393 | 1.43 | 13.2 | 12.79 | 108.79 | 57.94 |
| A/3010/ENV | 2.4 | 272 | 1.16 | 12.5 | 11.15 | 76.12 | 38.88 |
| A/303/ENV | 1.9 | 179 | 1.33 | 9.9 | 9.19 | 40.26 | 22.63 |
| A/309/ENV | 2.4 | 237 | 1.29 | 13.2 | 9.95 | 62.21 | 31.96 |
| Mean | 3.10 | 313.14 | 1.30 | 9.38 | 10.13 | 82.77 | 43.80 |
| StDev | 1.50 | 128.24 | 0.09 | 4.92 | 1.94 | 39.35 | 20.40 |
| %Variance | 48.3 | 44.4 | 7.3 | 52.4 | 19.2 | 47.5 | 46.6 |
| AFEN* | 2.90 | 660 | 1.8 | 2.5 | 23.4 | 179 | 72 |
| Vinson West ENV 5 | 3.5 | 228 | 1.4 | 3.6 | 6.51 | 68 | 30 |
| Vinson West Mean | 3.7 | 162 | 1.5 | 4.3 | 5.21 | 51 | 24 |
| Vinson West SD | 2.0 | 40 | 0.1 | 0.6 | 2.42 | 10 | 4 |
| Nimrod Mean | 3.7 | 310 | 0.99 | 3.35 | 8.31 | 70 | 55 |
| Nimrod SD | 0.3 | 50 | 0.05 | 0.55 | 1.11 | 13 | 12 |
| Toroa Mean | 8.7 | 650 | 1.12 | 4.27 | 7.55 | 224 | 166 |
| Toroa SD | 1.1 | 90 | 0.13 | 0.64 | 0.23 | 18 | 19 |

Historical data are reported in blue

* Average for deep water sediment (>1000m, AFEN, 1996).

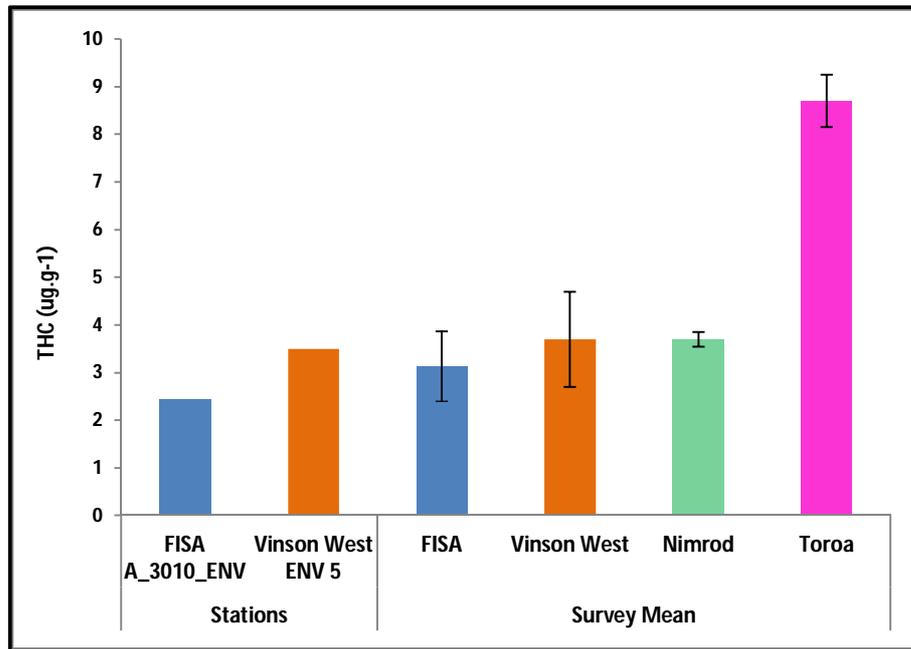
2.6.1.1 Historical Comparison with Total Hydrocarbon Concentration

The historical survey at the Vinson West prospect indicated an insignificantly higher THC value (mean 3.7±2.0SD; Figure 2.9) than recorded at FISA. A further comparison with the Nimrod site showed similar results and is indicative of a generally homogeneous THC level within this sediment type (i.e. gravelly silty sands), albeit that a slightly elevated standard deviation was recorded during these earlier studies. A further direct comparison between Vinson West

ENV5 and A/3010/ENV further revealed a similar elevation during the earlier survey ($3.5\mu\text{g.g}^{-1}$ compared to $2.4\mu\text{g.g}^{-1}$ respectively). This is a minor change between the two surveys. The total hydrocarbon level for the Toroa survey area was quite different which had a higher mean value of $8.7\pm 1.1\mu\text{g.g}^{-1}$. These results were in accordance with the high percentage of fine in this area, indicative of higher sedimentation and stronger assimilation of this hydrocarbon component in the surface sediments. As with TOC, this is likely to be a result of increased sediment fines (77.8%) at this shallower site.

The presence of hydrocarbons in the sediments is due to a range of predominantly natural sources. The likely source of this material is discussed in greater details in Section 2.6.2.

Figure 2.9 - Comparison of Sediment Hydrocarbon Content with Historical Data



2.6.2 Saturate/Aliphatic Hydrocarbons

All of the sample stations were analysed for n-alkanes using gas chromatography with flame ionisation detection (GC-FID). The results are summarised in Table 2.3 and individually listed in Table 2.4, which gives a breakdown of consecutive n-alkane content from nC₁₀ through to nC₃₇, together with the isoprenoid hydrocarbons Pristane (Pr) and Phytane (Ph).

Similar to THC, the total n-alkane concentrations were low overall, ranging from 155ng.g^{-1} to 731ng.g^{-1} (mean $313\pm 0.128\text{SD}$; Figure 2.10). The overall concentration of alkanes typically made up around 10% of the total THC recovered. This is quite low and is expected for uncontaminated marine sediments where background hydrocarbons are continuously being replenished by a low but chronic source of alkanes, usually from allochthonous sources.

Figure 2.10 - Total Saturate Alkanes

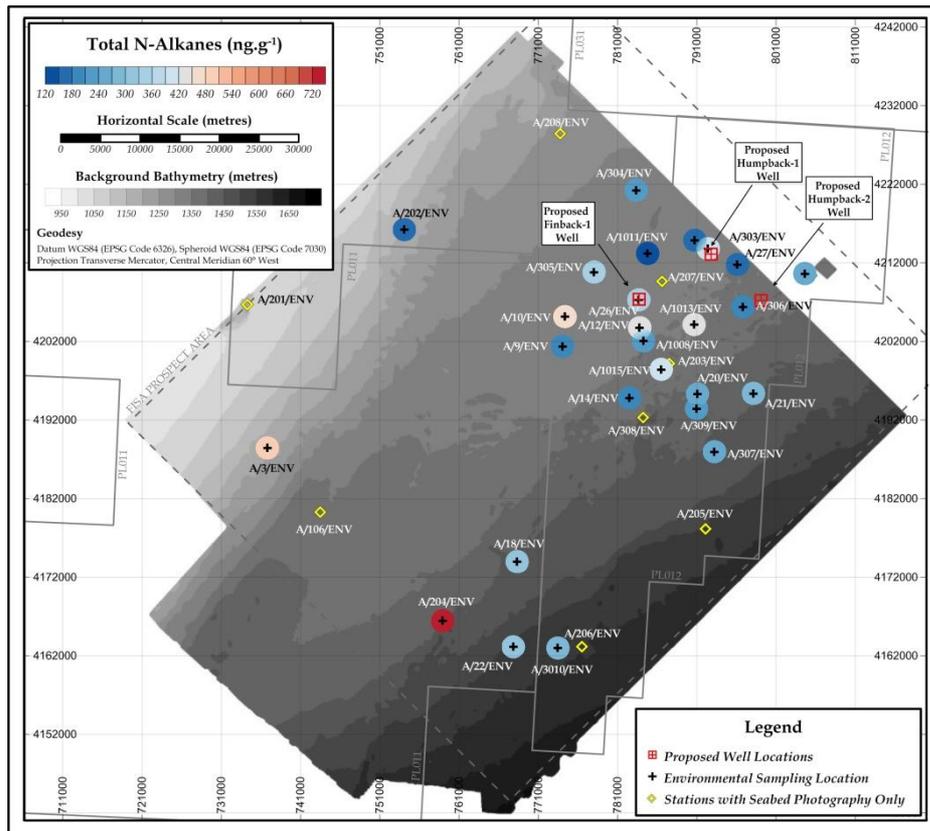
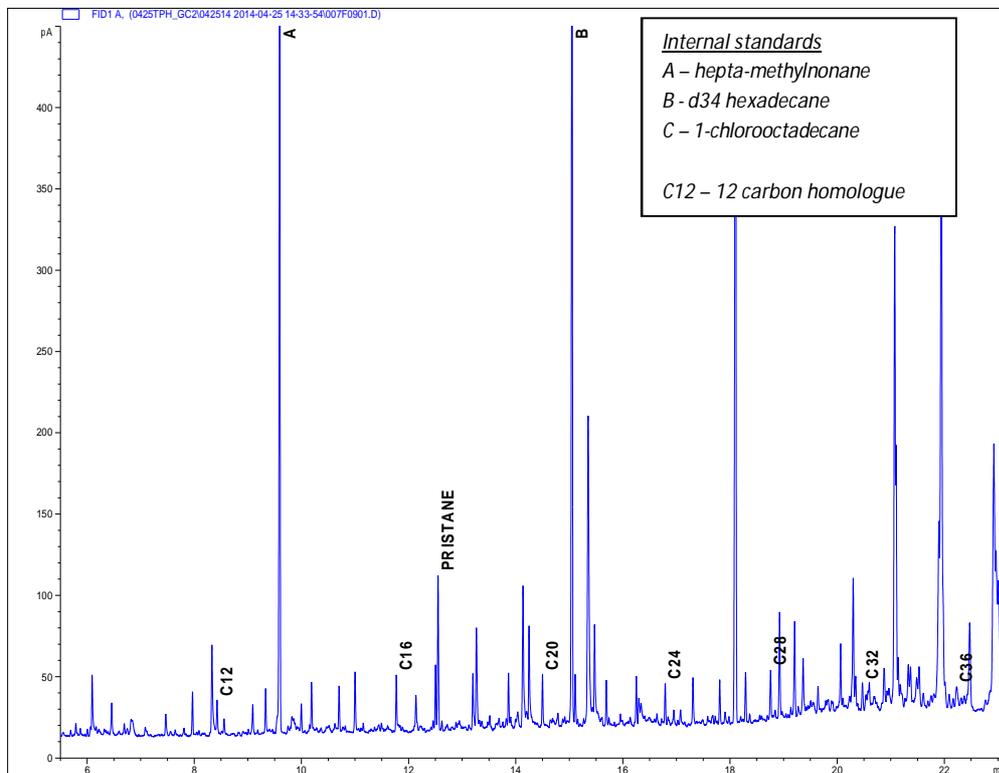


Figure 2.11 - Example Gas Chromatograms for Saturate Hydrocarbons Analysis (A/12/ENV)



Inspection of the individual gas chromatograms at all stations (Figure 2.11 and Appendix IV) indicated similar forms with little or no trends seen, other than those of natural background alkanes recorded in sediments of this type and region. Chromatograms showed a consistent homologous series of saturates throughout the range, but without a significant background “noise”, with the exception of station A/12/ENV (Figure 2.11) and A/204/ENV (Appendix IV), albeit at a relatively low level. All stations can be considered as typical examples of this deep water sedimentary environment.

A closer review of the different proportions of n-alkanes recorded can sometimes identify trends within the data or the source from which the different organic components derive. Even though the overall level of saturates is extremely low, the following ratios were further reviewed:

Carbon Preference Index (CPI)

The carbon preference index (CPI), is associated with the preference of biogenic n-alkanes (i.e. that of a preference for odd-carbon numbered homologues, particularly around nC₂₇₋₃₃; Sleeter *et al.*, 1980), derived from fatty acids, alcohols, esters and land plant waxes. The CPI was calculated for all stations and was very consistent ranging from 1.03 to 1.48 (mean 1.30±0.09SD) for the full saturate range (nC₁₀- nC₃₇; Table 2.3 and Figure 2.12). Results show only a very small dominance by terrigenous biogenic compounds although it is not clear if this is all allochthonous in nature. This is expected for an offshore environment where the impacts from terrestrial organic sources and plant matter are extremely limited.

Figure 2.12 - Hydrocarbon Analysis – Carbon Preference Index

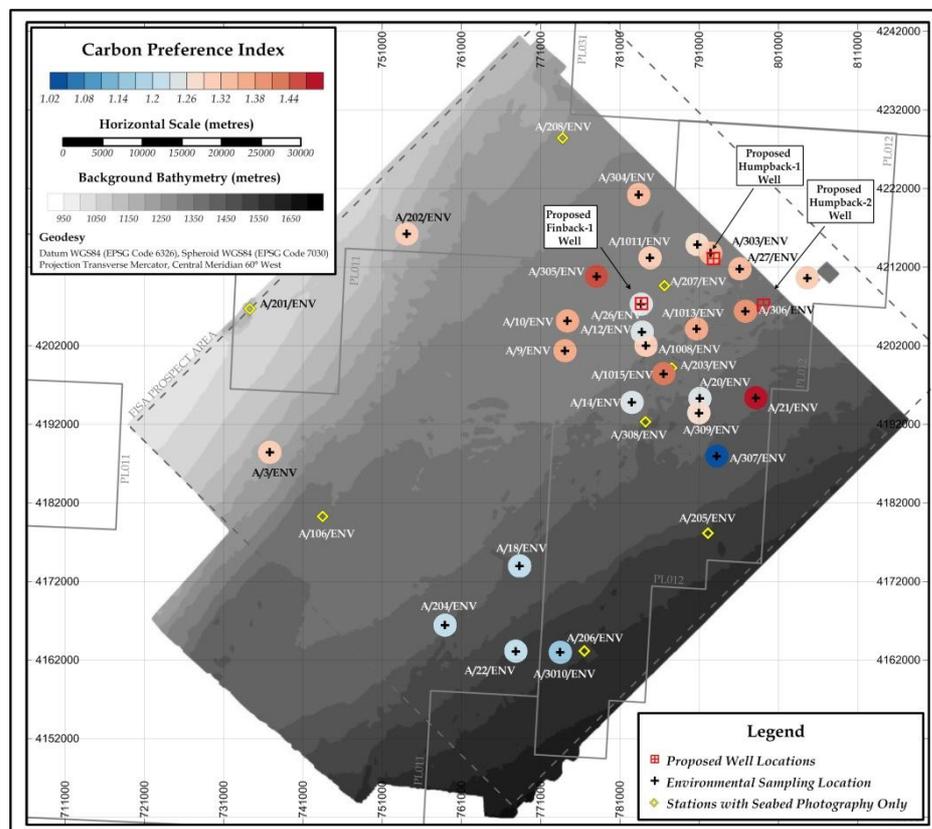


Table 2.4 - Total Aliphatic Concentrations (ng.g⁻¹)

| Station | A/9/ENV | A/12/ENV | A/14/ENV | A/18/ENV | A/20/ENV | A/202/ENV | A/204/ENV | A/21/ENV | A/22/ENV | A/26/ENV | A/27/ENV | A/3/ENV | A/301/ENV |
|------------------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|
| nC10 | <1 | 1.8 | <1 | <1 | <1 | 1.1 | 2.7 | <1 | <1 | <1 | <1 | 1.4 | <1 |
| nC11 | 1.8 | 10.8 | <1 | 4.9 | 3.3 | 1.9 | 11.3 | 1.1 | 3.4 | <1 | 5.3 | 6.8 | <1 |
| nC12 | 4.1 | 16.7 | 3.5 | 5.5 | 5.2 | 4.9 | 17.6 | 5.3 | 4.5 | 9.3 | 11.1 | 12.4 | 4.4 |
| nC13 | 6.1 | 16.0 | 6.8 | 10.8 | 8.7 | 4.4 | 22.8 | 7.8 | 10.0 | 10.5 | 14.0 | 16.9 | 4.6 |
| nC14 | 7.6 | 16.6 | 9.8 | 11.9 | 9.5 | 7.0 | 27.6 | 11.6 | 13.2 | 14.3 | 15.0 | 21.6 | 5.5 |
| nC15 | 11.9 | 25.2 | 12.3 | 19.9 | 15.7 | 8.0 | 43.7 | 15.2 | 17.9 | 22.7 | 23.1 | 33.1 | 8.0 |
| nC16 | 10.7 | 19.4 | 11.5 | 18.2 | 13.7 | 7.0 | 40.5 | 14.4 | 16.3 | 23.2 | 21.9 | 28.7 | 7.8 |
| nC17 | 12.7 | 24.3 | 12.9 | 20.5 | 16.3 | 8.6 | 44.3 | 17.6 | 17.8 | 24.1 | 24.0 | 35.9 | 11.2 |
| pristane | 31.2 | 62.5 | 34.2 | 54.3 | 41.2 | 20.7 | 122 | 49.2 | 49.2 | 63.2 | 57.7 | 85.3 | 23.0 |
| nC18 | 10.9 | 19.6 | 13.3 | 20.0 | 15.0 | 8.3 | 43.5 | 14.0 | 17.8 | 22.0 | 21.7 | 31.8 | 8.2 |
| phytane | 3.0 | 17.7 | 3.0 | 2.9 | 12.0 | 3.8 | 21.4 | 8.2 | 9.4 | 27.5 | 11.3 | 24.9 | 3.7 |
| nC19 | 9.3 | 17.1 | 11.6 | 16.9 | 13.7 | 6.8 | 43.8 | 16.5 | 15.2 | 16.7 | 21.4 | 27.2 | 7.2 |
| nC20 | 9.6 | 20.5 | 10.7 | 16.7 | 13.9 | 7.1 | 44.7 | 14.7 | 16.1 | 18.7 | 18.8 | 26.3 | 6.9 |
| nC21 | 7.8 | 15.6 | 10.0 | 15.2 | 10.8 | 6.1 | 37.9 | 12.4 | 15.1 | 16.3 | 16.0 | 23.1 | 6.0 |
| nC22 | 7.3 | 14.8 | 8.5 | 12.9 | 9.8 | 5.7 | 31.8 | 9.9 | 12.2 | 13.8 | 14.6 | 20.7 | 5.5 |
| nC23 | 7.3 | 14.7 | 8.3 | 14.4 | 10.1 | 6.1 | 29.5 | 12.1 | 13.2 | 14.2 | 14.5 | 22.6 | 5.8 |
| nC24 | 7.7 | 14.0 | 8.3 | 19.1 | 9.3 | 5.6 | 30.3 | 9.4 | 11.4 | 12.5 | 14.4 | 16.8 | 5.7 |
| nC25 | 8.6 | 14.8 | 9.5 | 15.8 | 9.2 | 6.5 | 29.9 | 10.2 | 13.7 | 15.7 | 16.7 | 21.0 | 6.5 |
| nC26 | 6.5 | 15.6 | 7.6 | 11.0 | 9.3 | 5.5 | 26.6 | 8.9 | 10.6 | 12.8 | 13.6 | 19.0 | 5.5 |
| nC27 | 9.3 | 17.6 | 10.9 | 15.3 | 10.9 | 7.3 | 36.5 | 13.5 | 13.4 | 17.9 | 18.7 | 25.2 | 7.7 |
| nC28 | 7.3 | 18.7 | 6.9 | 9.3 | 9.1 | 5.6 | 23.4 | 6.8 | 18.6 | 9.7 | 12.8 | 14.1 | 7.4 |
| nC29 | 14.1 | 41.1 | 19.0 | 23.2 | 17.5 | 14.3 | 49.8 | 23.0 | 29.6 | 30.8 | 33.6 | 37.8 | 14.8 |
| nC30 | 3.2 | 10.5 | 4.2 | 7.4 | 5.1 | 3.9 | 15.5 | 7.9 | 6.5 | 5.6 | 9.7 | 10.6 | 4.8 |
| nC31 | 9.4 | 19.8 | 12.1 | 15.9 | 10.2 | 13.6 | 33.0 | 20.9 | 11.9 | 16.6 | 23.4 | 25.8 | 11.3 |
| nC32 | 5.1 | 10.9 | 5.4 | 9.0 | 7.1 | 4.3 | 20.4 | 3.1 | 8.3 | 10.8 | 10.8 | 13.0 | 4.6 |
| nC33 | 8.2 | 17.2 | 2.5 | 5.5 | 8.9 | 4.1 | 19.0 | 7.7 | 3.7 | 9.8 | 17.0 | 10.4 | 3.9 |
| nC34 | <1 | 13.0 | <1 | 2.4 | 1.1 | 1.6 | 3.1 | 3.1 | 2.3 | 2.9 | 4.8 | 3.8 | 1.4 |
| nC35 | 1.1 | 3.8 | <1 | <1 | 1.5 | <1 | <1 | 1.2 | 2.6 | 2.4 | 1.9 | 2.4 | <1 |
| nC36 | <1 | 2.1 | 3.0 | 2.0 | 1.1 | <1 | 2.0 | 1.1 | <1 | 1.9 | 2.6 | 1.1 | <1 |
| nC37 | 1.4 | 3.0 | <1 | <1 | <1 | <1 | <1 | 3.8 | <1 | <1 | <1 | <1 | <1 |
| Total n-alkanes | 189 | 435 | 208 | 324 | 246 | 155 | 731 | 273 | 306 | 355 | 401 | 510 | 155 |

...continuing next page

Table 2.4 - Total Aliphatic Concentrations (ng.g⁻¹) (2 of 2)

| Station | A/302/ENV | A303/ENV | A/304/ENV | A/305/ENV | A/306/ENV | A/307/ENV | A/309/ENV | A/10/ENV | A/1008/ENV | A/1011/ENV | A/1013/ENV | A/1015/ENV | A/3010/ENV |
|-----------------|-----------|----------|-----------|-----------|-----------|-----------|-----------|----------|------------|------------|------------|------------|------------|
| nC10 | <1 | <1 | <1 | <1 | <1 | <1 | 1.3 | 4.5 | <1 | <1 | 3.3 | 1.1 | <1 |
| nC11 | 3.4 | <1 | 3.1 | 2.5 | 2.4 | 1.9 | 4.7 | 6.4 | 4.6 | <1 | 6.5 | 4.8 | 4.5 |
| nC12 | 6.0 | 4.8 | 4.4 | 7.1 | 2.7 | 6.2 | 3.9 | 8.0 | 3.0 | 2.9 | 8.5 | 6.3 | 5.8 |
| nC13 | 12.4 | 6.4 | 7.2 | 19.7 | 6.7 | 7.4 | 8.3 | 18.1 | 7.5 | 5.4 | 14.1 | 14.7 | 9.5 |
| nC14 | 11.1 | 7.5 | 9.5 | 13.2 | 8.6 | 10.3 | 9.8 | 16.5 | 10.4 | 6.6 | 17.2 | 14.9 | 12.7 |
| nC15 | 16.2 | 10.4 | 13.9 | 15.0 | 10.8 | 13.7 | 17.0 | 29.7 | 13.8 | 7.7 | 25.9 | 24.6 | 16.9 |
| nC16 | 12.5 | 8.9 | 12.1 | 15.6 | 9.8 | 12.7 | 13.6 | 24.8 | 13.1 | 6.8 | 22.5 | 22.8 | 15.3 |
| nC17 | 16.5 | 11.9 | 15.7 | 21.3 | 11.4 | 14.8 | 15.7 | 31.1 | 16.5 | 7.8 | 25.2 | 25.1 | 17.7 |
| pristane | 41.4 | 26.6 | 36.4 | 51.5 | 29.7 | 38.7 | 40.0 | 74.4 | 41.8 | 19.7 | 64.6 | 66.4 | 47.4 |
| nC18 | 14.4 | 8.7 | 12.8 | 20.0 | 9.1 | 14.8 | 15.0 | 22.8 | 14.7 | 7.1 | 18.1 | 18.2 | 17.1 |
| phytane | 2.5 | 2.7 | 2.2 | 11.7 | 3.9 | 2.1 | 3.0 | 6.7 | 4.7 | 1.7 | 5.1 | 5.0 | 3.8 |
| nC19 | 11.6 | 9.1 | 12.7 | 17.8 | 8.7 | 12.3 | 12.5 | 23.9 | 12.3 | 5.7 | 20.2 | 19.9 | 14.4 |
| nC20 | 11.9 | 8.5 | 13.4 | 20.4 | 8.8 | 12.0 | 12.1 | 23.2 | 11.9 | 6.5 | 23.6 | 19.0 | 14.5 |
| nC21 | 11.5 | 7.0 | 10.3 | 15.1 | 8.2 | 9.5 | 10.9 | 21.3 | 10.3 | 5.5 | 19.0 | 17.4 | 14.5 |
| nC22 | 13.7 | 6.5 | 8.8 | 13.1 | 7.1 | 8.6 | 9.4 | 19.5 | 9.2 | 5.5 | 17.6 | 14.8 | 11.7 |
| nC23 | 10.8 | 6.2 | 8.9 | 15.5 | 7.0 | 9.6 | 10.3 | 19.9 | 10.1 | 5.5 | 17.7 | 16.7 | 12.8 |
| nC24 | 10.0 | 6.5 | 7.6 | 11.2 | 7.3 | 25.7 | 8.7 | 17.4 | 8.6 | 4.9 | 14.0 | 14.8 | 10.6 |
| nC25 | 12.3 | 7.9 | 8.9 | 15.4 | 8.1 | 8.1 | 10.5 | 18.6 | 10.4 | 6.2 | 17.1 | 19.7 | 15.0 |
| nC26 | 9.9 | 6.2 | 7.5 | 13.0 | 6.6 | 7.8 | 8.7 | 15.3 | 8.1 | 5.0 | 14.4 | 12.7 | 10.0 |
| nC27 | 11.5 | 8.3 | 10.2 | 19.4 | 9.9 | 9.7 | 10.7 | 21.1 | 10.8 | 6.8 | 20.4 | 18.2 | 12.5 |
| nC28 | 8.5 | 8.7 | 8.3 | 10.2 | 4.9 | 8.0 | 7.2 | 14.3 | 7.6 | 4.6 | 11.8 | 12.3 | 9.4 |
| nC29 | 21.6 | 16.5 | 17.6 | 30.5 | 16.3 | 15.9 | 15.1 | 37.5 | 17.8 | 13.0 | 38.9 | 33.5 | 15.9 |
| nC30 | 3.5 | 4.2 | 5.1 | 9.2 | 5.1 | 5.0 | 5.7 | 10.0 | 6.6 | 3.0 | 9.2 | 8.5 | 8.9 |
| nC31 | 9.1 | 10.4 | 11.7 | 26.1 | 11.3 | 9.9 | 12.3 | 25.9 | 13.2 | 9.4 | 25.5 | 26.8 | 9.7 |
| nC32 | 4.6 | 5.1 | 6.4 | 6.4 | 5.7 | 6.8 | 6.9 | 14.3 | 7.8 | 3.9 | 14.2 | 12.2 | 8.4 |
| nC33 | 5.5 | 6.8 | 9.8 | 4.9 | 8.1 | 8.3 | 5.2 | 13.6 | 4.4 | 5.5 | 9.8 | 8.6 | 2.7 |
| nC34 | 1.8 | 1.3 | 1.4 | 2.2 | 1.7 | <1 | 1.2 | 1.2 | <1 | 1.6 | 3.7 | 2.5 | <1 |
| nC35 | <1 | 1.2 | 1.5 | 2.2 | <1 | <1 | <1 | <1 | <1 | <1 | 3.1 | 1.6 | <1 |
| nC36 | 1.9 | <1 | 1.1 | 1.6 | 1.0 | 1.1 | <1 | 1.8 | 1.1 | 2.1 | 1.3 | 1.5 | 1.5 |
| nC37 | 1.1 | <1 | <1 | 3.5 | <1 | 1.3 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Total n-alkanes | 253 | 179 | 230 | 352 | 187 | 241 | 237 | 461 | 234 | 139 | 423 | 393 | 272 |

Petrogenic/Biogenic or (P/B) Ratio

The P/B ratio compares the lighter, more petrogenic aliphatics with the heavier, and more biogenic aliphatics. Results were calculated for all stations showing a consistent but moderate ratio ranging from 0.69 to 0.93 (mean 0.80±0.06SD) with no clear pattern of distribution. These levels may indicate some minor influence from mixed hydrocarbon source with a possible petrogenic component.

The Pristane/Phytane Ratio

Pristane and phytane are both isoprenoidal alkanes commonly found as constituents within crude oils (Berthou and Friocourt, 1981). However, in biogenic environments, only pristane is commonly found in the marine environment as naturally biosynthesised and a

product of phytol moiety of chlorophyll. Phytane is generally absent or only present at low levels in uncontaminated natural systems (Blumer and Snyder, 1965). A presence of both isoprenoids at similar levels is typically taken as an indication of petroleum contamination.

The pristane/phytane ratio ranged from 3.40 to 18.70 (mean $9.38 \pm 4.92SD$). This would indicate a clear pristane dominance of biogenic origin. This is likely attributed to a significant biogenic influence on the sediments as a result of significant planktonic contribution. This plankton influence could possibly be from the highly productive surrounding surface waters of the Patagonian marine ecosystem. It should be noted that pristane/phytane ratio can often be difficult to interpret due to its erratic nature and should be used mainly to substantiate other interpretations. The use of the ratio in interpretative discourse is open to criticism, mainly owing to the natural occurrence of phytane in some older sediments and the confusing variation of sedimentary pristane, induced by the variability of phytoplankton numbers (Blumer and Snyder, 1965). This may be the case with the current study where high levels of background phytane (petrogenic in origin) are masked by even higher levels of pristane due to significant plankton influence.

2.6.2.1 Historical Comparison with Saturate/Alkanes Hydrocarbons

Previous levels of historical saturates at sites surveyed at the Vinson West, Nimrod and Toroa prospects registered mean total alkanes of 162, 310 and 650ng.g⁻¹, equivalent to 5.2%, 8.3% and 7.6% of the THC, respectively (Figure 2.13). A further direct comparison between Vinson West ENV5 and A/3010/ENV further revealed a similar concentration (228ng.g⁻¹ compared to 270ng.g⁻¹ respectively).

A general comparison of the saturate chromatograms indicated broad similarities based on uncontaminated hydrocarbon signatures.

For the CPI, the mean value of 1.30 recorded within the current study can be compared to a mean value of 1.50 ($\pm 0.10SD$) previously recorded at Vinson West, both being slightly higher than 1.12 and 0.99 recorded at Toroa and Nimrod, respectively (Figure 2.14). The values close to unity and the general lack of variance between sites indicates these values are typical for this depth and region of the South Atlantic, located outside of significant influences from terrestrial plants. Terrigenous influence is normally reflected in a higher CPI value which is due to elevated long chain odd numbered alkanes in wax cuticles of higher plants (Eglinton *et al.*, 1962). Further direct comparison between Vinson West ENV5 and A/3010/ENV also revealed a similar result (1.4 compared to 1.16, respectively).

Figure 2.13 - Comparison of Total Saturate Alkanes with Historical Data

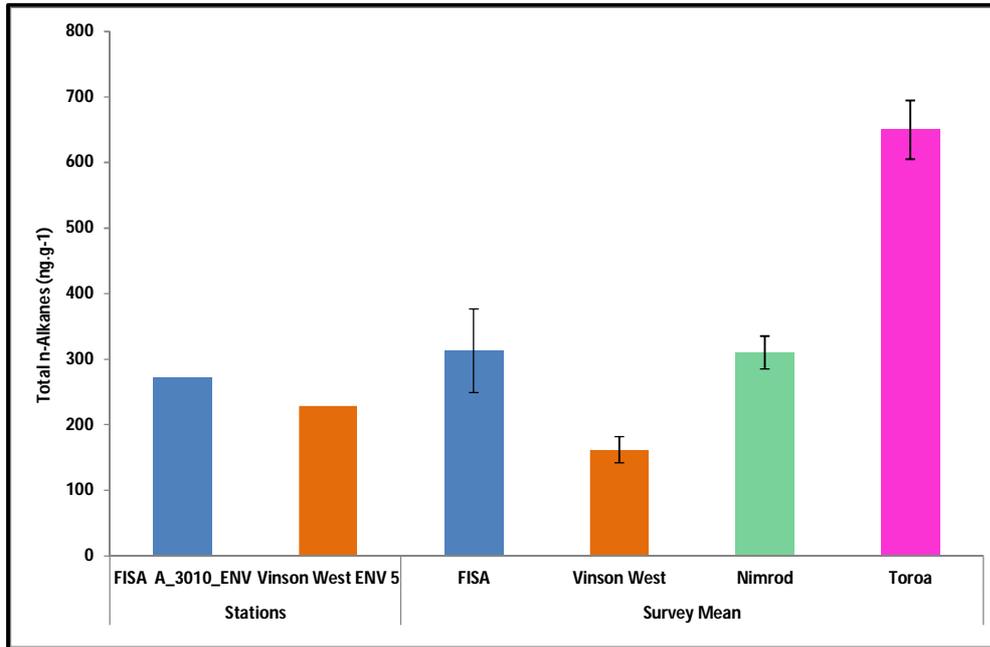
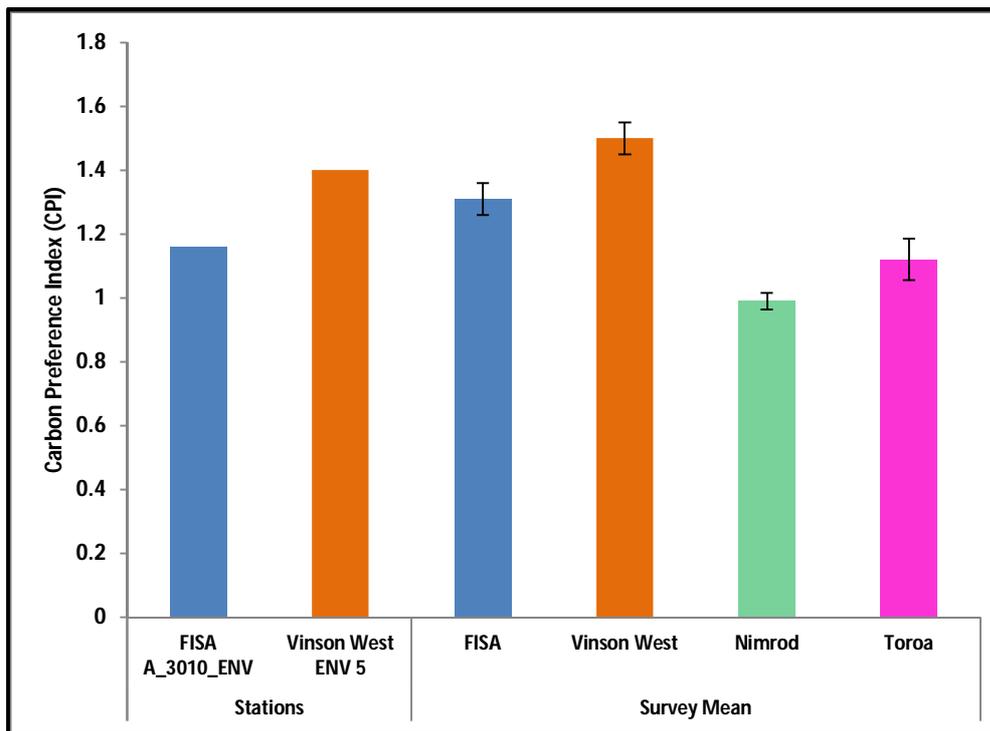


Figure 2.14 - Comparison of Carbon Preference Index with Historical Data



2.6.3 Polycyclic Aromatic Hydrocarbons (PAH)

Quantitative PAHs were analysed at each station using Gas Chromatography-Mass Spectrometry (GC-MS).

Results of the single ion current (SIC) analyses are summarised in Table 2.3, and detailed in Table 2.5, showing concentrations for both parent compounds and their alkyl derivatives. The concentrations of 16 PAH priority pollutants listed by US Environmental Protection Agency (EPA) are listed in Appendix V. The EPA 16 are used globally in assessment of contamination relating to both environmental and human health studies.

PAHs and their alkyl derivatives have been recorded in a wide range of marine sediments (Laflamme and Hites, 1978) with the majority of compounds produced from what is thought to be pyrolytic sources. These are the combustion of organic material such as forest fires (Youngblood and Blumer, 1975), the burning of fossil fuels and, in the case of offshore oilfields, flare stacks, etc. The resulting PAHs, rich in the heavier weight 4-6 ring aromatics, are normally transported to the sediments via atmospheric fallout or river run-off. Another PAH source is petroleum hydrocarbon, often associated with localised drilling activities. These are rich in the lighter, more volatile 2 and 3 ring PAHs (NPD; naphthalene (128), phenanthrene, anthracene (178) and dibenzothiophene (DBT) with their alkyl derivatives).

Total PAH concentrations (2-6 compounds) were moderately low for all sites analysed, ranging from 35.18ng.g⁻¹ to 207.65ng.g⁻¹ (mean 82.77±39.35SD; Table 2.3, Figure 2.15). Total PAHs indicated no real pattern of distribution within the surface sediments, although an elevated level was measured at station A/204/ENV (207.65ng.g⁻¹), and a weak correlation with % fines recorded throughout (P<0.05, Appendix XI).

Figure 2.15 - Total Polycyclic Aromatic Hydrocarbons (2-6 Ring)

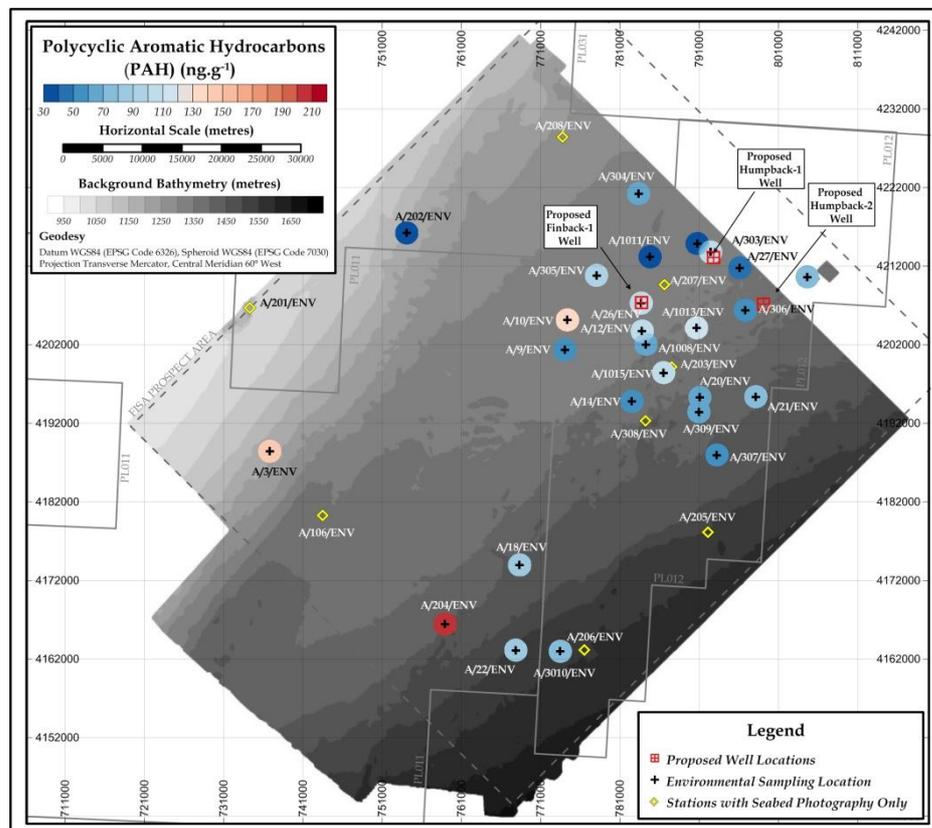


Table 2.5 - Polyaromatic Hydrocarbon Concentrations (Single Ion Currents, ng.g⁻¹)

| Station | A/9/ENV | A/12/ENV | A/14/ENV | A/18/ENV | A/20/ENV | A/202/ENV | A/204/ENV | A/21/ENV | A/22/ENV | A/26/ENV | A/27/ENV | A/3/ENV | A/301/ENV |
|---|---------|----------|----------|----------|----------|-----------|-----------|----------|----------|----------|----------|---------|-----------|
| Naphthalene | 1.0 | 2.0 | <1 | 1.0 | 2.5 | <1 | 2.7 | <1 | <1 | 1.2 | 1.3 | 1.7 | <1 |
| C1 Naphthalenes | 2.1 | 3.4 | 2.1 | 3.3 | 2.7 | 1.7 | 7.0 | 2.3 | 2.5 | 3.4 | 3.5 | 4.7 | 1.5 |
| C2 Naphthalenes | 3.6 | 5.8 | 3.7 | 5.4 | 4.3 | 2.9 | 12.4 | 4.3 | 4.6 | 6.4 | 6.7 | 8.9 | 2.8 |
| C3 Naphthalenes | 5.4 | 7.7 | 5.9 | 8.7 | 6.1 | 4.2 | 18.1 | 7.3 | 8.3 | 7.2 | 10.1 | 13.6 | 3.8 |
| C4 Naphthalenes | 3.5 | 5.5 | 4.2 | 6.5 | 4.5 | 3.0 | 14.1 | 5.7 | 5.4 | 8.5 | 6.3 | 10.2 | 3.0 |
| Sum Naphthalenes | 15.6 | 24.3 | 15.8 | 25.0 | 20.1 | 11.7 | 54.3 | 19.6 | 21.0 | 26.8 | 27.9 | 39.1 | 11.1 |
| Phenanthrene / Anthracene | 2.1 | 4.4 | 2.3 | 3.5 | 3.1 | 1.7 | 8.6 | 2.7 | 2.9 | 4.1 | 3.8 | 5.9 | 1.7 |
| C1 178 | 4.1 | 8.9 | 4.4 | 6.4 | 5.2 | 3.1 | 15.1 | 5.3 | 5.8 | 7.7 | 7.2 | 11.2 | 3.1 |
| C2 178 | 4.0 | 7.6 | 4.1 | 6.2 | 4.9 | 3.0 | 15.5 | 5.5 | 6.2 | 8.3 | 7.6 | 11.4 | 3.0 |
| C3 178 | 3.1 | 6.5 | 3.2 | 5.3 | 4.1 | 2.5 | 9.9 | 4.2 | 4.4 | 6.8 | 5.7 | 8.4 | 2.2 |
| Sum 178 | 13.3 | 27.4 | 14.0 | 21.4 | 17.3 | 10.4 | 49.1 | 17.7 | 19.4 | 26.9 | 24.3 | 36.9 | 9.9 |
| Dibenzthiophene | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| C1 Dibenzthiophenes | <1 | 1.2 | <1 | <1 | <1 | <1 | 1.8 | <1 | <1 | 1.3 | 1.2 | 1.8 | <1 |
| C2 Dibenzthiophenes | <1 | <1 | <1 | <1 | <1 | <1 | 1.8 | <1 | <1 | 1.0 | <1 | 1.4 | <1 |
| C3 Dibenzthiophenes | <1 | <1 | <1 | <1 | <1 | <1 | 1.1 | <1 | <1 | <1 | <1 | 1.2 | <1 |
| Sum Dibenzthiophenes | 0.0 | 1.2 | 0.0 | 0.0 | 0.0 | 0.0 | 4.7 | 0.0 | 0.0 | 2.3 | 1.2 | 4.3 | 0.0 |
| Fluoranthene / pyrene | 1.1 | 2.1 | 1.1 | 1.6 | 1.2 | <1 | 4.5 | 1.4 | 1.5 | 1.9 | 1.7 | 3.6 | <1 |
| C1 202 | 2.2 | 4.1 | 2.1 | 3.5 | 2.6 | 1.5 | 7.2 | 3.0 | 3.5 | 3.9 | 3.8 | 5.7 | 1.5 |
| C2 202 | 2.5 | 5.4 | 2.8 | 4.2 | 3.4 | 1.8 | 10.1 | 3.9 | 4.3 | 5.0 | 5.2 | 7.6 | 2.1 |
| C3 202 | 2.3 | 5.3 | 2.2 | 4.0 | 3.0 | 1.5 | 8.8 | 3.0 | 3.8 | 4.7 | 4.6 | 6.3 | 1.8 |
| Sum 202 | 8.1 | 16.9 | 8.3 | 13.3 | 10.2 | 4.8 | 30.6 | 11.3 | 13.1 | 15.4 | 15.3 | 23.2 | 5.4 |
| Benanthracene / chrysene | 1.4 | 2.7 | 1.5 | 2.3 | 1.8 | 1.0 | 6.4 | 1.8 | 2.1 | 2.6 | 2.5 | 3.7 | <1 |
| C1 228 | 2.4 | 4.4 | 2.4 | 3.6 | 2.8 | 1.9 | 8.3 | 3.7 | 3.7 | 4.5 | 4.2 | 6.0 | 1.7 |
| C2 228 | 2.3 | 5.3 | 2.6 | 4.2 | 3.4 | 1.7 | 9.3 | 3.2 | 4.4 | 5.2 | 4.5 | 6.3 | 1.8 |
| Sum 228 | 6.1 | 12.4 | 6.5 | 10.1 | 8.0 | 4.6 | 24.0 | 8.8 | 10.3 | 12.3 | 11.2 | 16.0 | 3.5 |
| Benzfluoranthenes / benzopyrenes | 1.3 | 4.6 | 2.6 | 3.5 | 2.7 | <1 | 11.0 | 3.5 | 3.7 | 4.1 | 4.4 | 6.1 | <1 |
| C1 252 | 3.3 | 5.6 | 3.6 | 5.6 | 3.7 | 2.6 | 10.2 | 5.3 | 5.3 | 6.6 | 6.5 | 8.2 | 2.4 |
| C2 252 | 2.9 | 5.3 | 3.0 | 5.5 | 2.9 | 2.2 | 11.0 | 4.7 | 4.4 | 5.9 | 6.5 | 8.3 | 1.7 |
| Sum 252 | 7.5 | 15.5 | 9.2 | 14.6 | 9.3 | 4.8 | 32.2 | 13.5 | 13.4 | 16.6 | 17.4 | 22.6 | 4.1 |
| Aranthanthrenes / indeno- pyrene / benzperylene | 1.1 | 2.6 | 1.2 | 1.8 | 1.3 | <1 | 5.5 | 1.9 | 1.9 | 2.1 | 2.5 | 3.0 | <1 |
| C1 276 | 1.7 | 2.6 | 1.7 | 2.6 | 1.8 | 1.4 | 5.3 | 2.0 | 2.2 | 3.4 | 3.0 | 1.1 | 1.2 |
| C2 276 | <1 | <1 | <1 | <1 | <1 | <1 | 1.9 | <1 | 1.1 | <1 | 1.1 | 1.2 | <1 |
| Sum 276 | 2.9 | 5.1 | 2.9 | 4.4 | 3.1 | 1.4 | 12.8 | 3.9 | 5.2 | 5.5 | 6.6 | 5.3 | 1.2 |
| Sum of NPD fraction | 29.0 | 52.9 | 29.8 | 46.4 | 37.4 | 22.1 | 108 | 37.4 | 40.3 | 55.9 | 53.4 | 80.3 | 21.0 |
| % NPD | 54.1 | 51.4 | 52.5 | 52.3 | 54.9 | 58.6 | 52.1 | 49.9 | 49.0 | 52.9 | 51.4 | 54.5 | 59.8 |
| Parent to derivative ratio | 1.2 | 1.1 | 1.1 | 1.1 | 1.2 | 1.4 | 1.1 | 1.0 | 1.0 | 1.1 | 1.1 | 1.2 | 1.5 |

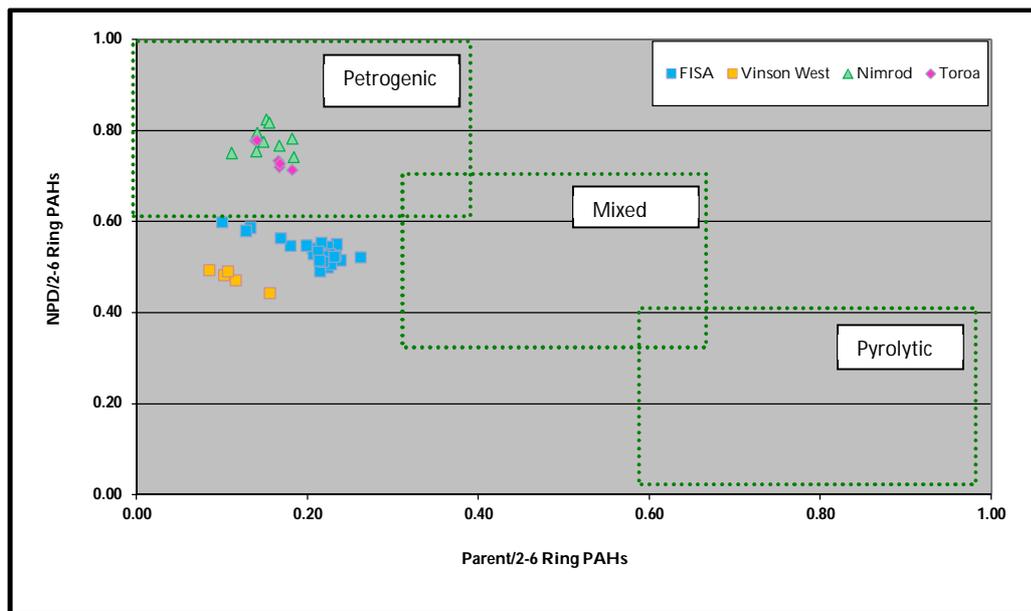
Table 2.5 - Polyaromatic Hydrocarbon Concentrations (Single Ion Currents, ng.g⁻¹) (2 of 2)

| Station | A/302/ENV | A/304/ENV | A/305/ENV | A/306/ENV | A/307/ENV | A/10/ENV | A/1008/ENV | A/1011/ENV | A/1013/ENV | A/1015/ENV | A/3010/ENV | A/303/ENV | A/309/ENV |
|---|-----------|-----------|-----------|-----------|-----------|----------|------------|------------|------------|------------|------------|-----------|-----------|
| Naphthalene | 1.2 | <1 | <1 | 1.7 | 1.1 | 2.3 | <1 | 1.2 | 2.7 | 1.8 | 1.0 | <1 | 1.0 |
| C1 Naphthalenes | 2.4 | 2.1 | 2.9 | 2.6 | 2.2 | 5.0 | 2.3 | 1.6 | 4.5 | 3.8 | 2.2 | 1.7 | 2.0 |
| C2 Naphthalenes | 4.3 | 3.9 | 5.4 | 3.9 | 3.7 | 8.6 | 4.2 | 2.7 | 7.6 | 6.7 | 4.7 | 2.8 | 3.9 |
| C3 Naphthalenes | 6.9 | 5.7 | 8.2 | 5.6 | 5.8 | 12.4 | 6.6 | 4.3 | 12.2 | 10.6 | 7.5 | 4.4 | 5.7 |
| C4 Naphthalenes | 5.1 | 4.5 | 5.7 | 3.8 | 4.3 | 9.6 | 4.7 | 2.7 | 8.5 | 6.9 | 5.3 | 3.1 | 4.9 |
| Sum Naphthalenes | 19.9 | 16.2 | 22.2 | 17.7 | 17.2 | 37.8 | 17.8 | 12.5 | 35.5 | 29.8 | 20.8 | 12.0 | 17.5 |
| Phenanthrene / Anthracene | 3.0 | 2.3 | 3.5 | 2.2 | 2.5 | 5.2 | 2.6 | 1.8 | 4.7 | 4.1 | 2.9 | 1.9 | 2.3 |
| C1 178 | 5.5 | 4.7 | 6.6 | 3.9 | 4.8 | 9.7 | 4.9 | 3.3 | 8.2 | 7.8 | 5.5 | 3.4 | 4.3 |
| C2 178 | 5.2 | 4.8 | 6.7 | 4.3 | 4.7 | 9.2 | 4.7 | 3.1 | 8.2 | 7.8 | 5.6 | 3.0 | 4.3 |
| C3 178 | 3.8 | 3.8 | 5.9 | 3.1 | 3.4 | 7.7 | 3.3 | 2.3 | 6.7 | 6.2 | 4.1 | 2.4 | 3.5 |
| Sum 178 | 17.6 | 15.5 | 22.7 | 13.5 | 15.4 | 31.9 | 15.6 | 10.5 | 27.7 | 25.9 | 18.1 | 10.6 | 14.5 |
| Dibenzthiophene | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| C1 Dibenzthiophenes | <1 | <1 | 1.1 | <1 | <1 | 1.4 | <1 | <1 | 1.3 | 1.2 | <1 | <1 | <1 |
| C2 Dibenzthiophenes | <1 | <1 | <1 | <1 | <1 | 1.2 | <1 | <1 | 1.2 | 1.0 | <1 | <1 | <1 |
| C3 Dibenzthiophenes | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Sum Dibenzthiophenes | 0.0 | 0.0 | 1.1 | 0.0 | 0.0 | 2.5 | 0.0 | 0.0 | 2.5 | 2.3 | 0.0 | 0.0 | 0.0 |
| Fluoranthene / pyrene | 1.4 | 1.2 | 1.7 | <1 | 1.1 | 2.3 | 1.2 | <1 | 1.9 | 1.8 | 1.4 | <1 | 1.1 |
| C1 202 | 3.0 | 2.4 | 3.7 | 2.3 | 2.3 | 5.5 | 2.6 | 1.7 | 4.6 | 4.3 | 2.9 | 1.6 | 2.4 |
| C2 202 | 3.2 | 3.0 | 4.6 | 3.0 | 3.0 | 6.1 | 3.2 | 2.0 | 5.3 | 5.3 | 3.8 | 2.0 | 3.3 |
| C3 202 | 3.0 | 2.6 | 3.3 | 2.7 | 2.8 | 6.0 | 2.8 | 1.7 | 4.8 | 4.6 | 3.0 | 1.7 | 2.9 |
| Sum 202 | 10.6 | 9.2 | 13.3 | 8.0 | 9.2 | 19.9 | 9.7 | 5.5 | 16.6 | 16.0 | 11.0 | 5.3 | 9.7 |
| Benzenanthracene / chrysene | 1.8 | 1.5 | 2.4 | 1.4 | 1.5 | 3.4 | 1.7 | 1.1 | 2.9 | 2.7 | 2.0 | 1.1 | 1.6 |
| C1 228 | 3.1 | 2.7 | 4.4 | 2.5 | 2.8 | 5.6 | 2.7 | 1.8 | 4.5 | 4.5 | 3.4 | 1.8 | 2.7 |
| C2 228 | 2.9 | 3.1 | 4.3 | 2.6 | 2.9 | 6.4 | 3.8 | 2.0 | 4.7 | 4.9 | 4.2 | 1.9 | 3.4 |
| Sum 228 | 7.8 | 7.3 | 11.0 | 6.5 | 7.2 | 15.4 | 8.3 | 5.0 | 12.1 | 12.0 | 9.6 | 4.9 | 7.8 |
| Benzfluoranthenes / benzopyrenes | 3.0 | 2.6 | 4.2 | 1.3 | 1.4 | 5.9 | 2.8 | <1 | 4.8 | 4.5 | 3.4 | 1.0 | 2.7 |
| C1 252 | 5.1 | 3.4 | 6.4 | 4.2 | 3.3 | 8.0 | 3.9 | 2.8 | 7.3 | 6.3 | 5.0 | 2.8 | 3.7 |
| C2 252 | 3.9 | 3.2 | 5.0 | 3.2 | 2.9 | 8.6 | 2.9 | 2.0 | 5.6 | 5.9 | 4.2 | 2.3 | 3.4 |
| Sum 252 | 12.0 | 9.2 | 15.6 | 8.8 | 7.6 | 22.5 | 9.6 | 4.9 | 17.7 | 16.7 | 12.5 | 6.1 | 9.8 |
| Aranthanthrenes / indeno- pyrene / benzperylene | 1.6 | 1.0 | 2.1 | 1.2 | 1.2 | 3.0 | 1.3 | <1 | 2.4 | 2.1 | 1.8 | <1 | 1.2 |
| C1 276 | 1.6 | 1.8 | 2.9 | 1.6 | 1.8 | 4.1 | 1.6 | 1.5 | 3.2 | 2.6 | 2.3 | 1.3 | 1.7 |
| C2 276 | <1 | <1 | <1 | <1 | <1 | 1.0 | <1 | <1 | 1.1 | 1.3 | <1 | <1 | <1 |
| Sum 276 | 3.1 | 2.8 | 5.0 | 2.8 | 2.9 | 8.1 | 2.9 | 1.5 | 6.7 | 6.1 | 4.1 | 1.3 | 2.9 |
| Sum of NPD fraction | 37.5 | 31.7 | 46.0 | 31.2 | 32.5 | 72.2 | 33.4 | 23.1 | 65.8 | 57.9 | 38.9 | 22.6 | 32.0 |
| % NPD | 52.8 | 52.7 | 50.6 | 54.6 | 54.7 | 52.3 | 52.2 | 57.9 | 55.3 | 53.3 | 51.1 | 56.2 | 51.4 |
| Parent to derivative ratio | 1.1 | 1.1 | 1.0 | 1.2 | 1.2 | 1.1 | 1.1 | 1.4 | 1.2 | 1.1 | 1.0 | 1.3 | 1.1 |

The NPD fraction, like total PAH, was low ranging from 21ng.g⁻¹ to 108ng.g⁻¹ (mean 43.8±20.4SD; Table 2.3), but moderately high and consistent by proportion which ranged from 49.0% to 59.8%. This reflects a low but stable petrogenic influence to the sediments. Like total PAH, the NPD fraction similarly correlated weakly with % fines.

Further information on the source(s) of PAH in the sediment may be obtained from a study of their alkyl homologue distributions (i.e. the degree of methyl, ethyl, substitution of the parent compounds). Pyrolytically derived PAHs are predominantly unalkylated whereas petrogenically derived PAHs are formed at relatively low temperatures (<150°C), and contains mainly alkylated species. The distribution of parent 2-6 ring PAH compounds also reflects whether the source is petrogenic or pyrolytic. The trend is represented graphically in Appendix V. These are three-dimensional plots which show the PAH concentrations, the parent compound distribution and the alkyl homologue distribution of the aromatic material in each of the sediments analysed. Predominantly, mixed to pyrolytic hydrocarbons were recorded within the aromatic materials, with approximately half or more of all PAHs represented by parent compounds. These results, combined with the moderate proportion of more petrogenically derived naphthalenes, phenanthrenes and dibenzothiophenes is demonstrated further in Figure 2.16. This indicates that the PAHs are predominantly petrogenic in origin; although an influence from mixed components was apparent throughout the lighter aliphatics (NPD).

Figure 2.16 - PAH Source Assignment for FISA and Historical Data

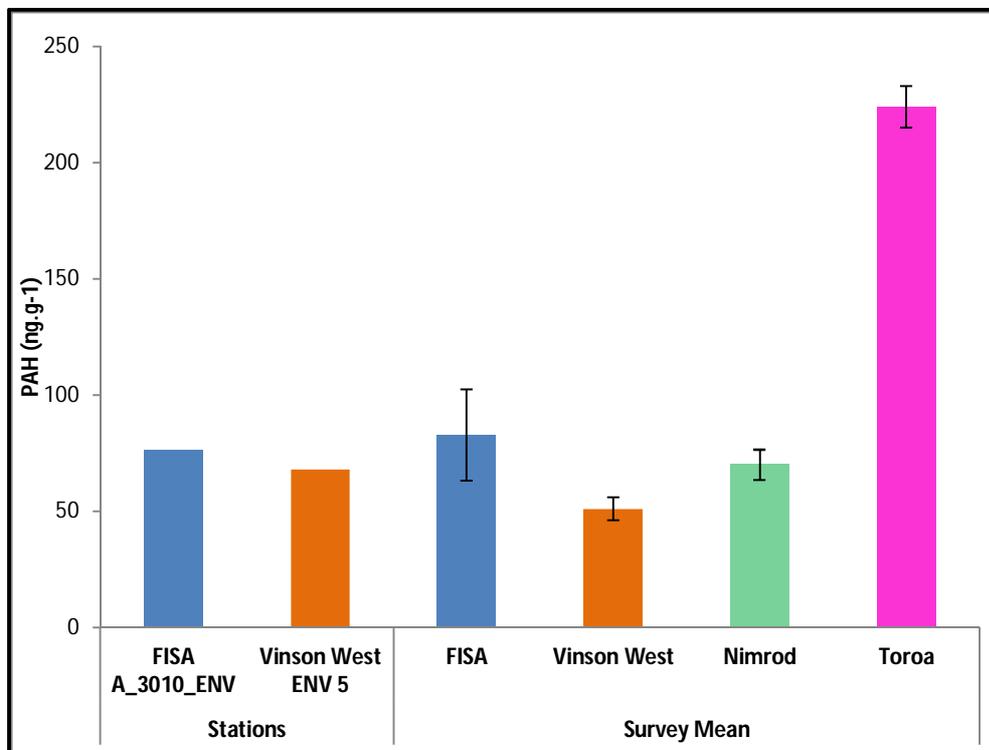


Overall, the amplitude of the hydrocarbon signature within the FISA survey area substrates was low and as generally expected for this region of the South Atlantic. However, the distribution of aromatics components suggests a very limited terrigenous source of material, with most fractions dominated by petrogenic material with only a minor pyrolytic component. Given the remote location of these sites along with their limited exposure to anthropogenic sources for thermogenic hydrocarbons, the most likely source of this material will be natural hydrocarbon seeps. The low THC and saturates levels suggest that this relates to a low level chronic supply, possibly from a remote origin.

2.6.3.1 Historical Comparison with Aromatic Hydrocarbons

The historical survey at the Vinson West and Nimrod prospects indicated similar or marginally lower concentrations of 51ng.g^{-1} and 70ng.g^{-1} , respectively, compared to 82.8ng.g^{-1} recorded at FISA (Figure 2.17). A further direct comparison between Vinson West ENV5 and A/3010/ENV revealed a similar level during the earlier survey (68ng.g^{-1}). A similar analysis carried out further south at the Toroa site, recorded a higher mean of 224ng.g^{-1} ($\pm 18\text{SD}$). This result is in accordance with the higher percentage of fines recorded in this area (77.8%), indicative of higher sedimentation and stronger assimilation of this hydrocarbon component in the surface sediments. Furthermore, this may also give some geographical indication of the direction from which natural petrogenic material may be transported, with elevated levels recorded towards the south and west.

Figure 2.17 - Comparison of Total Polycyclic Aromatic Hydrocarbons with Historical Data



2.7 HEAVY AND TRACE METAL CONCENTRATIONS

Results for heavy and trace metal analysis are given in Table 2.6 and Figures 2.18 to 2.27. All of the heavy and trace metals analysed (Al, Ba, Sn, As, Fe, Cd, Cr, Cu, Ni, Pb, V and Zn), with the exception of mercury (Hg), underwent a single hydrofluoric (HF) digestion and extraction for total sediment metals. The question of bioavailability of metals to marine organisms is a complex one, as sediment granulometry and the interface between waters and sediment all affect the bioavailability and subsequently toxicity. Therefore, even if a metal is found in higher concentrations it does not necessarily follow that this will have a detrimental effect on the environment, if present in an insoluble state. Historically, several extraction techniques have been applied to metal analysis in the past, with the most common applying to an HF/perchloric extraction for total metals, and a weaker nitric or aqua regia extraction. The latter techniques have shown close correlation to metal burdens in the tissues of benthic organisms (Luoma and Davies, 1983; Bryan and Langston, 1992). However, the overall extent to which a particular digests reflects bioavailability is still not well understood. A further fusion analysis was not considered necessary for Ba due to the unlikely occurrence of insoluble barium typically recorded in areas where previous drilling activities have occurred.

Metals occur naturally in the marine environment, and are widely distributed in both dissolved and sedimentary forms. Some are essential to marine life while others may be toxic to numerous organisms (Paez-Osuna and Ruiz-Fernandez, 1995). Rivers, coastal discharges, and the atmosphere are the principal modes of entry for most metals into the marine environment (Schaule and Patterson, 1983), with anthropogenic inputs occurring primarily as components of industrial and municipal wastes. Historically, several heavy and trace metals are found in elevated concentrations where drilling fluids or produced waters have been discharged by oil and gas installations. These include intentional additives (such as metal based salts and organo-metallic compounds in the fluids) as well as impurities within the drilling mud systems such as clays (e.g. bentonites; a gelling and viscosifying agent) and metal lignosulphates (viscosity controllers; McCourt *et al.*, 1991). Metals most characteristic for offshore contamination of marine sediments are Ba, Cr, Pb and Zn (Neff, 2005), although these may vary greatly dependent upon the constituents used.

Table 2.6 - Total Heavy and Trace Metal Concentrations ($\mu\text{g.g}^{-1}$ or ppm)

| | Arsenic (As) | Cadmium (Cd) | Chromium (Cr) | Copper (Cu) | Lead (Pb) | Mercury (Hg) | Nickel (Ni) | Tin (Sn) | Vanadium (V) | Zinc (Zn) | Aluminium (Al) mg.g^{-1} | Barium (Ba) | Iron (Fe) mg.g^{-1} |
|-------------------|--------------|--------------|---------------|-------------|-----------|--------------|-------------|----------|--------------|-----------|-----------------------------------|-------------|------------------------------|
| A/09/ENV | 11 | <0.1 | 136.7 | 16.8 | 5.6 | 0.02 | 9.8 | 1.2 | 77.2 | 69.8 | 25.9 | 306 | 118 |
| A/12/ENV | 10.2 | <0.1 | 133 | 13.9 | 6.1 | 0.01 | 15.3 | 1.4 | 76.2 | 140 | 25.4 | 291 | 112 |
| A/14/ENV | 11.1 | <0.1 | 136.3 | 14.5 | 6.2 | 0.01 | 10.6 | 1.1 | 74.8 | 70.7 | 24.5 | 250 | 131 |
| A/18/ENV | 11.1 | <0.1 | 140.2 | 15.7 | 5.9 | 0.01 | 10.0 | 1.3 | 72.5 | 74.0 | 20.7 | 253 | 135 |
| A/20/ENV | 9.6 | <0.1 | 144.5 | 19.6 | 5.9 | 0.01 | 13.2 | 2.1 | 75.4 | 83.7 | 22.8 | 285 | 116 |
| A/202/ENV | 13.2 | <0.1 | 113.9 | 16.2 | 6.1 | 0.01 | 9.3 | 1.5 | 75.0 | 70.4 | 22.4 | 279 | 134 |
| A/204/ENV | 3.2 | <0.1 | 34.9 | 17.0 | 4.2 | 0.02 | 10.2 | 1.5 | 52.7 | 43.3 | 27.2 | 529 | 24.7 |
| A/21/ENV | 8.9 | <0.1 | 156.8 | 13.7 | 6.9 | 0.01 | 14.2 | 1.5 | 76.3 | 86.2 | 28.1 | 327 | 117 |
| A/22/ENV | 10.1 | <0.1 | 126.1 | 14.7 | 6.2 | 0.01 | 10.1 | 1.2 | 70.0 | 71.0 | 22.9 | 508 | 122 |
| A/26/ENV | 6.4 | <0.1 | 92.4 | 17.1 | 5.9 | 0.02 | 12.2 | 1.5 | 69.4 | 64.3 | 32.6 | 414 | 49.2 |
| A/27/ENV | 10.9 | <0.1 | 110.1 | 15.8 | 6.1 | 0.01 | 10.5 | 2.1 | 74.0 | 71.5 | 26.5 | 313 | 102 |
| A/3/ENV | 12.3 | <0.1 | 89.4 | 15.5 | 7.4 | 0.03 | 10.1 | 1.1 | 72.1 | 69.6 | 28.4 | 415 | 95 |
| A/301/ENV | 7.6 | <0.1 | 127.9 | 20.1 | 6.3 | 0.03 | 17.6 | 1.3 | 71.6 | 88.0 | 26.7 | 321 | 93.8 |
| A/302/ENV | 9.1 | <0.1 | 170.1 | 17.2 | 5.2 | 0.01 | 14.9 | 1.4 | 82.4 | 84.5 | 23.5 | 268 | 116 |
| A/304/ENV | 10.1 | <0.1 | 147.5 | 14.3 | 5.2 | 0.01 | 10.4 | 1.6 | 77.4 | 72.7 | 25.5 | 311 | 121 |
| A/305/ENV | 11.3 | <0.1 | 143.1 | 11.8 | 6.6 | 0.01 | 10.9 | 1.4 | 78.1 | 75.4 | 25.9 | 317 | 118 |
| A/306/ENV | 7.3 | <0.1 | 140.3 | 18.1 | 5.8 | 0.02 | 20.4 | 2.3 | 89.4 | 85.0 | 31.5 | 338 | 89.5 |
| A/307/ENV | 9.9 | <0.1 | 148.9 | 15.9 | 6.1 | 0.01 | 12.9 | 1.7 | 80.9 | 78.5 | 25.0 | 442 | 128 |
| A/10/ENV | 11.1 | <0.1 | 143.2 | 11.2 | 4.9 | 0.01 | 10.6 | 1.3 | 84.1 | 73.0 | 27.0 | 340 | 10.7 |
| A/1008/ENV | 8.6 | <0.1 | 132.3 | 17.4 | 5.4 | 0.01 | 14.4 | 1.3 | 84.0 | 88.4 | 25.1 | 351 | 93.9 |
| A/1011/ENV | 11.4 | <0.1 | 130.3 | 13.5 | 6.1 | 0.02 | 11.6 | 1.2 | 83.8 | 78.6 | 25.8 | 296 | 104 |
| A/1013/ENV | 4.7 | <0.1 | 74.1 | 15.8 | 6.7 | 0.02 | 12.6 | 1.2 | 63.2 | 57.9 | 24.3 | 365 | 37.8 |
| A/1015/ENV | 6.8 | <0.1 | 82.6 | 19 | 13.3 | 0.02 | 14.9 | 1.5 | 80.3 | 73.2 | 38.2 | 338 | 55.7 |
| A/3010/ENV | 12.1 | <0.1 | 158.4 | 15.6 | 6.6 | 0.01 | 11.1 | 1.3 | 84.5 | 81.8 | 21.1 | 233 | 137 |
| A/303/ENV | 10.3 | <0.1 | 130.1 | 12.8 | 5.8 | 0.02 | 12.0 | 1.4 | 85.3 | 80.8 | 27.1 | 290 | 102 |
| A/309/ENV | 10.3 | <0.1 | 185.9 | 18.3 | 5.3 | 0.02 | 13.9 | 1.4 | 87.9 | 83.9 | 21.1 | 245 | 128 |
| Mean | 9.56 | <0.1 | 128.0 | 15.8 | 6.22 | 0.02 | 12.5 | 1.45 | 76.9 | 77.5 | 26.0 | 332 | 99.7 |
| StDev | 2.36 | - | 32.15 | 2.27 | 1.59 | 0.01 | 2.67 | 0.30 | 7.96 | 16.2 | 3.79 | 75.7 | 35.3 |
| %Variance | 24.7 | - | 25.1 | 14.3 | 25.5 | 43.2 | 21.5 | 20.8 | 10.4 | 20.8 | 14.6 | 22.8 | 35.4 |
| ERL* | 8.2 | 1.2 | 81 | 34 | 47 | 0.15 | 21 | - | - | 150 | - | - | - |
| AFEN** | 5 | <1 | 56 | 24 | 8 | <0.1 | 31 | - | 88 | 46 | 6.8 | 320 | 4.2 |
| Vinson West ENV 5 | 12.5 | 0.1 | 157.6 | 20 | 6.9 | 0.02 | 11 | <0.5 | 82.6 | 96.5 | 19.8 | 199 | 129 |
| Vinson West Mean | 11.2 | 0.1 | 139 | 17.9 | 6.4 | 0.02 | 10.6 | <0.5 | 75.3 | 87.1 | 20.0 | 222 | 123 |
| Vinson West SD | 0.8 | 0 | 11.6 | 4.6 | 0.6 | 0 | 0.5 | - | 4.2 | 7.4 | 1.86 | 30 | 5.31 |
| Nimrod Mean | 5.3 | 0.4 | 136.2 | 10.7 | 6.2 | <0.1 | 13.3 | 1.1 | 67 | 75.3 | 30.5 | 342 | 98.2 |
| Nimrod SD | 0.7 | 0.1 | 24.5 | 1.6 | 1.2 | - | 1.3 | 0.2 | 2.2 | 6.7 | 7.37 | 93 | 13.5 |
| Toroa Mean | 1 | 0.9 | 25.8 | 17 | 5.8 | <0.1 | 10.5 | 1 | 49 | 38.5 | 59.4 | 407 | 22.2 |
| Toroa SD | 0 | 0.1 | 1.7 | 2.3 | 1.2 | - | 0.8 | 0 | 1.3 | 8.7 | 2.54 | 9 | 0.49 |

Historical data are reported in blue

*lowest concentration of metal than can produce a harmful affect (Long et.al. 1995)

** Average for deep water sediment (>1000m, AFEN, 1996).

Trace metal contaminants in the marine environment tend to form associations with the non-residual phases of mineral matter, such as iron and manganese oxides and hydroxides, metal sulphides, organics, and carbonates. Metals associated with these non-residual phases are prone to various environmental interactions and transformations (physical, chemical and biological), potentially increasing their biological availability (Tessier *et al.*, 1979). Residual trace metals are defined as those which are part of the silicate matrix of the sediment and that are located mainly in the lattice structures of the component minerals. Non-residual trace metals are not part of the silicate matrix and have been incorporated into the sediment from aqueous solution by processes such as adsorption and organic complexes and may include trace metals originating from sources of pollution. Therefore, in monitoring trace metal contamination of the marine environment, it is important to distinguish these more mobile metals from the residual metals held tightly in the sediment lattice (Chester and Voutsinou, 1981), which are of comparatively little environmental significance. In this study, an analytical procedure involving digestion of sediment in hydrofluoric (HF) acid was employed to analyse the total elemental content of sediments retrieved across the survey area. The results constitute both residual and non-residual heavy metals concentration, much of which may not be available for biological uptake.

Although not directly related to the oil and gas industry, Cd levels consistently gave low concentrations in all samples with a mean concentration of $<0.1\mu\text{g.g}^{-1}$. There remains some debate as to toxicity of Cd to marine and terrestrial organisms. Some papers describe Cd as “very toxic” (Muniz *et al.*, 2004), whilst others consider this metal to have no negative effects (McLeese *et al.*, 1987). Other attempts to quantify the critical level of Cd toxification were carried out by Buchman (1999) and suggested ‘probable effect level’ of around $4.2\mu\text{g.g}^{-1}$.

Mercury (Hg) remained at low concentrations of $0.01\mu\text{g.g}^{-1}$ to $0.03\mu\text{g.g}^{-1}$ using ICP-MS at all of sites and depths sampled (mean $0.02\pm 0.01\text{SD}$).

Lead (Pb) was also recorded at a low level for the current FISA survey, ranging from $4.2\mu\text{g.g}^{-1}$ to $7.4\mu\text{g.g}^{-1}$, with the only relatively elevated values found at A/1015/ENV ($13.3\mu\text{g.g}^{-1}$; Figure 2.18).

Of particular relevance to the offshore oil and gas industry are metals associated with drilling related discharges. These can contain substantial amounts of barium sulphate (barites) as a weighting agent (NRC, 1983) and Ba is frequently used to detect the deposition of drilling fluids around offshore installations (Chow and Snyder, 1980; Gettleson and Laird, 1980; Tricine and Trefry, 1983). Barites also contain measurable concentrations of heavy metals as impurities, including Cd, Cr, Cu, Pb, Hg, and Zn (NRC, 1983). Heavy metals, either as impurities or additives are also present in other mud components. For this survey, natural Ba levels remained low and relatively consistent throughout the area ranging from $233\mu\text{g.g}^{-1}$ to $508\mu\text{g.g}^{-1}$ (mean $331.7\mu\text{g.g}^{-1}\pm 75.7\text{SD}$; Figure 2.19), with marginally higher levels recorded in the deeper sediments and a slight increase from north to south of the survey area. The majority of Ba is typically insoluble in the form of

a non-toxic sulphate (Gerrard *et al.*, 1999) - this metal is rarely of toxicological concern to the marine fauna.

Of the other metals, Cr, Ni, Cu, V and Zn all gave relatively low to moderate respective concentration means of $128\mu\text{g.g}^{-1}$ (Figure 2.20), $12.5\mu\text{g.g}^{-1}$ (Figure 2.21), $15.8\mu\text{g.g}^{-1}$ (Figure 2.22), $76.9\mu\text{g.g}^{-1}$ (Figure 2.23) and $77.5\mu\text{g.g}^{-1}$ (Figure 2.24). Overall, these metals have relatively low to moderate concentrations compared to other offshore environments (i.e. deep water (>1000m) Northeast Europe (AFEN, 1996, BSL unpublished)). The slightly elevated values recorded for Cr may be due to the occurrence of Tertiary volcanic rocks in the South Atlantic sediments, which are affected through the ice erosion pathways. Vanadium is often associated with the oil and gas industry as it is present in relatively high concentrations in most crude oils (Khalaf *et al.*, 1982). Most V enters seawater in suspension or colloidal form, passing quickly out of the water column and into silt deposition (Cole *et al.*, 1999). Consequently, as the natural background levels in the Falkland Islands region are relatively low, possible impacts from oil and gas explorations are likely to be detected from future surveys.

The crustal or matrix metal Al gave consistent results with a mean of 25.97mg.g^{-1} ($\pm 3.8\text{SD}$; Figure 2.25) whilst Fe indicated a very variable concentration 99.7mg.g^{-1} ($\pm 35.3\text{SD}$; Figure 2.26). This is equivalent to a respective percentage variance of 15% and 35%, with no clear pattern of distribution (Table 2.6 and Figure 2.29). Aluminium is often used as a normalisation metal in areas (i.e. standardising the proportion of metals against changes in sediment type), where significant changes in sediment parameters can mask relative changes in other metals. As there were no samples indicating significant metal elevations, this was not applied to the results for comparison. The variability of Fe is associated with the deposition of residual ice-rafted volcanic material in the form of granular basalts.

Iron is also an important metal as it is often associated with other elements, such as As to which they adsorb. Consequently, As was similarly variable and relatively moderate in concentration throughout this study (mean $9.56\mu\text{g.g}^{-1}$ $\pm 2.36\text{SD}$ and a variance of 25%; Figure 2.27). This is above the level recorded for similar depth sediments west of the British Isles (ca. 5 recorded, AFEN, 1996) and also above the ERL levels defined as the lowest concentration of a metal that produces adverse effects in 10% of the data reviewed (Long *et al.* 1995).

Figure 2.18 - Heavy Metal Concentration for Lead (Pb; mg.kg⁻¹)

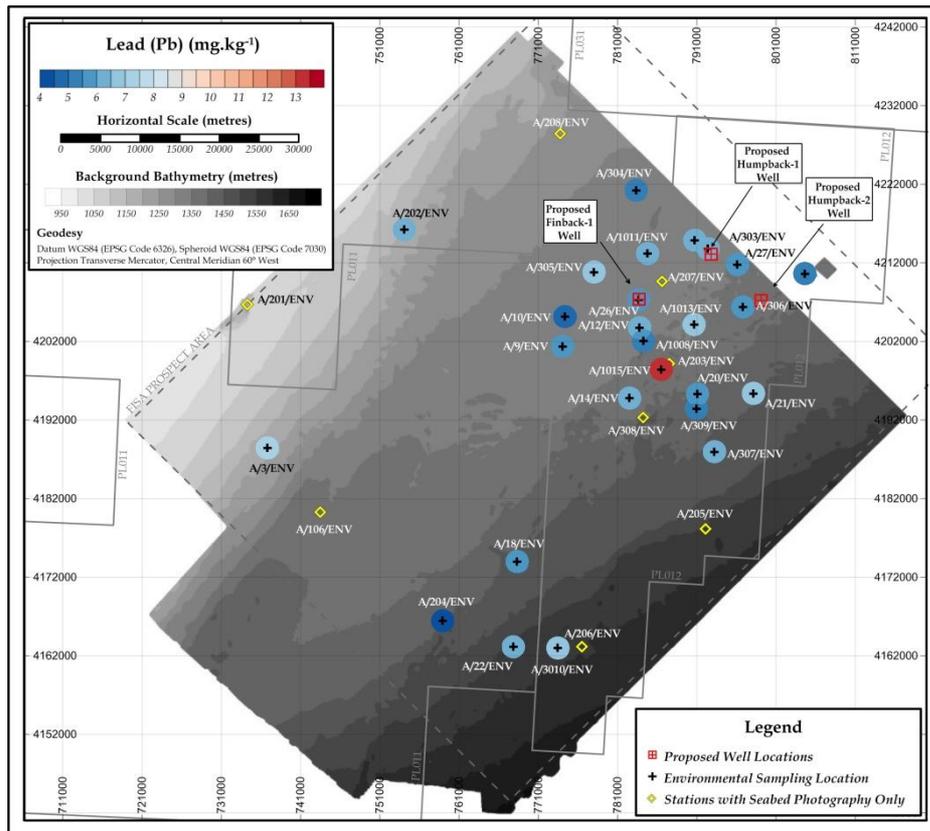


Figure 2.19 - Heavy Metal Concentration for Barium (Ba; mg.kg⁻¹)

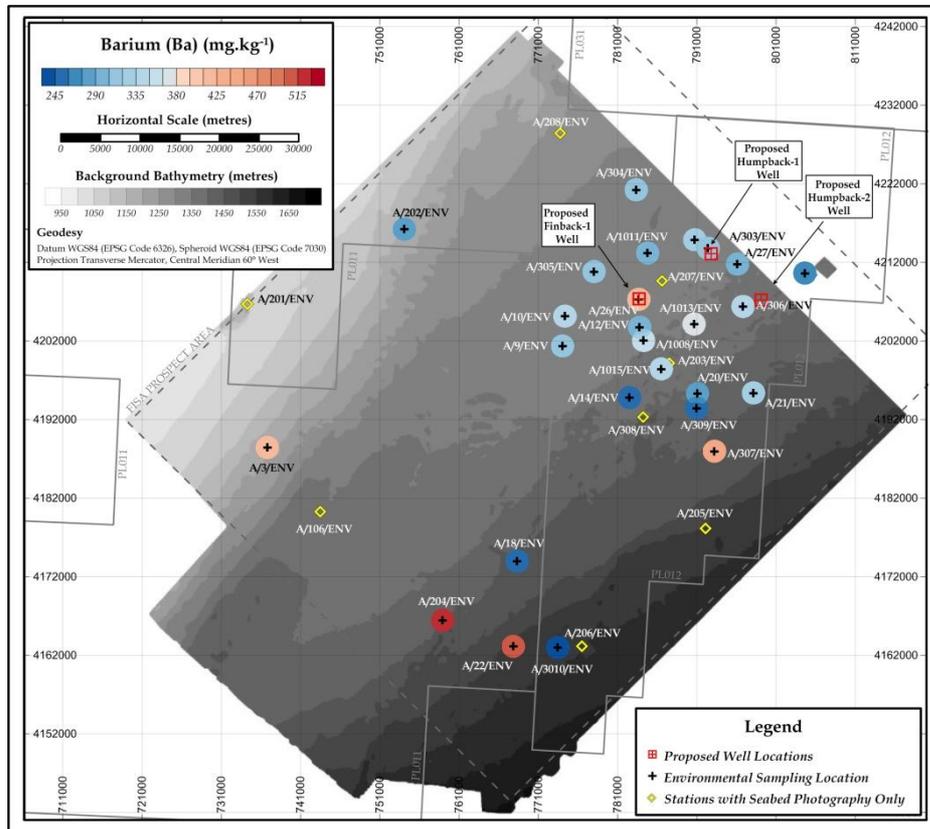


Figure 2.20 - Heavy Metal Concentration for Chromium (Cr; mg.kg⁻¹)

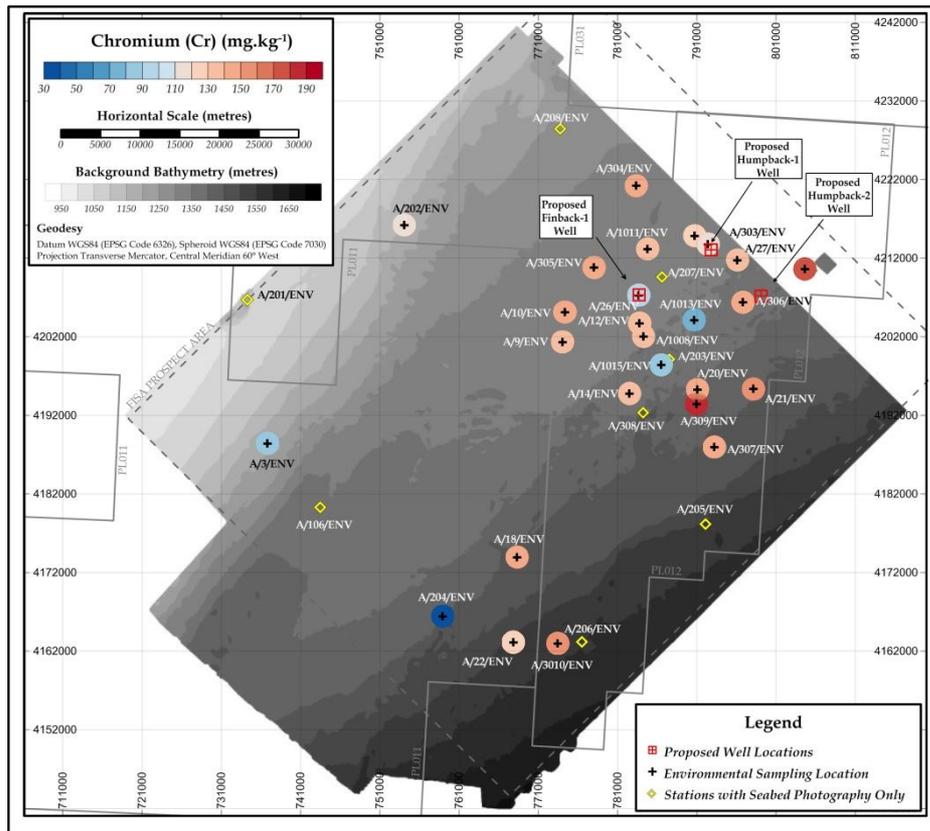


Figure 2.21 - Heavy Metal Concentration for Copper (Cu; mg.kg⁻¹)

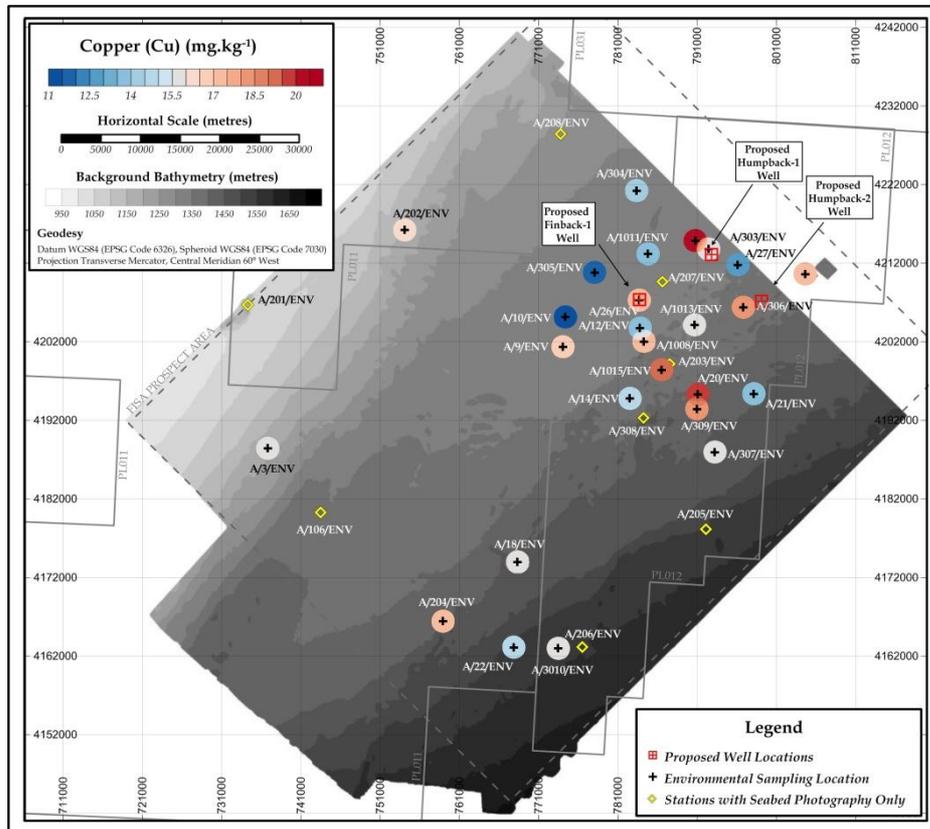


Figure 2.22 - Heavy Metal Concentration for Nickel (Ni; mg.kg⁻¹)

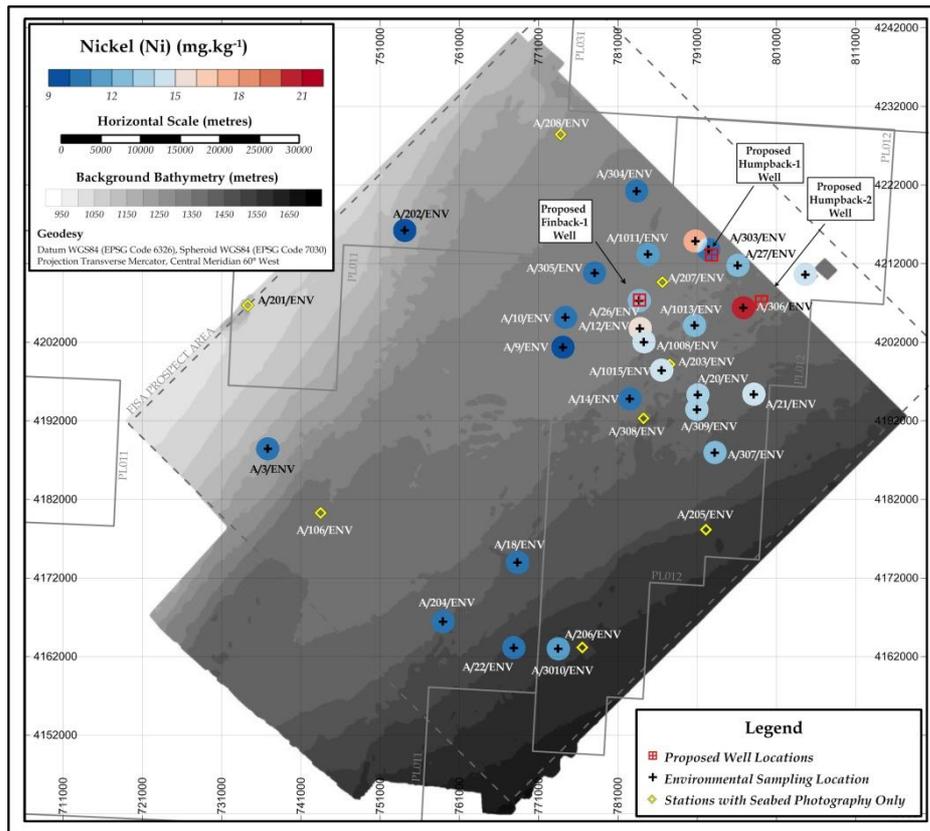


Figure 2.23 - Heavy Metal Concentration for Vanadium (V; mg.kg⁻¹)

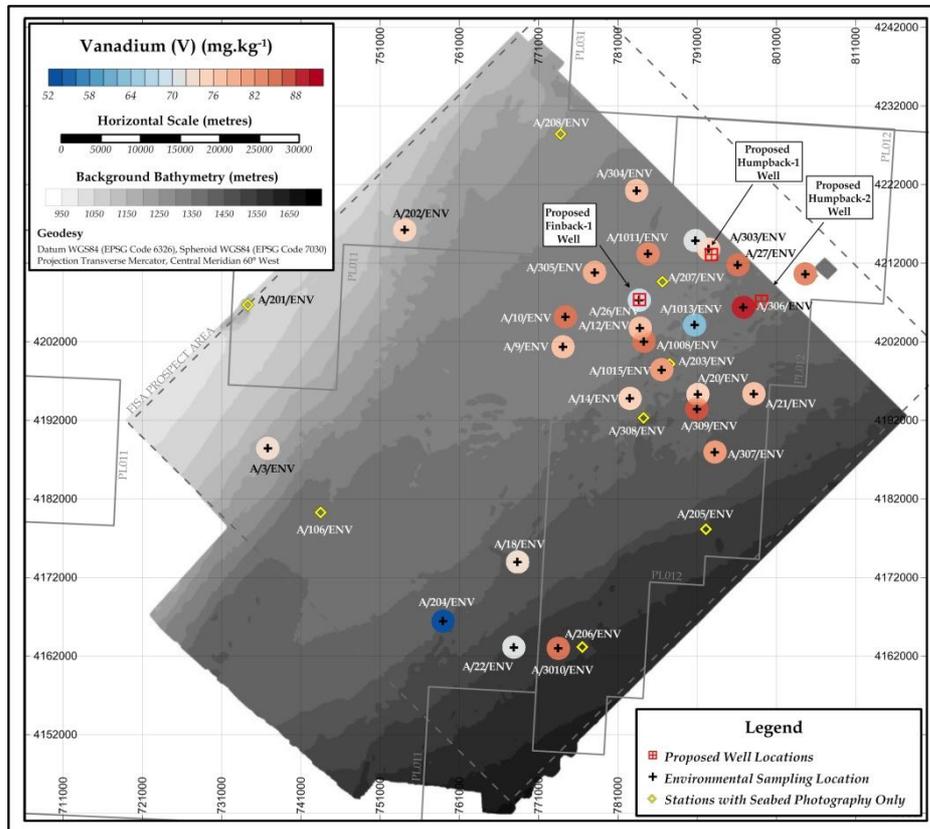


Figure 2.24 - Heavy Metal Concentration for Zinc (Zn; mg.kg⁻¹)

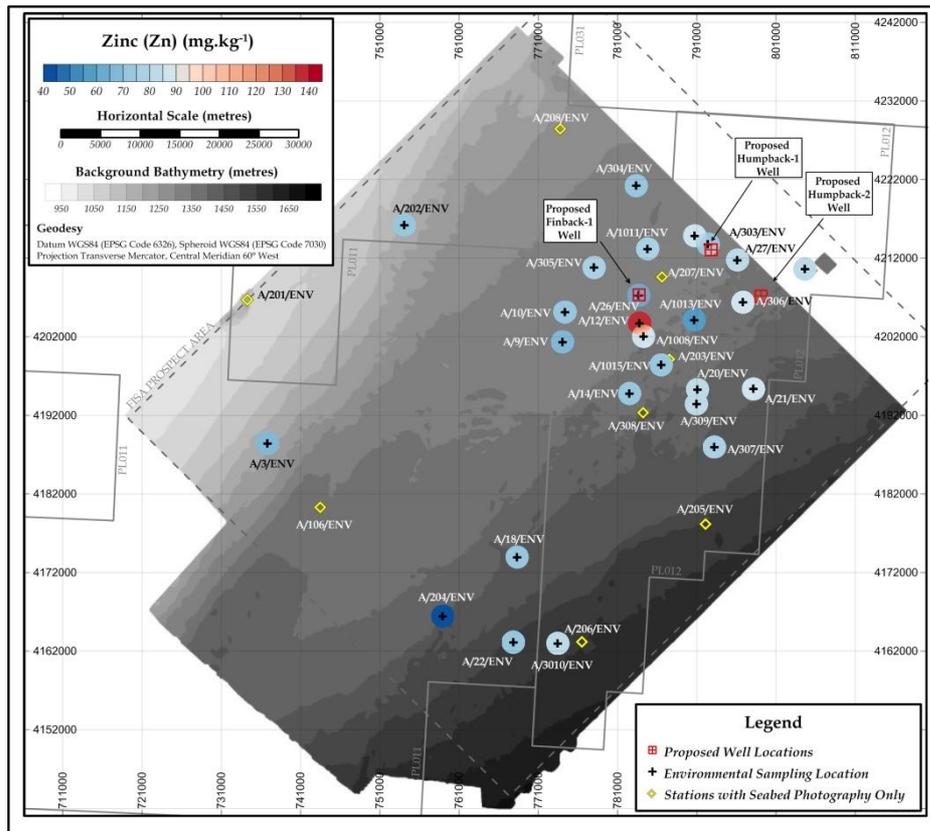


Figure 2.25 - Heavy Metal Concentration for Aluminium (Al; %)

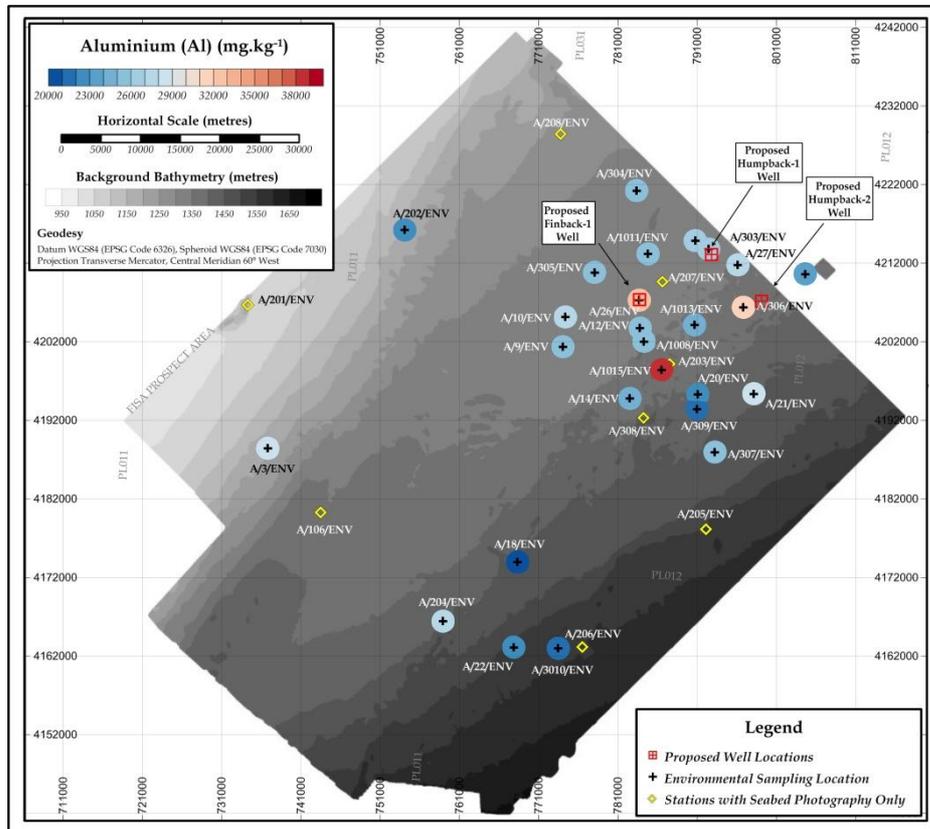


Figure 2.26 - Heavy Metal Concentration for Iron (Fe; %)

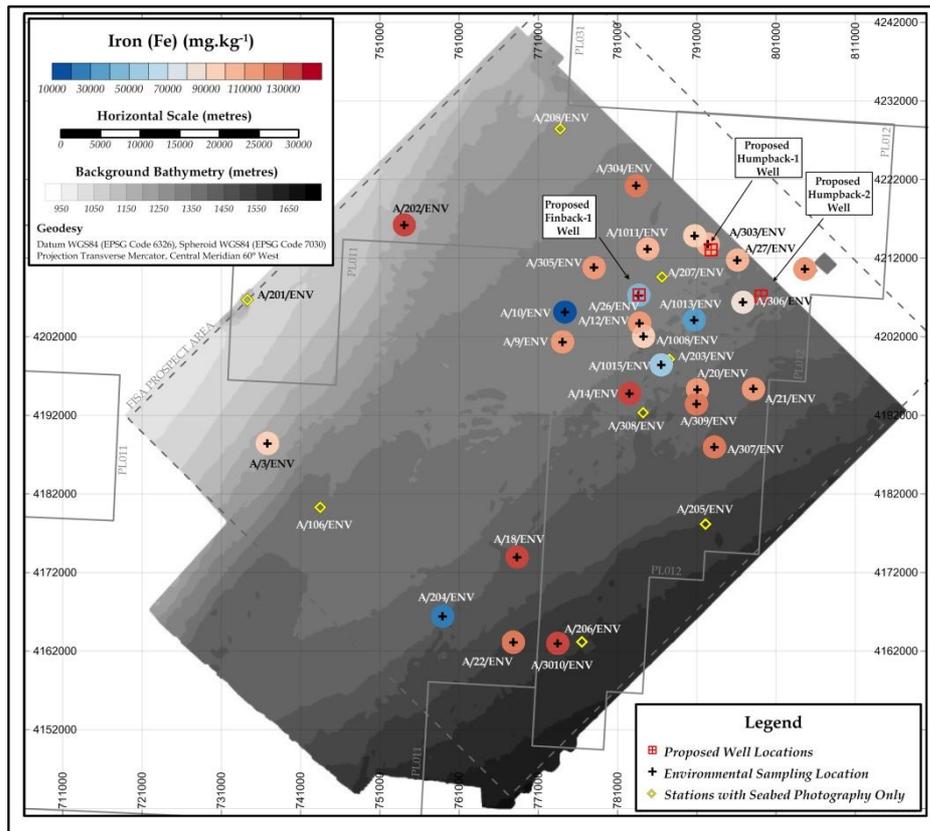
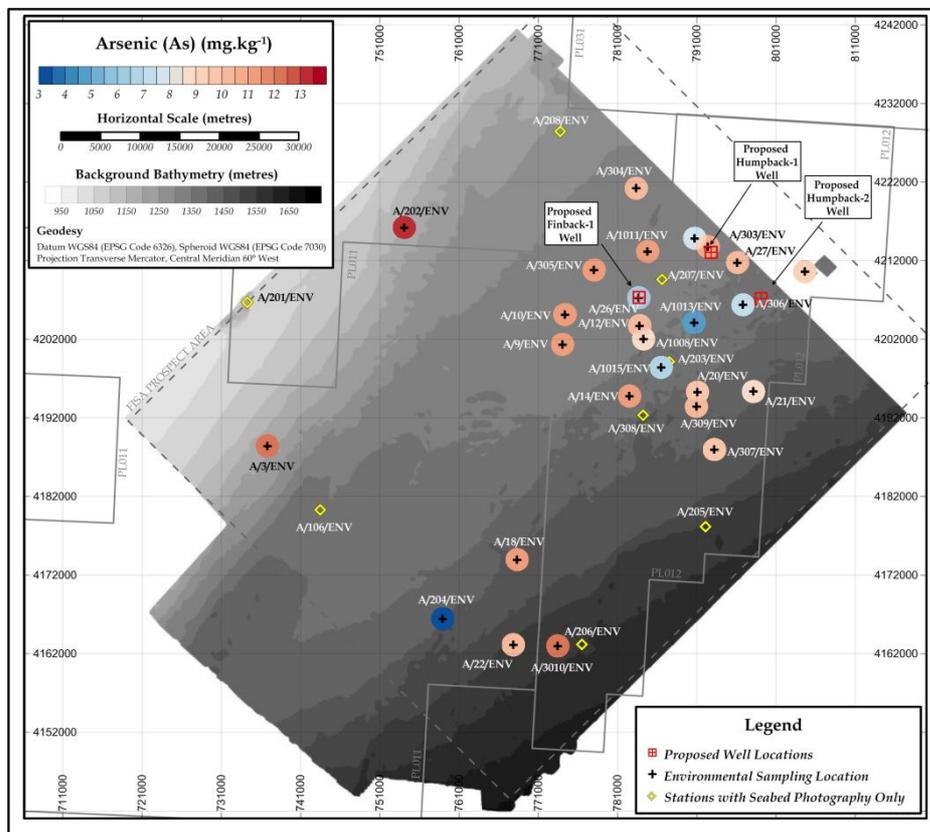


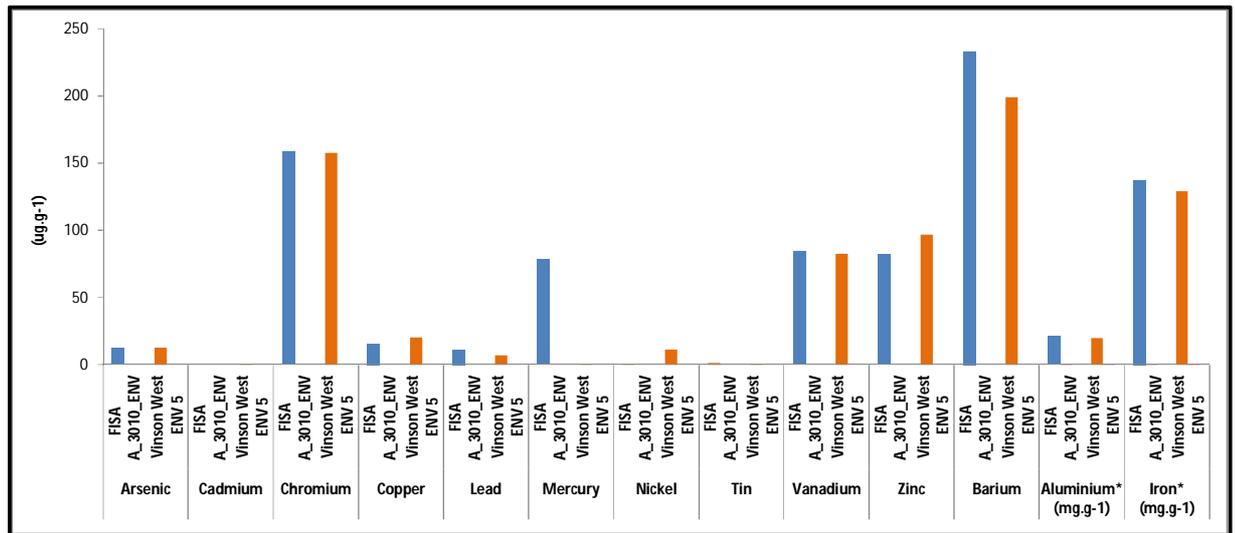
Figure 2.27 - Heavy Metal Concentration for Arsenic (As; mg.kg⁻¹)



2.7.1.1 Historical Comparison with Heavy Metals

A direct comparison was made between the results of station A/3010/ENV from the FISA area and Vinson West ENV5 sampled in 2011. Results are presented in Figure 2.28 which generally showed similar concentrations for all metals. Note that all values are based on $\mu\text{g.g}^{-1}$, with the exception of the crustal metals Fe and Al, which are presented at mg.g^{-1} .

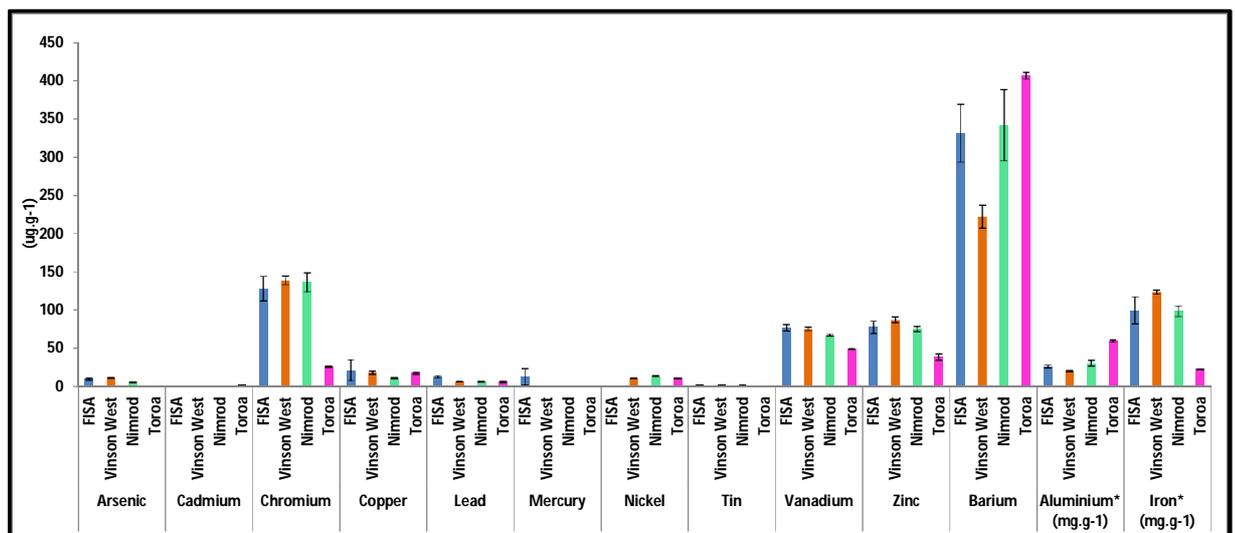
Figure 2.28 - Comparison of Heavy Metal for Stations FISA A/3010/ENV and Vinson West ENV 5



*Note that Aluminium and Iron are illustrated as mg.g^{-1} , while other heavy metals are $\mu\text{g.g}^{-1}$.

A further comparison between the mean values obtained for FISA and those obtained for the historical sites at Vinson West, Nimrod and Toroa Prospects are presented in Figure 2.29. These results are briefly discussed for each metal in Section 2.7.1.

Figure 2.29 - Comparison of Heavy Metal with Historical Data



*Note that Aluminium and Iron are illustrated as mg.g^{-1} , while other heavy metals are $\mu\text{g.g}^{-1}$.

Cadmium levels were similar to that previously recorded at the Vinson West ($0.1\mu\text{g.g}^{-1}$) but notably lower than levels obtained at Nimrod ($0.4\mu\text{g.g}^{-1}$) and Toroa ($0.9\mu\text{g.g}^{-1}$) sites (Table 2.6 and Figure 2.28).

Overall Hg levels were within the range previously recorded within the Vinson West area, which recorded a mean value of $0.02\mu\text{g.g}^{-1}$ via HF-digestion. Both of the historical sites Toroa and Nimrod displayed values of $<0.1\mu\text{g.g}^{-1}$ with both aqua regia and HF digest procedures (Table 2.6 and Figure 2.29).

With an overall mean value of $6.22\mu\text{g.g}^{-1}$ the FISA Pb concentrations were very similar to mean concentrations at the historical sites of Vinson West, Nimrod and Toroa, of $6.4\mu\text{g.g}^{-1}$, $6.2\mu\text{g.g}^{-1}$ and $5.8\mu\text{g.g}^{-1}$, respectively (Table 2.6 and Figure 2.29).

Previous surveys for Cr, Ni, Cu, V and Zn recorded similar values of $139\mu\text{g.g}^{-1}$, $10.9\mu\text{g.g}^{-1}$, $17.9\mu\text{g.g}^{-1}$, $75.3\mu\text{g.g}^{-1}$ and $87.1\mu\text{g.g}^{-1}$ for Vinson West; $136\mu\text{g.g}^{-1}$, $13.3\mu\text{g.g}^{-1}$, $10.7\mu\text{g.g}^{-1}$, $67\mu\text{g.g}^{-1}$ and $75.3\mu\text{g.g}^{-1}$ for Nimrod and $25.8\mu\text{g.g}^{-1}$, $10.5\mu\text{g.g}^{-1}$, $17.0\mu\text{g.g}^{-1}$, $49\mu\text{g.g}^{-1}$ and $38.5\mu\text{g.g}^{-1}$ for Toroa for the same metals, respectively (Table 2.6 and Figure 2.29).

A comparison with historical sites indicated a relatively high variability in both Fe and Al results. Vinson West and Nimrod displayed relatively similar data to that of FISA for Fe with 123.2mg.g^{-1} and 98.16mg.g^{-1} respectively as well as 18mg.g^{-1} and 30.52mg.g^{-1} for aluminium. In contrast, Toroa exhibited a decrease in Fe (22.38mg.g^{-1}) and an elevation in Al data (59.41mg.g^{-1}). This has been attributed the increased sedimentary regime found at this shallower site, where erosion has not exposed the older granular sediments (Table 2.6 and Figure 2.29).

The concentrations of As found at Vinson West, Nimrod and Toroa were also relatively variable ($11.2\mu\text{g.g}^{-1}$, $5.3\mu\text{g.g}^{-1}$ and $1.0\mu\text{g.g}^{-1}$) and therefore the result seen at FISA falls within the variability seen in this region (Table 2.6 and Figure 2.29).

2.8 WATER QUALITY

As little is known about the water column in the deeper waters of the proposed prospect area, several water quality profiles were undertaken within the FISA regional survey area. These replicate profiles provide a snapshot of the water column structure for the duration of the survey.

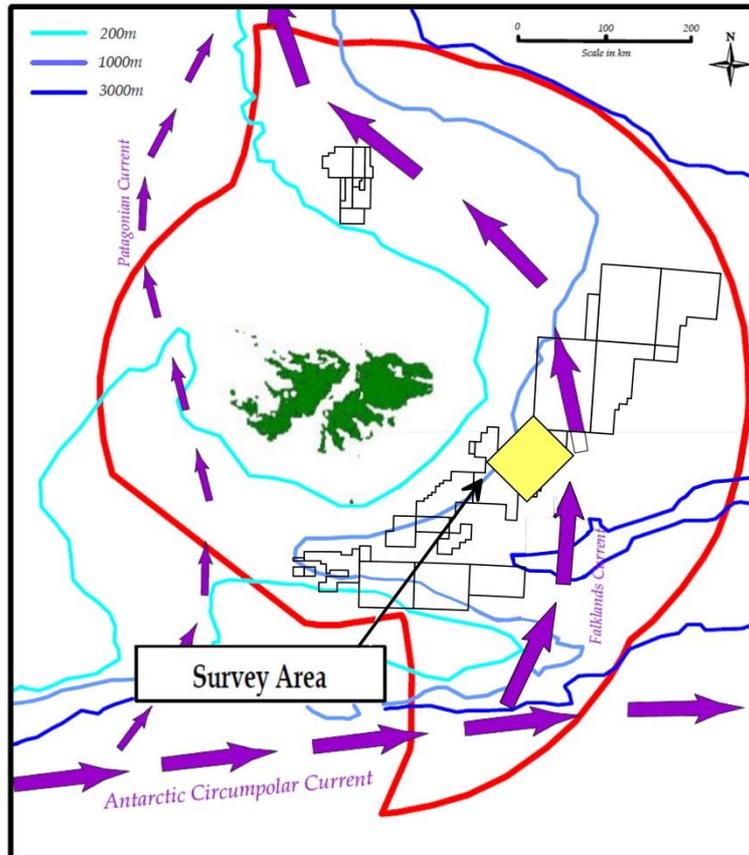
2.8.1 Regional Oceanography

This area is dominated by the temperate Malvinas current that follows the contour of the shelf slope from the south to the north (Falabella *et al.*, 2009). The sea water composition is strongly influenced by the sub-Antarctic waters and continental fresh waters (Acha *et al.*, 2004). With this sub-Antarctic water impact, the water columns in the Falkland Island Basin are usually cold, low in salinity and with a high dissolved oxygen and nutrient concentrations (Piola, 2008). The sea surface water temperature also follows a seasonal fluctuation resulting from a combination of multiple factors such as solar radiation, winds

and cloud cover. The average sea water temperature in the summer (January-March) is around 10°C and the annual salinity average in this area is around 33.7 psu (Palma *et al.*, 2008).

The general water circulation in the survey area is dominated by the Antarctic Circumpolar Current (ACC), which is located to the south of the Falklands in the Antarctic Polar Front or Antarctic Convergence where surface waters to the south meet warmer surface waters from the north. Travelling from west to east, the ACC passes around Cape Horn and then splits into two weaker current passing either side of the Falklands. These two northern components (collectively called the Malvinas current) are separated by the eastern flow following the southern edge of the Burdwood Bank before turning north to follow the eastern Falklands Continental Margin (Figure 2.30). The water masses influencing this site are therefore a result of this deviation of the ACC northwards, although it is unknown exactly how the current will affect the survey area within the South Falkland Basin, slightly west of this deviation. Evidence from the survey showed that softer deposited material only occurred in the shallower west, whilst elsewhere the seabed was either actively eroded or indicated were non-depositional sediments. This would suggest that stronger currents exist in the deeper waters to the east of the survey area. The presence on a non-zero bottom currents was directly verified by Harkema and Weatherly (1989, as cited in Garzoli, 1993). Their bottom current meters measured velocities of up to 10cm/s (Garzoli, 1993). Based on hydrographic data, it is believed that the Malvinas Current has a strong barotropic component and that it is well-mixed (Vivier and Provost 1999). A residual flow of water is expected to be from the south and west. This is supported by admiralty charts which show a surface flow east-north-east across the basin at velocities of up to 1.5 knots.

Figure 2.30 - General Current Circulation at the Proposed Survey Area



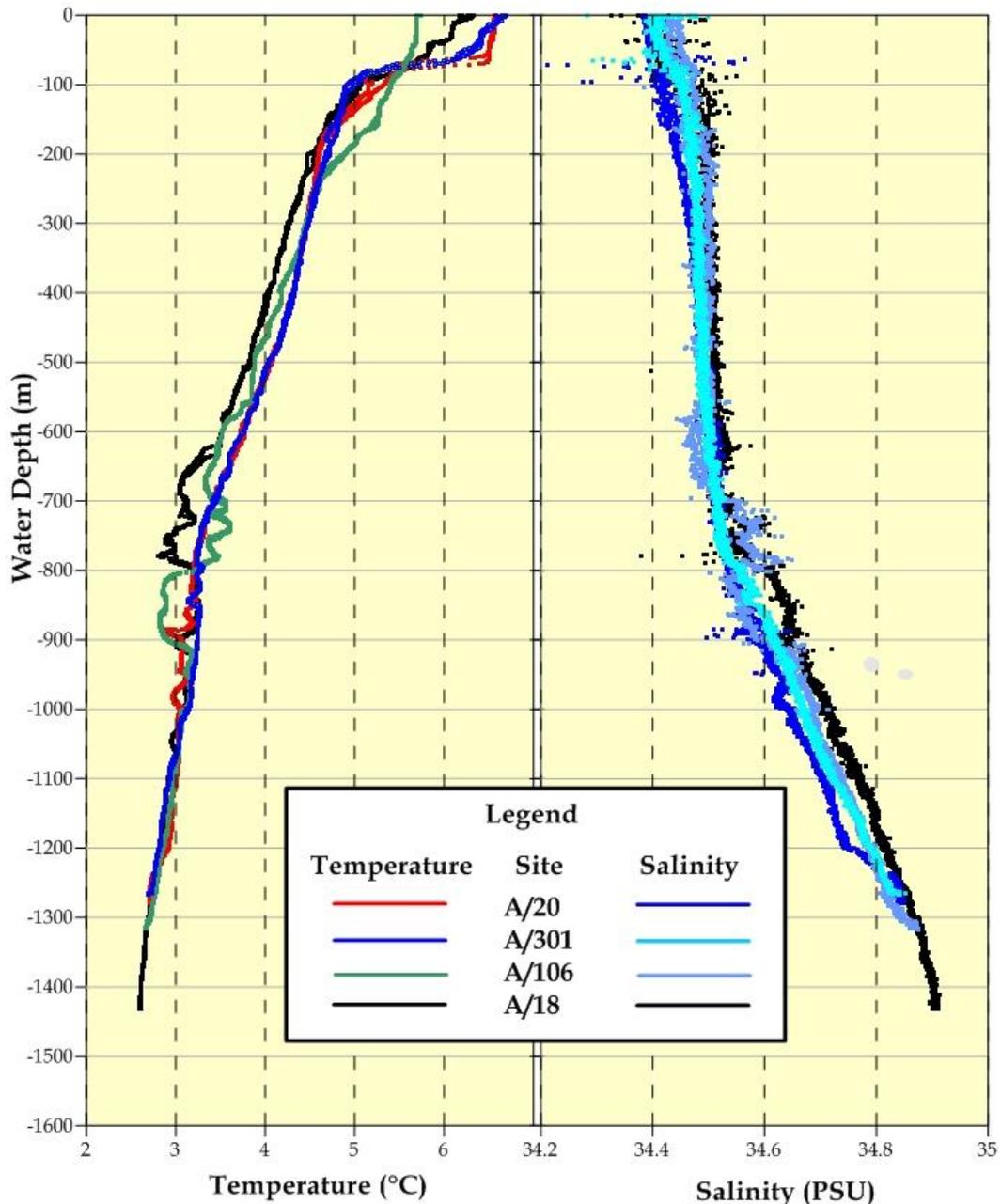
2.8.2 CTD Profiling (Conductivity-Temperature-Depth)

The structure of the water column within the FISA area was surveyed using a CTD probe with temperature and salinity sensors. A total of four profiles were recorded during the survey with the results presented in Figure 2.31, and the data extremes listed in Table 2.7. These datasets have been overlaid to show the consistency of the profile over depth and over a survey period of just over 1 month.

Table 2.7 - Water Quality Profile Extremes

| Profile | File name | Max Depth (m) | Date | Easting | Northing | Temp °C (Max/Min) | Salinity psu (Max/Min) |
|-----------|-----------|---------------|----------|---------|----------|-------------------|------------------------|
| A/20/ENV | FILE116 | 1055 | 03.01.14 | 791099 | 4195292 | 6.56/2.92 | 34.67/33.33 |
| A/301/ENV | FILE122 | 1267 | 08.01.14 | 790756 | 4214853 | 6.70/2.69 | 34.85/34.28 |
| A/106/ENV | FILE125 | 1317 | 16.01.14 | 743490 | 4180280 | 5.73/2.66 | 33.91/31.53 |
| A/18/ENV | FILE113 | 1432 | 01.02.14 | 768352 | 4173956 | 6.32/2.60 | 34.91/34.14 |

Figure 2.31 CTD Water Quality Profiles



Water profiles obtained from stations A/20/ENV, A/301/ENV, A/106/ENV and A/18/ENV all indicated similar and consistent profiles. Surface water temperature varied slightly from 5.73°C to 6.70°C with a small thermocline developing in the surface 80m. With increasing depth, water temperature continued to decline at a relatively consistent rate to around 2.60°C recorded at the deepest point (1432m). All profiles indicated a small area of mixed waters between 620m and 950m (Figure 2.31).

The salinity profiles for the same sites indicated a similar pattern, although the variability in salinity was limited to only 0.5psu throughout the whole water column. Whilst a subtle halocline was just perceptible in the surface 80m, the upper 600m of the water column remained relatively consistent, increasing in salinity by only 0.1psu. A slightly mixed zone

was then encountered between 620m and 950m, with a slightly greater increase in salinity recorded thereafter throughout the lower water column by around 0.4psu.

No significant differences were recorded between the four stations, indicative of one regional water column for this area and survey period.

2.8.2.1 Historical Comparison with Water quality data

Overall, a comparison with historical data from Vinson West, Nimrod and Toroa revealed similar results in regards to water temperature and salinity. Water temperature at the FISA site (~6°C) was lower than at the other three survey areas which recorded approx. 9°C at the surface. This could be due to a combination of seasonal effect and different water currents/masses mixing the surface layers. However, water temperature at the seabed was consistent throughout, recording values of ~2.6 °C. Higher seabed water temperature was measured at the Toroa site (~4.4°C) which is explained by the shallower bathymetry (~730m) at the seabed as opposed to around 1300-1600m in the other survey areas.

As with the temperature profiles, salinity profiles were relatively consistent throughout with no significant variations between survey areas. A negative correlation with temperature was observed in all profiles with the highest salinity encountered close to the seabed giving values of approximately 34.5psu.

2.8.3 Water Chemistry

Water quality at the proposed prospect area was good, as expected for an open 'blue-water' environment. Water samples were acquired at different water depths at two locations. Methods and analytical techniques are outlined in Appendix I and II with results presented in Table 2.8.

Table 2.8 - Water Quality Measurements for A/03/ENV

| Description | Detection limit (mg/l) | A/03/ ENV (1m) | A/03/ ENV (50m) | A/03/ ENV (400m) | A/03/ ENV (1134m) | A/23/ ENV (1m) | A/23/ ENV (50m) | A/23/ ENV (500m) | A/23/ ENV (1110m) |
|--|------------------------|----------------|-----------------|------------------|-------------------|----------------|-----------------|------------------|-------------------|
| Barium as Ba (Total) | 0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| Phosphorus as P (Total) | 0.1 | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 | <0.1 |
| Silicon as Si (Total) | 0.1 | 0.2 | 0.3 | 0.4 | 1 | 0.4 | 0.4 | 0.6 | 0.5 |
| Total Sulphur as SO ₄ (Dissolved) | 3 | 2320 | 2600 | 2040 | 2560 | 4796 | 4593 | 4222 | 4296 |
| Arsenic as As (Total) | 0.001 | 0.010 | 0.012 | 0.008 | 0.011 | 0.023 | 0.024 | 0.024 | 0.027 |
| Cadmium as Cd (Total) | 0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| Chromium as Cr (Total) | 0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Copper as Cu (Total) | 0.001 | 0.003 | 0.003 | 0.002 | 0.003 | 0.007 | 0.008 | 0.007 | 0.007 |
| Lead as Pb (Total) | 0.001 | <0.001 | <0.001 | <0.001 | 0.001 | <0.001 | 0.001 | 0.001 | <0.001 |
| Mercury as Hg (Total) | 0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 | 0.0004 | 0.0004 | 0.0003 | 0.0003 |
| Nickel as Ni (Total) | 0.001 | 0.002 | 0.002 | 0.001 | 0.002 | 0.006 | 0.005 | 0.004 | 0.004 |
| Zinc as Zn (Total) | 0.002 | 0.023 | 0.01 | 0.008 | 0.038 | 0.029 | 0.041 | 0.048 | 0.035 |
| Suspended Solids w | 5 | <5 | 9 | 5 | <5 | <5 | <5 | <5 | <5 |
| Total Alkalinity as CaCO ₃ | 2 | 116 | 116 | 124 | 126 | 112 | 114 | 114 | 120 |
| Total Acidity as CaCO ₃ | 2 | Nil | Nil | Nil | Nil | Nil | Nil | Nil | Nil |
| Nitrite as N | 0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 |
| Nitrate as N | 0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 |
| Total Oxidised Nitrogen as N | 0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 |
| Sulphide as S | 0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Total Organic Carbon | 0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 |
| Kjeldahl Nitrogen as N | 1 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 | <1.0 |
| Total Nitrogen as N | 1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Dissolved Organic Carbon | 0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 | <0.2 |
| Total Oil (µg/l) | 0.001 | 0.010 | 0.082 | 0.078 | 0.011 | 0.012 | 0.0095 | 0.0094 | 0.0096 |
| Total n alkanes | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 |
| Pristane | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 |
| Phytane | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 | 0.001 |

Note: All values are shown in mg/l (excluding alkanes)

All results showed very low levels or non-detectable concentration for all tests. As the water mass seems to be the same in the area (correlate with CTD casts), we can presume that these low values are representative of the entire FISA region.

2.9 MACROFAUNAL ANALYSIS

A macrofaunal analysis was carried out on all 59 replicates obtained at the 26 key baseline sediment sites, sampled within the FISA survey area with a particular concentration of stations around the proposed Finback-1 and Humpback-1 well locations. The sediments throughout the survey area generally conformed to silty SAND although gravel exposures and occasional drop stones were common, as identified in the seabed photography as well as sample residues. Macrofaunal samples were processed in the field using a 500µm mesh size. Subsequent macrofaunal taxonomy of all recovered fauna identified a total of 2,874 individuals (infauna and epifauna) from the 59 samples analysed. Faunal data for each sample are listed in Appendix VII, whilst univariate analyses are summarised in Tables 2.10 and 2.11. Of the 254 species recorded, 188 were infaunal, consisting of 69 annelid species and accounting for 48.4% of the total individuals. The molluscs were represented by 32 species (6.4% of total individuals), the crustaceans by 62 species (42.1%) and the echinoderms by only 13 species (and only 2.6% of total individuals), while all other groups (Cnidaria, Nemertea, Nematoda, Sipuncula, Echiurida, Turbellaria, Brachiopoda and Chaetognatha) accounted for the remaining 8.5%, or 12 species. A distribution of the different taxa is presented in Figures 2.32 and 2.33 by sample replicate, or Figure 2.34 by station.

With the exception of species that have been intentionally grouped into higher taxonomic levels (e.g. Nematoda, Nemertea, etc.), the majority of adult specimens were identified to genus level, or higher. This was approximately 88% of specimens (excluding juveniles and fragmented species) 18% of which were identified to species level. Only three juvenile specimens (Asterozoa, Holothurozoa and Isopoda) were recorded throughout the survey area (possibly reflecting the mid-summer timing of the sampling). Conversely, the polyp stage of Scyphozoa was recorded in reasonably high numbers, accounting for approximately 16% of all individuals recorded. The scyphozoan polyp stage is significantly suppressed within their life cycle, and can be classed as a juvenile form until the adult medusa stage is achieved. Juveniles are often excluded from community analyses (in accordance with OSPAR commission guidelines, 2004) due to having high mortality prior to reaching maturity, as well as difficulties in distinguishing species of the same genus and different life cycle stages. Consequently, they tend to induce a recruitment spike at certain times of the year due to rapid settlement and colonisation, but are essentially an ephemeral part of the population masking the underlying trends within the mature adults. Therefore, the two juveniles were excluded from the multivariate analyses.

Figure 2.32 Proportion of Individual Abundance by Main Group and Replicate

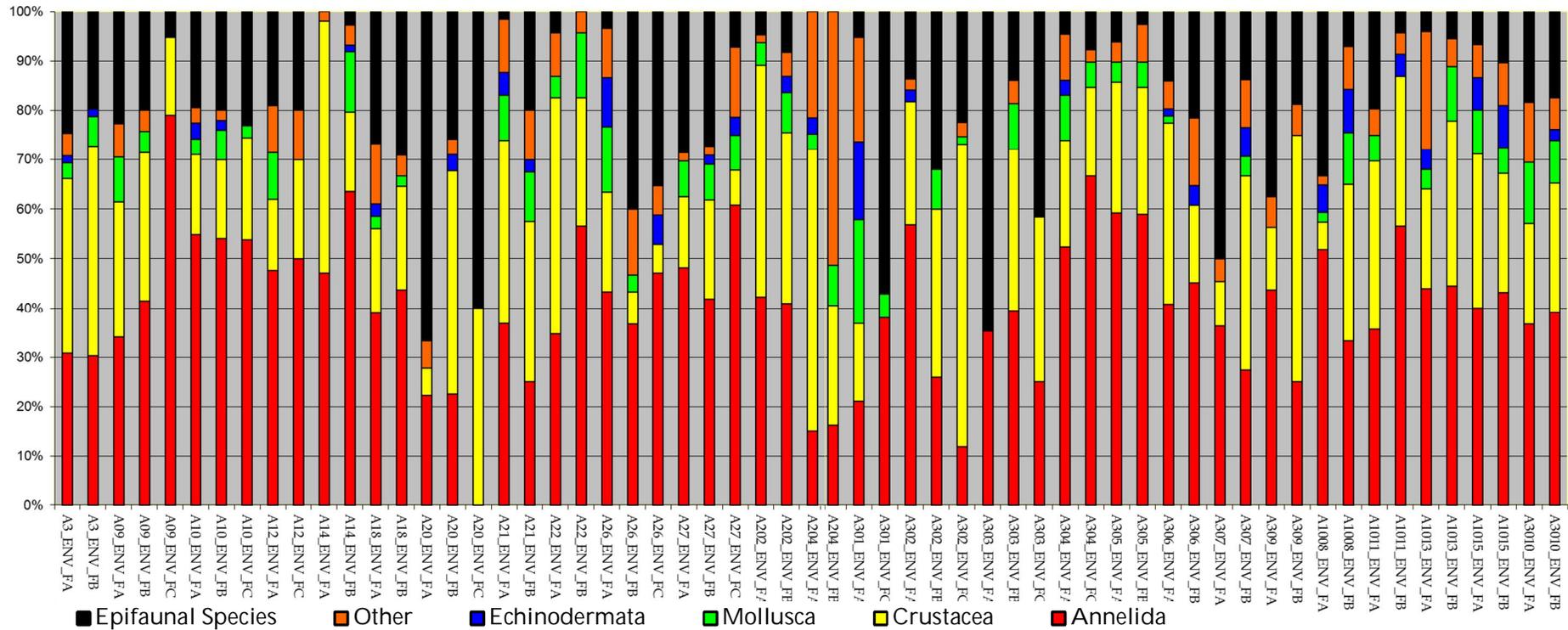


Figure 2.33 Proportion of Species Richness by Main Group and Replicate

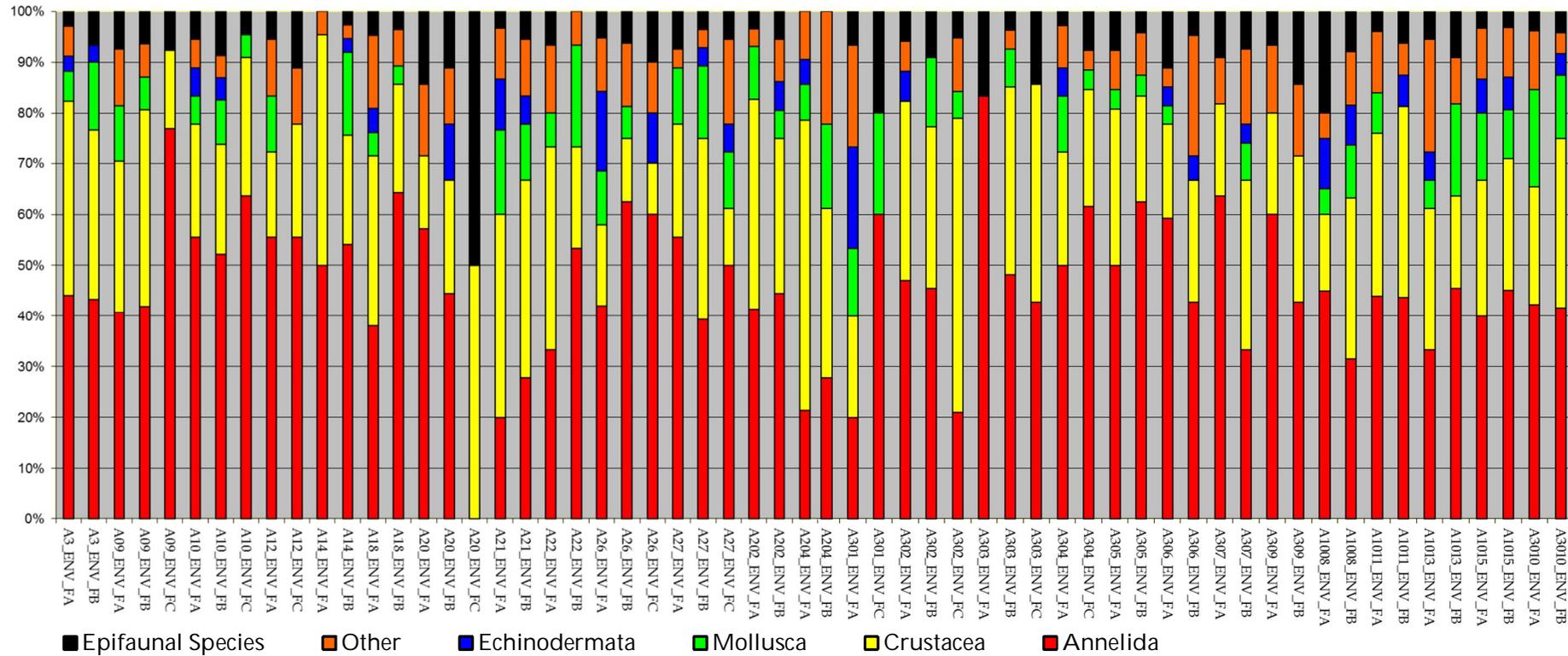
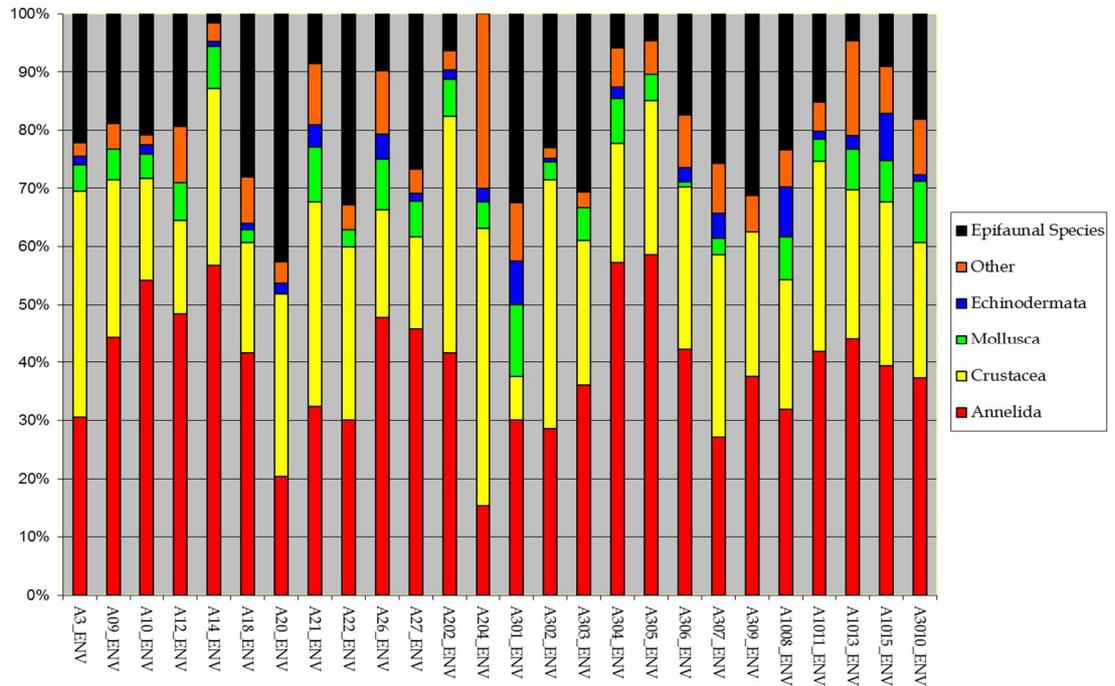
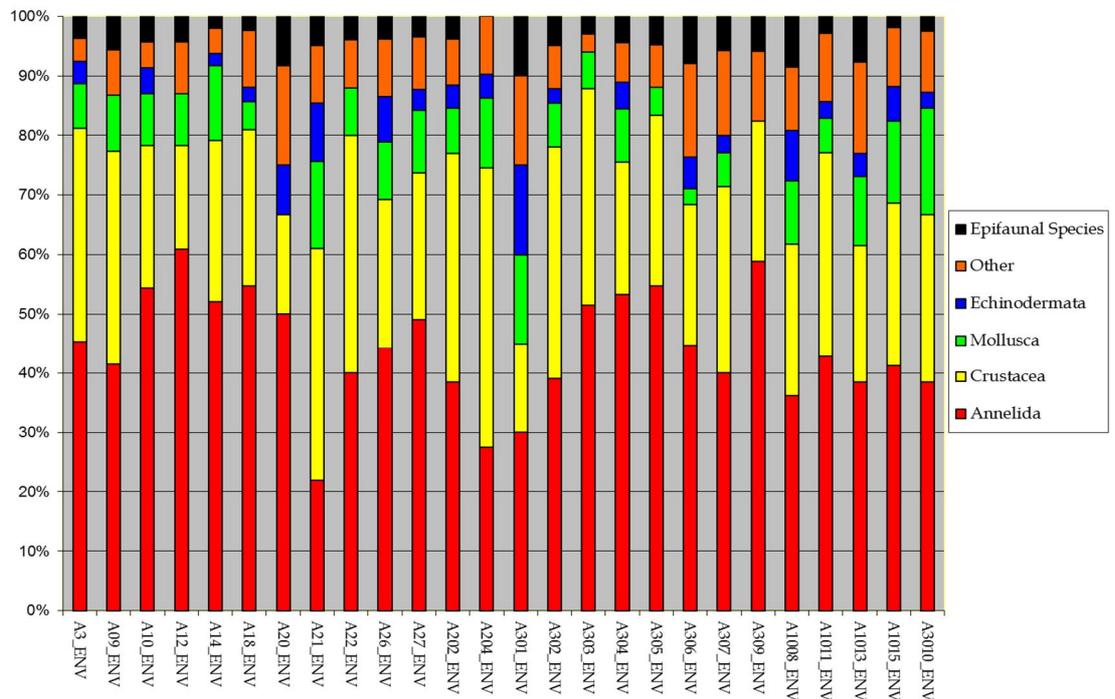


Figure 2.34 Proportion of Individual Abundance and Richness by Main Group and Station

(a) Individual Abundance



(b) Species Richness



Fragments of one species were also recorded accounting for only one specimen or 0.04% of the community recorded. This has also been excluded from the multivariate analyses as it is not often possible to differentiate the number of animals from which fragments are derived. The juveniles and fragmented species have been listed separately in Appendix VII.

2.9.1 Infaunal trends

The macrofauna for the FISA site was as expected for the South Falklands Basin of this sediment type and water depth. Unlike most deep-water low energy environments, the seabed exhibited a varied seabed with a mixed soft and granular sediment community generally dominated by small polychaetes. This dominance was shown in both abundance and richness, with six polychaetes recorded in the top ten numerically ranked species. This dominance was closely followed by crustaceans which were also well represented, having three species in the in the top ten numerically ranked taxa. In overall rank order, the key dominant species across the area were the Polychaete *Rhaphobranchium (Spinigerium) ehlersi*, the crustaceans *Ampelisca* and a Copepoda, separated by the polychaete *Gymnonereis fauveli* and followed by *Apistobranchus* sp I. A Sipuncula worm *Nephasoma (Nephasoma) diaphanes* was ranked 6th followed by the crustacean *Urothoe*. The polychaetes *Caulleriella*, Maldanidae and *Euchone pallida* were ranked 8th, 9th and 10th, respectively.

Table 2.9 - Overall Species Ranking (Top 15 Species)

| Overall Top 15 Rank | Species/Taxon | Total rank score (out of 590) | Numerical Abundance (59 replicates) | Numerical Top 15 rank |
|---------------------|--|-------------------------------|-------------------------------------|-----------------------|
| 1 | <i>Rhaphobranchium (Spinigerium) ehlersi</i> | 465 | 247 | 1 |
| 2 | <i>Ampelisca</i> sp. A | 186 | 60 | 4 |
| 3 | <i>Gymnonereis fauveli</i> | 183 | 63 | 3 |
| 4 | Copepoda | 183 | 106 | 2 |
| 5 | <i>Apistobranchus</i> sp I | 176 | 56 | 5 |
| 6 | <i>Nephasoma (Nephasoma) diaphanes</i> | 166 | 54 | 7 |
| 7 | <i>Urothoe</i> | 162 | 56 | 5 |
| 8 | <i>Caulleriella</i> | 154 | 51 | 8 |
| 9 | Maldanidae | 127 | 40 | 11 |
| 10 | <i>Euchone pallida</i> | 124 | 38 | 13 |
| 11 | <i>Leptognathia</i> | 122 | 42 | 10 |
| 12 | <i>Spiochaetopterus typicus</i> | 111 | 32 | 16 |
| 13 | <i>Melinna cristata</i> | 107 | 33 | 15 |
| 14 | Nematoda | 90 | 26 | 20 |
| 15 | <i>Eunoe</i> | 86 | 23 | 23 |

A measure of the overall dominance pattern in the sampling area was achieved by ranking the top species per sample replicate according to abundance, giving a rank score of 10 to the most abundant species, decreasing to 1 for the tenth most abundant species, and summing these scores for all 59 samples to provide an overall dominance score (Eleftheriou and Basford, 1989) for each species. The top 15 species are shown in Table 2.9. This ranking varied only slightly from that of the numerical ranking for the species overall with 12 out of the 15 species recorded in both ranks, further highlighting the general homogeneous yet broad nature of the community between sample replicates. Despite slight variations in order, the same top five species were recorded in both lists. The most dominant ranked species, the polychaete *R. ehlersi* recorded a score of 465 out of 590, identifying this species as the most dominant species throughout the replicates

acquired. Generally all phyla identified were typical for this deep sea region. While the species richness was high, individual counts were typically low, demonstrating a diverse benthic community.

Within the processed samples, residues consisted of black sand (of basaltic origin) with a varying amount of gravel and pebbles, mostly volcanic with many very rounded and polished by glacial transport. Many of these pebbles carried epifauna mostly consisting of sponges and Bryozoa and are discussed in Section 2.9.5. Biological residue was scarce or non-existent, with only very rare empty shells and on some stations dead *Lophelia* and Stylasteridae (Hydrozoa) remains. A discussion on the recovered faunal groups is discussed below. As a general comment, a bipolar distribution is quite common in deep sea fauna, especially in the polychaeta, with morphologically identical species found at both poles. Whether this morphological similarity extends towards the genetic material, is yet unproven. However a very wide to (in some cases) cosmopolitan distribution is a common feature of deep sea fauna.

Further comments relating to the macrofaunal population and their separate phylogenetic groups are presented below with example images of some specimens shown in Figure 2.25a-f:

Brachiopoda: Three species of Brachiopoda were recorded. An example species includes *Pelagodiscus atlanticus* which resembles a limpet, and has a virtually cosmopolitan distribution.

Sipunculids: Two species of Sipuncula were recorded. *Nephasoma diaphanes* (Figure 2.35g) is widespread in the deeper waters of the North and South Atlantic. *Golfingia margaretaea* ranges from the shallow waters of Europe to the Antarctic.

Polychaeta: This phylum provides the bulk of the infauna, with many common genera. Most prominent was the quill worm *Rhynchobranchium ehlersi* (Figure 2.35a), with frequently large number of specimens recorded in the samples, many of which reaching a good size. A couple of observations would point towards this species actually brooding its young. One specimen was found within a tube with juveniles tightly packed together. Another tube contained an adult followed by a chain of eggs/embryos filling the space behind. This brooding is found in many Antarctic species. The second most abundant polychaete was represented by *Apistobrachus* sp. I followed by *Gymnonereis fauveli* (Figure 2.35c).

Nothria conchylega is another quill worm with a bipolar distribution. Quill worms are predators, as are *Nephtys*, *Aaglaophamus* and *Lumbrineris* spp.. *Euphosine* spp. are short bodied polychaetes which

feed on sponges. This genus is generally rarely recorded but *Euphrosine cirrata* is a species common to the North and South Atlantic.

Most non-errantia polychaete species are sediment feeders, e.g. Cirratulidae (*Tharix*, *Caulleriella*, *Cirriformia* and *Cirratulus* spp.) and Maldanidae. Filter feeders included members of the family Sabellidae, of which the commonest was *Euchone pallida*.

Two Terebellid species should be mentioned here, *Terebellides stroemi* and *Melinna cristata*. These are further examples of species common to both hemispheres.

Crustacea:

Crustacea is one of the most common and diverse phyla recorded in the Antarctic and sub-Antarctic.

Amongst the barnacles, several specimens of a *Scalpellum* species (stalked or goose barnacle) were recorded. Another barnacle was *Altiterruca*, an asymmetrical raised species commonly found on *Lophelia* debris in the FISA area.

Amphipoda were very common; however notoriously difficult to identify to species level. It proved however possible to place a high proportion into genera or phyla, and to distinguish different species by characterising features. Common were *Ampelisca* (Figure 2.35b), *Urothoe* and Phoxocephalidae spp.. Some notably conspicuous specimens belonged to the Genus *Eusira*. While *Ampelisca* is known as a filtration feeder, most others are likely to feed on debris.

The Isopoda are very well represented by all the common deep sea genera, e.g. *Desmosoma*, *Haploniscus*, *Ischnomesus*, etc.. Most of the species are without eyes, given lack of light in this habitat. Despite comparatively recent keys it was not possible to go further than to genus level for this group; with some estimations quoting five out of six species as undescribed. Little is known about their life history and style.

The family Serolidae, typical of Antarctic and sub Antarctic waters, was not found although this species was recorded in a number of the seabed photographs acquired during the survey. An example image is shown in Section 2.10.1 in Figure 2.51 (poss. *Acutiserolis neaera*).

Tanaidacea were common in the samples, often very small specimens, with bigger specimens belonging to the genera *Apseudes* and similar *Pseudospyrapus*, the latter lacking an antennal scale.

Cumacea were somewhat scarce, but nevertheless a number of species could be identified to species level. They are suspension feeders burrowed in the upper layers of the sediment. The genera have a very wide distribution, and can be found at varying water depth.

Mollusca: This group proved fairly diverse, with some interesting material. The most interesting find is of a Monoplacophoran, likely *Neopilina* (Figure 2.35e). This group was thought to be extinct for 400 million years until the Danish zoologist Henning Lemche discovered and described living specimens from the deep sea, obtained during the Galathea expedition. These are acceptably placed on a side branch of evolution and thought to be related to the chitons. These animals are very rare, and so far only about 150 specimens have ever been found worldwide, belonging to about 25 species. Only one specimen was found at station A/21/ENV, which is about 20km away from the proposed well locations.

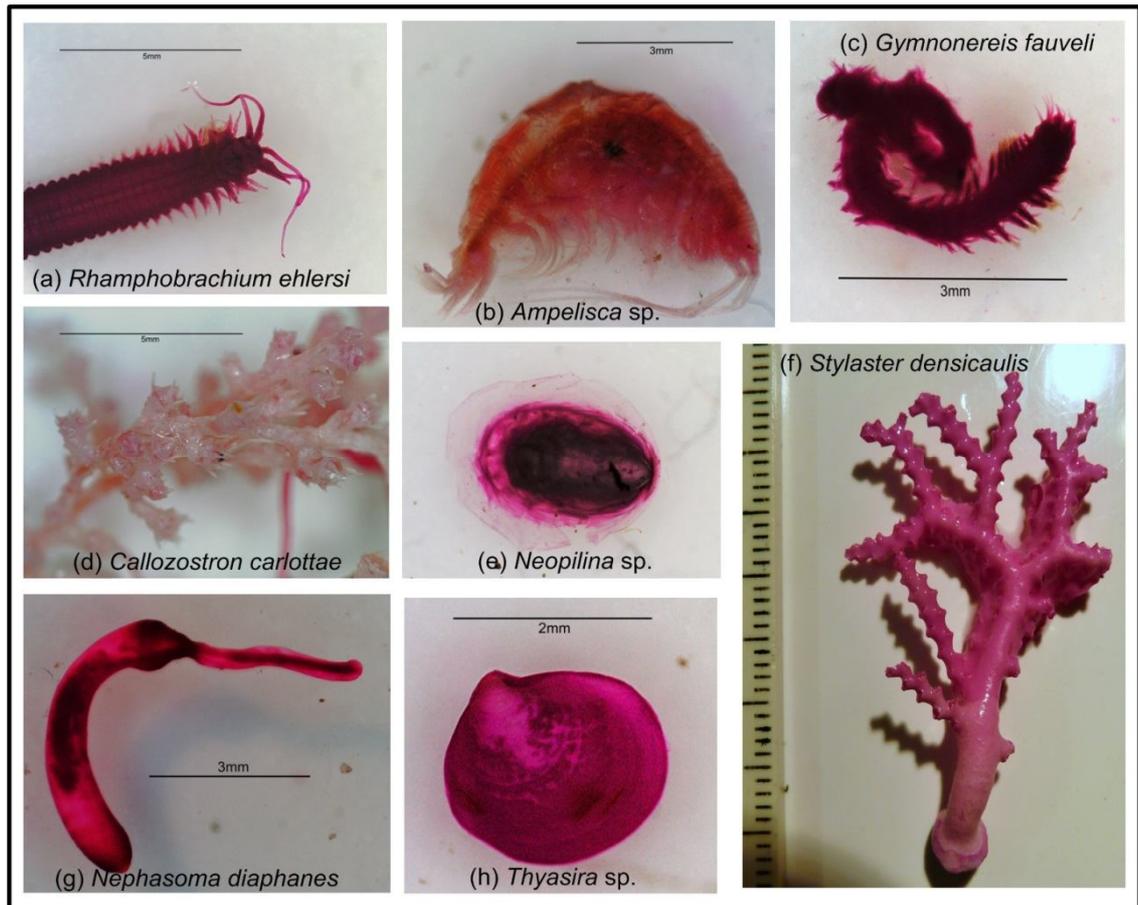
The species of Mollusca recorded are typical members of deep sea or deep water fauna, e.g. *Thyasira* (Figure 2.35h) *Skenea*, *Limopsis*, *Dacrydium*, *Limatula*, *Nuculana* and *Lyonsiella* etc. The fauna of this zoogeographical area known as the Magellanic region is poorly known.

Echinodermata: As expected, Ophiuroidea are the most common, in this case *Amphiura belgica* (named after a research vessel). A number of other species were recorded, most common of these was *Ophiozonella falklandica*.

Typical of deep sea fauna are very small holothurians, *Myriotrochus* sp. (with wheel like spicules in the dermis) and another species entirely without any calcareous bodies. These species are extremely small, with complete specimens rarely more than one mm in length.

Overall, the benthic community is of a typical Deep Sea habitat, with diverse species, low numbers and high variation between samples, with many species typical of bathyal and abyssal depth.

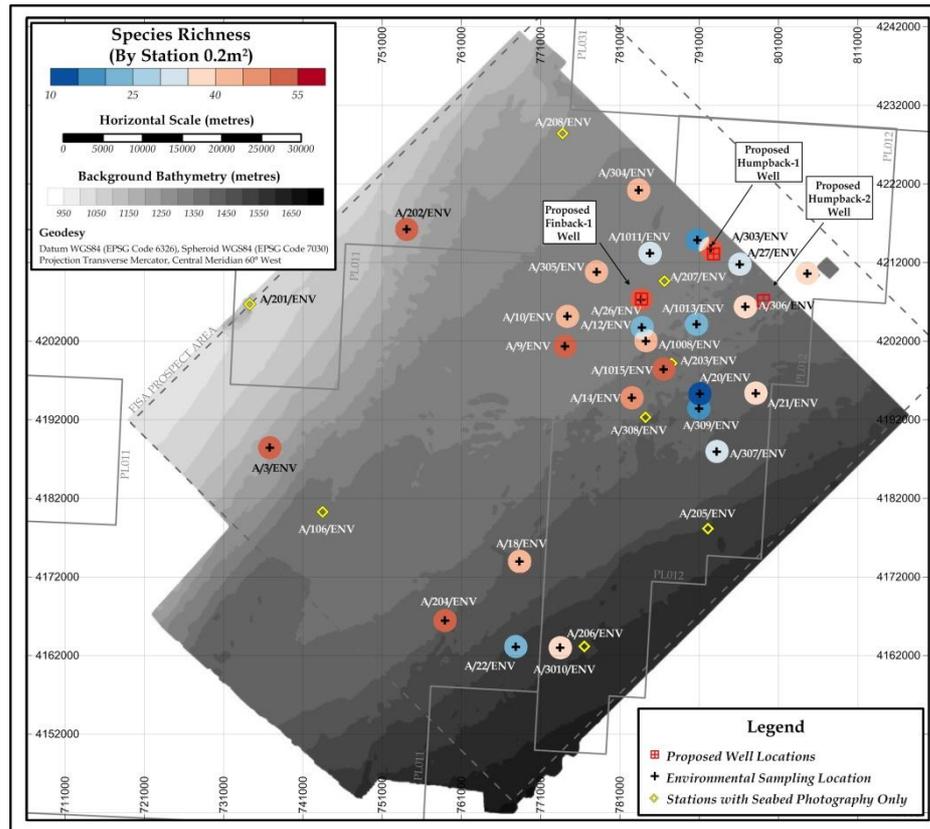
Figure 2.35 Example Macro-invertebrate Specimens Recorded during the Survey



2.9.2 Univariate Parameters

The primary and univariate parameters are listed for individual macrofaunal replicates, together with aggregated stations in Tables 2.10 (by replicates) and Table 2.11 (by stations), respectively. The average number of species per 0.1m² replicate for the FISA survey area is 20 ($\pm 9SD$), these are comparative to the other surveys performed in the South Falklands Basin which ranged from an average of 20 to 29 species per 0.1m² (Figures 2.36 and 2.37). The abundance of individuals recorded during this study, per m², was generally as expected at 351 ($\pm 185SD$), comparative to historical sites in the South Falklands Basin. This varied between 305, 407 and 417 per m² for Nimrod, Toroa and Vinson West, respectively (Figures 2.38 and 2.39).

Figure 2.36 Macrofauna – Species Richness (per 0.1m²)



The 2011 Vinson West survey area appears to have a marginally higher mean species abundance and richness than the current survey, although remained very similar compared to Nimrod and Toroa. This may be due to the large variability of habitats recorded during the FISA study, not encountered during the earlier surveys which were limited to a much smaller area. This is supported by the larger variance in standard deviation (shown as an error bars on Figure 2.37). As a direct comparison sites A/3010/ENV and Vinson West ENV 5 have a closer similarity in species abundance of 78 and 85 and richness of 39 and 45 per 0.2m², respectively. This equates to only a marginally lower survey mean of around 13.3% for abundance and 8.3% richness, for the current survey overall. The relationship between the number of taxa and the individuals is shown in Figure 2.40.

Figure 2.37 Macrofauna – Species Richness Comparison (per 0.1m²)

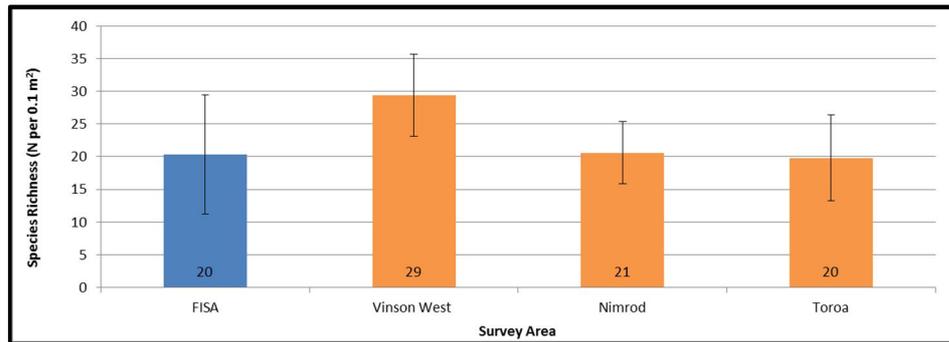


Figure 2.38 Macrofauna – Species Abundance (per m²)

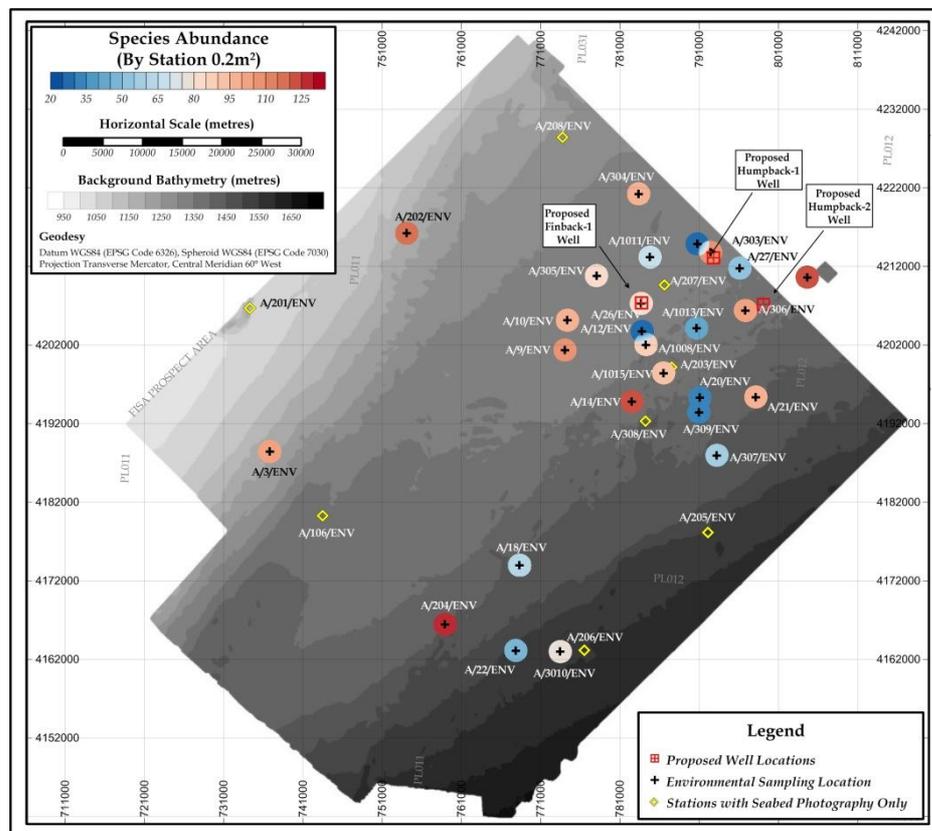


Figure 2.39 Macrofauna – Species Abundance Comparison (per m²)

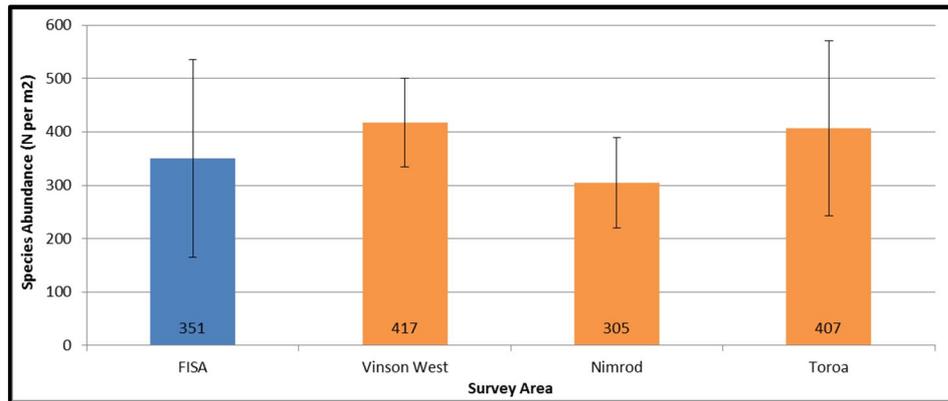
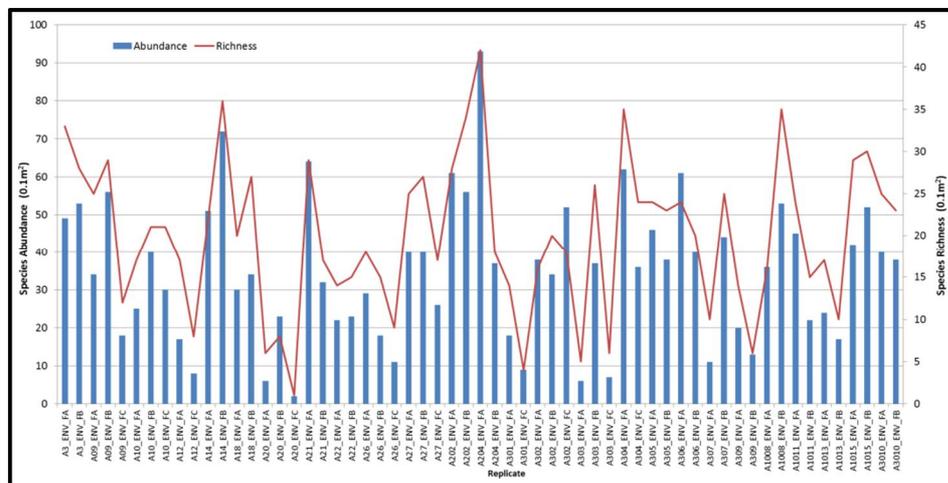
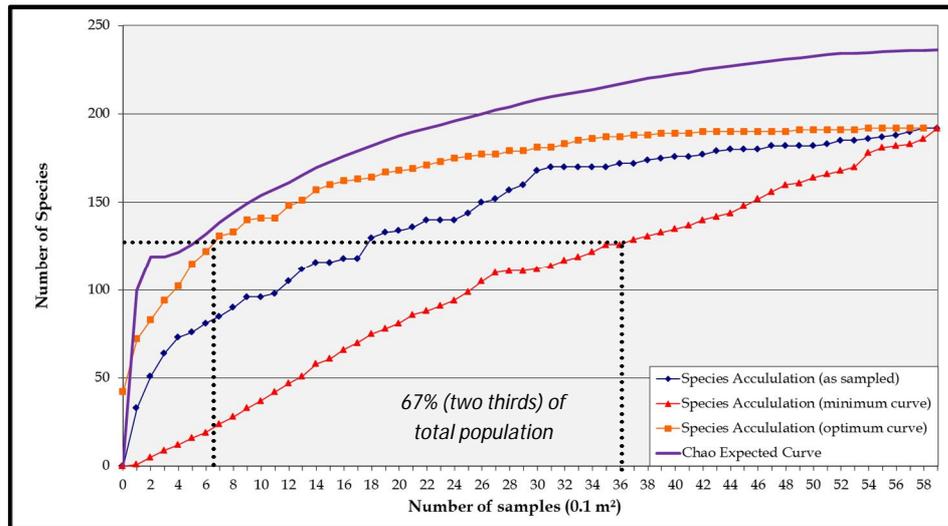


Figure 2.40 Species Abundance and Richness by Replicate (0.1m²)



The consistent accumulation of taxa with each replicate is demonstrated in a bioaccumulation curve shown in Figure 2.41. The minimum curves in this figure demonstrate the slow incremental increase in species accumulation, as additional replicates were acquired. This suggests that the population is quite diverse and representative of different habitats with a consistent but relatively low species richness being recorded in every new sample. By interpolation, this shows that between 6.5 and 36 x 0.1m² replicates are required to recover a representative proportion (i.e. 67% or 129 species) of the overall population.

Figure 2.41 Bioaccumulation Curve of the FISA Survey Area



By further interpolation, the theoretical total species populations and accrual by replicates was assessed using Chao-1 analysis (see Appendix II). This analysis estimates the maximum species accumulation for FISA site to be 236, compared to the actual 188 infaunal species recorded during the survey. This large degree of variability is due to the relatively low species richness recovered throughout the survey, requiring a significant number of samples to achieve asymptote.

The Shannon-Wiener Diversity remained at a relatively high mean level by replicate at 3.801 and slightly variable with a 24.7% variance (i.e. the proportion of standard deviation against the mean). By station, this mean increased to 4.64 and a variance of 13.8% (Figure 2.42). The Pielou's Equitability by station was low with a mean of 0.899 (± 0.05 SD or 6.5% variance) indicating only a weak species dominance across the sampling template. Margalef's Index (Species Richness) by station recorded a mean of 8.45 (± 2.7 SD or 26.62% variance) whilst Simpson's dominance illustrated a quite diverse community and low dominance at 0.945 (± 0.05 SD or 5.6% variance; Figure 4.39). All results conclude a relative low richness suggestive of a broad community with no particularly dominant species.

Figure 2.42 Macrofauna – Shannon-Wiener Diversity (H(s))

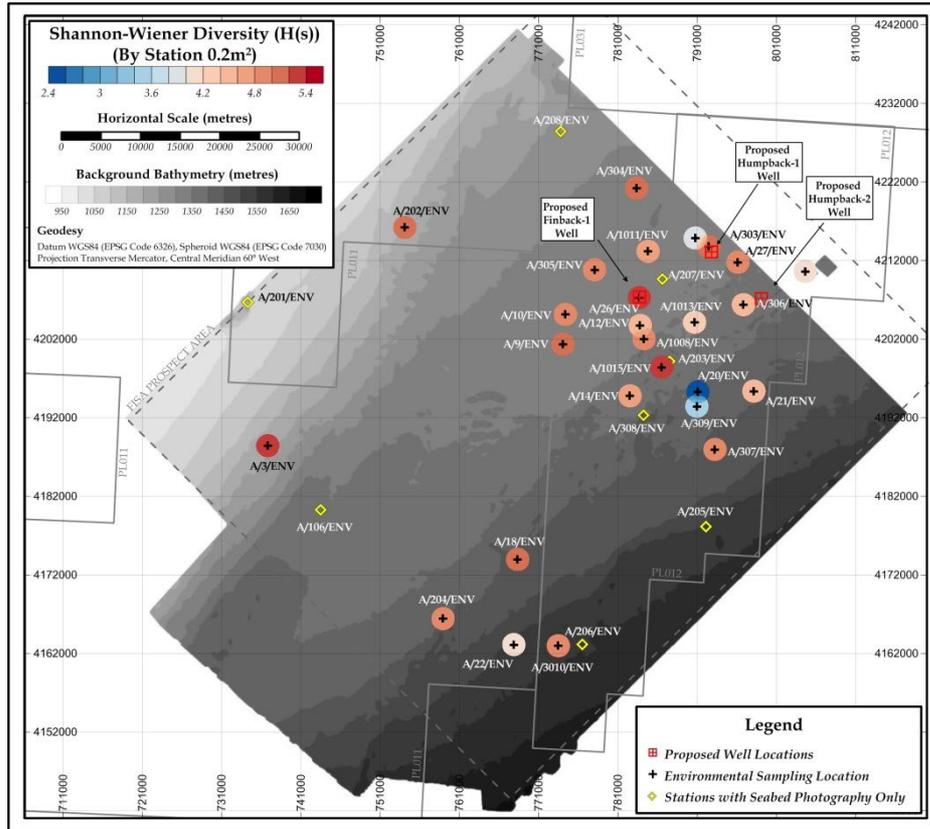
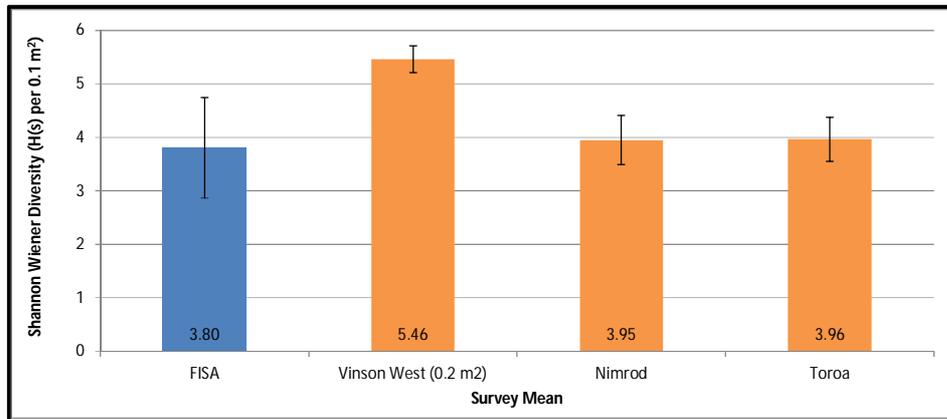


Figure 2.43 Macrofauna – Shannon-Wiener Diversity (H(s)) Comparison

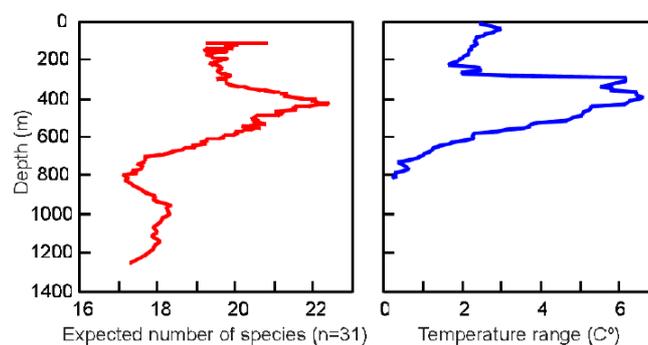


The large diversity within the FISA area, containing a relatively high species richness compared with abundance is typical of deep sea benthic community in this region and remains within the mean abundance and richness of the previous surveys completed in the region at Vinson West, Toroa and Nimrod.

The Atlantic Frontier Environmental Network or AFEN surveys (2000) showed that there was a general decline in diversity (or increase in dominance) with depth, consistent with

the low diversity of Arctic and Norwegian Basin deep-sea benthos. A close link between the hydrography was shown to exist within the macrobenthos for intermediate depths within the Faroe-Shetland Channel (Figure 2.44). Diversity was shown to be enhanced in the region of maximal temperature variation (~400m and 6°C), where the mixing of water masses supports both sets of fauna, increased niche availability and biotic interaction over well-developed epifaunal communities. Based on this model, where the depths of the survey area are recorded (~1000 – 1300m), we would expect a species richness of around 17 to 18 taxon/0.1m², indicating the average of 20 taxon/0.1m² observed in this survey to be higher than that observed at a similar bi-polar depths.

Figure 2.44 Illustration of correlation between diversity and water temperature, west of Shetlands



This relatively high diversity reflects the high number of species relative to generally low numbers of individuals recorded for many of those species. The most dominant species was represented by the polychaete *Rhamphobrachium (Spinigerium) ehlersi*, which was recorded in 52 of the 59 samples acquired at an average density of 42 per 1m², throughout the FISA survey area. This was followed by an amphipod *Ampelisca* sp. A and polychaete *Gymnonereis fauveli* which were present in 29 and 25 of the samples with an average density of 10.2 and 10.6 individuals per 1m², respectively. To further highlight the low dominance recorded throughout the survey only four taxa (*R. ehlersi*, *A. sp. A*, *G. fauveli* and a copepod), were recorded during the survey with an average density of more than one individual per sample (0.1m²), with 44 of the 188 infaunal taxa recorded represented by only a single specimen for the entire survey.

Table 2.10 - Univariate Faunal Parameters (0.1m² replicates)

| Station | Number of Species per 0.1m ² (S) | Number of Individuals per 0.1m ² (N) | Shannon-Wiener Diversity | Simpsons Diversity (1-Lambda') | Evenness (Pielou's Evenness) | Richness (Margalef) |
|---------------|---|---|--------------------------|--------------------------------|------------------------------|---------------------|
| A/3/ENV FA | 33 | 49 | 4.89 | 0.9821 | 0.9694 | 8.222 |
| A/3/ENV FB | 28 | 53 | 4.175 | 0.9267 | 0.8686 | 6.801 |
| A/09/ENV FA | 25 | 34 | 4.43 | 0.9715 | 0.9539 | 6.806 |
| A/09/ENV FB | 29 | 56 | 4.543 | 0.9604 | 0.9352 | 6.956 |
| A/09/ENV FC | 12 | 18 | 3.303 | 0.9216 | 0.9213 | 3.806 |
| A/10/ENV FA | 17 | 25 | 3.894 | 0.96 | 0.9526 | 4.971 |
| A/10/ENV FB | 21 | 40 | 4.015 | 0.941 | 0.9142 | 5.422 |
| A/10/ENV FC | 21 | 30 | 4.215 | 0.9701 | 0.9596 | 5.88 |
| A/12/ENV FA | 17 | 17 | 4.087 | 1 | 1 | 5.647 |
| A/12/ENV FC | 8 | 8 | 3 | 1 | 1 | 3.366 |
| A/14/ENV FA | 22 | 51 | 3.694 | 0.8894 | 0.8283 | 5.341 |
| A/14/ENV FB | 36 | 72 | 4.679 | 0.9476 | 0.905 | 8.184 |
| A/18/ENV FA | 20 | 30 | 4.006 | 0.9494 | 0.927 | 5.586 |
| A/18/ENV FB | 28 | 35 | 4.672 | 0.9832 | 0.9719 | 7.594 |
| A/20/ENV FA | 6 | 6 | 2.585 | 1 | 1 | 2.791 |
| A/20/ENV FB | 8 | 23 | 2.084 | 0.668 | 0.6947 | 2.233 |
| A/20/ENV FC | 1 | 2 | 0 | 0 | **** | 0 |
| A/21/ENV FA | 29 | 64 | 4.142 | 0.9092 | 0.8526 | 6.733 |
| A/21/ENV FB | 17 | 32 | 3.929 | 0.9577 | 0.9613 | 4.617 |
| A/22/ENV FA | 14 | 22 | 3.629 | 0.9524 | 0.9532 | 4.206 |
| A/22/ENV FB | 16 | 25 | 3.764 | 0.95 | 0.941 | 4.66 |
| A/26/ENV FA | 32 | 48 | 4.756 | 0.9743 | 0.9513 | 8.008 |
| A/26/ENV FB | 21 | 25 | 4.294 | 0.9833 | 0.9775 | 6.213 |
| A/26/ENV FC | 8 | 10 | 2.922 | 0.9556 | 0.974 | 3.04 |
| A/27/ENV FA | 27 | 41 | 4.439 | 0.961 | 0.9336 | 7.001 |
| A/27/ENV FB | 28 | 41 | 4.399 | 0.9488 | 0.9151 | 7.271 |
| A/27/ENV FC | 17 | 26 | 3.714 | 0.9262 | 0.9086 | 4.911 |
| A/202/ENV FA | 28 | 61 | 4.261 | 0.9355 | 0.8863 | 6.568 |
| A/202/ENV FB | 34 | 56 | 4.807 | 0.974 | 0.9449 | 8.198 |
| A/204/ENV FA | 43 | 94 | 4.827 | 0.9508 | 0.8896 | 9.244 |
| A/204/ENV FB | 18 | 37 | 3.424 | 0.8529 | 0.8211 | 4.708 |
| A/301/ENV FA | 14 | 18 | 3.684 | 0.9673 | 0.9675 | 4.498 |
| A/301/ENV FC | 4 | 9 | 1.447 | 0.5833 | 0.7233 | 1.365 |
| A/302/ENV FA | 16 | 38 | 3.43 | 0.8876 | 0.8576 | 4.124 |
| A/302/ENV FB | 20 | 34 | 4.146 | 0.9643 | 0.9594 | 5.388 |
| A/302/ENV FC | 18 | 52 | 2.639 | 0.6659 | 0.6328 | 4.302 |
| A/303/ENV FA | 5 | 6 | 2.252 | 0.9333 | 0.9697 | 2.232 |
| A/303/ENV FB | 26 | 37 | 4.574 | 0.9805 | 0.9731 | 6.923 |
| A/303/ENV FC | 6 | 7 | 2.522 | 0.9524 | 0.9755 | 2.569 |
| A/304/ENV FA | 35 | 62 | 4.865 | 0.972 | 0.9485 | 8.238 |
| A/304/ENV FC | 24 | 36 | 4.44 | 0.9762 | 0.9685 | 6.418 |
| A/305/ENV FA | 24 | 46 | 4.229 | 0.9478 | 0.9224 | 6.007 |
| A/305/ENV FB | 23 | 38 | 4.353 | 0.9701 | 0.9623 | 6.048 |
| A/306/ENV FA | 24 | 61 | 3.816 | 0.8923 | 0.8324 | 5.595 |
| A/306/ENV FB | 20 | 40 | 4.015 | 0.9436 | 0.9291 | 5.151 |
| A/307/ENV FA | 10 | 11 | 3.278 | 0.9818 | 0.9867 | 3.753 |
| A/307/ENV FB | 25 | 44 | 4.37 | 0.9609 | 0.9411 | 6.342 |
| A/309/ENV FA | 14 | 20 | 3.622 | 0.9526 | 0.9513 | 4.34 |
| A/309/ENV FB | 6 | 13 | 2.035 | 0.7179 | 0.7872 | 1.949 |
| A/1008/ENV FA | 16 | 36 | 3.224 | 0.8381 | 0.806 | 4.186 |
| A/1008/ENV FB | 35 | 53 | 4.963 | 0.9826 | 0.9676 | 8.564 |
| A/1011/ENV FA | 24 | 45 | 4.356 | 0.9646 | 0.95 | 6.042 |
| A/1011/ENV FB | 15 | 22 | 3.698 | 0.9524 | 0.9465 | 4.529 |
| A/1013/ENV FA | 17 | 24 | 3.887 | 0.9601 | 0.9509 | 5.035 |
| A/1013/ENV FB | 10 | 17 | 3.052 | 0.9044 | 0.9186 | 3.177 |
| A/1015/ENV FA | 29 | 42 | 4.576 | 0.9686 | 0.9421 | 7.491 |

| | | | | | | |
|--------------------|-------|-------|-------|--------|--------|-------|
| A/1015/ENV FB | 30 | 52 | 4.695 | 0.9744 | 0.9569 | 7.339 |
| A/3010/ENV FA | 25 | 40 | 4.463 | 0.9718 | 0.961 | 6.506 |
| A/3010/ENV FB | 23 | 38 | 4.217 | 0.9545 | 0.9323 | 6.048 |
| Mean | 20.4 | 35.1 | 3.804 | 0.916 | 0.920 | 5.409 |
| St Dev | 9.2 | 18.7 | 0.941 | 0.146 | 0.074 | 1.959 |
| % Variation | 45.3% | 53.1% | 24.7% | 16.0% | 8.1% | 36.2% |
| Nimrod Mean | 19.8 | 30.5 | 3.95 | 0.942 | 0.924 | 5.5 |
| Nimrod SD | 4.8 | 8.5 | 0.46 | 0.04 | 0.044 | 1.1 |
| Toroa Mean | 20.7 | 40.7 | 3.96 | 0.946 | 0.923 | 5.34 |
| Toroa SD | 6.6 | 16.4 | 0.41 | 0.024 | 0.037 | 1.27 |

Historical comparisons are in blue. Note: No historical data available for Vinson West

Table 2.11 - Univariate Faunal Parameters (for 0.2m² station)

| Station | Number of Species per 0.2m ² (S) | Number of Individuals per 0.2m ² (N) | Shannon-Wiener Diversity | Simpsons Diversity (1-Lambda') | Evenness (Pielou's Evenness) | Richness (Margalef) |
|--------------------------|---|---|--------------------------|--------------------------------|------------------------------|---------------------|
| A/3/ENV | 51 | 102 | 5.224 | 0.9703 | 0.9209 | 10.81 |
| A/09/ENV* | 50 | 108 | 5.081 | 0.9619 | 0.9002 | 10.47 |
| A/10/ENV* | 44 | 95 | 4.851 | 0.953 | 0.8885 | 9.443 |
| A/12/ENV | 22 | 25 | 4.404 | 0.99 | 0.9875 | 6.524 |
| A/14/ENV | 47 | 123 | 4.744 | 0.9351 | 0.854 | 9.559 |
| A/18/ENV | 42 | 65 | 5.04 | 0.9731 | 0.9346 | 9.822 |
| A/20/ENV* | 11 | 31 | 2.451 | 0.7183 | 0.7084 | 2.912 |
| A/21/ENV | 39 | 96 | 4.594 | 0.9366 | 0.8692 | 8.325 |
| A/22/ENV | 24 | 47 | 4.187 | 0.9482 | 0.9132 | 5.974 |
| A/26/ENV* | 50 | 83 | 5.359 | 0.9797 | 0.9496 | 11.09 |
| A/27/ENV* | 56 | 108 | 5.152 | 0.9486 | 0.8871 | 11.75 |
| A/202/ENV | 50 | 117 | 5.052 | 0.963 | 0.8951 | 10.29 |
| A/204/ENV | 52 | 131 | 4.868 | 0.9372 | 0.854 | 10.46 |
| A/301/ENV | 18 | 27 | 3.856 | 0.943 | 0.9248 | 5.158 |
| A/302/ENV* | 39 | 124 | 4.092 | 0.8752 | 0.7743 | 7.883 |
| A/303/ENV* | 32 | 50 | 4.823 | 0.9796 | 0.9647 | 7.924 |
| A/304/ENV | 44 | 98 | 5.119 | 0.9726 | 0.9377 | 9.378 |
| A/305/ENV | 41 | 84 | 4.922 | 0.9621 | 0.9186 | 9.028 |
| A/306/ENV | 36 | 101 | 4.471 | 0.936 | 0.8648 | 7.584 |
| A/307/ENV | 34 | 55 | 4.801 | 0.9717 | 0.9437 | 8.235 |
| A/309/ENV | 16 | 33 | 3.467 | 0.8958 | 0.8669 | 4.29 |
| A/1008/ENV | 44 | 89 | 4.907 | 0.9533 | 0.8988 | 9.58 |
| A/1011/ENV | 34 | 67 | 4.708 | 0.9643 | 0.9254 | 7.848 |
| A/1013/ENV | 24 | 41 | 4.211 | 0.9512 | 0.9184 | 6.193 |
| A/1015/ENV | 51 | 94 | 5.343 | 0.979 | 0.9418 | 11.01 |
| A/3010/ENV | 39 | 78 | 4.932 | 0.9657 | 0.9332 | 8.722 |
| Mean | 38.1 | 79.7 | 4.641 | 0.945 | 0.899 | 8.472 |
| St Dev | 12.4 | 32.5 | 0.643 | 0.053 | 0.058 | 2.224 |
| % Variation | 32.5% | 40.7% | 13.9% | 5.6% | 6.5% | 26.3% |
| Vinson West ENV 5 | 45 | 83 | 5.17 | 0.96 | 0.94 | - |
| Vinson West Mean | 53 | 39 | 5.46 | 0.97 | 0.95 | - |
| Vinson West SD | 8 | 12 | 0.25 | 0.01 | 0.01 | - |
| Nimrod Mean | 32.4 | 61.7 | 4.49 | 0.942 | 0.897 | 7.62 |
| Nimrod SD | 5.7 | 12.3 | 0.41 | 0.031 | 0.042 | 1.12 |
| Toroa Mean | 35.3 | 81.3 | 4.56 | 0.949 | 0.892 | 7.84 |
| Toroa SD | 7.9 | 23.6 | 0.41 | 0.023 | 0.046 | 1.48 |

* 3 replicates present (0.3m²)

Historical comparisons are in blue

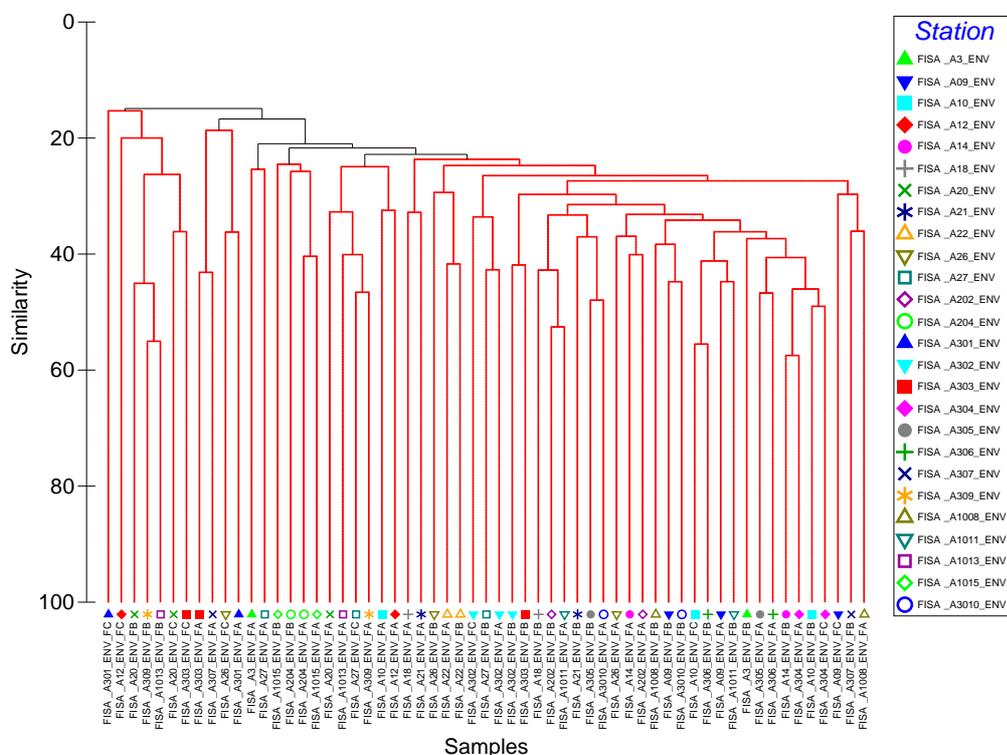
2.9.3 Multivariate Analyses

To provide a more thorough examination of the macrofaunal community, multivariate analyses was performed upon the data for both the replicate and aggregated stations using Plymouth Routines in Multivariate Ecological Research software (PRIMER; Clarke and Warwick 1994) to illustrate data trends. Unlike univariate parameters, multivariate analyses preserve the identity of the different species by assigning a similarity or dissimilarity between the samples. The analyses were undertaken on appropriately transformed data, relating to square-root for both replicates and stations.

2.9.3.1 Dendrogram – Group Average Method

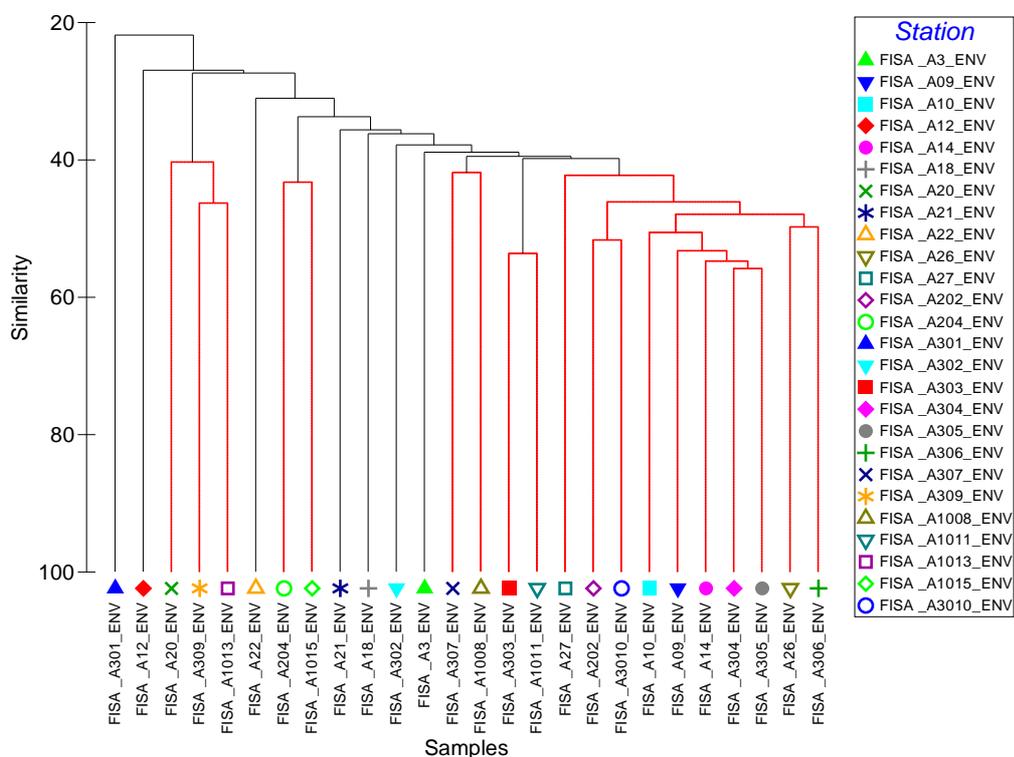
The similarity dendrogram is given for all replicates in Figure 2.45. This diagram shows that both inter-station and intra-station relationships are relatively weak and largely variable, despite this several sites showed stronger intra-station relationships by replicates of the same station clustering together. The SIMPROF test showed six significant structural groupings in the data (black branches) at between 15% to 23% similarity. Within these SIMPROF groupings all replicates had branched out at the 60% similarity mark. The lack of clustering within and between stations indicates a reasonably large degree of overlapping of species assemblages between replicates and stations, highlighting a broad homogeneous macro-invertebrate community. Given the large scale of this regional survey it is likely to encompass numerous variations of the same extensive habitat.

Figure 2.45 Dendrogram of Macrofaunal Replicates



This is further supported in Figure 2.46, which shows a similarity dendrogram by station made up of combined sites based on the two to three replicates (total surface area of 0.2m² - 0.3m²). Results indicate a slightly higher and narrower similarity range, with most sites branching off within the similarity range of 35%-55%. The SIMPROF routine concluded that there were 12 statistically significant differences between the 26 stations, with seven of these groups consisting just one station. Consequently, this plot shows that when lists are combined at a station level there appears to be more significant differences within the macro-invertebrate community.

Figure 2.46 Dendrogram of Macrofaunal Stations



2.9.3.2 MDS Ordination Plot

The replicate similarities were presented in a 2-dimensional representation with two axes of similarity. This multi-dimensional scaling (MDS) ordination is presented in Figure 2.47 for all 59 replicates and shows a general “cloud” of sample replicates, with the sites/groups that showed significant SIMPROF separation on the outskirts of the main cloud, particularly replicates of station A/20/ENV show the largest separation A/20/ENV/FC, which had only one infaunal individual (copepod).

However, the MDS plot is only of limited use due to its high stress level of 0.277, which provides an ordination that should be treated with scepticism, with points close to being

arbitrarily placed in the 2-D plot (see Appendix II, Table G). In this instance the 3-D plot was examined for clarifications on ordinations, which indicated a lower stress level of 0.211, yet showed a similar distribution.

Figure 2.47 MDS Ordination Plot by Replicate

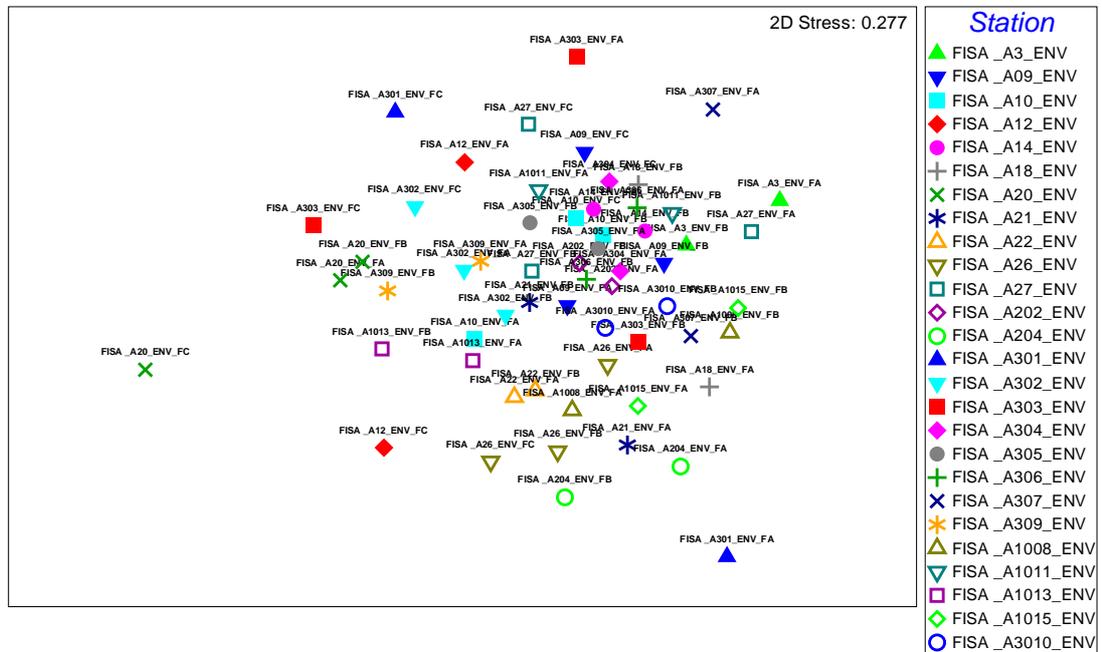
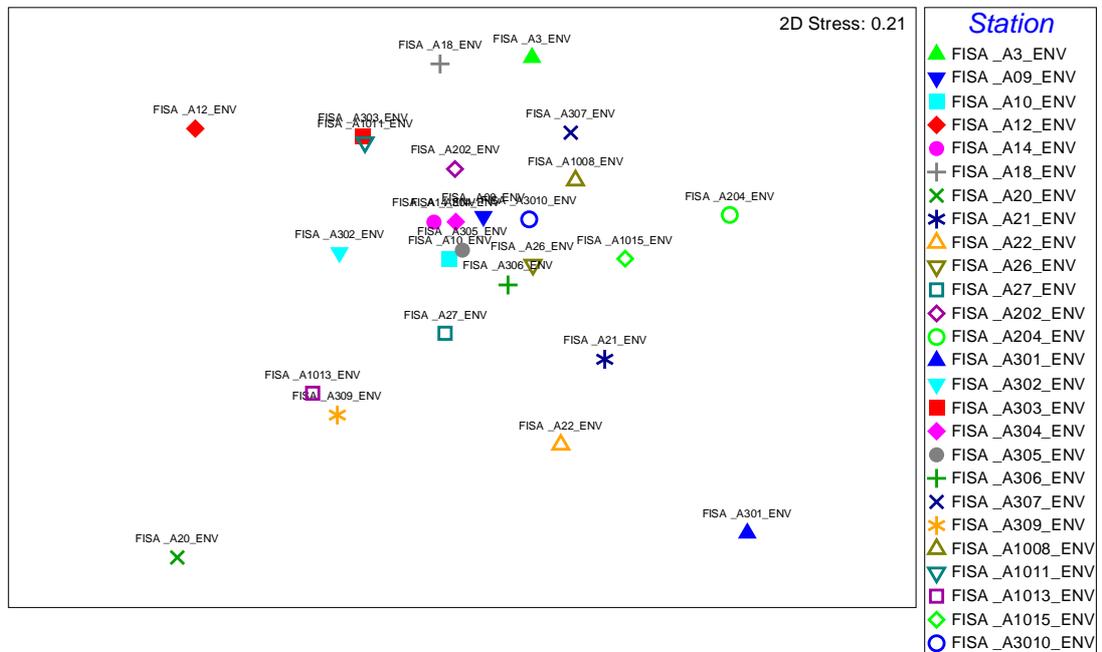


Figure 2.48 MDS Ordination Plot by Station



The MDS 2D ordination of samples by stations (Figure 2.48) indicated a similarly low stress score of 0.21, which means that the plot should be treated with some scepticism. Consequently, this plot has been interpreted with the use of the 3D plot which has a reduced stress level of 0.156 providing a more useful demonstration of the separation between station similarities. There is no particular trend to the data observed here however the sites that show significant SIMPROF structure show the greatest distance from the main cluster. In order to see the magnitude of the differences between the different SIMPROF groups an ANOSIM (analysis of similarity) test was performed, which showed a similarity R Value of 0.928 ($p < 0.01$), indicating a high degree of difference between the SIMPROF groups. From the data there appears to be no trend based on geographical location, with geographically close stations remaining biologically separated.

The most likely explanation for these differences is sediment type, however subsequent principle components analysis (PCA) and RELATE tests have failed to yield any significant correlation between particle size and biological community, with the exception of sites A/12/ENV, A/22/ENV and A/1015/ENV, which seemed to have the most Euclidean distance from the other sites using mean Phi particle size. The general lack of correlation here is likely due to the particle size analysis (PSA) itself. Although an industry standard, only a small proportion of the grab is sub-sampled for analysis, therefore often larger sized pebbles and gravels can be misrepresented. An unrepresentative proportion of larger sized sediments appears to be the case for some of the stations based on observation from the seabed photography. For instance the grab data for station A/20/ENV, clearly shows the presence of larger pebbles within the samples, that will not only reduce the grab volume but also a proportion of sediment suitable for infaunal inhabitation.

Due to the large amount of data and broad transition of sediment types identified, varying from sandy muds, through to gravelly sands, it appears that this may have created an underlying gradient of change in order to assign discrete biological communities. This results in a large amorphous grouping of station similarities, indicative of a single but broad macrofaunal assemblage.

In order to place this biological information into a regional context, the results have been included into a regional comparison: where samples from a BSL survey at Burdwood Bank, performed in 2008, have been ordinated in a MDS plot to assess any similarities/differences between the stations (Figure 2.49. BSL, 2009). The 2008 Burdwood Bank survey area is located approximately 250km southwest of the centre of the FISA survey area and exhibiting similar sediment properties to that of the FISA location. Multivariate analysis of the biology from Vinson West, Tora and Nimrod have not been used as these were analysed at different laboratories (i.e. Fugro Survey and the Natural History Museum) without any consistency being applied to putative species nomenclature.

The SIMPER calculations for the different clusters identifying the top five contributory species are presented in Table 4.11 along with their contribution percentages.

Table 2.12 - SIMPER analysis of the four different cluster formations and top Five contributing species and percentages

| | FISA 2014 | | FISA A/301 | | BB/s10 | |
|---------------|---------------------------------|-------|---------------------------------|-------|---------------------------------|-------|
| Burdwood Bank | Dissimilarity Percentage 75.21% | | Dissimilarity Percentage 85.19% | | Dissimilarity Percentage 85.47% | |
| | Nematoda | 2.11% | Nematoda | 3.29% | Nematoda | 3.16% |
| | Ostracoda | 2.09% | <i>Urothoe</i> | 2.26% | Ostracoda | 2.89% |
| | <i>Rhampobranchium</i> | 1.88% | Phoxocephalidae | 2.18% | <i>Rhampobranchium</i> | 2.8% |
| | <i>Aricidea oculata</i> | 1.52% | <i>Rhampobranchium ehlersi</i> | 2% | Pardaliscidae | 2.64% |
| | Copepoda | 1.5% | Ostracoda | 1.85% | <i>Urothoe</i> | 2.16% |
| FISA 2014 | | | Dissimilarity Percentage 78.18 | | Dissimilarity Percentage 88.51% | |
| | | | Copepoda | 2.84% | Pardaliscidae | 3.53% |
| | | | <i>Dacrydium</i> | 2.73% | <i>Rhampobranchium</i> | 2.78% |
| | | | Brachiopoda | 2.57% | Copepoda | 2.36% |
| | | | <i>Nephasoma diaphanes</i> | 2.35% | <i>Similipecten</i> | 2.06% |
| | | | <i>Ampelisca</i> | 2.13% | <i>Psolus</i> | 2.02% |
| FISA A/301 | | | | | Dissimilarity Percentage 82.41% | |
| | | | | | Pardaliscidae | 5.96% |
| | | | | | <i>Rhampobranchium</i> | 3.87% |
| | | | | | Nemertea | 3.77% |
| | | | | | <i>Eunoe</i> | 3.77% |
| | | | | | <i>Similipecten</i> | 3.77% |

The differences observed at the Burdwood Bank and FISA cluster are due to the higher abundance of nematodes, ostracods, *Rhampobranchium ehlersi* and *Aricidea oculata* at Burdwood Bank, however there were slightly higher counts of copepods at FISA. All these species were present in both surveys to some degree. The separation of A/301/ENV from the other clusters are generally attributed to the absence of copepods, nematodes, *Urothoe*, Phoxocephalidae and Pardaliscidae and generally low counts of Brachiopoda and *Dacrydium* with the prevalence of Copepoda, *Nephasoma diaphanes* and *Ampelisca*, amplified by the overall low species richness and abundance here. The differences observed at Burdwood Bank s10 were attributed to the relative high abundance of the crustacean Pardaliscidae which was either absent or had low counts in the other clusters, and the absence of *Eunoe*, Nematoda, *Urothoe* and Ostracoda combined with the relatively low counts of *Rhampobranchium ehlersi* yet prevalence of *Similipecten* all contributed to this locations biological separation. As this station only had one fauna sample this is sure to have exaggerated these differences in species counts.

The differences observed between the two main clusters appear to be an accumulation of subtle differences in species counts, mainly of the dominant species identified, which the RELATE test showed could partially be explained by the PSA data. The two outlying sites had low species richness's either attributed to the coarse substrate (A/301/ENV) or only having one faunal replicate (BB/s10), which seems to have amplified any differences observed.

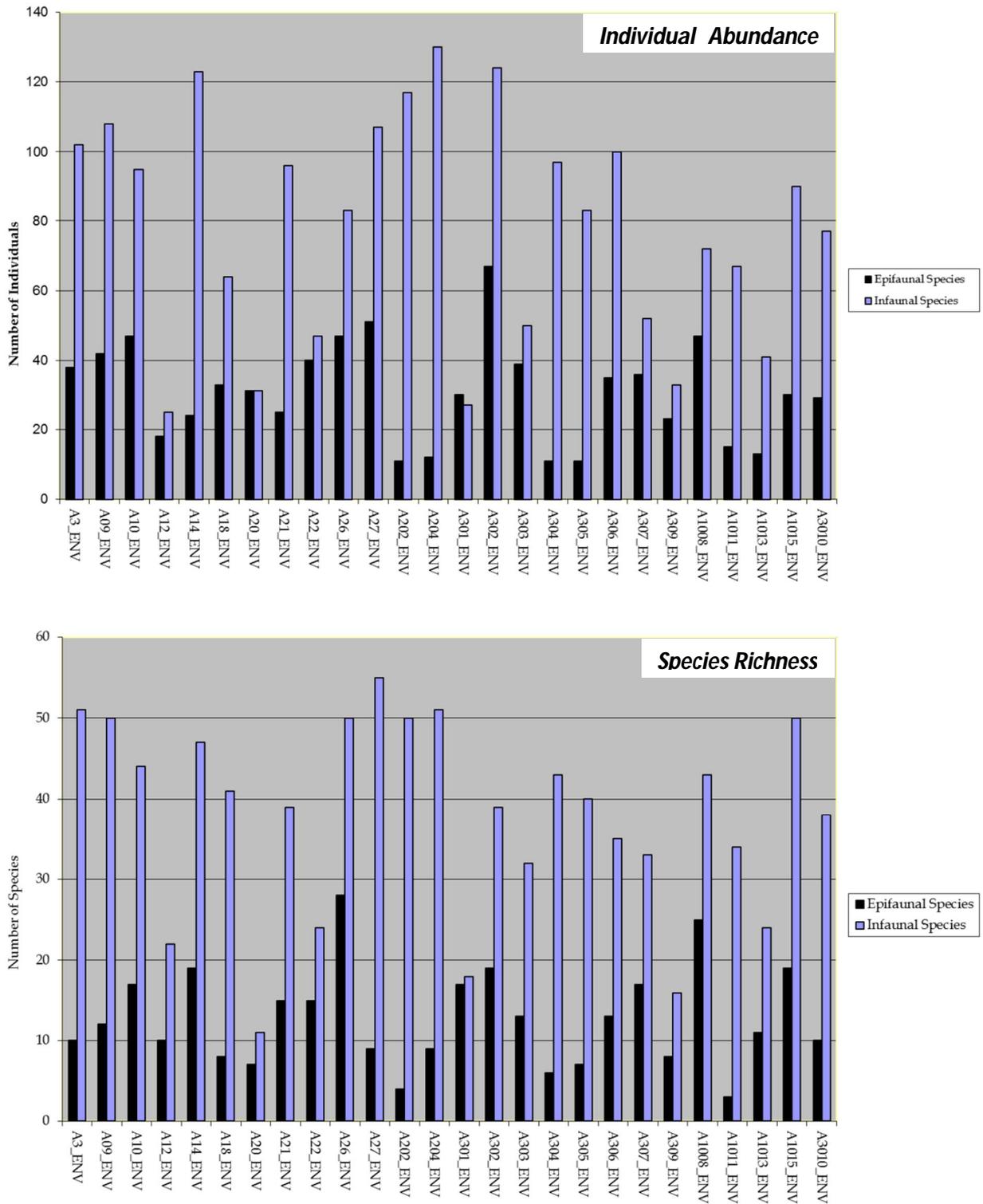
2.9.4 Environmental Variables

In order to assess whether any relationships between the biological community and chemical variables were present and whether these differences were significant, numerous principle components analyses and RELATE tests were performed. However no meaningful trends or conclusions were reached. Depending on the test, different stations showed differing separations. For instance, station A/1015/ENV, A/306/ENV, A/301/ENV and A/26/ENV, all showed separation from the other sites according to Ni, Zn and V, whereas station A/12/ENV and again A/26/ENV, separated from the other sites by % NPD. These individual variances are often not present when viewed from a different principle component dimension, and are therefore only weak relationships that lay in a particular category when forced into a PCA ordination. Subsequent RELATE tests have not shown any significant relationships of these environmental variables with biological separations. In addition to this a Pearson's correlation has been permuted of the macrofauna diversity indices with physico-chemical parameters. There are few significant weak correlations relating % fines and several of the metals with species richness, Shannon-Wiener diversity and Simpsons diversity (all $P < 0.05$), but no strong correlations were recorded (Appendix XI). Although such correlations can be regarded as negligible due to their co-incidence nature, % fines and AI both relate to the sediment parameters of particle size and may bare some relevance to the faunal distribution.

2.9.5 Epifaunal and other Biological Groups

All of the macrofaunal samples recorded the presence of invertebrate species that are generally considered to be epifaunal and are not statistically assessed within the infauna. Whilst the underwater photography showed that some habitats within the survey area are heterogeneous due to the intermittent exposures of underlying bedrock and sporadic drop-stones caused by ice modification (see Sections 2.3); the benthic sampling indicated varying quantities of these coarser admixtures. The consistency and importance of the epifaunal assemblages is demonstrated in Figure 2.50, highlighting the general proportion of infaunal and epifaunal species. Due to the presence/absence scale to which a lot of epifaunal species are identified, for the purpose of this chart and to highlight both the epifaunal abundance and richness, where epifaunal species have been recorded as present this has been given the numerical value of "1" only to represent the colony. Infauna and epifauna species are listed separately in Appendix VII.

Figure 2.50 Epifaunal Abundance and Richness versus Infauna (by Station)



Sessile epifauna were prevalent throughout the FISA macro-invertebrate samples. Some of the key groups are discussed below.

Porifera: The Class Hexactinellida was relatively well represented by many fragments, not sufficient for determination. In addition embryos in the form of little spheres were common in every sample. This is a feature of most deep water and deep sea sediments.

A single specimen of a *Sycon* species represented the Class Calcarea, with the rest made up of Demospongia, with siliceous spicules and varying amounts of spongin.

Most common were *Eurypon* spp., encrusting species with the spicules standing on the substrate giving a hairy appearance. These sponges are belonging to the order Hadromerida which have an extended planktonic larval life which allows for wide geographical dispersion. Most of the genera are typical for deeper water (e.g. *Hymedesmia* spp., *Crella* and *Lissodendoryx*). Many of the species inhabit a wide distribution in the Atlantic or are often cosmopolitan; in addition many species are only represented by a couple of specimens or small colonies.

Cnidaria: This group was regularly represented. Nine genera of thecate and athecate Hydrozoa and two genera of Stylasteridae were recorded. Live Stylasteridae were not infrequent, and the species *Stylaster densicaulis* (Figure 2.35f) was the most common, with the second species *Lepidopora* sp. only of sporadic occurrence. Stylasteridae are a group of Hydrozoa with a calcified skeleton resembling corals. The other Hydrozoa are common genera in the Atlantic but their species status could not be determined.

Scyphozoa were represented by their sessile polyp stage, likely of the order Coronatae (Deep Sea Medusae). These are trumpet shaped with very distinct annulation, and are known as voracious feeders and predators. They are very common in the deep water samples.

While some of the samples contained dead *Lophelia* and *Lophelia* debris, no live material was found. The material looked very weathered. The live solitary corals belong to the species *Flabellum curvatum*.

Several species of Octocorallia were found. Several specimens of Pennatulidae were too small to identify. Two species of Gorgonacea were not infrequent, *Callozostron carlotta* (Figure 2.35d) and the more common *Fannyella rossi*. The latter is a common and widespread species.

Bryozoa: This group was a common constituent on pebbles and stones, with many species endemic. While most specimens were encrusting, there are a number of upright forms, e.g. *Cornucopina*, *Isosecuriflustra* and *Notoplites*.

Other epibenthic taxa: *Rhabdopleura* is commonly found in polar waters both in the North and South Atlantic.

Further photographic examples of epifaunal species are given in Figure 4.49.

2.10 VIDEO/PHOTOGRAPHIC SURVEY OF SEDIMENT HABITATS

2.10.1 Video/Photographic survey

In addition to the analysis and interpretation of the benthic samples, additional photographic ground truthing data was also obtained at eight additional locations (total 34 stations) within the FISA survey area. These were selected based upon areas of bathymetric interest or on sediments representative of typical habitats encountered for the area to provide a more regional context. The bathymetric dataset indicated a generally habitat of silty sand, some areas targeted relate to features created by historical ice modifications. Occasional examples of larger clasts, such as boulders, were also frequently recorded. A list of the ground truthed sites and their location is given in Table 2.2 and summary photo pages in (Appendix VIII).

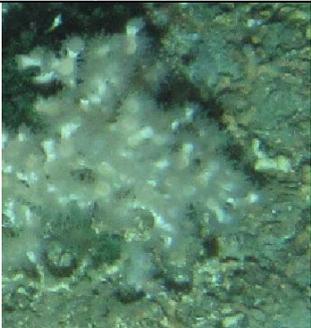
No environmentally sensitive habitats or benthic communities were recorded at or surrounding the proposed Finback-1 and Humpback well locations or within the FISA Block. This includes the absence of potential Annex I habitats such as gas escape feature, biogenic reefs or geological reefs, currently protected under the European Habitats Directive. Dead *Lophelia* fragments were identified within the fauna samples, and live *Lophelia* specimens were located in the photographs at several stations A/1008/ENV, A/205/ENV, A/206/ENV, A/301/ENV, A/303/ENV (Humpback -1, well location), A/304/ENV, and A/308/ENV. However where present, these were only as small sporadic colonies and do not represent the complex well developed structures that constitute reef status under the Annex I of the European Habitats Directive. Despite this lack of size or proliferations across the area, coral reefs, where found are classified by OSPAR as under threat and there may be instances along the edge of escarpment where larger aggregations may exist and should be protected from physical damage. The majority of evidence for the presence of this species relates to extensive areas of debris relating to relic residue populations located on the edge of the escarpment drop-off approximately 4.7km southeast of the proposed Finback-1, 2km southeast of Humpback-1 and approximately 5.7km northwest of the proposed Humpback-2 well locations.

Survey operations were carried out using a combined digital video and still camera system deployed in a drop-down frame (see Section 2). Results for each of the 34 ground truthing locations are summarised in Appendix VIII for the dominant habitats encountered, or Figure 4.49 for the common and/or conspicuous faunal groups encountered.

These photographs confirmed expectations acquired during the regional habitat assessment and observed on the bathymetric dataset (see Section 4.1). The seabed photography revealed evidence of ice modification mainly relating to cobbles and drop-stones sporadically placed throughout the survey area, as well as bedrock exposure also observed on escarpment outcrops. An assessment of the epifaunal assemblages shown within the seabed photographs, confirmed a relatively well populated community of species overall, with healthy aggregations colonising occasional drop-stones as well as

Lophelia debris, where present and some rooted into the sediment. Many of the species recorded are associated with deep water and/or cold water environments. A summary of the main taxa recorded, along with some example photographs is given in Figure 2.51.

Figure 2.51 Examples of Epifaunal and Megafauna Species Recorded at the FISA Site

| Species Examples from Seabed Photography | | |
|---|---|---|
|  |  |  |
| Porifera: Possible Demospongiae sp. | Porifera: Possible <i>Myxilla</i> sp. | Porifera: Possible <i>Mycale</i> sp. |
|  |  |  |
| Cnidaria: <i>Anthoptilum grandiflorum</i> | Cnidaria: Pennatulidae sp. | Cnidaria: Sea whip sp. |
|  |  |  |
| Cnidaria: <i>Lophelia pertusa</i> | Cnidaria: <i>Lophelia</i> debris | Cnidaria: Possible Stylasteridae sp. |
|  |  |  |
| Cnidaria: Gorgonian sp. | Cnidaria: Pennatulidae sp. | Cnidaria: Gorgonian sp. (Octocorallia) |

| | | |
|--|---|--|
|  <p>Cnidaria: Gorgonian sp. (Octocorallia)</p> |  <p>Cnidaria: Hydroid sp. (Surrounded by Porifera sp. and possible Bryozoa)</p> |  <p>Cnidarian sp.</p> |
|  <p>Cnidaria sp.</p> |  <p>Actiniaria sp.</p> |  <p>Actiniaria: Possible <i>Bolocera</i> sp.</p> |
|  <p>Actiniaria: Possible <i>Actinoscyphia aurelia</i></p> |  <p>Cnidaria: Alcyonium sp. With possible <i>Fannyella rossi</i></p> |  <p>Poss Cnidarian sp.</p> |
|  <p>Cnidaria: Possible <i>Flabellum</i> sp.</p> |  <p>Echinoderm: Crinoidea.</p> |  <p>Echinoderm: Crinoidea.</p> |

| | | |
|--|---|---|
|  <p>Bryozoan sp.</p> |  <p>Echinoid: Possible <i>Eucidaris</i> sp.</p> |  <p>Ophiuroidea: probably <i>Astrotoma agassizii</i>.</p> |
|  <p>Ophiuroidea sp.</p> |  <p>Asteroidea sp.</p> |  <p>Asteroidea: Possible <i>Culcita</i> sp.</p> |
|  <p>Holothuroidea sp.</p> |  <p>Holothuroidea sp.</p> |  <p>Isopoda: Possible <i>Acutiserolis neaera</i></p> |
|  <p>Decapoda: Caridea sp.</p> |  <p>Decapoda: Possible Galatheidae sp</p> |  <p>Decapoda: Nephropidae sp. probably <i>Thymops birsteini</i>.</p> |
|  <p>Decapoda: Lithodoidea sp probably <i>Paralomis</i> sp.</p> |  <p>Moridae sp.</p> | <p>No used</p> |

Overall, the phylogenetic make-up of the conspicuous megafauna observed was dominated by the cnidarians, with octocorals and Pennatulidae remaining prevalent throughout. Echinoderms were better represented in the seabed photography than the grab samples with ophiuroids, crinoids, asteroids and holothurians present. Where drop-stones were present, encrusting sponges were common along with anthozoans. Often rooted into soft sediments bryozoans and hydroids were observed as sparse tufts. There were also numerous burrows likely to be associated with crustacean and holothurian activity. Free-swimming megafauna included the demersal teleosts: Macrouridae, Grenadier, hake and batoids.

Previous assessments of the epifaunal component from seabed photographic records taken at the Vinson West, Nimrod and Toroa similar epifaunal communities, where recorded. One main difference recorded would be that of the tube-dwelling onuphid worms, *Onuphis pseudoirrescens*, and *Kinbergonuphis oligobranchiata* which were dominant within the Toroa and Nimrod surveys but absent from this survey, with occasional *K. oligobranchiata* specimens observed at Vinson West. Due to the large area covered by this FISA survey, with seabed photography undertaken at 34 stations, the epifaunal species are more extensive than previous surveys undertaken in the region. This is due the numerous colonised hard substrates identified, compared with that of Toroa and Nimrod which didn't undertake seabed photography. Toroa especially had poor representations of epifauna due to the survey area being comprised of softer sandy silt. The stations surveyed at Vinson West also observed only limited footage of boulders.

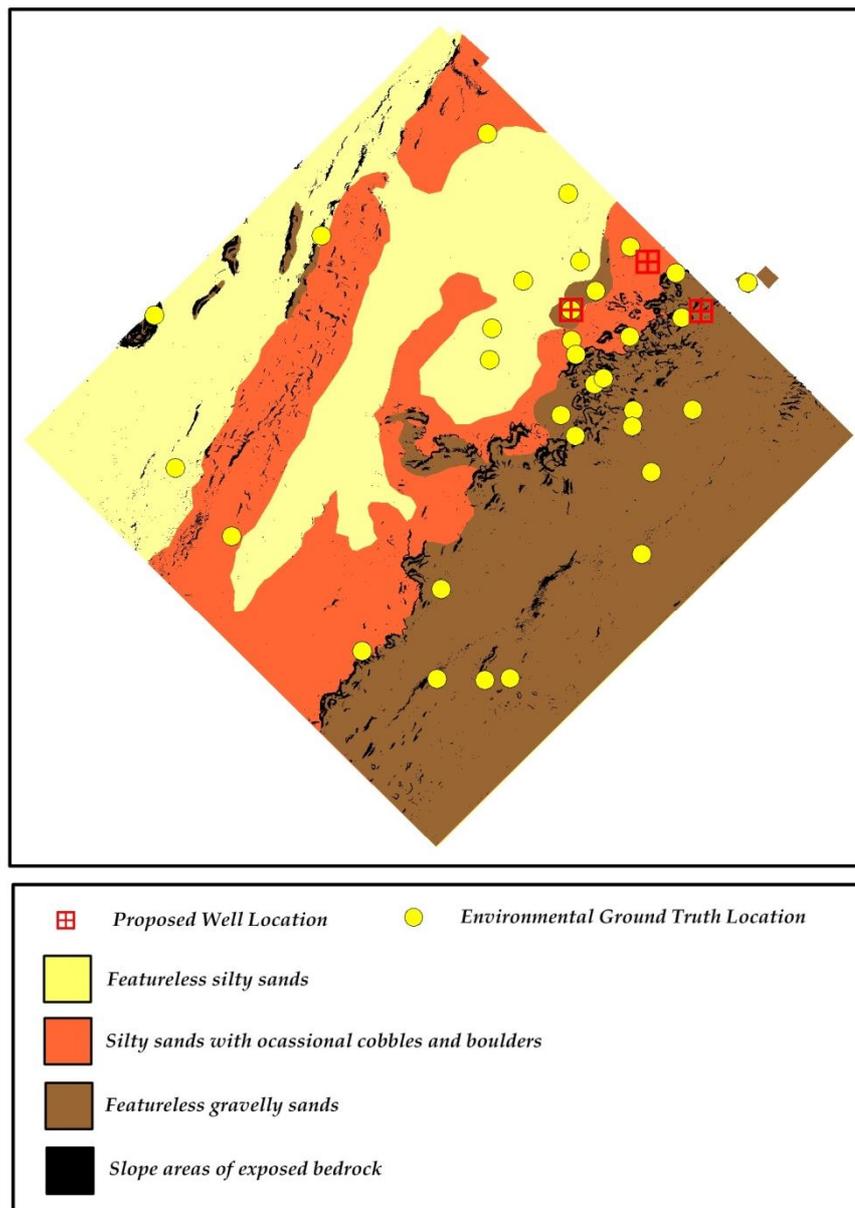
The remainder sites of the current survey area did not yield any evidence of particularly sensitive habitats, especially within the immediate vicinity of the proposed exploration wells. The habitats in these areas were considered to be homogenous, gravelly or slightly gravelly silty sands with limited sensitivity to the proposed operations. A review of both stills and video footage indicate that some potential CITES Appendix II coral species (such as *Flabellum* sp.) were observed.

2.10.2 Environmental Habitats

Within the survey area, the majority of the shallower sediments were dominated by a relatively homogeneous and featureless slightly gravelly silty sand, associated with a weak sedimentary environment throughout. The deeper sediments to the south and east were associated with a similar sediment type, but with greater erosion and significant gravel component, indicative of ice modification from drop stones throughout. The central part of the survey area is represented by a complex bathymetry indicative of a number of underlying geological structures forming an irregular escarpment running from the southwest to the northeast. These have been interpreted as steep sloped exposures of bedrock or a hard underlying formation (such as a concretion) revealing a low energy rock exposure. In the shallower, northwestern part of the survey area, this underlying formation appears to have either collapsed or has been eroded leaving steep sided depressions at a number of locations leaving slump material (boulder field) at its base.

Separating the shallower slightly gravelly silty sands from the deeper gravelly sands is the rocky escarpment along with an intermediate habitat type which marks the transition from homogeneous sands to the rock outcrops. This is interpreted as generally homogeneous sand punctuated by occasional to numerous isolated rock outcrops and boulders. This area is the dominant sediment type along the top of the escarpment and over a sub-cropping geological feature, observed in the bathymetry, running at a depth of 1150m along the length of the survey area in a northeast-southwest orientation. Here the sands thin to a veneer.

Figure 2.52 - Summary Habitat Classifications



Whilst the homogeneous sands showed relatively few conspicuous species, the diversity of fauna increased slightly in the deeper gravelly sediments to the southeast and significantly on the intermittent or escarpment bedrock exposures. On this hard substrate, the dominant fauna consists mostly of suspension feeding anthozoans, in particular both soft

and hard corals, although communities were generally small in size. Present within these areas was the reef-building coral *Lophelia pertusa*, although live examples were only found in small colonies, not sufficient to constitute an Annex I reef, as designated under the European Habitats Directive. However, the skeletal debris from this species was also found in relatively high concentrations in small isolated patches on flat rocky areas and along the top and on the escarpment slope, presumed to be of historical residues and the aggregation of eroded material produced over a significant period of time. The base of the escarpment is marked by slump deposits from the escarpment erosion in the form of a boulder field. The surrounded granular sediments with this area indicate the influence from currents in the form of localised scour and some sand rippling.

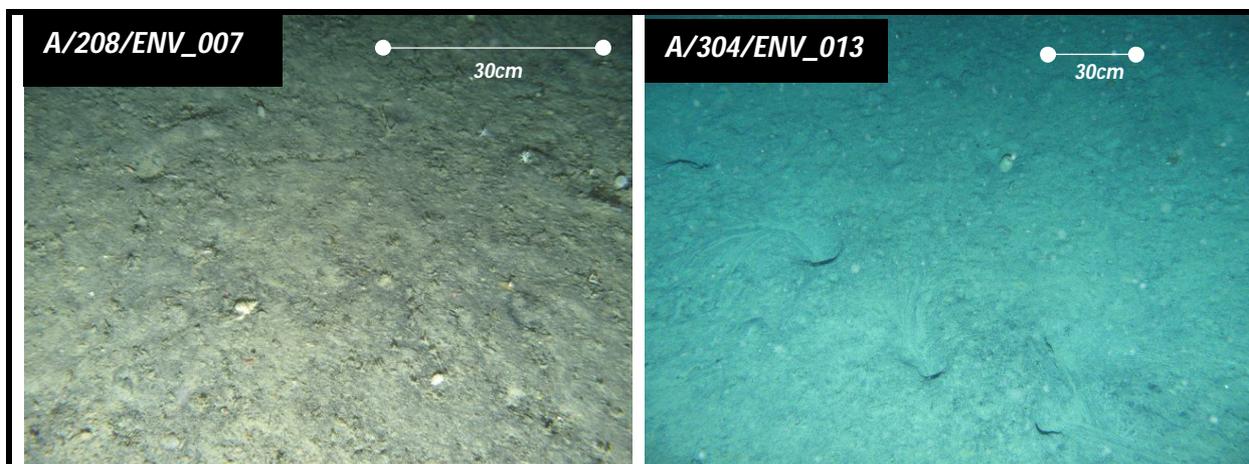
Environmental ground-truthing using both cameras and seabed sampling confirmed the presence of these sediment changes although the boundaries of these main areas are estimated based on the coverage by the ground truth stations or interpretation from the regional bathymetry.

A summary of the key habitat variations recorded during ground truthing within the FISA Survey area are outlined below in Figure 2.52 (and Appendix VIII). Greater detail regarding the habitat variations are presented in Volume 2 of this report (MG3, 2014b).

2.10.2.1 Habitat: Holocene Sedimentary Slightly Gravelly Silty Sand:

The majority of the seabed is described as featureless slightly gravelly silty sand. This is broadly similar to an offshore circalittoral sand (SS.SSa.OSa; Connor *et al.*, 2004). This material will be related to granular Holocene sedimentary material and granular Pleistocene residues (i.e. drop-stones, recorded during faunal sieving). Seabed photography recorded a presence of bioturbation in the form of burrows from various crustaceans, as well as other "lebensspuren" (animal tracks and furrows) likely to be produced by echinoderms (in particular spatangoids) observed in both the grab sampling and seabed imagery. An example of this habitat type is shown in Figure 2.53.

Figure 2.53 - Example Images of Slightly Gravelly Silty Sand Habitat



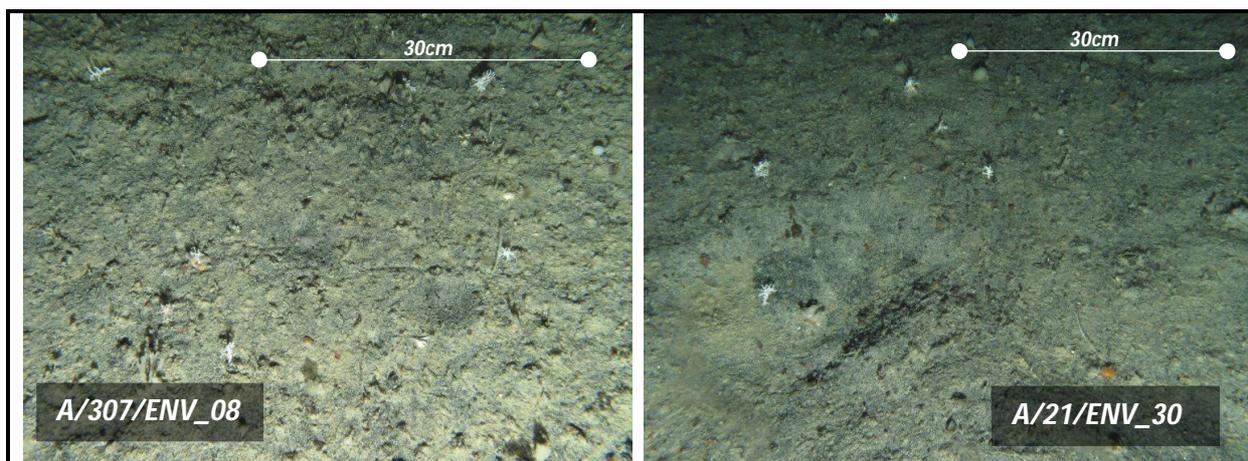
Relatively few conspicuous fauna were recorded in this habitat by seabed photography, although the presence of some bioturbation would signify a relatively rich infaunal community. The seabed was generally featureless with an absence of current related bedforms indicative of a sedimentary regime and limited hydrodynamic reworking of sediments. Conspicuous fauna observed included the isopod Serolidae (possibly the deep water species *Acutiserolis neaera*), occasional ophiuroids, the potential CITES Appendix II solitary cup coral (*Flabellum* sp.) and the striped shrimp (*Nauticaris* sp.).

2.10.2.2 Habitat: Featureless Gravelly Silty Sands:

The seabed to the southeast of the escarpment indicated a similar featureless, slightly mixed seabed to that described in Section 2.10.2.1, with the exception that coarser gravel admixtures were recorded on the surface of the seabed (as observed by seabed photography).

As with the shallower sediments this is broadly similar to an offshore circalittoral sand (SS.SSa.OSa; Connor *et al.*, 2004), but with the addition of fine gravels, although not sufficient to alter the general habitat to an offshore circalittoral mixed sediment (SS.SMx.OMx; Connor *et al.*, 2004). Nevertheless, the presence of fine pebbles at the surface has introduced a small epifaunal community of small isolated stone coral-like Hydrozoa *Stylaster* sp., usually in the form of a single branched 'sprig' only a few centimetres high. Other species recorded were echinoderms (a cushion star, the pencil urchin (*Cidaris* sp.) and a burrowing holothurian) and the same Serolidae isopod as recorded elsewhere on the FISA site (possibly *A. neaera*). The small pebble surfaces also support the occasional foliose bryozoan, ascidians and solitary sponge (*Suberites* sp.), although the solitary cup corals (*Flabellum* sp.) previously recorded on the Holocene slightly gravelly silty sands, appeared to be absent.

Figure 2.54 - Example Images of Gravelly Silty Sand



This sediment will be related to granular Holocene sedimentary material and coarser granular Pleistocene residues (drop-stones observed at the surface and during faunal sieving). Seabed photography indicated a presence of bioturbation in the form of

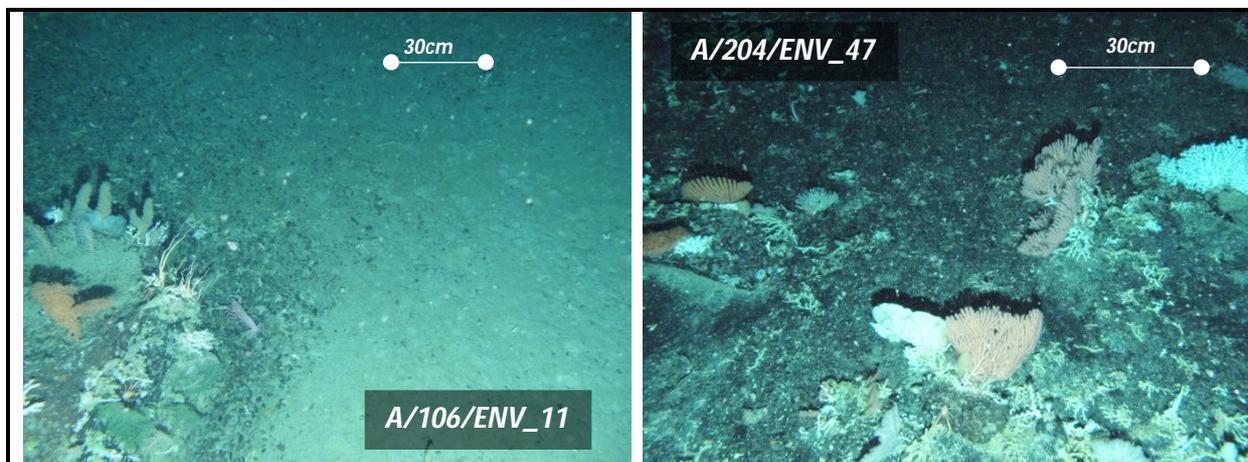
spatangoid furrows and surface “lebensspuren” (animal tracks). An example of this habitat type is shown in Figure 2.54.

The Finback-1 and Humpback-2 proposed well location is located in this sediment type.

2.10.2.3 Habitat: Silty Sands with Occasional Cobbles and Boulders:

This habitat type is recorded in areas where underlying hard geology pinches out close to the surface and along the peripheral edges of the bedrock exposures where the surface granular sediments thin to a veneer. This creates a mixed habitat of featureless sands punctuated by rock outcrops, large cobbles and boulders all populated with small but conspicuous epifaunal communities, all associated with the hard substrate. An example of this habitat type is shown in Figure 2.55.

Figure 2.55 - Example Images of Silty Sands with Cobble and Boulder Outcrops



As with the previous habitat, this sediment is broadly similar to an offshore circalittoral sand (SS.SSa.OSa; Connor *et al.*, 2004), but with localised outcrops of circalittoral low energy rock (CR.LCR; Connor *et al.*, 2004). These isolated hard substrates are well populated by developed epifaunal communities based on both hard and soft corals. Examples of species recorded include the hard branched stone coral-like Hydrozoa *Stylaster* sp., occasionally developed into fan shaped morphology of 20-30cm in diameter. Other corals include soft octocorals (similar to *Callogorgia* sp.), and solitary cup corals, solitary sponges (*Suberites* sp.), and some foliose bryozoans. Living around these rocks are a number of echinoderms, including ophiuroids spread amongst the corals, the pencil urchin (*Cidaris* sp.), whilst several examples of larger boulders observed a resident small lobster as well as several small squat lobsters (*Munida* sp.).

Some of these boulders indicated the presence of current scour including localised areas of ripples indicative of turbulence around the feature and the subsequent reworking of the sediments by the seabed water movements.

The Humpback-1 proposed well location (A/27/ENV) is located in this sediment type.

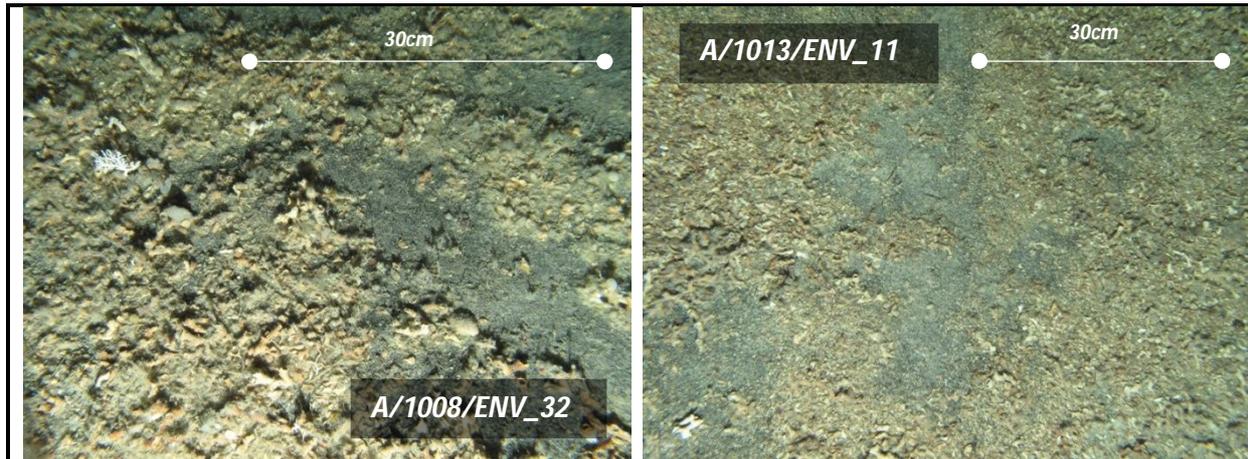
2.10.2.4 Bedrock Exposure and Localised Patches of Relic Coral Debris

Several exposures of bedrock were recorded within the FISA survey area, predominantly associated with the steeper slopes of the central escarpment and on the edges of collapsed depressions in the northwest. For the most part, the epifaunal coverage of exposed bedrock, including the areas of slumped boulders found at the base of most steep sloped areas, was limited to a low lying faunal turf of bryozoans, hydroids and the occasional soft coral. Some larger examples of well-developed *Stylaster* sp. hydrozoans were also recorded, but these were low in number. However, the most dominant species recorded in these areas was that of the reef-building stone coral *Lophelia pertusa*. This was in localised areas of high density, associated with the sides of boulders at the base of the escarpment, or low level coverage at the top of the escarpment, at the edge of the drop-off. Both areas are likely to be associated with the strongest water movement in the area.

Generally, the coverage by *L. pertusa* was predominantly made up of relic debris of whole and fragmented material no larger than 10cm in length. Relatively few examples of live material were recorded and these were all less than a few decimetres in diameter. This would suggest that larger reefs created by *L. pertusa* are absent from the area. A low ambient seabed water temperature of approximately 2.5°C is thought to inhibit the growth rate and development of *Lophelia*, restricting its size and proliferation in the polar regions (Zibrowius, 1980; Cairns, 1994). Corals preferred temperature range is thought to be between 6°C and 8°C (Frederiksen *et al.*, 1992; Freiwald, 1998).

The extensive coverage of *Lophelia* debris appears to be a relic feature of considerable age. An example of this habitat type is shown in Figure 2.56. As the area of proliferation is localised and limited to a rocky, non-sedimentary environment, the residues of earlier colonies are not incorporated within a carbonate mound or bioherm. Consequently, the presence of *Lophelia* (live or dead) in these areas is not thought to be of major significance to the local biodiversity in the region and therefore are not thought to be of notable conservational importance. These corals are generally considered to be vulnerable due to their sensitivity to trawling damage and their slow growth rate. The FISA area is not currently impacted by trawlers at the depth of this populations (1300m to 1400m), and the closest populations are expected to be around 3.8km south of the nearest proposed well (Humpback-1).

Figure 2.56 - Example Images of Coral Debris from *Lophelia pertusa* (Scleractinia)



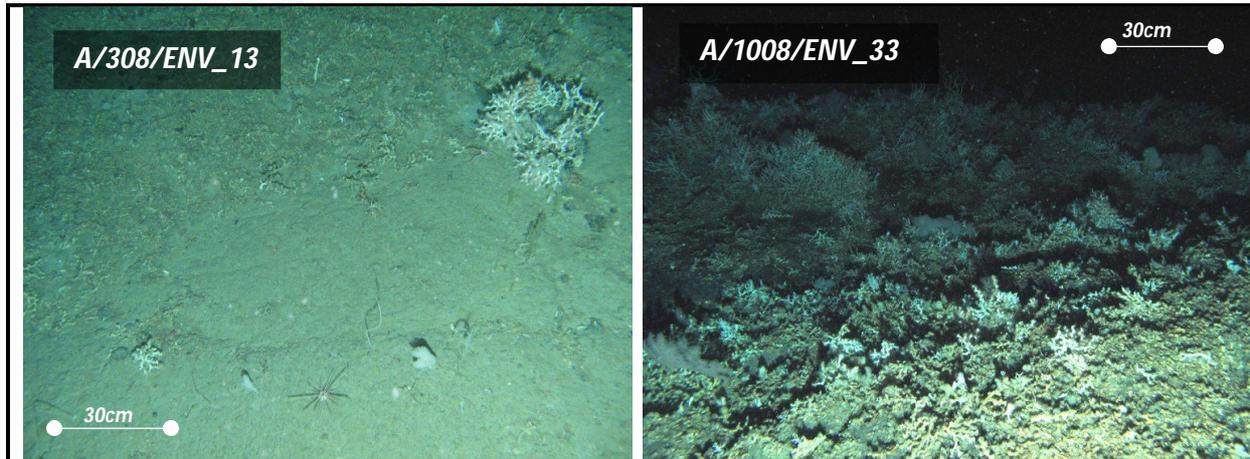
2.10.2.5 Potential Annex I Habitats and Other Sensitive Habitats

The majority of evidence for the presence of *L. pertusa* relates to extensive areas of debris relating to relic residue populations located on the edge of the escarpment drop-off approximately 3.8km southeast of Humpback-1 and approximately 5.7km northwest of the proposed Humpback-2 well locations. Examples of live populations are presented in Figure 2.36. These mostly show localised 'sprigs' of *Lophelia* (a few decimetres in length) recorded on the edges of prominent boulders or at the very edge of the escarpment drop-off where the current movements are greatest. No 'thickets' (aggregations large than 1m) of *L. pertusa* were recorded during the survey.

Despite the lack of size or proliferations across the area, whilst the recorded aggregations of live *Lophelia* are not generally classified as Annex I habitats, there may be instances along the edge of escarpment where larger aggregations may exist and should be protected from physical damage. However, as these areas are located several kilometres from the proposed well locations, they are not expected to be impacted by drilling related discharges from either of the two proposed well sites. Should the final well locations move, then these should not be brought within 500m from the edge of the escarpment.

The remainder of the survey area did not yield any evidence of particularly sensitive habitats, particularly within the immediate vicinity of the proposed exploration wells. The habitats in these areas were considered to be homogenous, gravelly or slightly gravelly silty sands with limited sensitivity to the proposed operations. A review of both stills and video footage indicate that some potential CITES Appendix II coral species (such as *Flabellum* sp.) were observed.

Figure 2.57 - Example Images of Live *Lophelia pertusa* Corals (Scleractinia) Found in the FISA Survey Area



3 CONCLUSION

The bathymetry and geology of the surveyed area showed a deep water environment with low sedimentation rate and a seabed of gravelly silty sands with historical influences from glacial activity in the form of some coarser sediment admixtures. The bathymetry indicated a shallower seabed to the northwest with the gradient in this area punctuated by a number of well-defined escarpments where the slope of the seabed increased from $<2^\circ$ to in excess of 45° , indicative of almost vertical rock faces. The base of these features was marked by slump deposits of boulders eroded from the escarpment. Ground truthing of sediments across the site indicates relatively homogenous slightly gravelly silty sand throughout. Below the escarpment to the southeast of the survey area, the proportion of surface gravels increased slightly, whilst intermediate sediments at the top of the escarpment were represented by a thin veneer of silty sands punctuated by regular large cobbles, rock outcrops and boulders.

The main habitats recorded related to an offshore circalittoral sand which was separated into two areas of greater or lesser surface gravels (a result of historical glacial deposits in the area). The deeper waters indicate a lower depositional rate. The proposed well locations of Finback-1, Humpback-1 and 2 were all located within the gravelly sands, with sediments at Humpback-1 proposed well location also comprising localised exposures of cobbles or boulders.

Environmental baseline sampling undertaken within the FISA regional survey area acquired physico-chemical and biological macro-invertebrate samples from 26 stations and further seabed photography from 34 stations. Observations for each of the key parameters are summarised as follows:

In line with the general habitat variations recorded across the survey area, the particle size analysis indicated some variation in the different proportions of sands, fines (below $63\mu\text{m}$) and gravels ($>2\text{mm}$) which indicated some variability across the survey area. The average for the sediment composition each of the three main size fractions was sand (60.3%), fines (32.7%) and gravels (7.0%). The proportion of these different sizes ranged from a sandy mud and muddy sand through to a gravelly sand, but with the dominant sediment type representative of a slightly gravelly muddy sand as denominated by the Folk characterisation (Folk, 1954).

Analytical results mostly indicated a uni-modal distributions peaking around the medium sand to fine sand with a consistent tail of fines varying through silts and clays, or occasionally a bi-modal distribution with a slightly elevated proportion of coarse silts, and/or gravels. The mean particle size ranged from $45\mu\text{m}$ to $354\mu\text{m}$, demonstrating the slightly elevated proportions of silts and clays at two sites and gravels at four sites. With the exception of these sites, the sorting coefficient generally ranged from 2.3 to 2.9 based on poorly sorted slightly gravelly muddy sand. The mean proportion of gravels was typically around 7%, but elevated levels between 16% to 27.3% were recorded at the four stations. However, where mixed sediment occurs, the sampling process itself can be biased against coarser material due to a small sub-sampling procedures or the inability to sample mixed

sediments by the samplers themselves. In these cases, coarser clasts are likely to be underestimated, with cobbles and boulders frequently recorded by the seabed photography.

Only two locations indicated proportion of fines greater than 50%, with a peak 66.7% recorded at station A/26/ENV at the same location as the Humpback well prospect. The mean proportion of sediment fines was 32%. The distribution of fines indicated a pattern of distribution with fewer fines in the deeper sediments. This pattern is considered unusual for a deep-water environment where, typically, the deeper waters indicates a higher rates of pelagic sedimentation due to decreased water movement. In FISA the majority of softer sedimentary are recorded above the escarpment feature north and west of the proposed well locations and follows a similar trend identified with the neighbouring sites of Toroa and Nimrod which indicated softer sedimentary material on the continental slope, but heavily eroded and non-depositional bedrocks and glacial sands below. This may be a combination of the East Falklands current coupled with the rugose seabed created by discordant or irregular formation erosion rates. This has been observed in some of the video footage on this and other neighbouring sites (i.e. FINA).

Some of the chemical parameters surveyed followed patterns observed within the particle size analysis. Total organic carbon (TOC), total hydrocarbon content (THC) and saturates all indicated relatively low background concentrations with a subtle pattern of distribution relative to the mean particle size or the proportion of fines. This relates to the increased surface area provided by silts and clays which sorbs and adsorbs organic and some inorganic components within the sediments.

The total hydrocarbon content (THC) concentrations gave a mean of $3.1\mu\text{g.g}^{-1}$ whilst alkanes indicated a mean of 313ng.g^{-1} , approximately 10% of the total THC recovered. Inspection of the individual gas chromatograms at all stations revealed no background fingerprints throughout with no specific envelopes or peaks relating to anthropogenic or petrogenic. Whilst a review of the alkanes showed that terrestrial influences to the saturates were also minimal at the site, indicative of an offshore site influenced by Antarctic water masses, and a relatively low pyrolytic polycyclic aromatic hydrocarbon influence, the presence of 2 and 3 ring PAHs (naphthalene, phenanthrene and dibenzothiophene or NPD) and the high incident of alkylated PAHs suggests a clear and chronic input from petrogenic sources, albeit at a trace level. This is likely to be from natural hydrocarbon seeps previously recorded within the region. Historical datasets on the Burdwood Bank (BSL, 2008) and surveys at Toroa have similarly recorded background petrogenic influences to the surface environmental samples with a suggestion that the source are south and west of the FISA location. Total PAH concentrations (2-6 compounds) recorded a mean of 82.8 ng.g^{-1} whilst the mean of the NPD fraction 43.8 ng.g^{-1} .

The concentrations of heavy and trace metals were consistent with historical surveys within the area. The distribution of the metals often correlated with themselves and occasionally other sediment factors. The most obvious of these was that of aluminium which indicated a strong correlation with % gravels and sorting coefficient. Several other metals indicated auto-correlations with each other (that is to say that they followed similar

trends relating to the same parameter but not necessarily each other). One clear correlation between metal was recorded between iron and arsenic. Iron indicated a huge variability across the survey area ranging from 10.7 to 135mg.g⁻¹. This has been interpreted as a function of the variability of the residual glacial materials recorded at some sites, in particular the black basalt sands reworked by the currents in some areas. Arsenic, which is often adsorbed onto iron, indicated modest but quite variable concentrations ranging from 3.2 to 12.3 µg.g⁻¹. All metals were processed for total content using a hydrofluoric acid digestion. Results recorded trace levels of cadmium and mercury, whilst the means of lead (6.2µg.g⁻¹), barium (331.7µg.g⁻¹), chromium (128µg.g⁻¹), nickel (12.5µg.g⁻¹), copper (15.8µg.g⁻¹), vanadium (76.9µg.g⁻¹) and zinc (77.5µg.g⁻¹) all gave background concentrations.

Multiple water column profiles taken over a time period of a calendar month indicated a consistent water column with a weak thermocline in the surface 80m followed by a slow cooling to the seabed from 5.73 to 6.70°C at the surface to around 2.60°C at 1432m. All profiles indicated a small area of mixed waters between 620m and 950m. The salinity only varied by 0.5psu throughout the whole water column, showing similar structure to that of the thermocline but an increase from 34.4 to 34.9psu with depth. Water chemistry, recorded at four depths at two sites, indicated trace or undetectable concentrations expected for open 'blue-water' environment.

A macrofaunal analysis was carried out on all 59 replicates obtained at 26 baseline sediment sites, sampled within the FISA survey and processed in the field using a 500µm mesh size. Subsequent macrofaunal taxonomy of all recovered fauna identified a total of 2,874 individuals. Of the 254 species recorded, 188 were infaunal dominated by small polychaetes. This dominance was by both abundance and richness, with six polychaetes recorded in the top ten numerically ranked species. This dominance was closely followed by crustaceans which were also well represented, having three species in the in the top ten numerically ranked taxa. In overall rank order, the top 5 key dominant species across the area were the Polychaete *Rhamphobrachium (Spinigerium) ehlersi* the crustaceans *Ampelisca* and a Copepoda, separated by the polychaete *Gymnonereis fauveli* and followed by another polychaete *Apistobranchus* sp I. The species list was very diverse due to the number of sites, with around 88% of specimens identified to genus or species. Several undescribed species were recorded, but around 45% of the species list was similar to the population recorded on the Burdwood Bank in 2009 on similar sediment type (BSL, 2008). Whilst the deep water coral *Lophelia pertusa* was recorded in the samples, all incidents were of relic debris with no live animals recovered. Camera operations, however, did identify small examples of this species, but of a morphology insufficient to be considered as an Annex I habitat under the European Habitats Directive, or the OSPAR list of threatened habitats. Also of note from the macrofaunal samples was one group of solitary corals (*Flabellum* sp.) and a very rare mollusc of the group Monoplacophora (*Neopilina*). This group was thought to be extinct for 400 million years until a recent discovery of a living specimen from the deep sea. These animals are very rare, and so far only about 150 specimens have ever been found worldwide, belonging to about 25 species.

Univariate analysis of the data indicated that the species richness and abundance were similar to previous surveys in the area by surface area. This was 20 species for 0.1m² and a mean of 351 individuals per m², although the variability of the sediments has shown significant variability between samples. The mean Shannon-Weiner diversity was 4.64 for the station (0.2m²). The bioaccumulation curves indicated that the potential number of species calculated from asymptote was estimated to be 236 species (using Chao-1 analysis).

Multivariate analysis indicated no clearly separated faunal populations, although results showing clustered stations at the 35%-55% similarity range producing 12 statistically distinct groups within the population, seven of which represented by a single station. An ordination of these results failed to produce a clear 2D plot due to high stress within the calculation, but a 3D representation indicated no significant trend based on geographical location, sediment type or other environmental factors.

Due to the large amount of data and broad transition of sediment types identified, varying from sandy muds, though to gravelly sands, it appears that this may have created an underlying gradient of change from to assign discrete biological communities. This results in a large amorphous grouping of station similarities, indicative of a single but broad macrofaunal assemblage. A multivariate comparison with a similar dataset acquired at the Burdwood Bank showed significant separation in the populations despite the similar species list. A review of these differences using the software showed that differences observed between the two main clusters appear to be a subtle accumulation of differences in species counts within the dominant species identified, rather than any fundamental population change.

In addition to the infaunal community the qualitative presence of the epifaunal community was reviewed from the seabed samples, and provided a rich faunal assemblage, especially in areas where drop-stones were common. Key faunal groups were the sponges, class Hexactinellida, Calcarea and Demospongia; many of the genera were typical for deeper water (e.g. *Hymedesmia* spp., *Crella* and *Lissodendoryx*), with a wide and sometime cosmopolitan distribution. The Cnidaria were represented by nine genera of thecate and athecate Hydrozoa and two genera of Stylasteridae. In particular *Stylaster densicaulis* which was also recorded in the photography. The live solitary coral belonging to the species *Flabellum curvatum* (a potential CITES Appendix II coral species) was also recorded. Several Octocorallia were found, including the sea pens Pennatulidae (generally too small to identify) and two species of Gorgonacea. Bryozoa were also very common constituent on pebbles and stones, with many species endemic.

A further assessment of the epifaunal assemblages shown within the seabed photographs, confirmed a relatively well populated community of species overall, with healthy aggregations colonising occasional drop-stones as well as *Lophelia* debris, where present and some rooted into the sediment. Many of the species recorded were associated with deep water and/or cold water environments. Overall, the phylogenetic make-up of the conspicuous megafauna observed was dominated by the cnidarians, with octocorals and Pennatulidae remaining prevalent throughout. Echinoderms were better represented in

the seabed photography than the grab samples with ophiuroids, crinoids, asteroids and holothurians present. Where drop-stones were present, encrusting sponges were common along with anthozoans. Often rooted into soft sediments bryozoans and hydroids were observed as sparse tufts. There were also numerous burrows likely to be associated with crustacean and holothurian activity. Free-swimming megafauna included the demersal teleosts: Macrouridae, Grenadier, hake and batoids.

The remainder of the survey area did not yield any evidence of particularly sensitive habitats, especially within the immediate vicinity of the proposed exploration wells. The habitats in these areas were considered to be homogenous, gravelly or slightly gravelly silty sands with limited sensitivity to the proposed operations. A review of both stills and video footage indicate that some

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APPENDIX I – FIELD OPERATIONS AND SURVEY METHODS

I.1 SEABED PHOTOGRAPHY AND VIDEO

Seabed photography was undertaken using a “Sea Bug” system developed by Subsea Technology and Rentals (STR). The system is based on a housed G10 Canon Camera which can output up to a 14.7 megapixel quality (five megapixel used for the current project to reduce operational upload time). The system is mounted in a fish with a skid and drop-down frame which can be towed as a sled on, or just above the seabed for the camera. The output from the camera is controlled from the surface using a video stream supplied from the camera and number of function controls operated through a multiplexer system and armoured 3km sonar cable. Once on the seabed, the system was towed for at least 100m and a minimum of 5 digital stills photographs acquired at each site.



Camera system configuration

The key acquisition parameters of the system used are as follows:

| Standard Features | Comment |
|---------------------------------|--|
| Image Resolution | 5 to 14.7 megapixel (up to 4,416 x 3,312 pixels) |
| Light Sensitivity setting | ISO 60-1600 Auto/Manual Selected |
| Sensor Type | 1 / 1.8" format high density CCD sensor |
| Light source | 6 x 1000lumen controllable LED lamps Stills strobe TTL controller |
| Typical settings on this survey | Aperture priority at F8, Shutter speed typically 1/125 th second, Auto flash mode (TTL) |
| Framing Video Used | 320 Line / 50 Hz PAL |
| Control System | STR Multiport DTS |
| Other sensors | Roll, Pitch, Heading and Altimeter |

The system was deployed from the stern of the vessel using the vertical drop-down frame technique and lowered to the seabed using the inbuilt altimeter. The online video was used to control the frames height and attitude once at the seabed. The seabed was surveyed using a combination of video and digital still photographs (minimum of 5 shots per site).

Positioning of the system was maintained throughout by using a USBL positioning beacon attached to the cable above the camera frame. The vessel maintained this positioning or followed a target transect using its dynamic positioning (DP) and was allowed to move at a tow speed of approximately 0.5 - 1 knot (< 0.5m/sec). As no environmentally sensitive sites (such as biogenic reefs or sponges), were encountered within the main survey area, evasive measures to prevent contact with the seabed were not necessary. Operations over the area of hard corals were carried out at a controlled height just off the seabed where possible. The frame was also fitted with a laser scale set to a 30cm separation.

Stills images have been reviewed for larger and obvious mega-fauna and epifaunal species to assist in diagnosing sediment changes across the sites and to assist in the macrofaunal interpretation. The camera system is a particularly useful tool in recording larger contacts, hard substrates or occasionally seabed features impossible to sample using the Box corer or grab samplers.

I.2 BENTHIC ENVIRONMENTAL SAMPLING

A combination of benthic sampling devices were mobilised to allow for the varied surface geological conditions, as previously encountered at neighbouring sites throughout the South Falklands Basin (BSL, 2008). The unusual and variable nature of the surface geology encountered at the site necessitated a flexible approach to sample recovery, without which little or no material would have been recovered at many of the locations. MG3 mobilised a 0.25m² USNEL box corer as the primary seabed sampler. Additional sample acquisition contingency measures were in place in the form of alternative sampling devices, in the event that unsuitable geology was encountered by the primary seabed sampler. A unique double Van Veen grab sampler was mobilised (designed for operation in very soft sediments and compacted sands), along with a 0.1m² Hamon grab sampler (for operation in mixed sediments) as contingency devices. In the event, all three devices had to be utilised in order to recover material in the compacted and sometimes gravelly sediments encountered within the FISA survey area. Details of the sampling equipment and associated operations are as follows:

I.2.1 Box Corer Operations

The primary seabed sampler was a 0.25m² USNEL type box corer, requiring a single deployment at each of the proposed locations. Pre-deployment procedures included cleaning of the inner stainless steel box, cable and blocks to be generally grease free. A record of the samplers touch down at deployment depth was monitored by means of a load cell (within the winch) which recorded tension on the cable. Samples were subject to quality control on retrieval and were retained in the following circumstances:

- Water above sample was undisturbed;
- Spade closure complete allowing no sediment washout;
- Penetration of the box was sufficient to maintain a seal at the base of the core but not over penetrated and allowing water to remain above the sample when recovered;
- Sampler was retrieved perfectly upright and had not fouled in any way;
- Inspection/access doors had closed properly enclosing the sample;
- No disruption of the sample through striking the side of the vessel;
- Sample were taken inside the acceptable target range; and
- The sample was acceptable to the principle scientist.



For each sample, the whole core was inspected, described and sub-sampled for two 0.1m² replicates before being photographed, and then processed onboard using a Wilson Auto-Siever over a 500µm aperture mesh. [Key observations from samples were colour, sediment classification, layering (including RDLs), smell (including the presence of H₂S), obvious fauna and evidence of bioturbation]. The separation of the macrofaunal samples was undertaken following area segregation and siphoning the supernatant surface waters prior to sediment extractions down to a depth of 20cm. The remainder of the core sample (0.05m²) was surface sub-sampled for physico-chemistry. An additional hand core (89mm id) was also taken to preserve the surface structure of the samples for possible later geotechnical analysis. This was pushed into the full penetration of the box core, and complete core extracted and stored upright.

Pre-deployment procedures included the cleaning of the inner stainless Box corer buckets, cable and blocks so that they were generally grease-free. On retrieval, the whole sample was inspected, described and photographed prior to processing. Key observations from samples were colour, sediment classification, layering (including RDLs), smell (including the presence of H₂S), obvious fauna and evidence of bioturbation and evidence of anthropogenic debris. To obtain sufficient replication, this device was deployed a total of four times to recover a suitable surface area for both macro-invertebrate and an additional sample for sediment physico-chemistry.

I.2.2 Double Van Veen Operations

Where box corer penetration failed (usually in compacted sands limiting penetration) the double grab sampler was employed. This device has two samplers in a ballasted frame and acquires a seabed sample area of 2 x 0.1m² on each deployment. Pre-deployment procedures included the cleaning of the inner stainless grab buckets, cable and blocks so that they were generally grease free. Samples were subject to quality control on retrieval and were retained in the following circumstances:



- Water above sample was undisturbed;
- Bucket closure complete allowing no sediment washout;
- Penetration of the grab was sufficient to maintain a seal at the base;
- Sampler was retrieved perfectly upright and had not fouled in any way;
- Sampler access doors had closed properly enclosing the sample;
- No disruption of the sample through striking the side of the vessel;
- Sample was taken within the acceptable target range; and
- The sample was acceptable to the principle scientist.

I.2.3 Hamon Grab Operations

If ground truthing revealed mixed coarse sediment, a 0.1m² Hamon grab was used to recover sediments including; gravels, pebbles and cobbles. This device has a single 0.1m² scoop attached to a pivoting arm, and allows retention of a sample even if the grab is not completely closed. While requiring a minimum of three deployments for this survey and recovering a relatively disturbed sample, it does however have a high success rate in sampling mixed coarse sediments where other samplers would fail. Pre-deployment procedures included the cleaning of the inner stainless grab bucket, cable and blocks so that they were generally grease free. Samples were subject to quality control on retrieval and were retained in the following circumstances.

- Water above sample was undisturbed;
- Penetration of the grab was sufficient to maintain a seal at the base;
- Sampler was retrieved perfectly upright and had not fouled in any way;
- Sampler access doors had closed properly enclosing the sample;
- No disruption of the sample through striking the side of the vessel;
- Sample was taken within the acceptable target range; and
- The sample was acceptable to the principle scientist.

On recovery, the whole sample was inspected, described and photographed prior to processing. A minimum of three 0.1m² replicates were required per sample location to acquire enough material for two fauna replicates and sub-sampling of physico-chemistry. Faunal replicates were processed onboard using a Wilson Auto-Siever over a 500µm aperture mesh. Key observations from samples were colour, sediment classification, layering (including RDLs), smell (including the presence of H₂S),

obvious fauna and evidence of bioturbation and any anthropogenic debris. The remaining sample (0.1m²) was sub-sampled for physico-chemistry.

I.2.4 Sample Processing

Sub-sampling of physico-chemistry was undertaken from the sampler with the following material retrieved from the surface 2cm:

- Hydrocarbon analysis (collected with stainless steel scoop or directly in a prewashed foil capped glass jar);
- Heavy and trace metals (collected with plastic scoop and stored in doubled lined ziplock plastic bag);
- Particle Size Analysis (PSA) (collected with plastic/stainless steel scoop and stored in doubled lined ziplock plastic bag);
- Total organic matter and total organic carbon (TOM & TOC)(collected with plastic scoop and stored in doubled lined ziplock plastic bag); and
- Duplicate samples from all of the above.

The preservation of materials was undertaken using standard techniques. All physico-chemical samples were stored in appropriate containers (i.e. glass for hydrocarbons, and plastics for metals and PSA) and immediately frozen and stored (<-18°C) for later transportation (frozen) to the laboratory on demobilisation. Faunal samples were processed onsite using a Wilson Auto-Siever over a 500µm aperture mesh, with all residues retained, fixed and stained in 5% buffered formalin and a vital stain (Rose Bengal) for storage and transportation. This material was later transferred to IMS. All biological samples were double-labelled, with internal tags. For samples that retained minor amounts of clays a small amount of additional di-sodium hexametaphosphate was included within the fixative onboard to induce clay separation during storage. Photographs from the field sampling operations are given in the final environmental baseline report.

I.2.5 Water Sampling using the Niskin Bottle

Water samples were taken at four different depths – at the surface, thermocline, mid-depth and at the seabed to obtain a representative vertical profile. Pre-deployment procedures included a thorough check of the sampling device and all rubber seals, the firm attachment of the Niskin Bottle to the water sampling wire as well as correct setup and function of all mechanisms.

I.2.6 Sample Processing

On recovery, samples were sampled for the following material and stored appropriately depending on the analysis:

- 1 x glass bottle for hydrocarbons analysis;
- 1 x plastic bottle for heavy metals analysis;
- 1 x plastic bottle for nutrients analysis; and
- 1 x plastic bottle as a spare.

APPENDIX II – LABORATORY ANALYSES AND STATISTICAL ANALYSES

II.1 PARTICLE SIZE DISTRIBUTION

The samples recovered from each site were analysed by Benthic Solutions Limited which is accredited under the National Marine Biological Association Quality Control scheme (NMBAQC) for PSA analysis.

The sample was homogenised and split into a small sub-sample for laser diffraction and the remaining material was sieved through stainless steel sieves with mesh apertures of 8000µm, 4000µm and 2000µm. In most cases almost the entire sample would pass through the sieve stack, but any material retained on the sieve, such as small shells, shell fragments and stones were removed and the weight was recorded.

The smaller sub-sample was wet screened through a 2000µm and determined using a Malvern Mastersizer 2000 particle sizer according to Standard Operating Procedures (SOP). The results obtained by a laser sizer have been previously validated by comparison with independent assessment by wet sieving (Hart, 1996). The range of sieve sizes, together with their Wentworth classifications, is given in Table A. For additional quality control, all datasets were run through the mastersizer in triplicate and the variations in sediment distributions assessed to be within the 95% percentile.

The separate assessments of the fractions above and below 2000µm were combined using a computer programme. This followed a manual input of the sieve results for fractions 16-8mm, 8-4mm and 4-2mm fractions and the electronic data captured by the Mastersizer below 2000µm.

This method defines the particle size distributions in terms of Phi mean, median, fraction percentages (i.e. coarse sediments, sands and fines), sorting (mixture of sediment sizes) and skewness (weighting of sediment fractions above and below the mean sediment size; Folk 1954).

Formulae and classifications for particle calculations made are given below:

- **Graphic Mean (M)** - a very valuable measure of average particle size in Phi units (Folk and Ward, 1957).

$$M = \frac{\phi 16 + \phi 50 + \phi 84}{3}$$

Where M = The graphic mean particle size in Phi
 ϕ = the Phi size of the 16th, 50th and 84th percentile of the sample

Table A - Phi and Sieve Apertures with Wentworth Classifications

| Aperture in microns | Aperture in Phi Unit | Sediment Description | |
|---------------------|----------------------|----------------------|---------------|
| 2000 | -1 | Granule | Gravel |
| 1400 | -0.5 | Very Coarse Sand | Sands |
| 1000 | 0 | | |
| 710 | 0.5 | Coarse Sand | |
| 500 | 1 | | |
| 355 | 1.5 | Medium Sand | |
| 250 | 2 | | |
| 180 | 2.5 | Fine Sand | |
| 125 | 3 | | |
| 90 | 3.5 | Very Fine Sand | |
| 63 | 4 | | |
| 44 | 4.5 | Coarse Silt | Fines (Silts) |
| 31.5 | 5 | | |
| 22 | 5.5 | Medium Silt | |
| 15.6 | 6 | | |
| 11 | 6.5 | Fine Silt | |
| 7.8 | 7 | | |
| 5.5 | 7.5 | Very Fine Silt | |
| 3.9 | 8 | | |
| 2 | 9 | Clay | Fines (Clays) |
| 1 | 10 | | |

- **Sorting (D)** – the inclusive graphic standard deviation of the sample is a measure of the degree of sorting (Table B).

$$D = \frac{\phi_{84} + \phi_{16}}{4} + \frac{\phi_{95} + \phi_5}{6.6}$$

where

D = the inclusive graphic standard deviation

φ = the Phi size of the 84th, 16th, 95th and 5th percentile of the sample

Table B - Sorting Classifications

| Sorting Coefficient (Graphical Standard Deviation) | Sorting Classifications |
|--|-------------------------|
| 0.00 < 0.35 | Very well sorted |
| 0.35 < 0.50 | Well sorted |
| 0.50 < 0.71 | Moderately well sorted |
| 0.71 < 1.00 | Moderately sorted |
| 1.00 < 2.00 | Poorly sorted |
| 2.00 < 4.00 | Very poorly sorted |
| 4.00 + | Extremely poorly sorted |

- **Skewness (S)** – the degree of asymmetry of a frequency or cumulative curve (Table C).

$$S = \frac{\phi_{84} + \phi_{16} - (\phi_{50})}{2(\phi_{84} - \phi_{16})} + \frac{\phi_{95} + \phi_5 - 2(\phi_{50})}{2(\phi_{95} - \phi_5)}$$

where

S = the skewness of the sample
 ϕ = the Phi size of the 84th, 16th, 50th, 95th and 5th percentile of the sample

Table C - Skewness Classifications

| Skewness Coefficient | Mathematical Skewness | Graphical Skewness |
|----------------------|-----------------------|------------------------|
| +1.00 > +0.30 | Strongly positive | Strongly coarse skewed |
| +0.30 > +0.10 | Positive | Coarse skewed |
| +0.10 > -0.10 | Near symmetrical | Symmetrical |
| -0.10 > -0.30 | Negative | Fine skewed |
| -0.30 > -1.00 | Strongly negative | Strongly fine skewed |

- **Graphic Kurtosis (K)** – The degree of peakedness or departure from the ‘normal’ frequency or cumulative curve (Table D).

$$K = \frac{\phi_{95} - \phi_5}{2.44(\phi_{75} - \phi_{25})}$$

Where

K = Kurtosis
 ϕ = the Phi size of the 95th, 5th, 75th and 25th percentile of the sample

Table D - Kurtosis Classifications

| Kurtosis Coefficient | Kurtosis Classification | Graphical meaning |
|----------------------|-------------------------|---|
| 0.41 < 0.67 | Very Platykurtic | Flat-peaked; the ends are better sorted than the centre |
| 0.67 < 0.90 | Platykurtic | |
| 0.90 < 1.10 | Mesokurtic | Normal; bell shaped curve |
| 1.11 < 1.50 | Leptokurtic | Curves are excessively peaked; the centre is better sorted than the ends. |
| 1.50 < 3.00 | Very Leptokurtic | |
| 3.00 + | Extremely Leptokurtic | |

II.2 SEDIMENT TOTAL ORGANIC CARBON (TOC)

Organic and carbon sediments are analysed using a combination of tests. These include Total Carbon (TC), analysed using a known weight of dried soil and combusted at 1300° C and the amount of Carbon determined by Infra-Red detection, and Total Organic Carbon (TOC; see below). In addition to the standard accreditation as outlined below, additional analytical quality control (AQC), is carried

out with every batch where a soil of known value is determined (every batch of 20 samples or part thereof). Blank determinations are also carried out routinely where required.

Total Inorganic Carbon (TIC) is determined by calculation: $TC - TOC = TIC$

TOC was analysed using an Eltra combustion method. This method is used for total carbon analysis of dried, crushed rock powder and environmental soil samples. The samples are previously treated with 10% HCl to remove inorganic carbon (Carbonates) before washing to remove residual acids and further dried. The Carbon Analyser heats the sample in a flow of oxygen and any carbon present is converted to carbon dioxide which is measured by infra-red absorption. The percentage carbon is then calculated with respect to the original sample weight. The range for the method is 0.01 - 100%. The method is current being evaluated under the UKAS accreditation scheme.

II.3 HYDROCARBON CONCENTRATIONS (TOTAL HYDROCARBON CONCENTRATIONS AND ALIPHATICS)

II.3.1 General Precautions

High purity solvents were used throughout the analyses. Solvent purity was assessed by evaporating an appropriate volume to 1ml and analysing the concentrate by GC for general hydrocarbons, target n-alkanes and aromatics. All glassware and extraction sundries were cleaned prior to use by thorough rinsing with hydrocarbon-free deionised water followed by two rinses with dichloromethane. All glassware was heated in a high temperature oven at 450°C for 6 hours.

II.3.2 Extraction Procedure for Hydrocarbons

Each analytical sample (15±0.1g) was spiked with an internal standard solution containing the following components: aliphatics - heptamethylnonane, 1-chlorooctadecane and squalane. The sample was then wet vortex extracted using three successive aliquots of DCM/Methanol. The extracts were combined and water partitioned to remove the methanol and any excess water from the sample.

Solvent extracts were chemically dried and then reduced to approximately 1ml using a Kuderna Danish evaporator with micro Snyder.

II.3.3 Column fractionation for Aliphatic and Aromatic Fractions

The concentrated extract was transferred to a pre-conditioned flash chromatography column containing approximately 1g of activated Silica gel. The compounds were eluted with 3ml of Pentane/DCM (2:1). An aliquot of the extract was then taken and analysed for total hydrocarbon (THC) content and individual n-alkanes by large volume injection GC-FID.

II.3.4 Quality Control Samples

The following quality control samples were prepared with the batches of sediment samples:

- A method blank comprising 15±0.1g of baked anhydrous sodium sulphate (organic free) treated as a sample.
- A matrix matched standard sample consisting of 15±0.1g baked sand spiked with Florida mix and treated as sample.
- A sample duplicate - any one sample from the batch, dependent upon available sample mass, analysed in duplicate.

II.3.5 Hydrocarbon Analysis

Analysis of total hydrocarbons and aliphatics was performed by using an Agilent 6890 with an FID detector. Appropriate column and GC conditions were used to provide sufficient chromatographic separation of all analytes and the required sensitivity.

II.3.5.1 Carbon Preference Index

The carbon preference index is calculated as follows:

$$CPI = \frac{\text{odd homologues (nC}_{11}\text{ to nC}_{35})}{\text{even homologues (nC}_{10}\text{ to nC}_{34})}$$

II.3.5.2 Petrogenic/Biogenic or (P/B) Ratio

The Petrogenic/Biogenic Ratio is calculated as follows:

$$P/B \text{ Ratio} = \frac{P = \text{sum of nC}_{10}\text{ to nC}_{20}}{B = \text{sum of nC}_{21}\text{ to nC}_{35}}$$

II.3.6 Calibration and Calculation

GC techniques require the use of internal standards in order to obtain quantitative results. The technique requires addition of non-naturally occurring compounds to the sample, allowing correction for varying recovery.

Target analytes concentrations were calculated by comparison with the nearest eluting internal standards. A relative response factor was applied to correct the data for the differing responses of target analytes and internal standards. Response factors were established prior to running samples, from solutions containing USEPA(16) PAHs + Dibenzothiophene for the GCMS, Florida mix (even n-Alkanes nC10-nC40) for individual GCFID targets and a Diesel/Mineral Oil mix for total oil determination.

The mean detection limits used for the sediment total hydrocarbons and n-alkanes were:

1. n-alkane - 1ng.g^{-1} (ppb)
2. Total Hydrocarbons - 100ng.g^{-1} (ppb)

II.4 HEAVY AND TRACE METAL CONCENTRATIONS

Sediment samples were homogenised and a 50g portion of each sample was air dried at room temperature. Each sample was then ground down to a fine powder ($<100\mu\text{m}$) by hand using a metal free mortar and pestle. A clean sand sample was hand ground prior to preparation of the field samples as a blank.

II.4.1 Sample Digestion Procedure

Total Metals by ICPOES (Hydrofluoric /Boric acid Extractable Metals - Fe, Ba, Sr & Al)

Approximately 0.20g of the sediment sample was accurately weighed and placed in a PTFE bottle and 2.5mls of hydrofluoric acid was added. The bottle was then placed in an oven at $105\pm 5^\circ\text{C}$ for approximately 30 minutes and then allowed to air cool in a fume cupboard. A further 65mls of 4% boric acid was then added to the bottle and the contents were then mixed thoroughly and placed in a polypropylene flask. The solution was then made up to 100ml with deionised water and analysed by ICP-OES.

Total Metals by ICPMS (Hydrofluoric /Nitric acid Extractable Metals - Cr, Cu, Ni, Zn, As, Pb, Sn, V & Cd).

Approximately 0.10g of the sediment sample was accurately weighed and placed in a PTFE bottle. Approximately 1ml of hydrofluoric acid, 1ml of nitric acid and 1ml of water were added and the bottle placed in an oven at $105\pm 5^\circ\text{C}$ for approximately 60 minutes. The bottle was then allowed to air cool in a fume cupboard. The extract was transferred to a plastic beaker and evaporated to dryness. The residue was then cooled and dissolved in 2ml of nitric acid. This was transferred to a 100ml volumetric flask and made up to volume with deionised water. The metals concentrations in the extract were determined by ICP-MS.

The mean detection limits are given in Table E for acid leachable (AL) and hydrofluoric acid (HF) digestions.

Table E - Heavy Metals - Mean Detection Limits (MDL)

| Analyte | Unit | MDL |
|---------|---------------------------------|------|
| Ni | $\mu\text{g}\cdot\text{g}^{-1}$ | 5 |
| V | $\mu\text{g}\cdot\text{g}^{-1}$ | 5 |
| Al | $\mu\text{g}\cdot\text{g}^{-1}$ | 10 |
| Zn | $\mu\text{g}\cdot\text{g}^{-1}$ | 5 |
| Fe | $\mu\text{g}\cdot\text{g}^{-1}$ | 10 |
| Cu | $\mu\text{g}\cdot\text{g}^{-1}$ | 5 |
| Ba | $\mu\text{g}\cdot\text{g}^{-1}$ | 5 |
| Cr | $\mu\text{g}\cdot\text{g}^{-1}$ | 5 |
| As | $\mu\text{g}\cdot\text{g}^{-1}$ | 1 |
| Cd | $\mu\text{g}\cdot\text{g}^{-1}$ | 1 |
| Pb | $\mu\text{g}\cdot\text{g}^{-1}$ | 1 |
| Sn | $\mu\text{g}\cdot\text{g}^{-1}$ | 0.5 |
| Hg | $\mu\text{g}\cdot\text{g}^{-1}$ | 0.01 |

| | | |
|-------|--------|------|
| ICPMS | ICPOES | TMMS |
|-------|--------|------|

II.4.2. Mercury Digestion Procedure

Approximately 1g of the sediment was accurately weighed and transferred to a beaker. Hydrogen peroxide (10ml of 30 volumes) was added, and the covered sample left to digest for 0.5 hour in the fume cupboard. 10ml of nitric acid was added and the sample placed on the hotplate for 1 hour.

After digestion, the sample was filtered through a Whatman 542 filter paper into a 100ml standard flask. The watch-glass and beaker were rinsed thoroughly, transferring the washings to the filter paper. The filter paper was rinsed until the volume was approximately 90ml. Subsequently, the filter funnel was rinsed into the flask and then the flask was made up to 100ml volume and mixed well. The filtrate was then analysed by ICP-MS.

II.4.3 Analytical Methodology

Inductively Coupled Plasma Optical Emission Spectrometry

The instrument is calibrated using dilutions of the 1ml (=10mg) spectroscopic solutions. The final calibration solutions are matrix matched with the relevant acids. The calibration line consists of five standards.

Inductively Coupled Plasma- Mass Spectrometry

The instrument is calibrated using dilutions of the 1ml (=10mg) spectroscopic solutions. The calibration line consists of seven standards.

The analytes are scaled against internal standards to take account of changes in plasma conditions as a result of matrix differences for standards and samples. The internal standards have a similar mass and ionisation properties to the target metals.

II.5 MACRO-INVERTEBRATE ANALYSIS

II.5.1 Methodology

All macrofaunal determination was carried by MG3 subcontractor BSL using a specialist taxonomist. This senior taxonomist has a wealth of experience in macrofaunal identification in temperate deep water environments (such as Ireland, Scotland, Faroes and sub-Antarctic waters).

Benthic sediment samples were thoroughly washed with freshwater on a 500µm sieve to remove traces of formalin, placed in gridded, white trays and then hand sorted by eye followed by binocular microscope, to remove all fauna. Sorted organisms were preserved in 70% IMS and 5% glycerol. Where possible, all organisms were identified to species level according to appropriate keys for the region. Colonial and encrusting organisms were recorded by presence alone and, where colonies could be identified as a single example, these were also recorded, although these datasets have not been considered in the overall statistical analysis of the material. The presence of anthropogenic components was also recorded where relevant.

All taxa were distinguished to species level and identified to at least family level where possible, although information is limited for the area, many of the species that could not be fully identified were separated putatively. Whilst some of the groups were only partially separated in this document, ongoing analysis with further site-specific well sites will increase our knowledge of the area and a more definitive faunal matrix will be provided at a later point in time. Nomenclature for species names were allocated either when identity was confirmed, allocated as "cf." when apparently identifying to a known species but confirmation was not possible (for example, incomplete specimens or descriptions), or allocated as "aff." when close to but distinct from a described species. The terms "indet." refers to being unable to identify to a lower taxon and "juv" as a juvenile to that species, genus or family. Species lists for the twenty six stations (56 samples (typically 2 replicates per station)), together with univariate parameters for both sample replicates and stations, are given within Section 2.9.2 and Appendix VII.

II.5.2 Quality Assurance

Benthic Solutions is committed to total quality control from the start of a project to its completion. All samples taken or received by the company were given a unique identification number. All analytical methods were carried out according to recognised standards for marine analyses. All taxonomic staff are fully qualified to post-doctorate level. Documentation is maintained that indicates the stage of analysis that each sample has reached. A full reference collection of all specimens has been retained for further clarification of putative species groups where/if required. BSL is a participant in the National Marine Biological Association Quality Control (NMBAQC) quality assurance scheme.

Furthermore, as requested by the Noble Energy Falklands, a reconciliation process has been applied on the macrofaunal species collected by verifying identified species with specimens curated at the NHM in London. This was carried out at the NHM on 13th to 15th August 2014, with taxonomists from BSL and NHM participating and RPS overseeing the reconciliation process. The results of this

process confirmed that the standard of taxonomy undertaken for the FISA site was very high. A statement from the NHM confirmed that 'both teams (BSL and NHM) were working to the same standards and arriving at similar identification levels (i.e. the same species were being recognised by both team). There was also a useful exchange of experience and literature used in the identification process. For the most part, identifications were similar and so there is comparability between the different studies in that the diversity of the fauna is being captured' (email from Gordon Patterson to RPS 9th Sept 2014).

Where previous putative species have previously been assigned to species, the nomenclature for this has been included in the faunal matrix for future continuity and to aid curation of the reference material. One consistent observation made by the NHM taxonomists during the workshop was of the excellent condition of the material available. This is anticipated to be due to a combination of the following:

- Experienced and well-trained staff in the field;
- Use of the Wilson Auto-siever during field processing;
- Control and buffering of preservation fluids; and
- Sample sorting techniques by very experienced personnel.

Further analytical quality control on the sorting efficiency of macrofaunal samples was carried out by Aquatic Environments with a certificate provided in Appendix VIII.

Digital datasets are kept for all sites in the form of excel spreadsheets (by sample and by station) on MG3 archive and within the BSL's archive. This system is duplicated onto a second archive drive in case of electronic failure. These datasets will be stored in this way for a minimum of three years, or transferred to storage disk (data CD or DVD).

II.5.3 Data Standardisation and Analyses

In accordance to OSPAR Commission (2004) guidelines, all species falling into juvenile, colonial, planktonic or meiofaunal taxa are excluded from the full analyses within the dataset (this is discussed further within the text of Section 2.9). This helps to reduce the variability of data undertaken during different periods within the year, or where minor changes may occur or where some groups may only be included in a non-quantitative fashion, such as presence/absence. Certain taxa, such as the Nematoda, normally associated with meiofauna, were included where individuals greater than 10mm were recorded. The following primary and univariate parameters were calculated for each all data by stations and sample (Table F).

Table F - Primary and Univariate Parameter Calculations

| Variable | Parameter | Formula | Description |
|-----------------------|-----------------|---|--|
| Total Species | S | Number of species recorded | Species richness |
| Total Individuals | N | Number of individuals recorded | Sample abundance |
| Shannon-Wiener Index | H(s) | $H(s) = -\sum_{i=1}^s (P_i) (\log_2 P_i)$ where s = number of species & P _i = proportion of total sample belonging to i th species. | Diversity: using both richness and equitability, recorded in log 2. |
| Simpsons Diversity | 1-Lambda | $\text{Lambda} = \sum \left(\frac{n_i(n_i-1)}{N(N-1)} \right)$ where n _i = number of individuals in the i th species & N = total number of individuals | Evenness, related to dominance of most common species (Simpson, 1949) |
| Pielou's Equitability | J | $J = \frac{H(s)}{(\log S)}$ where s = number of species & H(s) = Shannon-Wiener diversity index. | Evenness or distribution between species (Pielou, 1969) |
| Margalefs Richness | D _{Mg} | $D_{Mg} = \frac{(S-1)}{(\log N)}$ where s = number of species & N = number of individuals. | Richness derived from number of species and total number of individuals (Clifford and Stevenson, 1975) |

In addition to univariate methods of analysis, data for both sample replicates and stations were analysed using multivariate techniques. These serve to reduce complex species-site data to a form that is visually interpretable. A multivariate analyse was based on transformed data (double square root) to detect any improved relationships when effects of dominance were reduced. The basis for multivariate analyses was based upon the software PRIMER (Plymouth Routines In Multivariate Ecological Research).

Similarity Matrices and Hierarchical Agglomerative Clustering

A similarity matrix is used to compare every individual sample replicate and/or stations with each other. The coefficient used in this process is based upon Bray Curtis (Bray and Curtis, 1957), considered to be the most suitable for community data. These are subsequently assigned into groups of replicates and/or stations according to their level of similarity and clustered together based upon a Group Average Method into a dendrogram of similarity.

Non-Metric Multidimensional Scaling (nMDS): nMDS is currently widely used in the analysis of spatial and temporal change in benthic communities (e.g. Warwick and Clarke, 1991). The recorded observations from data were exposed to computation of triangular matrices of similarities between all pairs of samples. The similarity of every pair of sites was computed using the Bray-Curtis index on transformed data. Clustering was by a hierarchical agglomerative method using group average sorting, and the results are presented as a dendrogram and as a two-dimensional ordination plot. The degree of distortion involved in producing an ordination gives an indication of the adequacy of the nMDS representation and is recorded as a stress value as outline in Table G.

Table G - Inference from nMDS Stress Values

| nMDS Stress | Adequacy of Representation for Two-Dimensional Plot |
|--------------|--|
| ≤0.05 | Excellent representation with no prospect of misinterpretation. |
| >0.05 to 0.1 | Good ordination with no real prospect of a misleading interpretation. |
| >0.1 to 0.2 | Potentially useful 2-d plot, though for values at the upper end of this range too much reliance should not be placed on plot detail; superimposition of clusters should be undertaken to verify conclusions. |
| >0.2 to 0.3 | Ordination should be treated with scepticism. Clusters may be superimposed to verify conclusions, but ordinations with stress values >2.5 should be discarded. A 3-d ordination may be more appropriate. |
| >0.3 | Ordination is unreliable with points close to being arbitrarily placed in the 2-d plot. A 3-d ordination should be examined. |

SIMPER: the nMDS clustering program is used to analyse differences between sites. SIMPER enables those species responsible for differences to be identified by examining the contribution of individual species to the similarity measure. As all sites grouped within a single cluster, this program was subsequently not used.

Bioaccumulation Curve Estimates are undertaken using **Chao-1 (S^*_1)**. This is a formula that estimates how many additional species would be needed to sample all of the asymptotic species richness of a region, based on the samples acquired. It calculates this by comparing the number of species that occur in one sample with those that occur in two samples where;

$$S^*_1 = S_{obs} + (a^2/2b)$$

S_{obs} is the number of species observed

a is the number of species observed just once

b is the number of species observed just twice

RELATE – Is non-parametric Mantel test that looks at the relationship between 2 matrices (often biotic and environmental). This shows the degree of seriation, an alternative to cluster analysis, which looks for a sequential pattern in community change. The test computes Spearman's rank correlation coefficient (R) between the corresponding elements of each pair of matrices to produce a correlation statistic present between the two datasets, the significance of the correlation determined by a permutation procedure (Clarke and Gorley, 2006).

ANOSIM (Analysis of Similarity) – Non parametric, multivariate test often used in community ecology that calculates Bray-Curtis coefficient (for biological data) or Euclidean distance (for environmental data) based on permutations of ranked data. It produces an R value which is an effect level on a scale of 0-1, $R=1$ where all differences between sites are greater than any differences within site, $R=0$ when there is no separation between groups. P value (<5%) is the likelihood of arriving at that R value by chance, this significance value is determined by a permutation procedure (Clarke and Gorley, 2006).

SIMPROF (similarity profile) test - analyses data for significant clusters that show evidence of a multivariate pattern in data that are *a priori* unstructured, i.e. single samples from each site, this differs from the ANOSIM tests which permutes data based on a grouping factor such as 'site' or 'year'. The test works by comparing samples which have been ranked and ordered by resemblance against an expected profile which is obtained by permuting random species (variables) across the set of samples, a mean of 1000 permutations is taken to produce an expected result for null structure with rare and common species displaying the same pattern. If the actual data deviates outside the 95% limits of the expected profile then there is evidence for significant structure and vice versa. The 'significant structure' is well represented on a dendrogram which will also show the clusters containing that lack significant differentiation (null structure), (Clarke and Gorley, 2006).

II.6 WATER QUALITY

Water profiles were collected using a Valeport CTD continuous reading water quality profiler depth rated to 3000m. This was fitted with sensors to record measurements throughout the water column from sea surface down to the seabed. The probe was pre-programmed and set to recording mode and lowered at an approximate rate of ca. 65m/min. This was a sufficient time to allow the onboard sensors (conductivity, temperature and pressure) to equilibrate to ambient conditions during the cast. The sampling frequency of the instrument was set to 1 reading per second. Data was recorded during both down and up casts. A total of nine data casts were acquired throughout the FISA survey area.

Profile sensors and derived parameters are as follows:

- Conductivity: mS/cm
- Temperature: °C
- Depth: Metres
- Density: g/cm³ calculated
- Sound Velocity: m/sec calculated
- Salinity: ppt calculated

To accompany these profiles, water samples were recovered using a Niskin bottle at four depth strata. Samples were collected at the surface, thermocline, mid-depth and at the seabed. Due to the 'blue water' nature of the survey area, spatial variation of water samples was deemed to be negligible. Therefore two full sets of water samples were acquired from the approximate centre of the survey area with temporal separation of approximately 10 days.

II.7 BIBLIOGRAPHY

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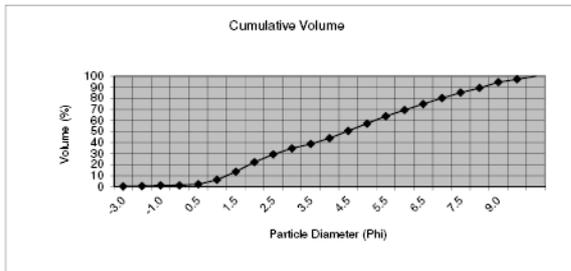
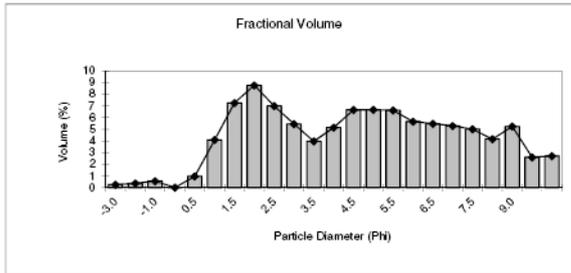
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APPENDIX III - PARTICLE SIZE DISTRIBUTION

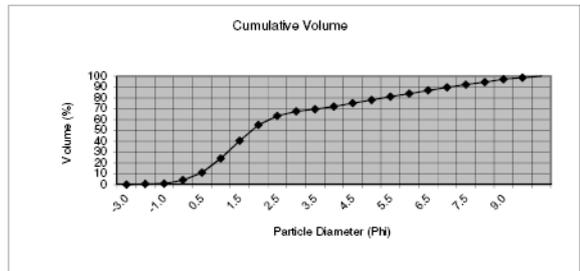
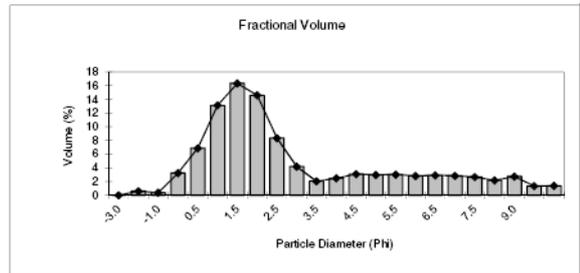
Sample No.: A_3_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 12:37



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.27 | 0.3 | Pebble |
| 4.000 | -2.0 | 0.37 | 0.6 | |
| 2.000 | -1.0 | 0.57 | 1.2 | Granule |
| 1.000 | 0.0 | 0.02 | 1.2 | V.Coarse Sand |
| 0.710 | 0.5 | 0.98 | 2.2 | |
| 0.500 | 1.0 | 4.09 | 6.3 | Coarse Sand |
| 0.355 | 1.5 | 7.26 | 13.5 | |
| 0.250 | 2.0 | 8.76 | 22.3 | Medium Sand |
| 0.180 | 2.5 | 7.00 | 29.3 | |
| 0.125 | 3.0 | 5.45 | 34.8 | Fine Sand |
| 0.900 | 3.5 | 3.97 | 38.7 | |
| 0.063 | 4.0 | 5.16 | 43.9 | V.Fine Sand |
| 0.044 | 4.5 | 6.66 | 50.6 | |
| 0.032 | 5.0 | 6.68 | 57.2 | Coarse Silt |
| 0.022 | 5.5 | 6.62 | 63.8 | |
| 0.016 | 6.0 | 5.66 | 69.5 | Medium Silt |
| 0.011 | 6.5 | 5.48 | 75.0 | |
| 0.008 | 7.0 | 5.29 | 80.3 | Fine silt |
| 0.006 | 7.5 | 5.01 | 85.3 | |
| 0.004 | 8.0 | 4.16 | 89.4 | V.Fine Silt |
| 0.002 | 9.0 | 5.24 | 94.7 | |
| 0.001 | 10.0 | 2.61 | 97.3 | Coarse Clay |
| <0.001 | >10.0 | 2.71 | 100.0 | Medium Clay |
| | | | | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.045 | 0.216 | 4.48 |
| Median | 0.046 | | 4.46 |
| Sorting Coefficient | Value: 2.68 | Inference: Very Poorly Sorted | |
| Skewness | 0.06 | Symmetrical | |
| Kurtosis | 0.78 | Platykurtic | |
| % Fines | 56.11% | Coarse Silt | |
| % Sands | 42.69% | | |
| % Gravel | 1.20% | | |

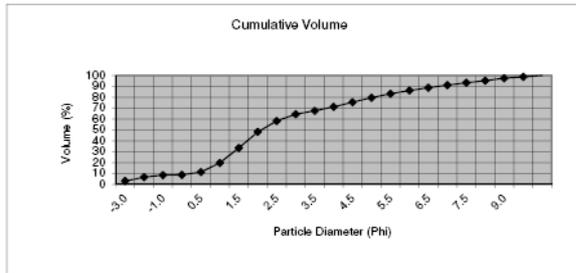
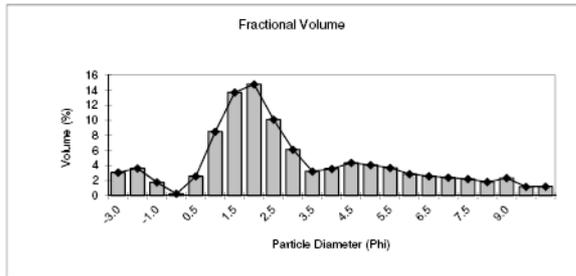
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| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.58 | 0.6 | |
| 2.000 | -1.0 | 0.36 | 0.9 | Granule |
| 1.000 | 0.0 | 3.24 | 4.2 | V.Coarse Sand |
| 0.710 | 0.5 | 6.86 | 11.0 | |
| 0.500 | 1.0 | 16.32 | 24.2 | Coarse Sand |
| 0.355 | 1.5 | 16.33 | 40.5 | |
| 0.250 | 2.0 | 14.60 | 55.1 | Medium Sand |
| 0.180 | 2.5 | 8.35 | 63.4 | |
| 0.125 | 3.0 | 4.18 | 67.6 | Fine Sand |
| 0.900 | 3.5 | 2.04 | 69.7 | |
| 0.063 | 4.0 | 2.47 | 72.1 | V.Fine Sand |
| 0.044 | 4.5 | 3.08 | 75.2 | |
| 0.032 | 5.0 | 2.98 | 78.2 | Coarse Silt |
| 0.022 | 5.5 | 3.02 | 81.2 | |
| 0.016 | 6.0 | 2.81 | 84.0 | Medium Silt |
| 0.011 | 6.5 | 2.90 | 86.9 | |
| 0.008 | 7.0 | 2.84 | 89.8 | Fine silt |
| 0.006 | 7.5 | 2.65 | 92.4 | |
| 0.004 | 8.0 | 2.17 | 94.6 | V.Fine Silt |
| 0.002 | 9.0 | 2.73 | 97.3 | |
| 0.001 | 10.0 | 1.34 | 98.6 | Coarse Clay |
| <0.001 | >10.0 | 1.36 | 100.0 | Medium Clay |
| | | | | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.141 | 0.369 | 2.82 |
| Median | 0.287 | | 1.80 |
| Sorting Coefficient | Value: 2.56 | Inference: Very Poorly Sorted | |
| Skewness | 0.57 | Very Positive (Coarse) | |
| Kurtosis | 0.96 | Mesokurtic | |
| % Fines | 27.87% | Fine Sand | |
| % Sands | 71.18% | | |
| % Gravel | 0.95% | | |

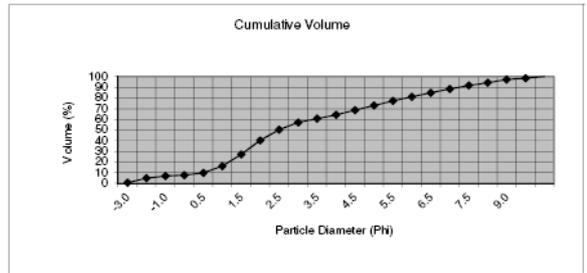
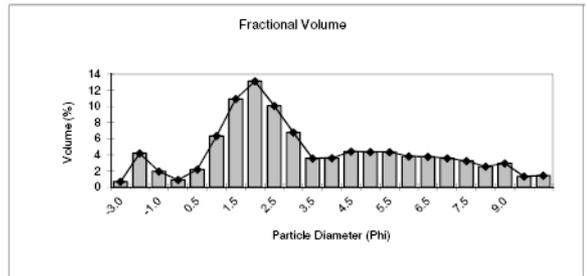
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| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 3.08 | 3.1 | Pebble |
| 4.000 | -2.0 | 3.63 | 6.7 | |
| 2.000 | -1.0 | 1.78 | 8.5 | Granule |
| 1.000 | 0.0 | 0.25 | 8.7 | V.Coarse Sand |
| 0.710 | 0.5 | 2.56 | 11.3 | |
| 0.500 | 1.0 | 8.49 | 19.8 | Coarse Sand |
| 0.355 | 1.5 | 13.69 | 33.5 | |
| 0.250 | 2.0 | 14.78 | 48.3 | Medium Sand |
| 0.180 | 2.5 | 10.10 | 58.4 | |
| 0.125 | 3.0 | 6.11 | 64.5 | Fine Sand |
| 0.900 | 3.5 | 3.22 | 67.7 | |
| 0.063 | 4.0 | 3.57 | 71.3 | V.Fine Sand |
| 0.044 | 4.5 | 4.35 | 75.6 | |
| 0.032 | 5.0 | 4.07 | 79.7 | Coarse Silt |
| 0.022 | 5.5 | 3.69 | 83.4 | |
| 0.016 | 6.0 | 2.88 | 86.3 | Medium Silt |
| 0.011 | 6.5 | 2.59 | 88.9 | |
| 0.008 | 7.0 | 2.39 | 91.2 | Fine silt |
| 0.006 | 7.5 | 2.21 | 93.5 | |
| 0.004 | 8.0 | 1.82 | 95.3 | V.Fine Silt |
| 0.002 | 9.0 | 2.34 | 97.6 | |
| 0.001 | 10.0 | 1.18 | 98.8 | Coarse Clay |
| <0.001 | >10.0 | 1.21 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|--------------------|------|
| Mean (MZ) | 0.143 | 1.894 | 2.81 |
| Median | 0.238 | | 2.07 |
| Sorting Coefficient | Value | Inference | |
| | 2.80 | Very Poorly Sorted | |
| Skewness | 0.29 | Positive(Coarse) | |
| Kurtosis | 1.32 | Leptokurtic | |
| % Fines | 28.73% | Fine Sand | |
| % Sands | 62.78% | | |
| % Gravel | 8.49% | | |

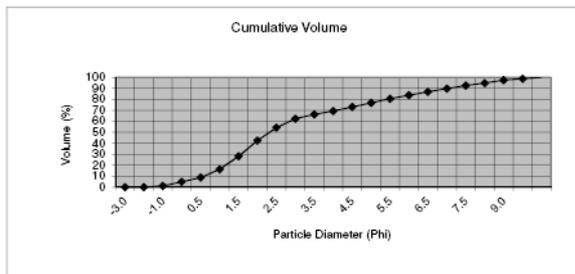
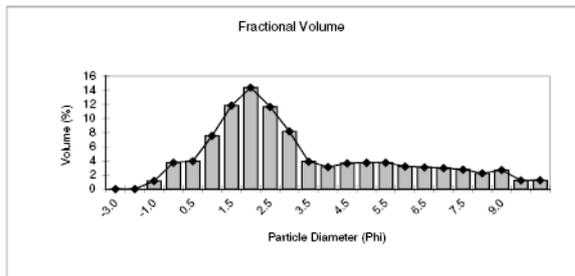
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| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.67 | 0.7 | Pebble |
| 4.000 | -2.0 | 4.18 | 4.8 | |
| 2.000 | -1.0 | 1.95 | 6.8 | Granule |
| 1.000 | 0.0 | 0.89 | 7.7 | V.Coarse Sand |
| 0.710 | 0.5 | 2.19 | 9.9 | |
| 0.500 | 1.0 | 6.34 | 16.2 | Coarse Sand |
| 0.355 | 1.5 | 10.91 | 27.1 | |
| 0.250 | 2.0 | 13.11 | 40.2 | Medium Sand |
| 0.180 | 2.5 | 10.07 | 50.3 | |
| 0.125 | 3.0 | 6.77 | 57.1 | Fine Sand |
| 0.900 | 3.5 | 3.56 | 60.6 | |
| 0.063 | 4.0 | 3.61 | 64.2 | V.Fine Sand |
| 0.044 | 4.5 | 4.41 | 68.6 | |
| 0.032 | 5.0 | 4.37 | 73.0 | Coarse Silt |
| 0.022 | 5.5 | 4.34 | 77.4 | |
| 0.016 | 6.0 | 3.81 | 81.2 | Medium Silt |
| 0.011 | 6.5 | 3.76 | 84.9 | |
| 0.008 | 7.0 | 3.58 | 88.5 | Fine silt |
| 0.006 | 7.5 | 3.24 | 91.7 | |
| 0.004 | 8.0 | 2.55 | 94.3 | V.Fine Silt |
| 0.002 | 9.0 | 2.96 | 97.2 | |
| 0.001 | 10.0 | 1.32 | 98.6 | Coarse Clay |
| <0.001 | >10.0 | 1.43 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|--------------------|------|
| Mean (MZ) | 0.104 | 1.232 | 3.27 |
| Median | 0.182 | | 2.46 |
| Sorting Coefficient | Value | Inference | |
| | 2.88 | Very Poorly Sorted | |
| Skewness | 0.29 | Positive(Coarse) | |
| Kurtosis | 1.09 | Mesokurtic | |
| % Fines | 35.77% | V.Fine Sands | |
| % Sands | 57.44% | | |
| % Gravel | 6.80% | | |

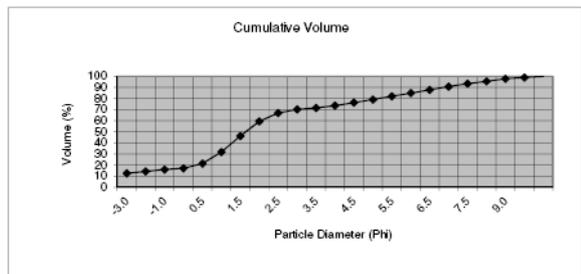
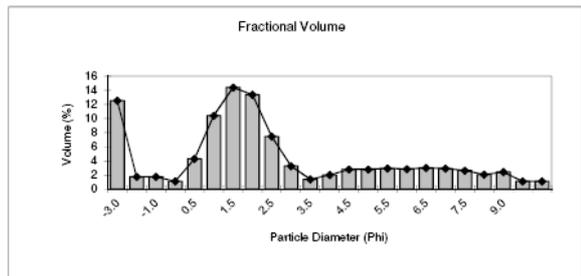
Sample No.: A_1011_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 12:24



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.00 | 0.0 | |
| 2.000 | -1.0 | 1.16 | 1.2 | Granule |
| 1.000 | 0.0 | 3.74 | 4.9 | V.Coarse Sand |
| 0.710 | 0.5 | 3.95 | 8.9 | |
| 0.500 | 1.0 | 7.54 | 16.4 | Coarse Sand |
| 0.355 | 1.5 | 11.82 | 28.2 | |
| 0.250 | 2.0 | 14.36 | 42.6 | Medium Sand |
| 0.180 | 2.5 | 11.64 | 54.2 | |
| 0.125 | 3.0 | 8.17 | 62.4 | Fine Sand |
| 0.900 | 3.5 | 3.90 | 66.3 | |
| 0.063 | 4.0 | 3.13 | 69.4 | V.Fine Sand |
| 0.044 | 4.5 | 3.65 | 73.1 | |
| 0.032 | 5.0 | 3.74 | 76.8 | Coarse Silt |
| 0.022 | 5.5 | 3.75 | 80.6 | |
| 0.016 | 6.0 | 3.20 | 83.8 | Medium Silt |
| 0.011 | 6.5 | 3.10 | 86.9 | |
| 0.008 | 7.0 | 2.97 | 89.8 | Fine silt |
| 0.006 | 7.5 | 2.76 | 92.6 | |
| 0.004 | 8.0 | 2.24 | 94.8 | V.Fine Silt |
| 0.002 | 9.0 | 2.69 | 97.5 | Coarse Clay |
| 0.001 | 10.0 | 1.22 | 98.8 | Medium Clay |
| <0.001 | >10.0 | 1.25 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.117 | 0.355 | 3.10 |
| Median | 0.205 | | 2.28 |
| Sorting Coefficient | Value: 2.48 | Inference: Very Poorly Sorted | |
| Skewness | 0.46 | Very Positive (Coarse) | |
| Kurtosis | 0.97 | Mesokurtic | |
| % Fines | 30.57% | V.Fine Sands | |
| % Sands | 68.27% | | |
| % Gravel | 1.16% | | |

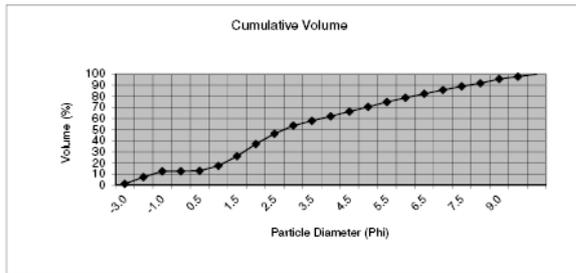
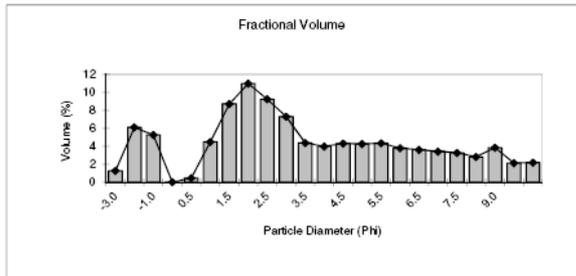
Sample No.: A_12_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 11:27



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 12.52 | 12.5 | Pebble |
| 4.000 | -2.0 | 1.73 | 14.2 | |
| 2.000 | -1.0 | 1.73 | 16.0 | Granule |
| 1.000 | 0.0 | 1.09 | 17.1 | V.Coarse Sand |
| 0.710 | 0.5 | 4.28 | 21.4 | |
| 0.500 | 1.0 | 10.40 | 31.8 | Coarse Sand |
| 0.355 | 1.5 | 14.37 | 46.1 | |
| 0.250 | 2.0 | 13.34 | 59.5 | Medium Sand |
| 0.180 | 2.5 | 7.43 | 66.9 | |
| 0.125 | 3.0 | 3.25 | 70.2 | Fine Sand |
| 0.900 | 3.5 | 1.37 | 71.5 | |
| 0.063 | 4.0 | 2.00 | 73.5 | V.Fine Sand |
| 0.044 | 4.5 | 2.78 | 76.3 | |
| 0.032 | 5.0 | 2.78 | 79.1 | Coarse Silt |
| 0.022 | 5.5 | 2.92 | 82.0 | |
| 0.016 | 6.0 | 2.83 | 84.9 | Medium Silt |
| 0.011 | 6.5 | 3.00 | 87.8 | |
| 0.008 | 7.0 | 2.91 | 90.8 | Fine silt |
| 0.006 | 7.5 | 2.62 | 93.4 | |
| 0.004 | 8.0 | 2.05 | 95.4 | V.Fine Silt |
| 0.002 | 9.0 | 2.41 | 97.8 | Coarse Clay |
| 0.001 | 10.0 | 1.08 | 98.9 | Medium Clay |
| <0.001 | >10.0 | 1.08 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.224 | 8.943 | 2.16 |
| Median | 0.325 | | 1.62 |
| Sorting Coefficient | Value: 3.61 | Inference: Very Poorly Sorted | |
| Skewness | 0.12 | Positive (Coarse) | |
| Kurtosis | 1.43 | Leptokurtic | |
| % Fines | 26.47% | Fine Sand | |
| % Sands | 57.55% | | |
| % Gravel | 15.98% | | |

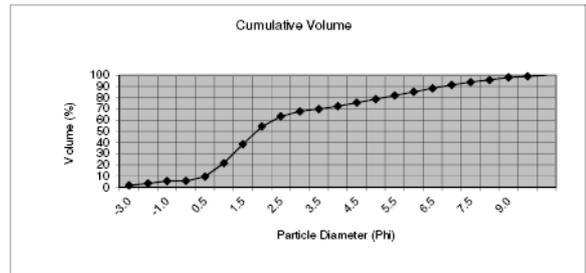
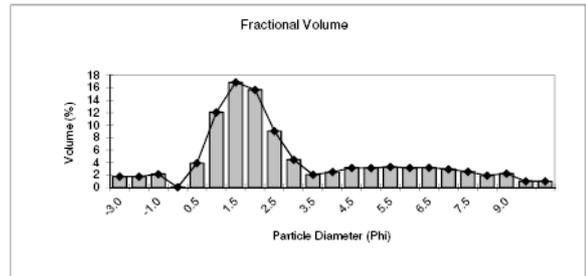
Sample No.: A_1013_ENV Operator PaulSil
Source Data: Nobel Energy Date&Time: 29/05/2014 13:43



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 1.27 | 1.3 | Pebble |
| 4.000 | -2.0 | 6.10 | 7.4 | |
| 2.000 | -1.0 | 5.25 | 12.6 | Granule |
| 1.000 | 0.0 | 0.00 | 12.6 | V.Coarse Sand |
| 0.710 | 0.5 | 0.46 | 13.1 | |
| 0.500 | 1.0 | 4.47 | 17.5 | Coarse Sand |
| 0.355 | 1.5 | 8.69 | 26.2 | |
| 0.250 | 2.0 | 10.97 | 37.2 | Medium Sand |
| 0.180 | 2.5 | 9.23 | 46.4 | |
| 0.125 | 3.0 | 7.29 | 53.7 | Fine Sand |
| 0.900 | 3.5 | 4.37 | 58.1 | |
| 0.063 | 4.0 | 3.98 | 62.1 | V.Fine Sand |
| 0.044 | 4.5 | 4.31 | 66.4 | |
| 0.032 | 5.0 | 4.25 | 70.6 | Coarse Silt |
| 0.022 | 5.5 | 4.34 | 75.0 | |
| 0.016 | 6.0 | 3.78 | 78.8 | Medium Silt |
| 0.011 | 6.5 | 3.61 | 82.4 | |
| 0.008 | 7.0 | 3.42 | 85.8 | Fine silt |
| 0.006 | 7.5 | 3.26 | 89.0 | |
| 0.004 | 8.0 | 2.81 | 91.9 | V.Fine Silt |
| 0.002 | 9.0 | 3.84 | 95.7 | Coarse Clay |
| 0.001 | 10.0 | 2.12 | 97.8 | Medium Clay |
| <0.001 | >10.0 | 2.18 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|-------------|--------|--------------------|------|
| Mean (MZ) | 0.094 | 1.933 | 3.41 |
| Median | 0.153 | | 2.71 |
| Sorting | Value | Inference | |
| Coefficient | 3.18 | Very Poorly Sorted | |
| Skewness | 0.22 | Positive(Coarse) | |
| Kurtosis | 1.12 | Leptokurtic | |
| % Fines | 37.93% | v.Fine Sands | |
| % Sands | 49.45% | | |
| % Gravel | 12.62% | | |

Sample No.: A_14_ENV Operator PaulSil
Source Data: Nobel Energy Date&Time: 29/05/2014 14:31

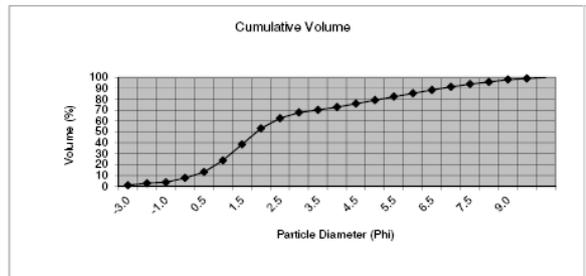
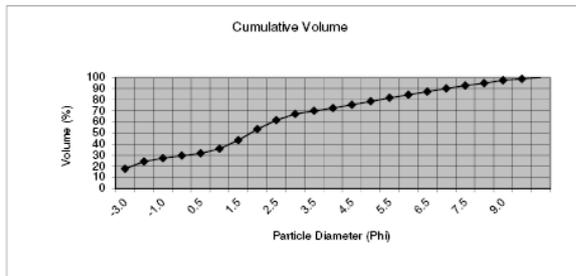
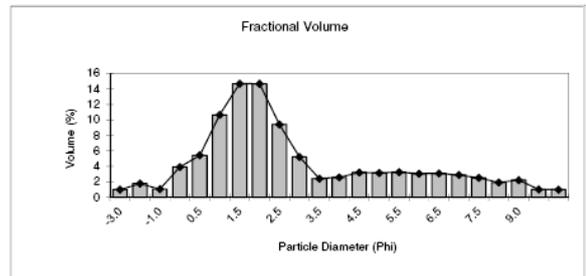
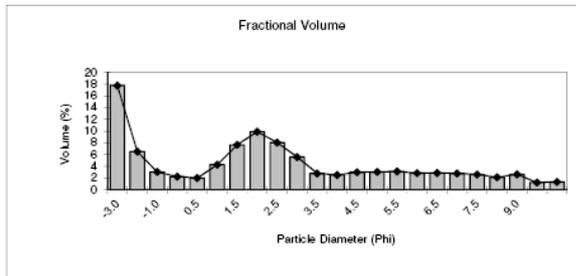


| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 1.76 | 1.8 | Pebble |
| 4.000 | -2.0 | 1.76 | 3.5 | |
| 2.000 | -1.0 | 2.14 | 5.7 | Granule |
| 1.000 | 0.0 | 0.04 | 5.7 | V.Coarse Sand |
| 0.710 | 0.5 | 3.92 | 9.6 | |
| 0.500 | 1.0 | 12.08 | 21.7 | Coarse Sand |
| 0.355 | 1.5 | 16.88 | 38.6 | |
| 0.250 | 2.0 | 15.69 | 54.3 | Medium Sand |
| 0.180 | 2.5 | 9.07 | 63.3 | |
| 0.125 | 3.0 | 4.46 | 67.8 | Fine Sand |
| 0.900 | 3.5 | 2.06 | 69.9 | |
| 0.063 | 4.0 | 2.49 | 72.4 | V.Fine Sand |
| 0.044 | 4.5 | 3.18 | 75.5 | |
| 0.032 | 5.0 | 3.13 | 78.7 | Coarse Silt |
| 0.022 | 5.5 | 3.30 | 82.0 | |
| 0.016 | 6.0 | 3.16 | 85.1 | Medium Silt |
| 0.011 | 6.5 | 3.20 | 88.3 | |
| 0.008 | 7.0 | 2.95 | 91.3 | Fine silt |
| 0.006 | 7.5 | 2.54 | 93.8 | V.Fine Silt |
| 0.004 | 8.0 | 1.93 | 95.8 | |
| 0.002 | 9.0 | 2.24 | 98.0 | Coarse Clay |
| 0.001 | 10.0 | 1.00 | 99.0 | Medium Clay |
| <0.001 | >10.0 | 1.00 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|-------------|--------|------------------------|------|
| Mean (MZ) | 0.144 | 0.832 | 2.80 |
| Median | 0.279 | | 1.84 |
| Sorting | Value | Inference | |
| Coefficient | 2.66 | Very Poorly Sorted | |
| Skewness | 0.43 | Very Positive (Coarse) | |
| Kurtosis | 1.13 | Leptokurtic | |
| % Fines | 27.64% | Fine Sand | |
| % Sands | 66.69% | | |
| % Gravel | 5.67% | | |

Sample No.: A_1015_ENV Operator: PaulSil
 Source Data: Nobel Energy Date & Time: 29/05/2014 15:50

Sample No.: A_18_ENV Operator: PaulSil
 Source Data: Nobel Energy Date & Time: 29/05/2014 13:21



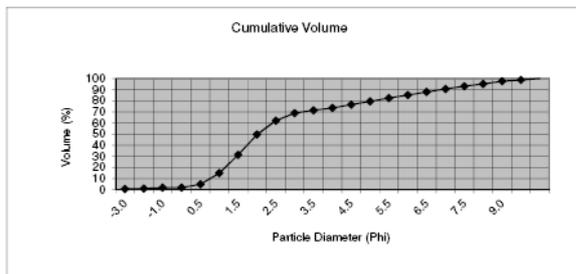
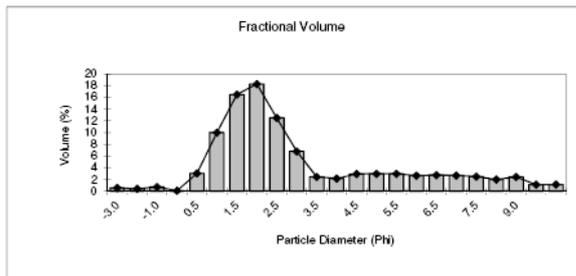
| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8,000 | -3.0 | 17.75 | 17.7 | Pebble |
| 4,000 | -2.0 | 6.52 | 24.3 | |
| 2,000 | -1.0 | 3.04 | 27.3 | Granule |
| 1,000 | 0.0 | 2.25 | 29.6 | V.Coarse Sand |
| 0,710 | 0.5 | 2.01 | 31.6 | |
| 0,500 | 1.0 | 4.28 | 35.8 | Coarse Sand |
| 0,355 | 1.5 | 7.67 | 43.5 | |
| 0,250 | 2.0 | 9.90 | 53.4 | Medium Sand |
| 0,180 | 2.5 | 8.05 | 61.5 | |
| 0,125 | 3.0 | 5.58 | 67.0 | Fine Sand |
| 0,900 | 3.5 | 2.80 | 69.8 | |
| 0,063 | 4.0 | 2.53 | 72.4 | V.Fine Sand |
| 0,044 | 4.5 | 3.00 | 75.4 | |
| 0,032 | 5.0 | 3.04 | 78.4 | Coarse Silt |
| 0,022 | 5.5 | 3.12 | 81.5 | |
| 0,016 | 6.0 | 2.83 | 84.4 | Medium Silt |
| 0,011 | 6.5 | 2.87 | 87.2 | |
| 0,008 | 7.0 | 2.80 | 90.0 | Fine silt |
| 0,006 | 7.5 | 2.62 | 92.7 | |
| 0,004 | 8.0 | 2.14 | 94.8 | V.Fine Silt |
| 0,002 | 9.0 | 2.64 | 97.4 | Coarse Clay |
| 0,001 | 10.0 | 1.24 | 98.7 | Medium Clay |
| <0.001 | >10.0 | 1.34 | 100.0 | Fine Clay |

| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8,000 | -3.0 | 1.00 | 1.0 | Pebble |
| 4,000 | -2.0 | 1.76 | 2.8 | |
| 2,000 | -1.0 | 1.06 | 3.8 | Granule |
| 1,000 | 0.0 | 3.91 | 7.7 | V.Coarse Sand |
| 0,710 | 0.5 | 5.42 | 13.2 | |
| 0,500 | 1.0 | 10.65 | 23.8 | Coarse Sand |
| 0,355 | 1.5 | 14.67 | 38.5 | |
| 0,250 | 2.0 | 14.65 | 53.1 | Medium Sand |
| 0,180 | 2.5 | 9.41 | 62.5 | |
| 0,125 | 3.0 | 5.22 | 67.8 | Fine Sand |
| 0,900 | 3.5 | 2.42 | 70.2 | |
| 0,063 | 4.0 | 2.58 | 72.8 | V.Fine Sand |
| 0,044 | 4.5 | 3.20 | 75.9 | |
| 0,032 | 5.0 | 3.14 | 79.1 | Coarse Silt |
| 0,022 | 5.5 | 3.24 | 82.3 | |
| 0,016 | 6.0 | 3.04 | 85.4 | Medium Silt |
| 0,011 | 6.5 | 3.09 | 88.5 | |
| 0,008 | 7.0 | 2.88 | 91.3 | Fine silt |
| 0,006 | 7.5 | 2.51 | 93.8 | |
| 0,004 | 8.0 | 1.91 | 95.8 | V.Fine Silt |
| 0,002 | 9.0 | 2.23 | 98.0 | Coarse Clay |
| 0,001 | 10.0 | 1.02 | 99.0 | Medium Clay |
| <0.001 | >10.0 | 1.00 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|------------------------------------|------|
| Mean (MZ) | 0.354 | 6.612 | 1.50 |
| Median | 0.286 | | 1.81 |
| Sorting Coefficient | Value: 4.13 | Inference: Extremely Poorly Sorted | |
| Skewness | -0.03 | Symmetrical | |
| Kurtosis | 0.80 | Platykurtic | |
| % Fines | 27.63% | Medium Sand | |
| % Sands | 45.06% | | |
| % Gravel | 27.30% | | |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.149 | 0.565 | 2.75 |
| Median | 0.272 | | 1.88 |
| Sorting Coefficient | Value: 2.58 | Inference: Very Poorly Sorted | |
| Skewness | 0.45 | Very Positive (Coarse) | |
| Kurtosis | 1.06 | Mesokurtic | |
| % Fines | 27.25% | Fine Sand | |
| % Sands | 68.93% | | |
| % Gravel | 3.82% | | |

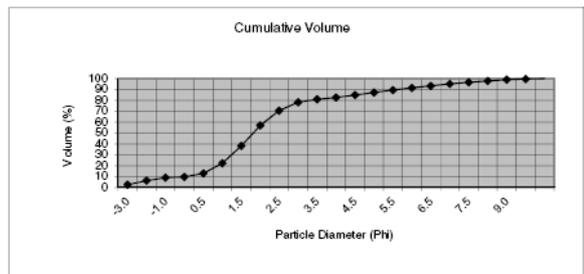
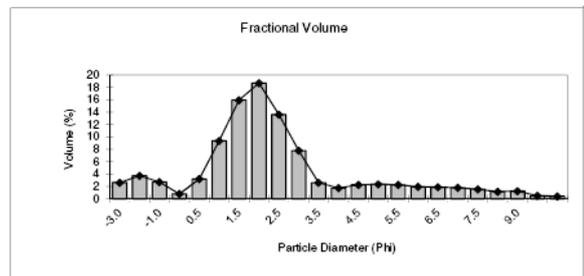
Sample No.: A_20_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 11:17



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.58 | 0.6 | Pebble |
| 4.000 | -2.0 | 0.44 | 1.0 | |
| 2.000 | -1.0 | 0.73 | 1.8 | Granule |
| 1.000 | 0.0 | 0.11 | 1.9 | V.Coarse Sand |
| 0.710 | 0.5 | 3.06 | 4.9 | |
| 0.500 | 1.0 | 10.00 | 14.9 | Coarse Sand |
| 0.355 | 1.5 | 16.48 | 31.4 | |
| 0.250 | 2.0 | 18.28 | 49.7 | Medium Sand |
| 0.180 | 2.5 | 12.51 | 62.2 | |
| 0.125 | 3.0 | 6.81 | 69.0 | Fine Sand |
| 0.900 | 3.5 | 2.46 | 71.5 | |
| 0.063 | 4.0 | 2.19 | 73.7 | V.Fine Sand |
| 0.044 | 4.5 | 2.95 | 76.6 | |
| 0.032 | 5.0 | 3.00 | 79.6 | Coarse Silt |
| 0.022 | 5.5 | 2.98 | 82.6 | |
| 0.016 | 6.0 | 2.68 | 85.3 | Medium Silt |
| 0.011 | 6.5 | 2.75 | 88.0 | |
| 0.008 | 7.0 | 2.71 | 90.7 | Fine silt |
| 0.006 | 7.5 | 2.51 | 93.2 | |
| 0.004 | 8.0 | 2.02 | 95.3 | V.Fine Silt |
| 0.002 | 9.0 | 2.45 | 97.7 | |
| 0.001 | 10.0 | 1.14 | 98.9 | Coarse Clay |
| <0.001 | >10.0 | 1.14 | 100.0 | Medium Clay |
| | | | | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|------------------------|------|
| Mean (MZ) | 0.131 | 0.278 | 2.93 |
| Median | 0.248 | | 2.01 |
| Sorting Coefficient | Value | Inference | |
| | 2.31 | Very Poorly Sorted | |
| Skewness | 0.59 | Very Positive (Coarse) | |
| Kurtosis | 1.04 | Mesokurtic | |
| % Fines | 26.34% | Fine Sand | |
| % Sands | 71.91% | | |
| % Gravel | 1.75% | | |

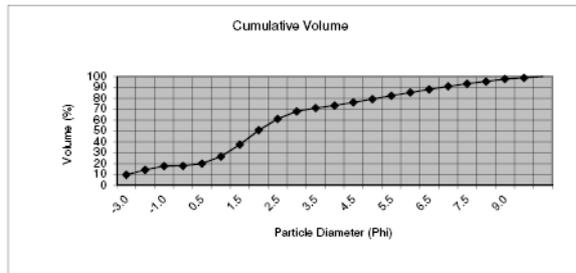
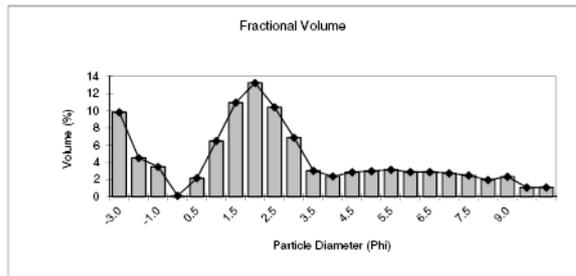
Sample No.: A_21_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 11:47



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 2.61 | 2.6 | Pebble |
| 4.000 | -2.0 | 3.72 | 6.3 | |
| 2.000 | -1.0 | 2.70 | 9.0 | Granule |
| 1.000 | 0.0 | 0.78 | 9.8 | V.Coarse Sand |
| 0.710 | 0.5 | 3.20 | 13.0 | |
| 0.500 | 1.0 | 9.36 | 22.4 | Coarse Sand |
| 0.355 | 1.5 | 15.94 | 38.3 | |
| 0.250 | 2.0 | 18.70 | 57.0 | Medium Sand |
| 0.180 | 2.5 | 13.60 | 70.6 | |
| 0.125 | 3.0 | 7.78 | 78.4 | Fine Sand |
| 0.900 | 3.5 | 2.60 | 81.0 | |
| 0.063 | 4.0 | 1.73 | 82.7 | V.Fine Sand |
| 0.044 | 4.5 | 2.26 | 85.0 | |
| 0.032 | 5.0 | 2.34 | 87.3 | Coarse Silt |
| 0.022 | 5.5 | 2.26 | 89.6 | |
| 0.016 | 6.0 | 1.92 | 91.5 | Medium Silt |
| 0.011 | 6.5 | 1.89 | 93.4 | |
| 0.008 | 7.0 | 1.77 | 95.2 | Fine silt |
| 0.006 | 7.5 | 1.54 | 96.7 | |
| 0.004 | 8.0 | 1.15 | 97.9 | V.Fine Silt |
| 0.002 | 9.0 | 1.24 | 99.1 | |
| 0.001 | 10.0 | 0.50 | 99.6 | Coarse Clay |
| <0.001 | >10.0 | 0.40 | 100.0 | Medium Clay |
| | | | | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|--------------------|------|
| Mean (MZ) | 0.214 | 1.732 | 2.23 |
| Median | 0.289 | | 1.79 |
| Sorting Coefficient | Value | Inference | |
| | 2.33 | Very Poorly Sorted | |
| Skewness | 0.23 | Positive (Coarse) | |
| Kurtosis | 2.30 | Very Leptokurtic | |
| % Fines | 17.27% | Fine Sand | |
| % Sands | 73.70% | | |
| % Gravel | 9.03% | | |

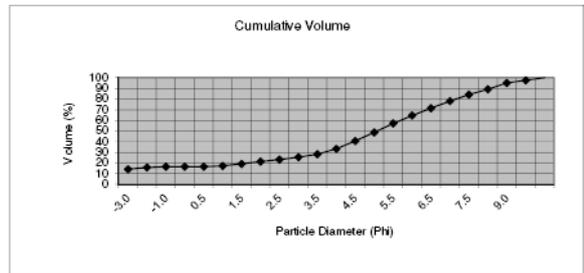
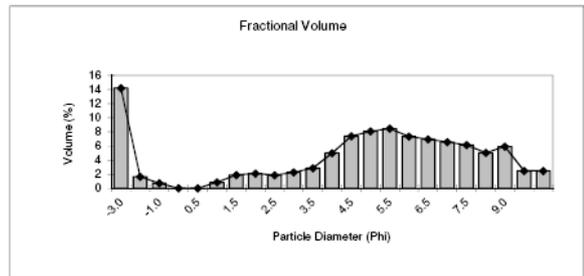
Sample No.: A_22_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 11:08



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 9.83 | 9.8 | Pebble |
| 4.000 | -2.0 | 4.53 | 14.4 | |
| 2.000 | -1.0 | 3.49 | 17.9 | Granule |
| 1.000 | 0.0 | 0.12 | 18.0 | V.Coarse Sand |
| 0.710 | 0.5 | 2.16 | 20.1 | |
| 0.500 | 1.0 | 6.48 | 26.6 | Coarse Sand |
| 0.355 | 1.5 | 10.95 | 37.6 | |
| 0.250 | 2.0 | 13.24 | 50.8 | Medium Sand |
| 0.180 | 2.5 | 10.40 | 61.2 | |
| 0.125 | 3.0 | 6.89 | 68.1 | Fine Sand |
| 0.900 | 3.5 | 3.03 | 71.1 | |
| 0.063 | 4.0 | 2.38 | 73.5 | V.Fine Sand |
| 0.044 | 4.5 | 2.86 | 76.4 | |
| 0.032 | 5.0 | 3.00 | 79.4 | Coarse Silt |
| 0.022 | 5.5 | 3.15 | 82.5 | |
| 0.016 | 6.0 | 2.88 | 85.4 | Medium Silt |
| 0.011 | 6.5 | 2.89 | 88.3 | |
| 0.008 | 7.0 | 2.75 | 91.0 | Fine silt |
| 0.006 | 7.5 | 2.48 | 93.5 | |
| 0.004 | 8.0 | 1.96 | 95.5 | V.Fine Silt |
| 0.002 | 9.0 | 2.34 | 97.8 | Coarse Clay |
| 0.001 | 10.0 | 1.09 | 98.9 | Medium Clay |
| <0.001 | >10.0 | 1.09 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|-------------------------|------|
| Mean (MZ) | 0.245 | 36.424 | 2.03 |
| Median | 0.256 | | 1.96 |
| Sorting Coefficient | Value | Inference | |
| | 4.06 | Extremely Poorly Sorted | |
| Skewness | -0.08 | Symmetrical | |
| Kurtosis | 1.78 | Very Leptokurtic | |
| % Fines | 26.49% | Fine Sand | |
| % Sands | 55.66% | | |
| % Gravel | 17.85% | | |

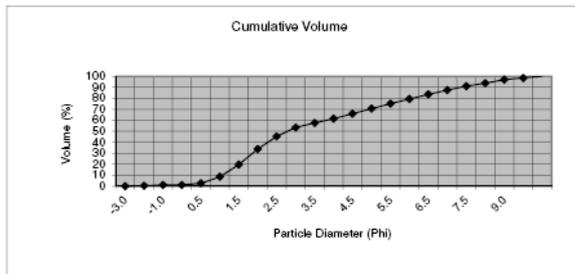
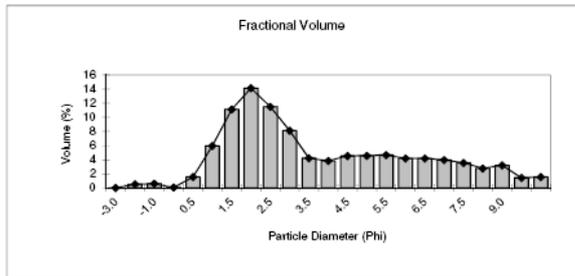
Sample No.: A_26_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 16:16



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 14.18 | 14.2 | Pebble |
| 4.000 | -2.0 | 1.64 | 15.8 | |
| 2.000 | -1.0 | 0.71 | 16.5 | Granule |
| 1.000 | 0.0 | 0.00 | 16.5 | V.Coarse Sand |
| 0.710 | 0.5 | 0.00 | 16.5 | |
| 0.500 | 1.0 | 0.85 | 17.4 | Coarse Sand |
| 0.355 | 1.5 | 1.88 | 19.3 | |
| 0.250 | 2.0 | 2.10 | 21.4 | Medium Sand |
| 0.180 | 2.5 | 1.85 | 23.2 | |
| 0.125 | 3.0 | 2.26 | 25.5 | Fine Sand |
| 0.900 | 3.5 | 2.86 | 28.3 | |
| 0.063 | 4.0 | 4.97 | 33.3 | V.Fine Sand |
| 0.044 | 4.5 | 7.38 | 40.7 | |
| 0.032 | 5.0 | 8.07 | 48.7 | Coarse Silt |
| 0.022 | 5.5 | 8.45 | 57.2 | |
| 0.016 | 6.0 | 7.33 | 64.5 | Medium Silt |
| 0.011 | 6.5 | 6.95 | 71.5 | |
| 0.008 | 7.0 | 6.55 | 78.0 | Fine silt |
| 0.006 | 7.5 | 6.13 | 84.1 | |
| 0.004 | 8.0 | 5.02 | 89.2 | V.Fine Silt |
| 0.002 | 9.0 | 5.91 | 95.1 | Coarse Clay |
| 0.001 | 10.0 | 2.46 | 97.5 | Medium Clay |
| <0.001 | >10.0 | 2.46 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|-------------------------|------|
| Mean (MZ) | 0.084 | 20874.306 | 3.58 |
| Median | 0.030 | | 5.06 |
| Sorting Coefficient | Value | Inference | |
| | 6.08 | Extremely Poorly Sorted | |
| Skewness | -0.58 | Very Negative(fine) | |
| Kurtosis | 2.62 | Very Leptokurtic | |
| % Fines | 66.71% | V.Fine Sands | |
| % Sands | 16.76% | | |
| % Gravel | 16.53% | | |

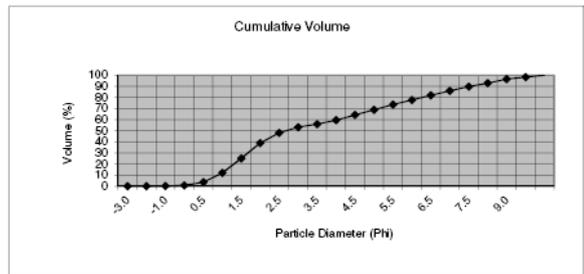
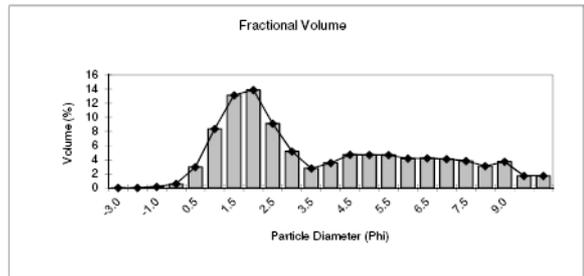
Sample No.: A_27_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 14:51



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.50 | 0.5 | |
| 2.000 | -1.0 | 0.59 | 1.1 | Granule |
| 1.000 | 0.0 | 0.06 | 1.2 | V.Coarse Sand |
| 0.710 | 0.5 | 1.55 | 2.7 | |
| 0.500 | 1.0 | 5.95 | 8.7 | Coarse Sand |
| 0.355 | 1.5 | 11.10 | 19.8 | |
| 0.250 | 2.0 | 14.11 | 33.9 | Medium Sand |
| 0.180 | 2.5 | 11.48 | 45.3 | |
| 0.125 | 3.0 | 8.12 | 53.5 | Fine Sand |
| 0.900 | 3.5 | 4.19 | 57.7 | |
| 0.063 | 4.0 | 3.83 | 61.5 | V.Fine Sand |
| 0.044 | 4.5 | 4.52 | 66.0 | |
| 0.032 | 5.0 | 4.55 | 70.6 | Coarse Silt |
| 0.022 | 5.5 | 4.65 | 75.2 | |
| 0.016 | 6.0 | 4.18 | 79.4 | Medium Silt |
| 0.011 | 6.5 | 4.17 | 83.6 | |
| 0.008 | 7.0 | 3.95 | 87.5 | Fine silt |
| 0.006 | 7.5 | 3.55 | 91.1 | |
| 0.004 | 8.0 | 2.77 | 93.8 | V.Fine Silt |
| 0.002 | 9.0 | 3.20 | 97.0 | |
| 0.001 | 10.0 | 1.43 | 98.5 | Coarse Clay |
| <0.001 | >10.0 | 1.54 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|-------------|--------|------------------------|------|
| Mean (MZ) | 0.086 | 0.240 | 3.54 |
| Median | 0.148 | | 2.75 |
| Sorting | Value | Inference | |
| Coefficient | 2.47 | Very Poorly Sorted | |
| Skewness | 0.45 | Very Positive (Coarse) | |
| Kurtosis | 0.82 | Platykurtic | |
| % Fines | 38.52% | V.Fine Sands | |
| % Sands | 60.40% | | |
| % Gravel | 1.09% | | |

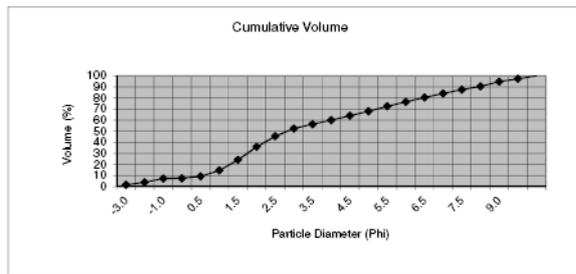
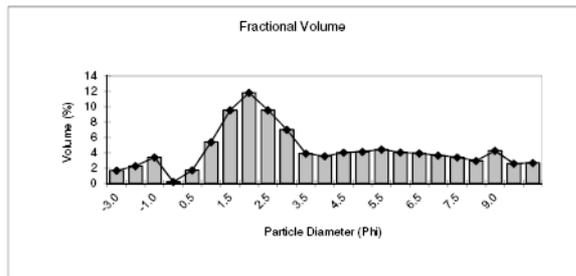
Sample No.: A_202_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 10:56



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.00 | 0.0 | |
| 2.000 | -1.0 | 0.15 | 0.2 | Granule |
| 1.000 | 0.0 | 0.57 | 0.7 | V.Coarse Sand |
| 0.710 | 0.5 | 2.96 | 3.7 | |
| 0.500 | 1.0 | 8.34 | 12.0 | Coarse Sand |
| 0.355 | 1.5 | 13.12 | 25.1 | |
| 0.250 | 2.0 | 13.85 | 39.0 | Medium Sand |
| 0.180 | 2.5 | 9.08 | 48.1 | |
| 0.125 | 3.0 | 5.15 | 53.2 | Fine Sand |
| 0.900 | 3.5 | 2.76 | 56.0 | |
| 0.063 | 4.0 | 3.54 | 59.5 | V.Fine Sand |
| 0.044 | 4.5 | 4.70 | 64.2 | |
| 0.032 | 5.0 | 4.67 | 68.9 | Coarse Silt |
| 0.022 | 5.5 | 4.65 | 73.6 | |
| 0.016 | 6.0 | 4.15 | 77.7 | Medium Silt |
| 0.011 | 6.5 | 4.20 | 81.9 | |
| 0.008 | 7.0 | 4.09 | 86.0 | Fine silt |
| 0.006 | 7.5 | 3.81 | 89.8 | |
| 0.004 | 8.0 | 3.08 | 92.9 | V.Fine Silt |
| 0.002 | 9.0 | 3.71 | 96.6 | |
| 0.001 | 10.0 | 1.70 | 98.3 | Coarse Clay |
| <0.001 | >10.0 | 1.71 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|-------------|--------|------------------------|------|
| Mean (MZ) | 0.088 | 0.266 | 3.51 |
| Median | 0.160 | | 2.65 |
| Sorting | Value | Inference | |
| Coefficient | 2.60 | Very Poorly Sorted | |
| Skewness | 0.47 | Very Positive (Coarse) | |
| Kurtosis | 0.78 | Platykurtic | |
| % Fines | 40.47% | V.Fine Sands | |
| % Sands | 59.38% | | |
| % Gravel | 0.15% | | |

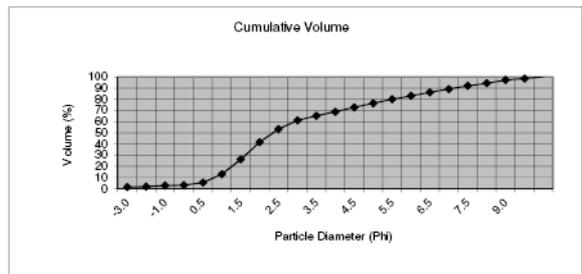
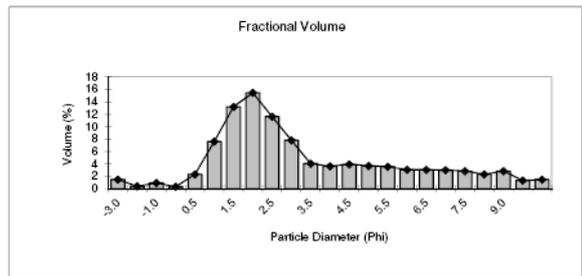
Sample No.: A_204_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 10:49



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 1.69 | 1.7 | Pebble |
| 4.000 | -2.0 | 2.27 | 4.0 | |
| 2.000 | -1.0 | 3.41 | 7.4 | Granule |
| 1.000 | 0.0 | 0.20 | 7.6 | V.Coarse Sand |
| 0.710 | 0.5 | 1.72 | 9.3 | |
| 0.500 | 1.0 | 5.39 | 14.7 | Coarse Sand |
| 0.355 | 1.5 | 9.53 | 24.2 | |
| 0.250 | 2.0 | 11.79 | 36.0 | Medium Sand |
| 0.180 | 2.5 | 9.53 | 45.5 | |
| 0.125 | 3.0 | 7.00 | 52.5 | Fine Sand |
| 0.900 | 3.5 | 3.90 | 56.4 | |
| 0.063 | 4.0 | 3.56 | 60.0 | V.Fine Sand |
| 0.044 | 4.5 | 4.01 | 64.0 | |
| 0.032 | 5.0 | 4.12 | 68.1 | Coarse Silt |
| 0.022 | 5.5 | 4.43 | 72.5 | |
| 0.016 | 6.0 | 4.04 | 76.6 | Medium Silt |
| 0.011 | 6.5 | 3.91 | 80.5 | |
| 0.008 | 7.0 | 3.64 | 84.1 | Fine silt |
| 0.006 | 7.5 | 3.41 | 87.5 | |
| 0.004 | 8.0 | 2.95 | 90.5 | V.Fine Silt |
| 0.002 | 9.0 | 4.26 | 94.8 | |
| 0.001 | 10.0 | 2.58 | 97.3 | Coarse Clay |
| <0.001 | >10.0 | 2.67 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.082 | 1.087 | 3.61 |
| Median | 0.145 | | 2.79 |
| Sorting Coefficient | Value: 3.12 | Inference: Very Poorly Sorted | |
| Skewness | 0.29 | Positive(Coarse) | |
| Kurtosis | 1.04 | Mesokurtic | |
| % Fines | 40.03% | v.Fine Sands | |
| % Sands | 52.61% | | |
| % Gravel | 7.36% | | |

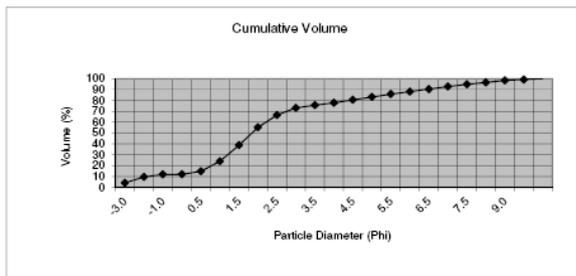
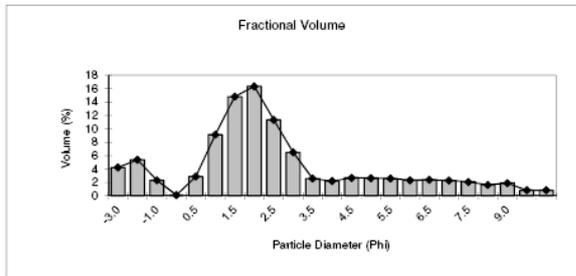
Sample No.: A_301_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 14:10



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 1.49 | 1.5 | Pebble |
| 4.000 | -2.0 | 0.41 | 1.9 | |
| 2.000 | -1.0 | 0.95 | 2.8 | Granule |
| 1.000 | 0.0 | 0.34 | 3.2 | V.Coarse Sand |
| 0.710 | 0.5 | 2.33 | 5.5 | |
| 0.500 | 1.0 | 7.61 | 13.1 | Coarse Sand |
| 0.355 | 1.5 | 13.19 | 26.3 | |
| 0.250 | 2.0 | 15.45 | 41.8 | Medium Sand |
| 0.180 | 2.5 | 11.61 | 53.4 | |
| 0.125 | 3.0 | 7.82 | 61.2 | Fine Sand |
| 0.900 | 3.5 | 4.06 | 65.2 | |
| 0.063 | 4.0 | 3.63 | 68.9 | V.Fine Sand |
| 0.044 | 4.5 | 3.94 | 72.8 | |
| 0.032 | 5.0 | 3.69 | 76.5 | Coarse Silt |
| 0.022 | 5.5 | 3.55 | 80.0 | |
| 0.016 | 6.0 | 3.07 | 83.1 | Medium Silt |
| 0.011 | 6.5 | 3.06 | 86.2 | |
| 0.008 | 7.0 | 3.01 | 89.2 | Fine silt |
| 0.006 | 7.5 | 2.85 | 92.0 | |
| 0.004 | 8.0 | 2.34 | 94.4 | V.Fine Silt |
| 0.002 | 9.0 | 2.85 | 97.2 | |
| 0.001 | 10.0 | 1.33 | 98.5 | Coarse Clay |
| <0.001 | >10.0 | 1.47 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.110 | 0.289 | 3.18 |
| Median | 0.200 | | 2.32 |
| Sorting Coefficient | Value: 2.44 | Inference: Very Poorly Sorted | |
| Skewness | 0.51 | Very Positive (Coarse) | |
| Kurtosis | 0.96 | Mesokurtic | |
| % Fines | 31.14% | V.Fine Sands | |
| % Sands | 66.02% | | |
| % Gravel | 2.84% | | |

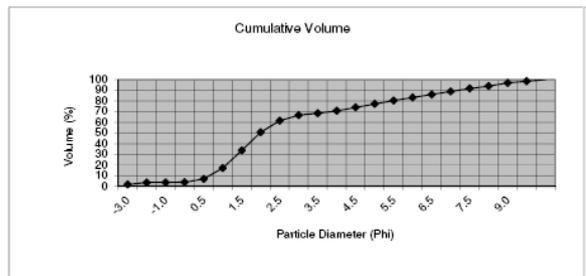
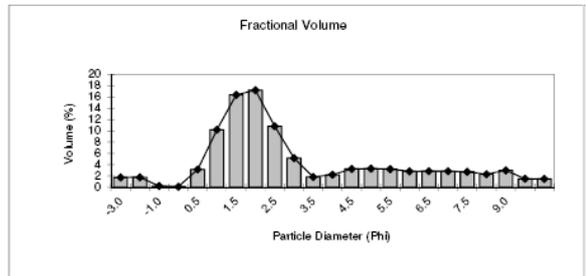
Sample No.: A_302_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 16:02



| Aperture (um) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8,000 | -3.0 | 4.24 | 4.2 | Pebble |
| 4,000 | -2.0 | 5.37 | 9.6 | |
| 2,000 | -1.0 | 2.33 | 11.9 | Granule |
| 1,000 | 0.0 | 0.11 | 12.0 | V.Coarse Sand |
| 0,710 | 0.5 | 2.87 | 14.9 | |
| 0,500 | 1.0 | 9.13 | 24.0 | Coarse Sand |
| 0,355 | 1.5 | 14.80 | 38.9 | |
| 0,250 | 2.0 | 16.35 | 55.2 | Medium Sand |
| 0,180 | 2.5 | 11.35 | 66.5 | |
| 0,125 | 3.0 | 6.49 | 73.0 | Fine Sand |
| 0,900 | 3.5 | 2.58 | 75.6 | |
| 0,063 | 4.0 | 2.20 | 77.8 | V.Fine Sand |
| 0,044 | 4.5 | 2.69 | 80.5 | |
| 0,032 | 5.0 | 2.64 | 83.1 | Coarse Silt |
| 0,022 | 5.5 | 2.60 | 85.7 | |
| 0,016 | 6.0 | 2.33 | 88.1 | Medium Silt |
| 0,011 | 6.5 | 2.38 | 90.4 | |
| 0,008 | 7.0 | 2.30 | 92.7 | Fine silt |
| 0,006 | 7.5 | 2.08 | 94.8 | |
| 0,004 | 8.0 | 1.63 | 96.4 | V.Fine Silt |
| 0,002 | 9.0 | 1.90 | 98.3 | |
| 0,001 | 10.0 | 0.82 | 99.2 | Coarse Clay |
| <0.001 | >10.0 | 0.83 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.177 | 2.539 | 2.50 |
| Median | 0.283 | | 1.82 |
| Sorting Coefficient | Value: 2.73 | Inference: Very Poorly Sorted | |
| Skewness | 0.27 | Positive(Coarse) | |
| Kurtosis | 1.85 | Very Leptokurtic | |
| % Fines | 22.18% | Fine Sand | |
| % Sands | 65.88% | | |
| % Gravel | 11.94% | | |

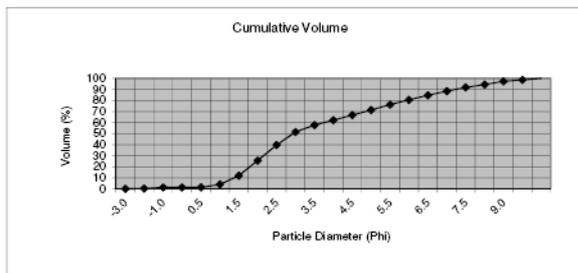
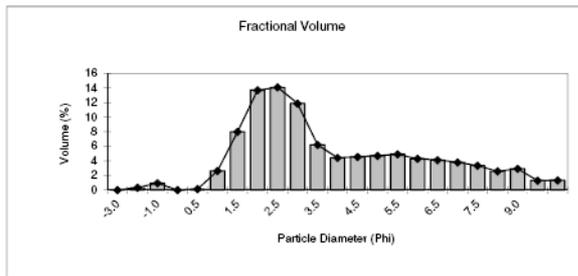
Sample No.: A_303_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 10:24



| Aperture (um) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8,000 | -3.0 | 1.75 | 1.7 | Pebble |
| 4,000 | -2.0 | 1.78 | 3.5 | |
| 2,000 | -1.0 | 0.21 | 3.7 | Granule |
| 1,000 | 0.0 | 0.12 | 3.9 | V.Coarse Sand |
| 0,710 | 0.5 | 3.16 | 7.0 | |
| 0,500 | 1.0 | 10.18 | 17.2 | Coarse Sand |
| 0,355 | 1.5 | 16.35 | 33.5 | |
| 0,250 | 2.0 | 17.19 | 50.7 | Medium Sand |
| 0,180 | 2.5 | 10.79 | 61.5 | |
| 0,125 | 3.0 | 5.18 | 66.7 | Fine Sand |
| 0,900 | 3.5 | 1.83 | 68.5 | |
| 0,063 | 4.0 | 2.21 | 70.7 | V.Fine Sand |
| 0,044 | 4.5 | 3.25 | 74.0 | |
| 0,032 | 5.0 | 3.29 | 77.3 | Coarse Silt |
| 0,022 | 5.5 | 3.22 | 80.5 | |
| 0,016 | 6.0 | 2.82 | 83.3 | Medium Silt |
| 0,011 | 6.5 | 2.84 | 86.2 | |
| 0,008 | 7.0 | 2.83 | 89.0 | Fine silt |
| 0,006 | 7.5 | 2.73 | 91.7 | |
| 0,004 | 8.0 | 2.31 | 94.0 | V.Fine Silt |
| 0,002 | 9.0 | 3.01 | 97.0 | |
| 0,001 | 10.0 | 1.49 | 98.5 | Coarse Clay |
| <0.001 | >10.0 | 1.47 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.125 | 0.329 | 3.00 |
| Median | 0.255 | | 1.97 |
| Sorting Coefficient | Value: 2.52 | Inference: Very Poorly Sorted | |
| Skewness | 0.57 | Very Positive (Coarse) | |
| Kurtosis | 0.97 | Mesokurtic | |
| % Fines | 29.26% | V.Fine Sands | |
| % Sands | 67.01% | | |
| % Gravel | 3.74% | | |

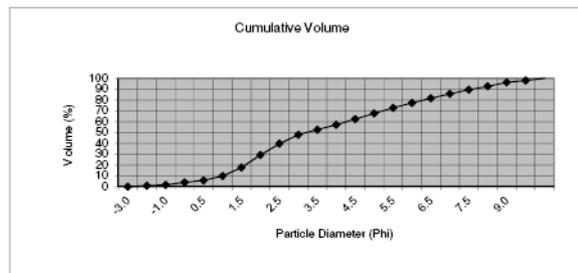
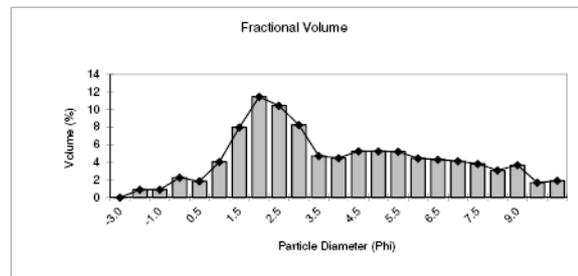
Sample No.: A_304_ENV Operator: PaulSil
 Source Data: Nobel Energy Date& Time: 29/05/2014 10:13



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.31 | 0.3 | |
| 2.000 | -1.0 | 0.94 | 1.3 | Granule |
| 1.000 | 0.0 | 0.00 | 1.3 | V.Coarse Sand |
| 0.710 | 0.5 | 0.12 | 1.4 | |
| 0.500 | 1.0 | 2.62 | 4.0 | Coarse Sand |
| 0.355 | 1.5 | 7.99 | 12.0 | |
| 0.250 | 2.0 | 13.68 | 25.7 | Medium Sand |
| 0.180 | 2.5 | 14.07 | 39.7 | |
| 0.125 | 3.0 | 11.86 | 51.6 | Fine Sand |
| 0.900 | 3.5 | 6.21 | 57.8 | |
| 0.063 | 4.0 | 4.41 | 62.2 | V.Fine Sand |
| 0.044 | 4.5 | 4.55 | 66.8 | |
| 0.032 | 5.0 | 4.70 | 71.5 | Coarse Silt |
| 0.022 | 5.5 | 4.89 | 76.4 | |
| 0.016 | 6.0 | 4.29 | 80.6 | Medium Silt |
| 0.011 | 6.5 | 4.11 | 84.8 | |
| 0.008 | 7.0 | 3.79 | 88.5 | Fine silt |
| 0.006 | 7.5 | 3.35 | 91.9 | |
| 0.004 | 8.0 | 2.57 | 94.5 | V.Fine Silt |
| 0.002 | 9.0 | 2.92 | 97.4 | |
| 0.001 | 10.0 | 1.29 | 98.7 | Coarse Clay |
| <0.001 | >10.0 | 1.33 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.080 | 0.186 | 3.65 |
| Median | 0.132 | | 2.92 |
| Sorting Coefficient | Value: 2.27 | Inference: Very Poorly Sorted | |
| Skewness | 0.47 | Very Positive (Coarse) | |
| Kurtosis | 0.86 | Platykurtic | |
| % Fines | 37.79% | V.Fine Sands | |
| % Sands | 60.95% | | |
| % Gravel | 1.26% | | |

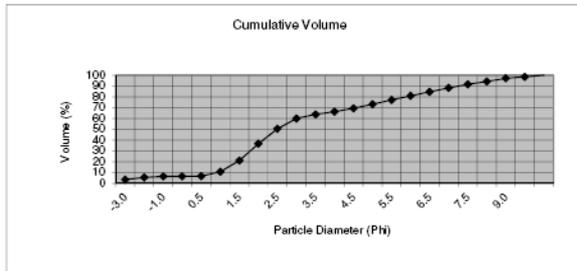
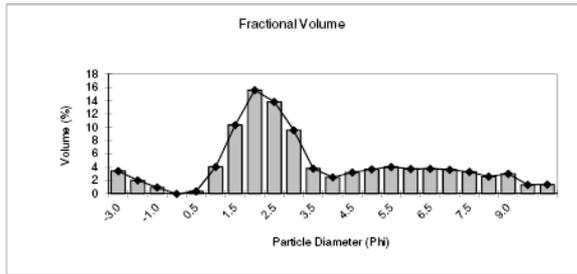
Sample No.: A_305_ENV Operator: PaulSil
 Source Data: Nobel Energy Date& Time: 29/05/2014 15:11



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.89 | 0.9 | |
| 2.000 | -1.0 | 0.89 | 1.8 | Granule |
| 1.000 | 0.0 | 2.26 | 4.0 | V.Coarse Sand |
| 0.710 | 0.5 | 1.84 | 5.9 | |
| 0.500 | 1.0 | 4.05 | 9.9 | Coarse Sand |
| 0.355 | 1.5 | 7.97 | 17.9 | |
| 0.250 | 2.0 | 11.44 | 29.3 | Medium Sand |
| 0.180 | 2.5 | 10.44 | 39.8 | |
| 0.125 | 3.0 | 8.24 | 48.0 | Fine Sand |
| 0.900 | 3.5 | 4.72 | 52.7 | |
| 0.063 | 4.0 | 4.48 | 57.2 | V.Fine Sand |
| 0.044 | 4.5 | 5.27 | 62.5 | |
| 0.032 | 5.0 | 5.25 | 67.7 | Coarse Silt |
| 0.022 | 5.5 | 5.20 | 72.9 | |
| 0.016 | 6.0 | 4.45 | 77.4 | Medium Silt |
| 0.011 | 6.5 | 4.31 | 81.7 | |
| 0.008 | 7.0 | 4.13 | 85.8 | Fine silt |
| 0.006 | 7.5 | 3.83 | 89.7 | |
| 0.004 | 8.0 | 3.09 | 92.7 | V.Fine Silt |
| 0.002 | 9.0 | 3.68 | 96.4 | |
| 0.001 | 10.0 | 1.68 | 98.1 | Coarse Clay |
| <0.001 | >10.0 | 1.90 | 100.0 | Medium Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.073 | 0.294 | 3.77 |
| Median | 0.110 | | 3.18 |
| Sorting Coefficient | Value: 2.60 | Inference: Very Poorly Sorted | |
| Skewness | 0.31 | Very Positive (Coarse) | |
| Kurtosis | 0.86 | Platykurtic | |
| % Fines | 42.79% | V.Fine Sands | |
| % Sands | 55.44% | | |
| % Gravel | 1.78% | | |

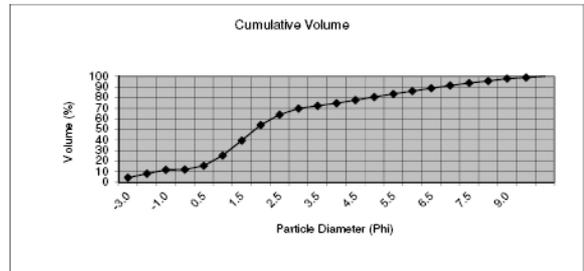
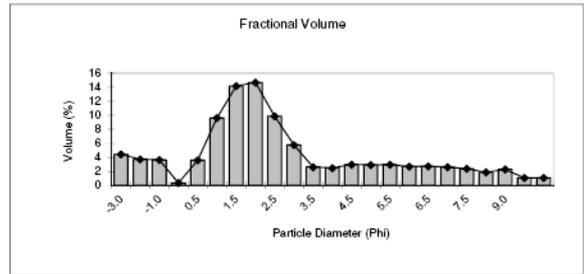
Sample No.: A_306_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 15:00



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 3.40 | 3.4 | Pebble |
| 4.000 | -2.0 | 2.01 | 5.4 | |
| 2.000 | -1.0 | 0.95 | 6.4 | Granule |
| 1.000 | 0.0 | 0.00 | 6.4 | V.Coarse Sand |
| 0.710 | 0.5 | 0.36 | 6.7 | Coarse Sand |
| 0.500 | 1.0 | 4.05 | 10.8 | |
| 0.355 | 1.5 | 10.33 | 21.1 | Medium Sand |
| 0.250 | 2.0 | 15.59 | 36.7 | |
| 0.180 | 2.5 | 13.83 | 50.5 | |
| 0.125 | 3.0 | 9.55 | 60.1 | Fine Sand |
| 0.900 | 3.5 | 3.80 | 63.9 | |
| 0.063 | 4.0 | 2.48 | 66.4 | V.Fine Sand |
| 0.044 | 4.5 | 3.20 | 69.6 | |
| 0.032 | 5.0 | 3.69 | 73.2 | Coarse Silt |
| 0.022 | 5.5 | 4.04 | 77.3 | |
| 0.016 | 6.0 | 3.73 | 81.0 | Medium Silt |
| 0.011 | 6.5 | 3.76 | 84.8 | |
| 0.008 | 7.0 | 3.62 | 88.4 | Fine silt |
| 0.006 | 7.5 | 3.29 | 91.7 | |
| 0.004 | 8.0 | 2.58 | 94.3 | V.Fine Silt |
| 0.002 | 9.0 | 3.00 | 97.3 | Coarse Clay |
| 0.001 | 10.0 | 1.34 | 98.6 | Medium Clay |
| <0.001 | >10.0 | 1.40 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.098 | 1.557 | 3.36 |
| Median | 0.183 | | 2.45 |
| Sorting Coefficient | Value: 2.87 | Inference: Very Poorly Sorted | |
| Skewness | 0.31 | Very Positive (Coarse) | |
| Kurtosis | 1.19 | Leptokurtic | |
| % Fines | 33.65% | V.Fine Sands | |
| % Sands | 59.99% | | |
| % Gravel | 6.36% | | |

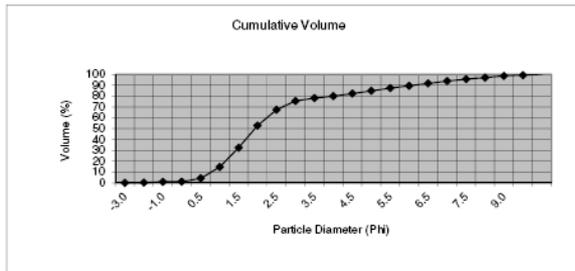
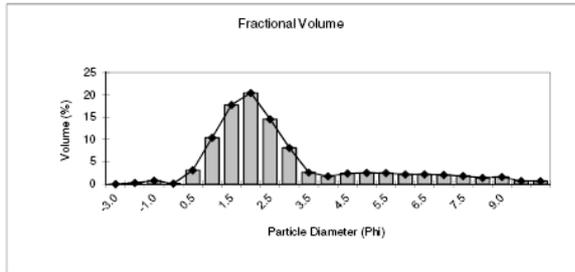
Sample No.: A_307_ENV Operator: PaulSil
 Source Data: Nobel Energy Date&Time: 29/05/2014 13:01



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 4.42 | 4.4 | Pebble |
| 4.000 | -2.0 | 3.70 | 8.1 | |
| 2.000 | -1.0 | 3.61 | 11.7 | Granule |
| 1.000 | 0.0 | 0.33 | 12.1 | V.Coarse Sand |
| 0.710 | 0.5 | 3.56 | 15.6 | Coarse Sand |
| 0.500 | 1.0 | 9.58 | 25.2 | |
| 0.355 | 1.5 | 14.15 | 39.4 | Medium Sand |
| 0.250 | 2.0 | 14.66 | 54.0 | |
| 0.180 | 2.5 | 9.85 | 63.9 | |
| 0.125 | 3.0 | 5.75 | 69.6 | Fine Sand |
| 0.900 | 3.5 | 2.60 | 72.2 | |
| 0.063 | 4.0 | 2.46 | 74.7 | V.Fine Sand |
| 0.044 | 4.5 | 2.96 | 77.6 | |
| 0.032 | 5.0 | 2.90 | 80.5 | Coarse Silt |
| 0.022 | 5.5 | 2.93 | 83.5 | |
| 0.016 | 6.0 | 2.67 | 86.1 | Medium Silt |
| 0.011 | 6.5 | 2.70 | 88.9 | |
| 0.008 | 7.0 | 2.59 | 91.4 | Fine silt |
| 0.006 | 7.5 | 2.34 | 93.8 | |
| 0.004 | 8.0 | 1.86 | 95.6 | V.Fine Silt |
| 0.002 | 9.0 | 2.25 | 97.9 | Coarse Clay |
| 0.001 | 10.0 | 1.06 | 98.9 | Medium Clay |
| <0.001 | >10.0 | 1.06 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|-------------|-------------------------------|------|
| Mean (MZ) | 0.160 | 2.452 | 2.65 |
| Median | 0.279 | | 1.84 |
| Sorting Coefficient | Value: 2.89 | Inference: Very Poorly Sorted | |
| Skewness | 0.30 | Positive (Coarse) | |
| Kurtosis | 1.44 | Leptokurtic | |
| % Fines | 25.32% | Fine Sand | |
| % Sands | 62.95% | | |
| % Gravel | 11.73% | | |

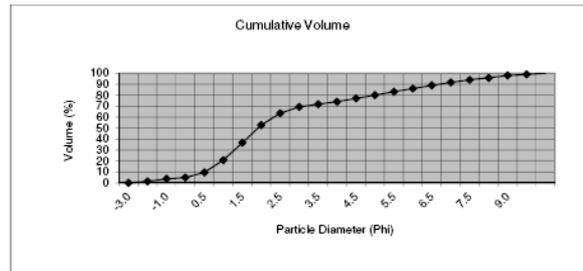
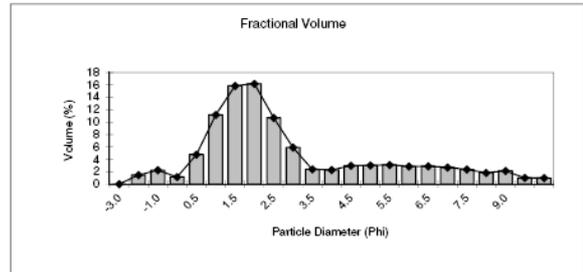
Sample No.: A-309-ENV Operator PaulSil
Source Data: Nobel Energy Date&Time: 29/05/2014 15:34



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 0.26 | 0.3 | |
| 2.000 | -1.0 | 0.79 | 1.0 | Granule |
| 1.000 | 0.0 | 0.13 | 1.2 | V.Coarse Sand |
| 0.710 | 0.5 | 3.13 | 4.3 | Coarse Sand |
| 0.500 | 1.0 | 10.40 | 14.7 | Coarse Sand |
| 0.355 | 1.5 | 17.74 | 32.4 | Medium Sand |
| 0.250 | 2.0 | 20.43 | 52.9 | Medium Sand |
| 0.180 | 2.5 | 14.55 | 67.4 | Fine Sand |
| 0.125 | 3.0 | 8.12 | 75.5 | Fine Sand |
| 0.900 | 3.5 | 2.62 | 78.2 | V.Fine Sand |
| 0.063 | 4.0 | 1.77 | 79.9 | V.Fine Sand |
| 0.044 | 4.5 | 2.41 | 82.3 | Coarse Silt |
| 0.032 | 5.0 | 2.52 | 84.9 | Coarse Silt |
| 0.022 | 5.5 | 2.48 | 87.4 | Medium Silt |
| 0.016 | 6.0 | 2.17 | 89.5 | Medium Silt |
| 0.011 | 6.5 | 2.18 | 91.7 | Fine silt |
| 0.008 | 7.0 | 2.07 | 93.8 | Fine silt |
| 0.006 | 7.5 | 1.83 | 95.6 | V.Fine Silt |
| 0.004 | 8.0 | 1.40 | 97.0 | V.Fine Silt |
| 0.002 | 9.0 | 1.62 | 98.6 | Coarse Clay |
| 0.001 | 10.0 | 0.70 | 99.3 | Medium Clay |
| <0.001 | >10.0 | 0.68 | 100.0 | Fine Clay |

| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|------------------------|------|
| Mean (MZ) | 0.167 | 0.265 | 2.58 |
| Median | 0.265 | | 1.92 |
| Sorting Coefficient | Value | Inference | |
| | 1.97 | Poorly Sorted | |
| Skewness | 0.56 | Very Positive (Coarse) | |
| Kurtosis | 1.65 | Very Leptokurtic | |
| % Fines | 20.07% | Fine Sand | |
| % Sands | 78.89% | | |
| % Gravel | 1.05% | | |

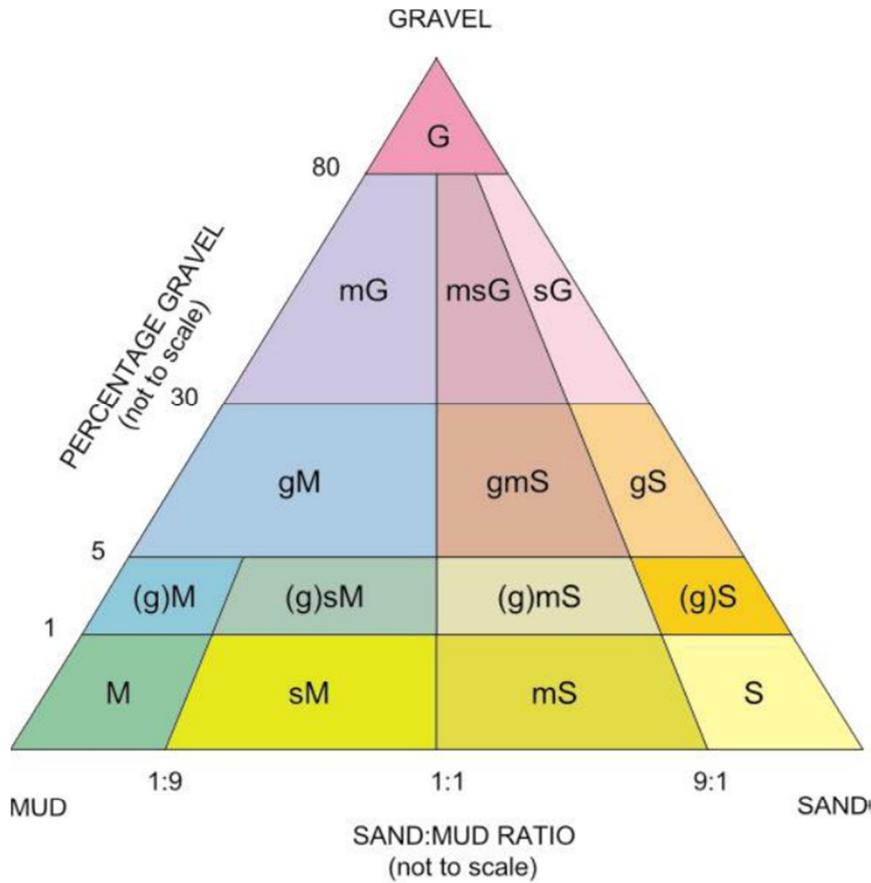
Sample No.: A_3010_ENV Operator PaulSil
Source Data: Nobel Energy Date&Time: 29/05/2014 13:56



| Aperture (µm) | Aperture (Phi unit) | Percentage Fractional | Cumulative | Sediment Description |
|---------------|---------------------|-----------------------|------------|----------------------|
| 8.000 | -3.0 | 0.00 | 0.0 | Pebble |
| 4.000 | -2.0 | 1.47 | 1.5 | |
| 2.000 | -1.0 | 2.24 | 3.7 | Granule |
| 1.000 | 0.0 | 1.16 | 4.9 | V.Coarse Sand |
| 0.710 | 0.5 | 4.77 | 9.6 | Coarse Sand |
| 0.500 | 1.0 | 11.17 | 20.8 | Coarse Sand |
| 0.355 | 1.5 | 15.85 | 36.7 | Medium Sand |
| 0.250 | 2.0 | 16.18 | 52.8 | Medium Sand |
| 0.180 | 2.5 | 10.69 | 63.5 | Fine Sand |
| 0.125 | 3.0 | 5.92 | 69.4 | Fine Sand |
| 0.900 | 3.5 | 2.39 | 71.8 | V.Fine Sand |
| 0.063 | 4.0 | 2.29 | 74.1 | V.Fine Sand |
| 0.044 | 4.5 | 2.99 | 77.1 | Coarse Silt |
| 0.032 | 5.0 | 3.03 | 80.1 | Coarse Silt |
| 0.022 | 5.5 | 3.11 | 83.3 | Medium Silt |
| 0.016 | 6.0 | 2.86 | 86.1 | Medium Silt |
| 0.011 | 6.5 | 2.88 | 89.0 | Fine silt |
| 0.008 | 7.0 | 2.69 | 91.7 | Fine silt |
| 0.006 | 7.5 | 2.37 | 94.1 | V.Fine Silt |
| 0.004 | 8.0 | 1.83 | 95.9 | V.Fine Silt |
| 0.002 | 9.0 | 2.15 | 98.0 | Coarse Clay |
| 0.001 | 10.0 | 1.00 | 99.0 | Medium Clay |
| <0.001 | >10.0 | 0.98 | 100.0 | Fine Clay |

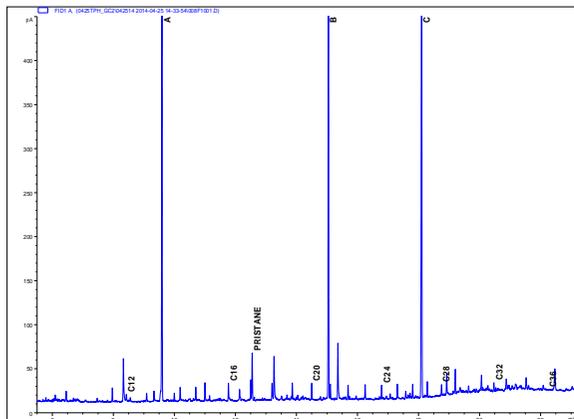
| Graphical | mm | StDev (mm) | Phi |
|---------------------|--------|------------------------|------|
| Mean (MZ) | 0.148 | 0.361 | 2.76 |
| Median | 0.268 | | 1.90 |
| Sorting Coefficient | Value | Inference | |
| | 2.39 | Very Poorly Sorted | |
| Skewness | 0.52 | Very Positive (Coarse) | |
| Kurtosis | 1.05 | Mesokurtic | |
| % Fines | 25.87% | Fine Sand | |
| % Sands | 70.42% | | |
| % Gravel | 3.71% | | |

Modified Folk classification

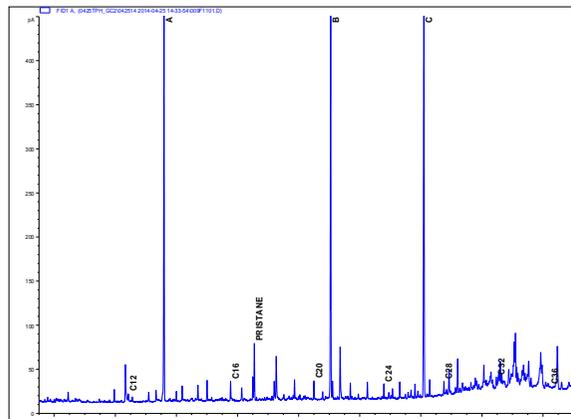


- M _____ Mud
- sM _____ Sandy mud
- (g)M _____ Slightly gravelly mud
- (g)sM _____ Slightly gravelly sandy mud
- gM _____ Gravelly mud
- S _____ Sand
- mS _____ Muddy sand
- (g)S _____ Slightly gravelly sand
- (g)mS _____ Slightly gravelly muddy sand
- gmS _____ Gravelly muddy sand
- gS _____ Gravelly sand
- G _____ Gravel
- mG _____ Muddy gravel
- msG _____ Muddy sandy gravel
- sG _____ Sandy gravel

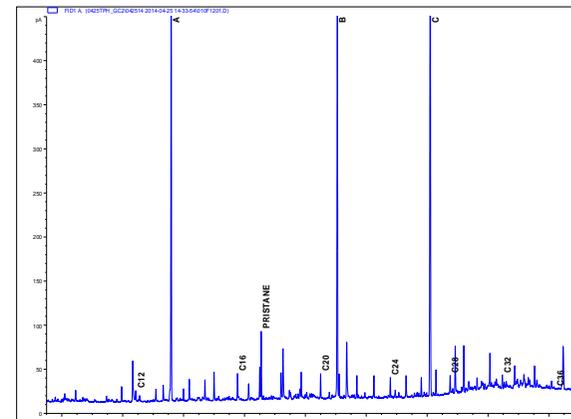
APPENDIX IV - GC-FID TRACES (SATURATES)



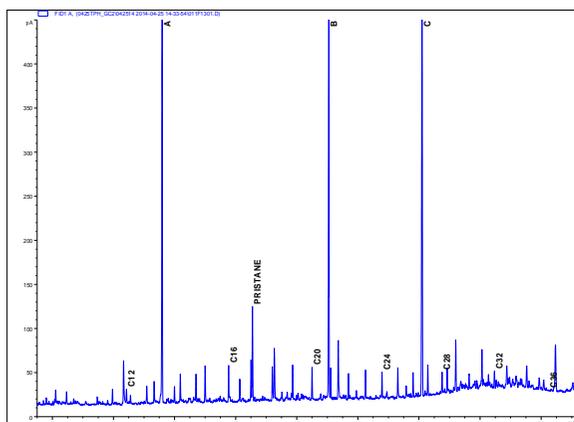
A/20/ENV



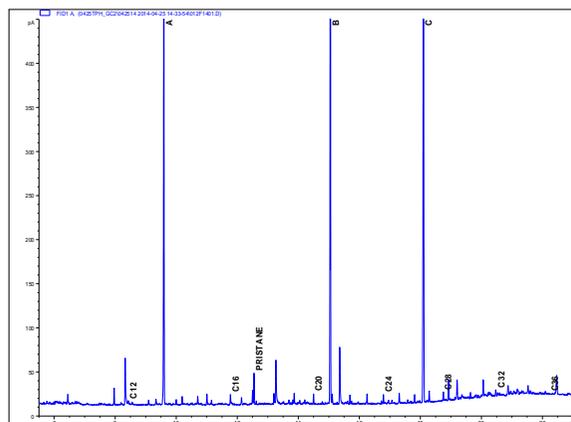
A/26/ENV



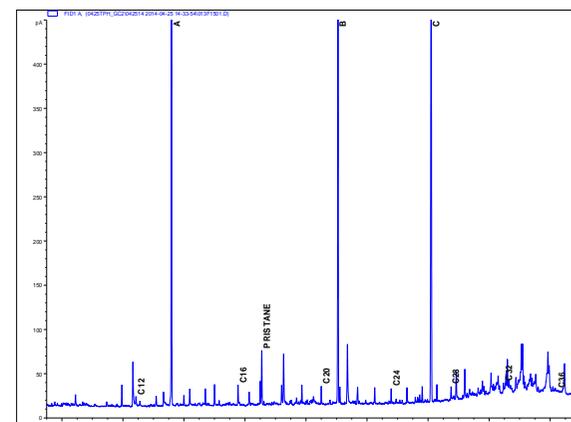
A/27/ENV



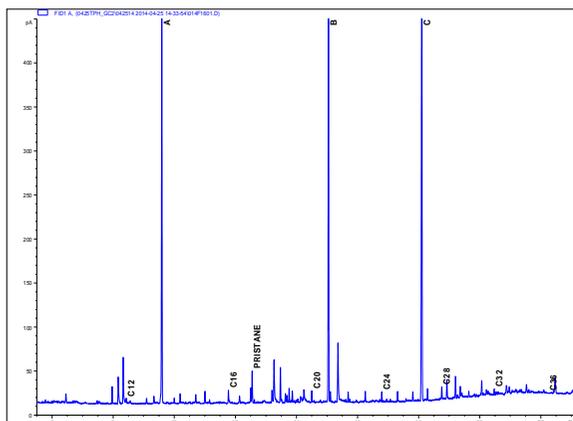
A/3/ENV



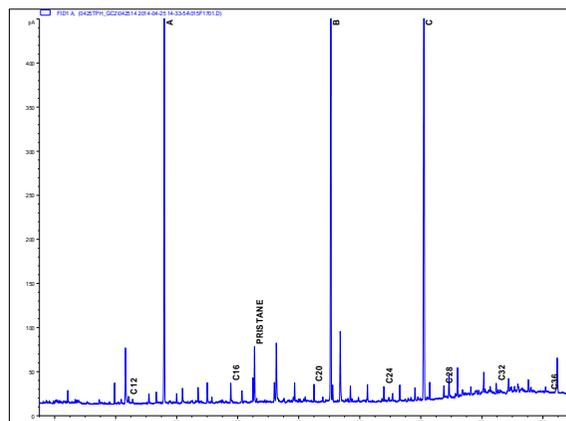
A/301/ENV



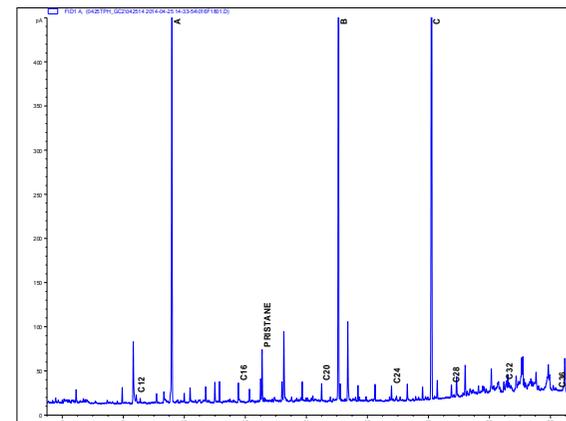
A/302/ENV



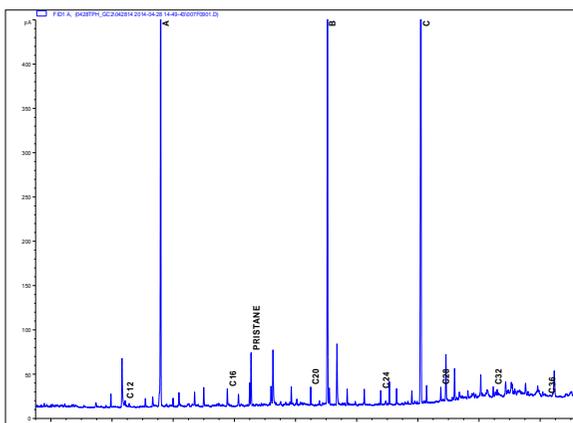
A/303/ENV



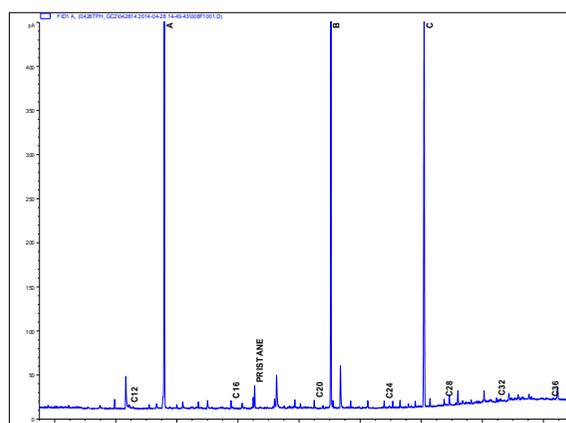
A/304/ENV



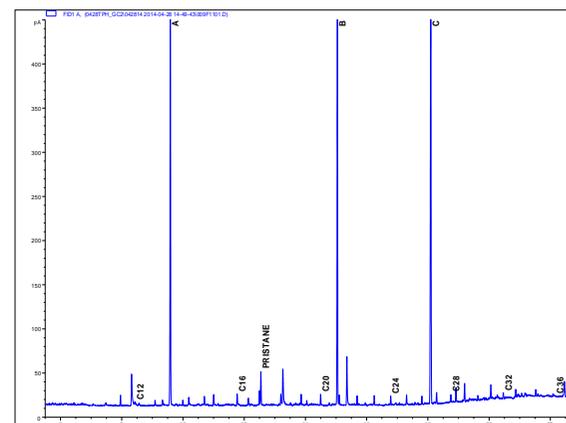
A/9/ENV



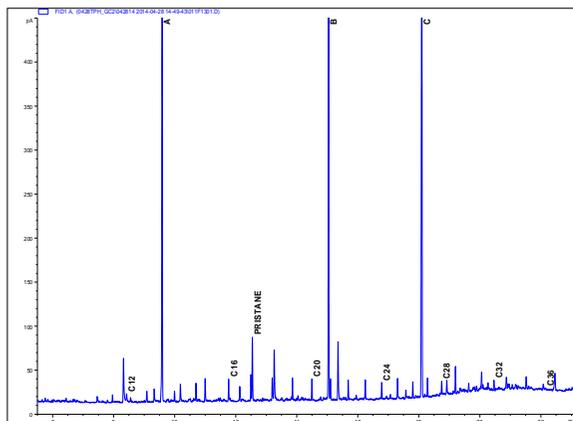
A/14/ENV



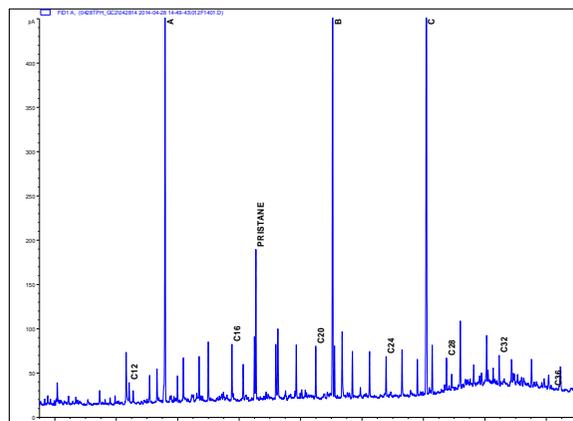
A/1011/ENV



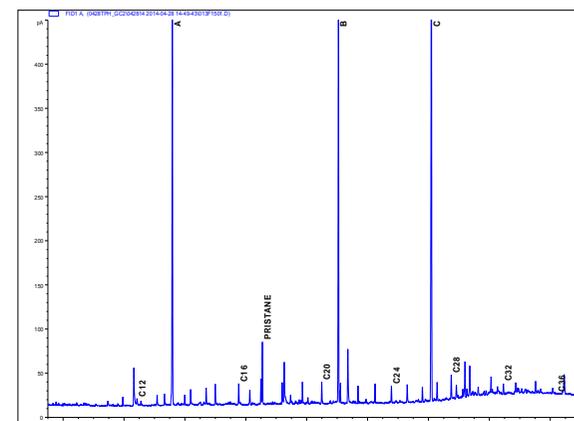
A/306/ENV



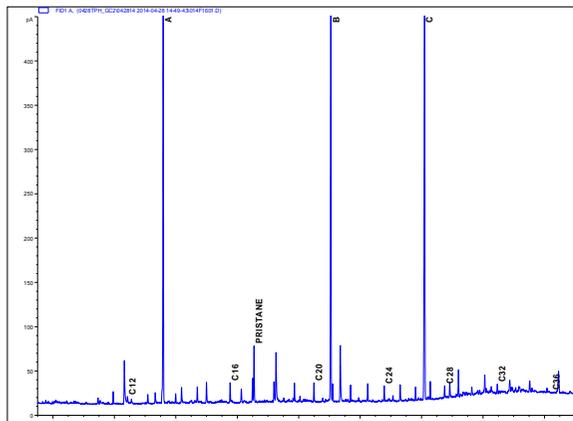
A/3010/ENV



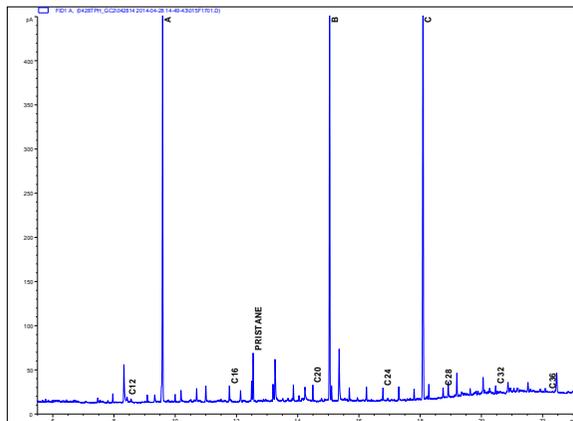
A/204/ENV



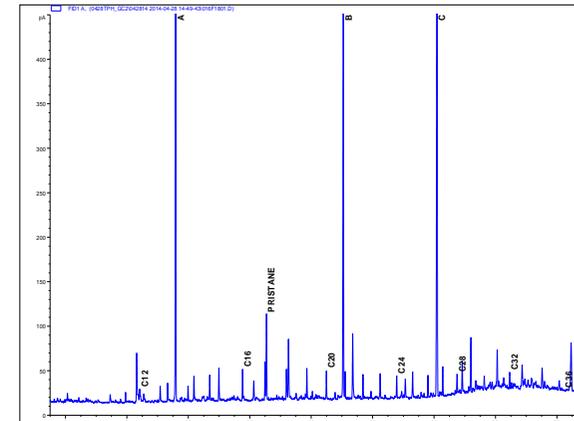
A/22/ENV



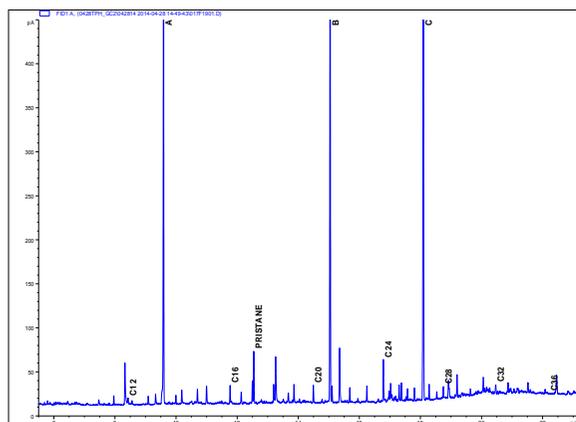
A/309/ENV



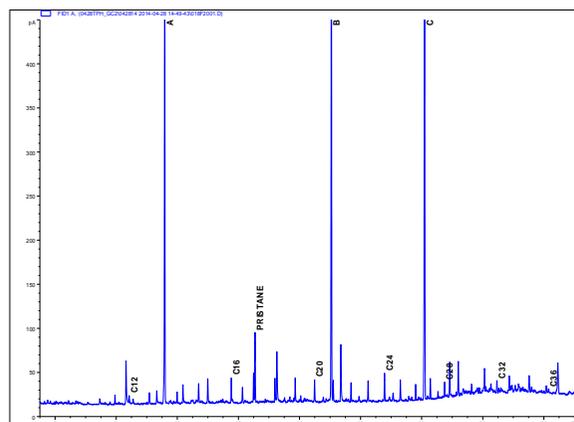
A/1008/ENV



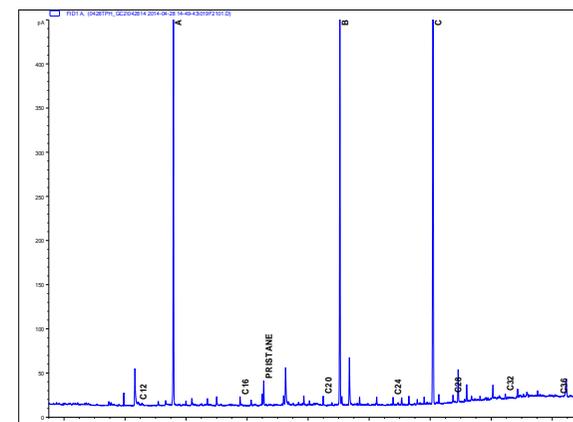
A/1015/ENV



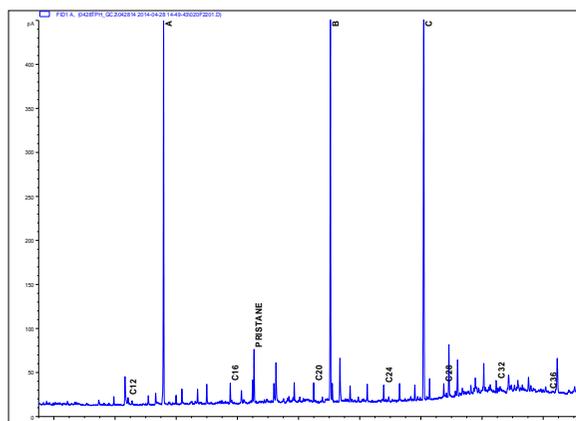
A/307/ENV



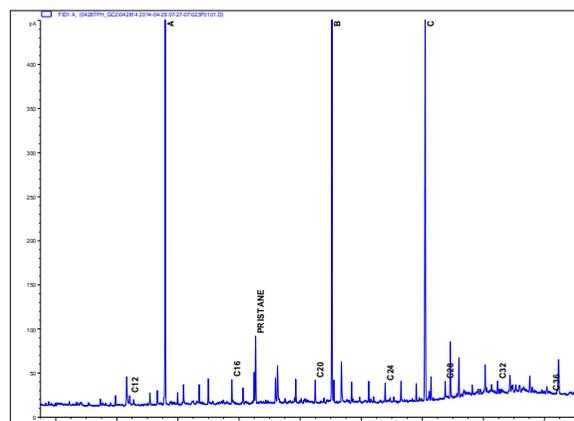
A/18/ENV



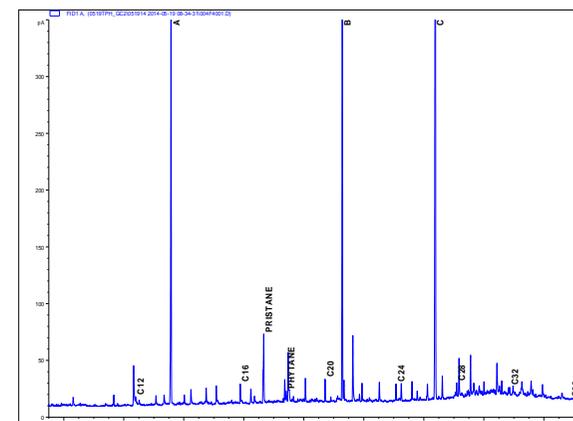
A/202/ENV



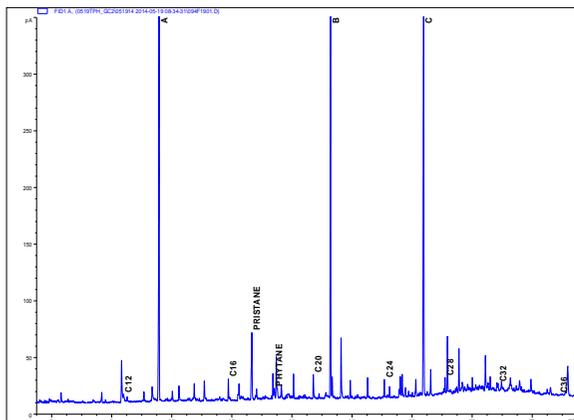
A/1013/ENV



A/10/ENV



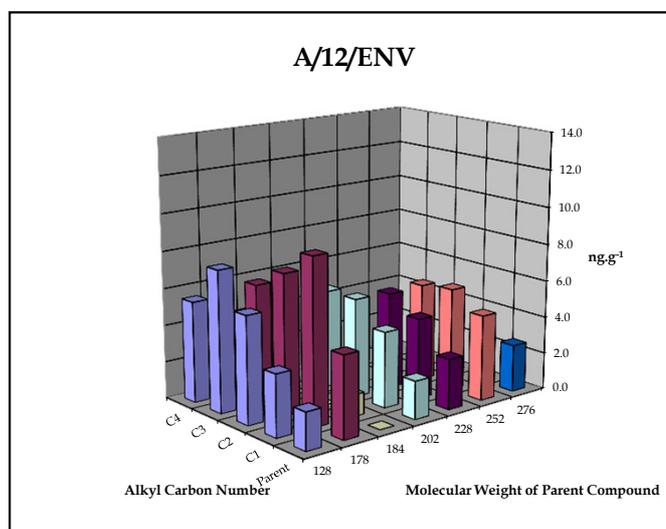
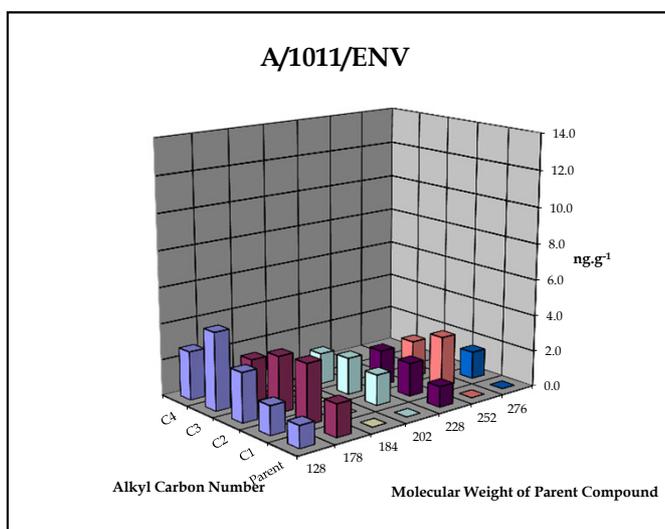
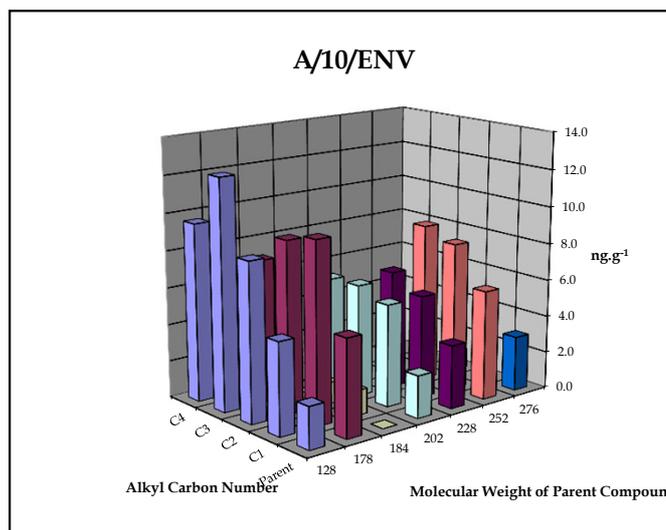
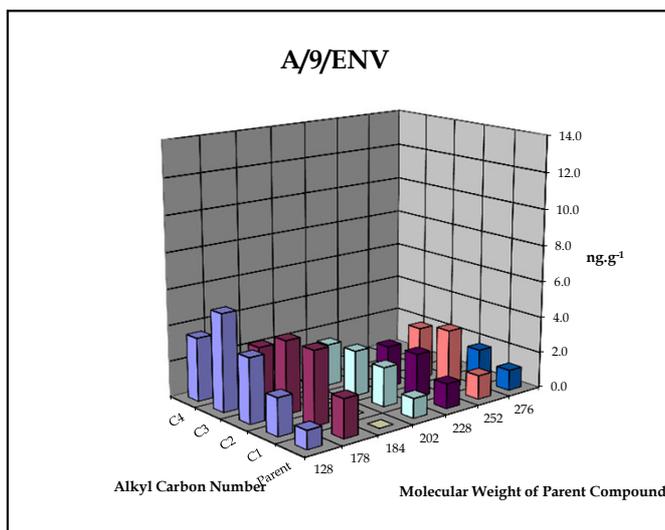
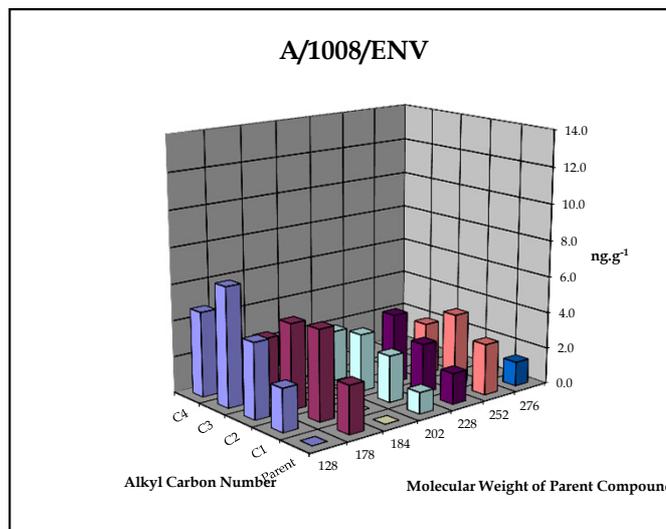
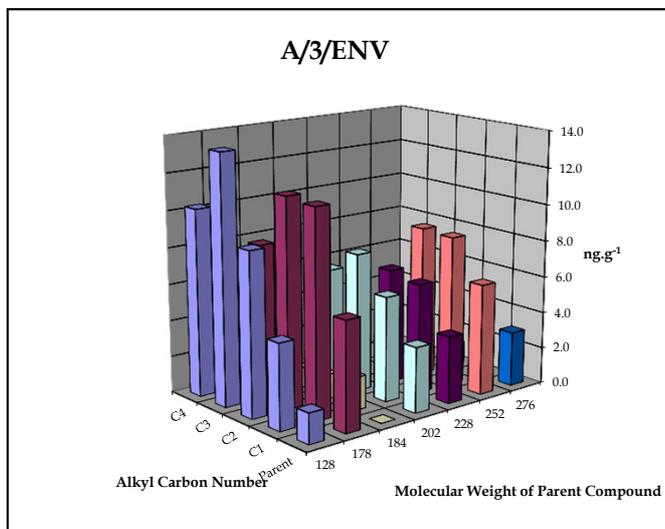
A/21/ENV



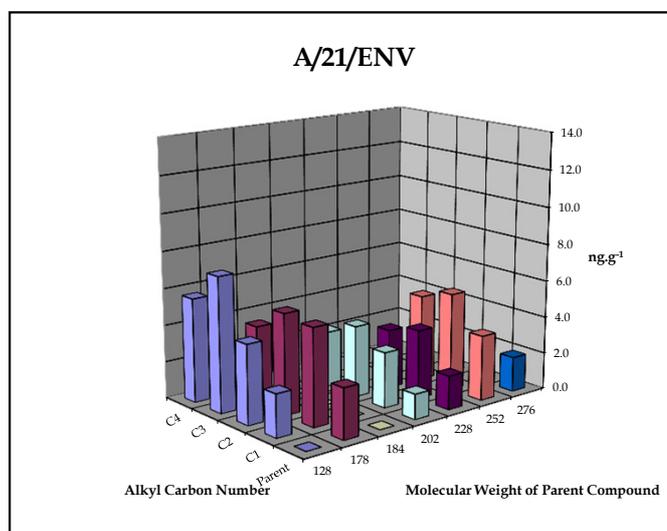
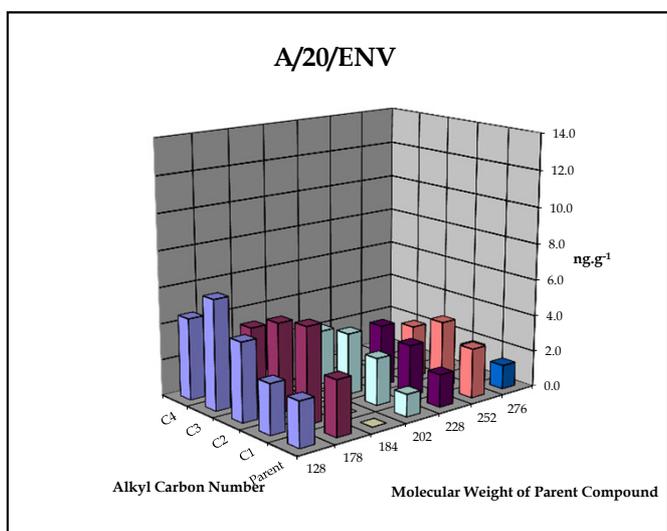
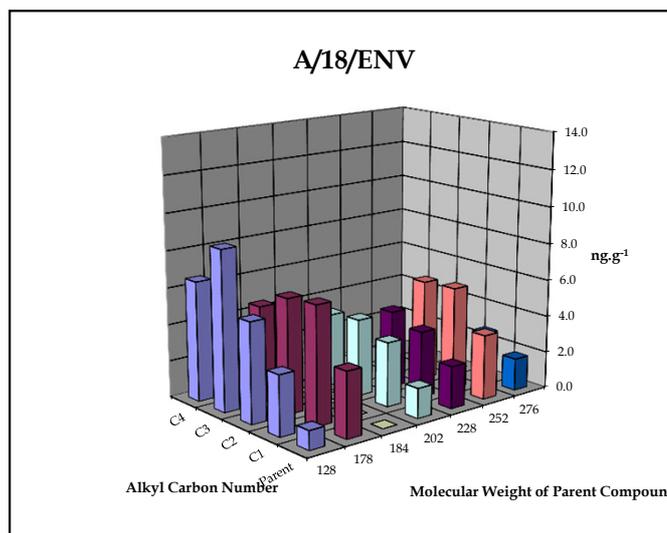
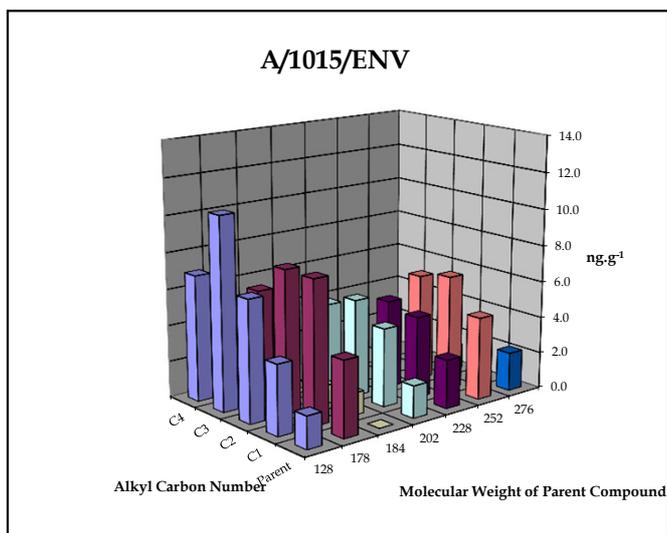
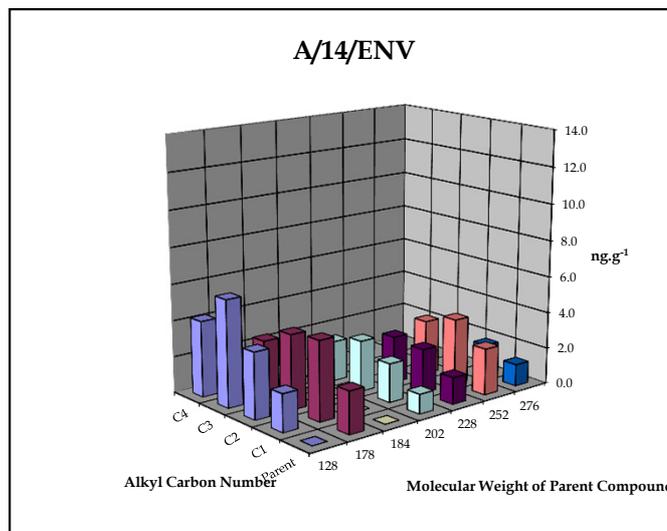
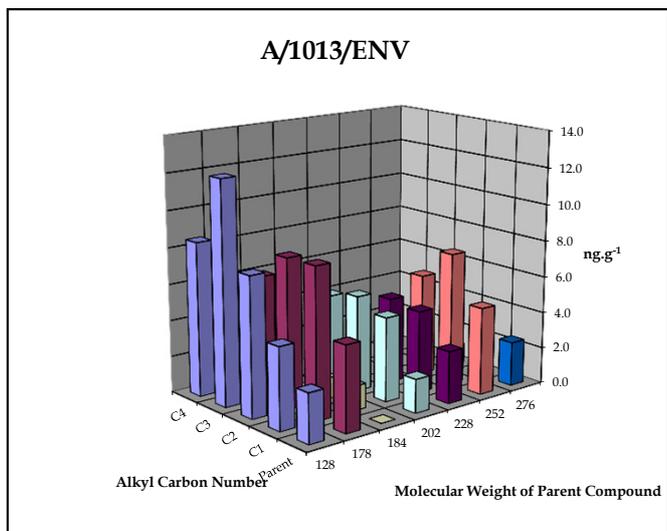
A/305/ENV

APPENDIX V - POLYCYCLIC AROMATIC HYDROCARBONS

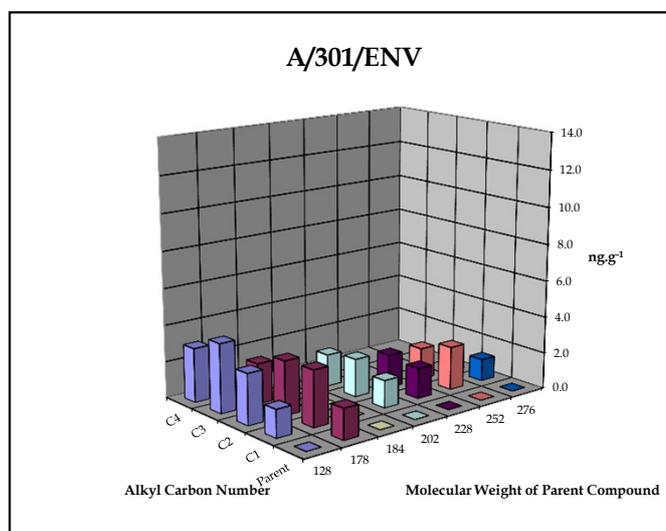
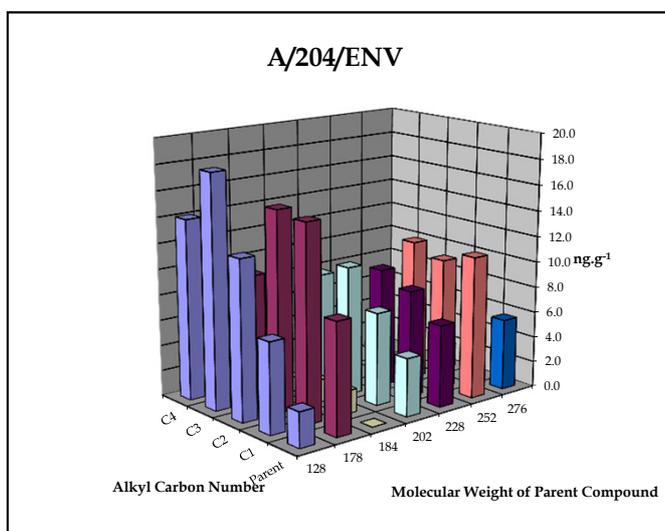
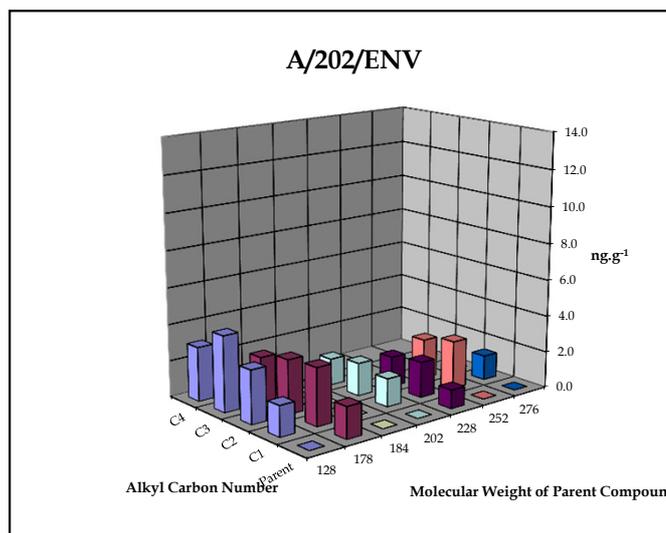
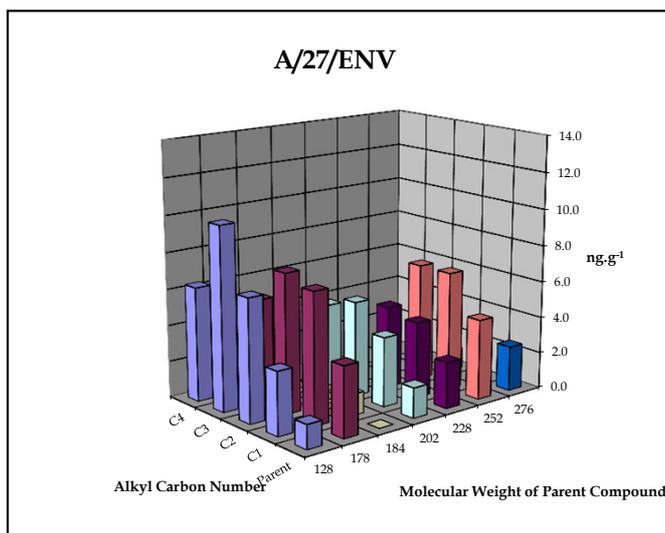
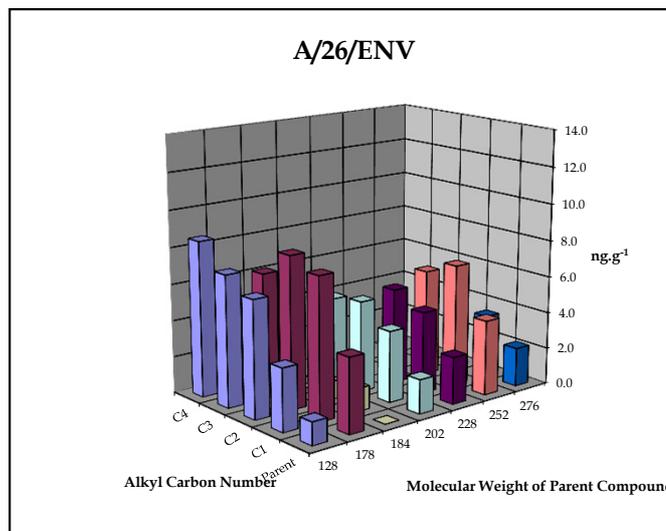
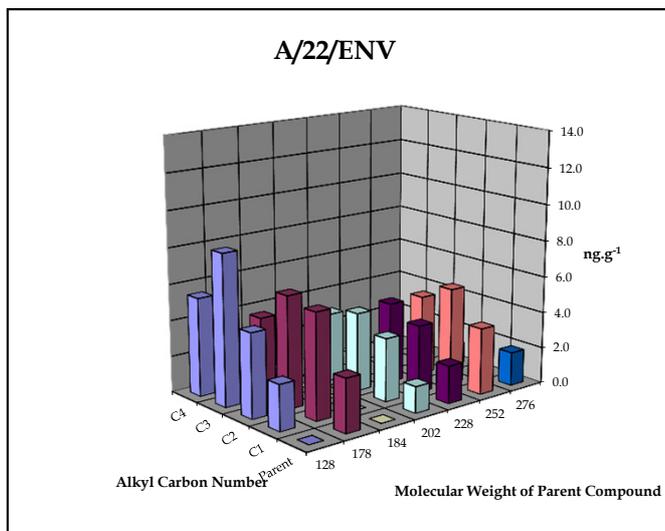
Polycyclic Aromatic Hydrocarbons - Parent and Alkylated Compounds



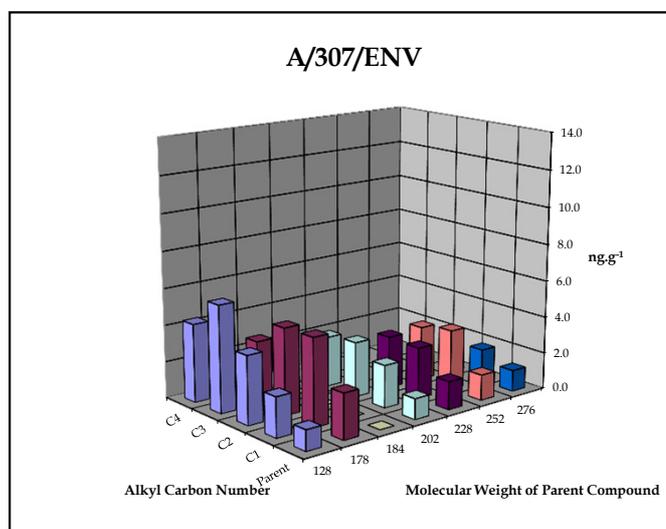
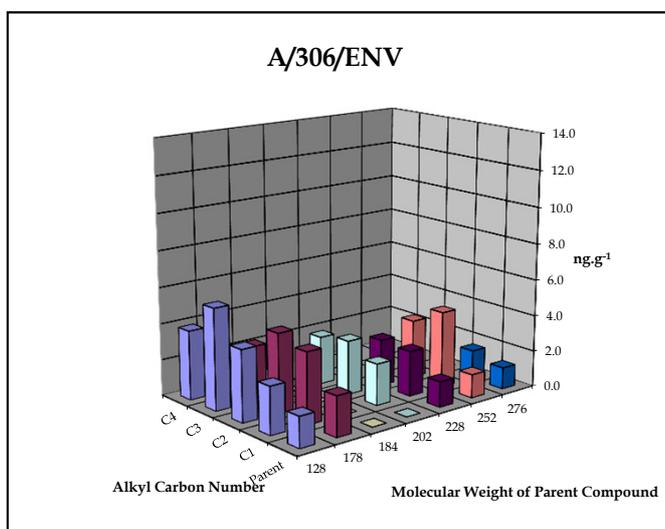
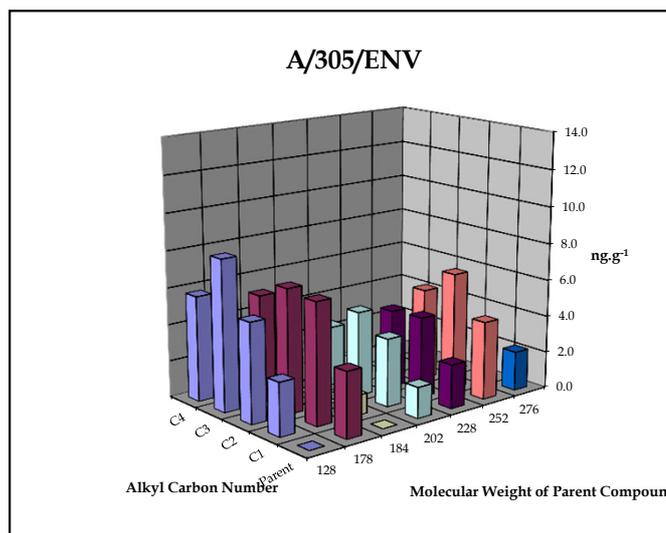
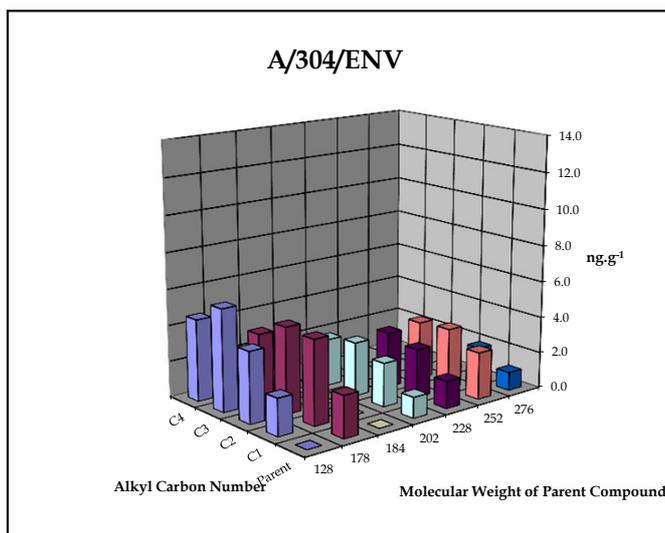
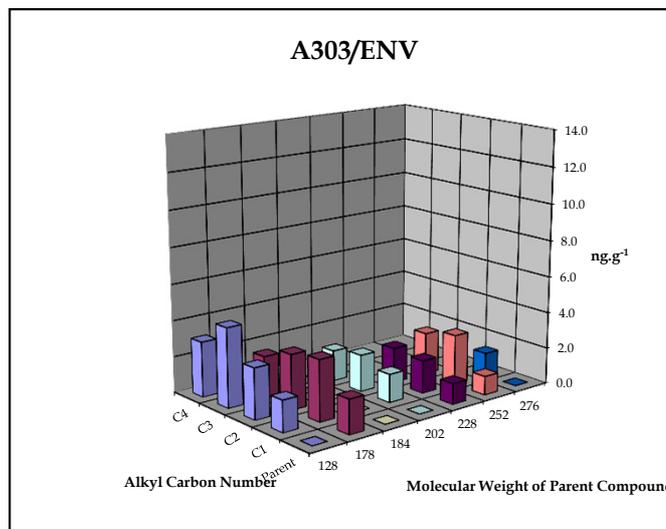
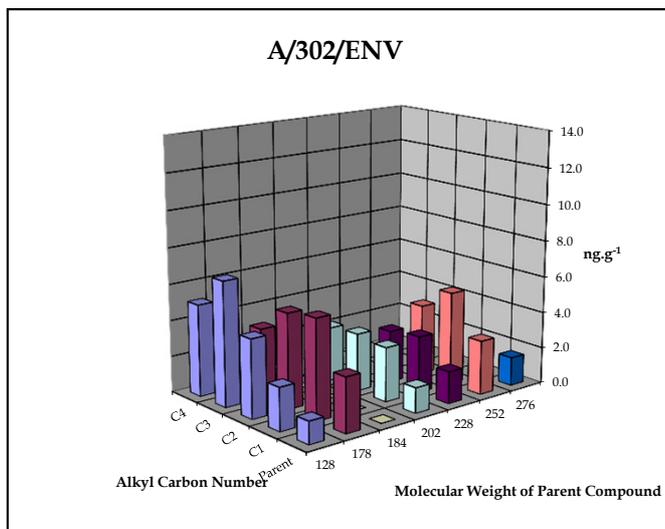
Polycyclic Aromatic Hydrocarbons - Parent and Alkylated Compounds



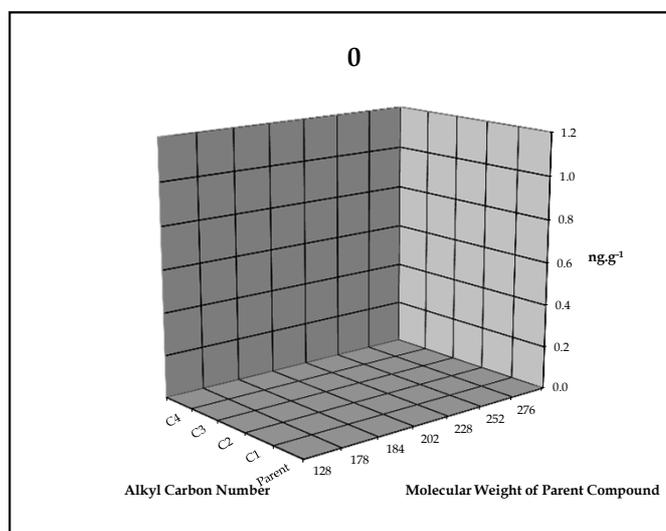
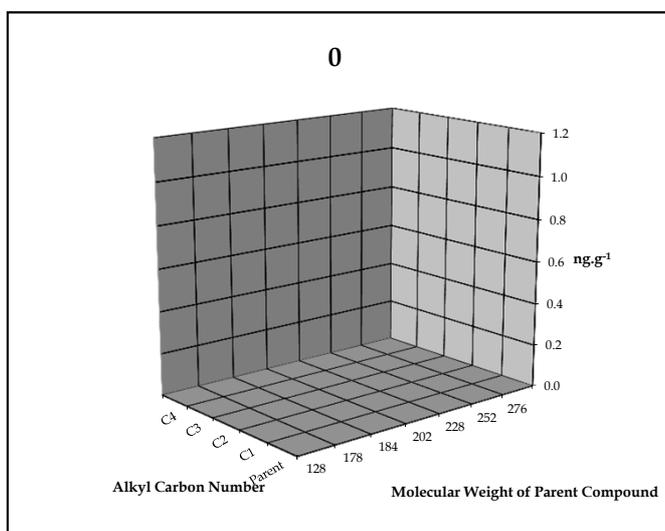
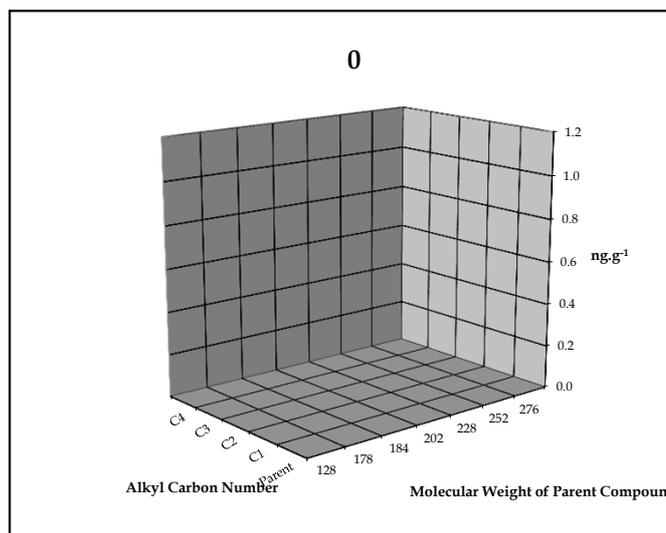
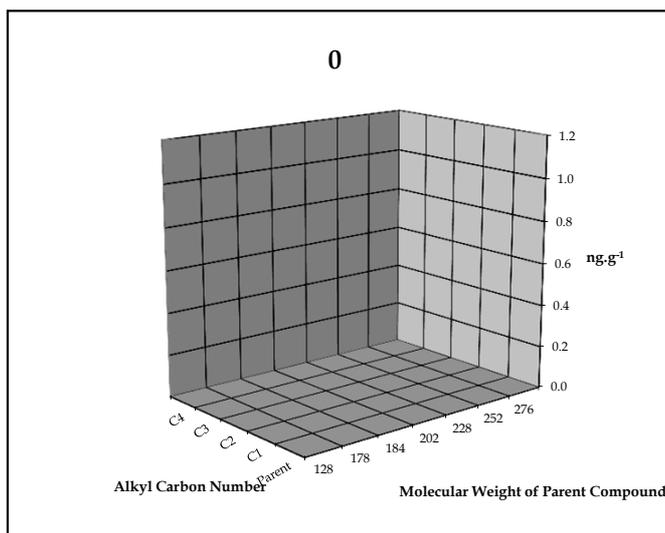
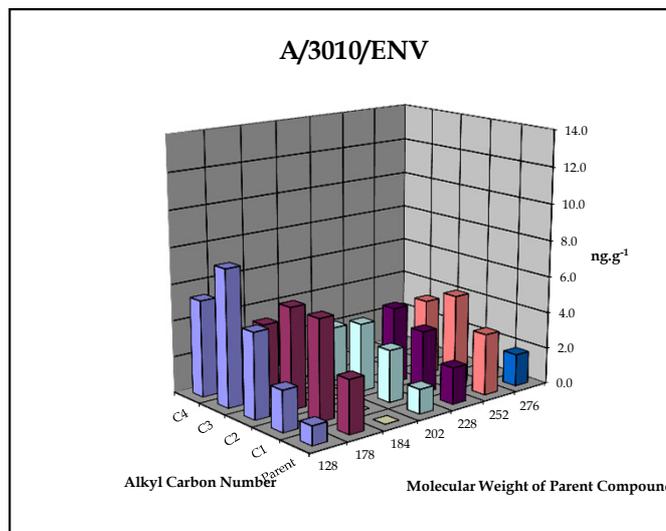
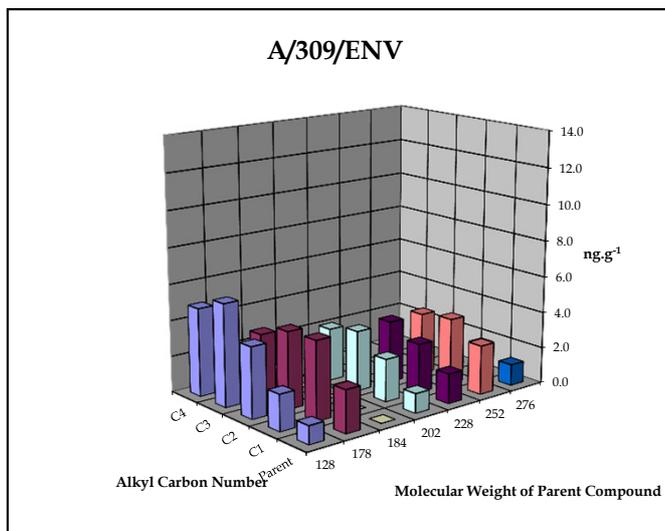
Polycyclic Aromatic Hydrocarbons - Parent and Alkylated Compounds



Polycyclic Aromatic Hydrocarbons - Parent and Alkylated Compounds



Polycyclic Aromatic Hydrocarbons - Parent and Alkylated Compounds





Polyaromatic Hydrocarbon Concentrations (ng/g dry weight basis) EPA 16 PAHs

| PAH Fraction | # PAH | Station : Mass | A/3/ENV | A/1008/EN | A/9/ENV | A/10/ENV | A/1011/ENV | A/12/EN | A/1013/EN | A/14/EN | A/1015/EN | A/18/EN | A/20/EN |
|---------------------------|--------------|---------------------------|---------|-----------|---------|----------|------------|---------|-----------|---------|-----------|---------|---------|
| Naphthalene | 1 | 128 | 1.7 | <1 | 1.0 | 2.3 | 1.2 | 2.0 | 2.7 | <1 | 1.8 | 1.0 | 2.5 |
| C1 Naphthalenes | | 142 | 4.7 | 2.3 | 2.1 | 5.0 | 1.6 | 3.4 | 4.5 | 2.1 | 3.8 | 3.3 | 2.7 |
| C2 Naphthalenes | | 156 | 8.9 | 4.2 | 3.6 | 8.6 | 2.7 | 5.8 | 7.6 | 3.7 | 6.7 | 5.4 | 4.3 |
| C3 Naphthalenes | | 170 | 13.6 | 6.6 | 5.4 | 12.4 | 4.3 | 7.7 | 12.2 | 5.9 | 10.6 | 8.7 | 6.1 |
| C4 Naphthalenes | | 184 | 10.2 | 4.7 | 3.5 | 9.6 | 2.7 | 5.5 | 8.5 | 4.2 | 6.9 | 6.5 | 4.5 |
| Sum Naphthalenes | | 39 | 18 | 16 | 38 | 13 | 24 | 36 | 16 | 30 | 25 | 20 | |
| Phenanthrene / Anthracene | 2 | 178 | 5.9 | 2.6 | 2.1 | 5.2 | 1.8 | 4.4 | 4.7 | 2.3 | 4.1 | 3.5 | 3.1 |
| C1 178 | | 192 | 11 | 5 | 4 | 10 | 3 | 9 | 8 | 4 | 8 | 6 | 5 |
| C2 178 | | 206 | 11 | 5 | 4 | 9 | 3 | 8 | 8 | 4 | 8 | 6 | 5 |
| C3 178 | | 220 | 8 | 3 | 3 | 8 | 2 | 6 | 7 | 3 | 6 | 5 | 4 |
| Sum 178 | | 37 | 16 | 13 | 32 | 11 | 27 | 28 | 14 | 26 | 21 | 17 | |
| Dibenzthiophene | | 184 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| C1 Dibenzthiophenes | | 198 | 2 | <1 | <1 | 1 | <1 | 1 | 1 | <1 | 1 | <1 | <1 |
| C2 Dibenzthiophenes | | 212 | 1 | <1 | <1 | 1 | <1 | <1 | 1 | <1 | 1 | <1 | <1 |
| C3 Dibenzthiophenes | | 226 | 1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| Sum Dibenzthiophenes | | 4 | 0 | 0 | 3 | 0 | 1 | 3 | 0 | 2 | 0 | 0 | |
| Fluoranthene / pyrene | 2 | 202 | 4 | 1 | 1 | 2 | <1 | 2 | 2 | 1 | 2 | 2 | 1 |
| C1 202 | | 216 | 6 | 3 | 2 | 6 | 2 | 4 | 5 | 2 | 4 | 4 | 3 |
| C2 202 | | 230 | 8 | 3 | 3 | 6 | 2 | 5 | 5 | 3 | 5 | 4 | 3 |
| C3 202 | | 244 | 6 | 3 | 2 | 6 | 2 | 5 | 5 | 2 | 5 | 4 | 3 |
| Sum 202 | | 23 | 10 | 8 | 20 | 5 | 17 | 17 | 8 | 16 | 13 | 10 | |
| Benzanthracene / chrysene | 2 | 228 | 4 | 2 | 1 | 3 | 1 | 3 | 3 | 1 | 3 | 2 | 2 |
| C1 228 | | 242 | 6 | 3 | 2 | 6 | 2 | 4 | 5 | 2 | 4 | 4 | 3 |
| C2 228 | | 256 | 6 | 4 | 2 | 6 | 2 | 5 | 5 | 3 | 5 | 4 | 3 |
| Sum 228 | | 16 | 8 | 6 | 15 | 5 | 12 | 12 | 7 | 12 | 10 | 8 | |
| Benzfluoranthenes / | 3 | 252 | 6 | 3 | 1 | 6 | <1 | 5 | 5 | 3 | 5 | 4 | 3 |
| C1 252 | | 266 | 8 | 4 | 3 | 8 | 3 | 6 | 7 | 4 | 6 | 6 | 4 |
| C2 252 | | 280 | 8 | 3 | 3 | 9 | 2 | 5 | 6 | 3 | 6 | 5 | 3 |
| Sum 252 | | 23 | 10 | 8 | 23 | 5 | 16 | 18 | 9 | 17 | 15 | 9 | |
| Aranthanthrenes / | 3 | 276 | 3 | 1 | 1 | 3 | <1 | 3 | 2 | 1 | 2 | 2 | 1 |
| C1 276 | | 290 | 1 | 2 | 2 | 4 | 2 | 3 | 3 | 2 | 3 | 3 | 2 |
| C2 276 | | 304 | 1 | <1 | <1 | 1 | <1 | <1 | 1 | <1 | 1 | <1 | <1 |
| Sum 276 | | 5 | 3 | 3 | 8 | 2 | 5 | 7 | 3 | 6 | 4 | 3 | |
| Sum of all fractions | | | 147 | 64 | 54 | 138 | 40 | 103 | 119 | 57 | 109 | 89 | 68 |
| Sum of NPD fraction | | | 80 | 33 | 29 | 72 | 23 | 53 | 66 | 30 | 58 | 46 | 37 |
| NPD / 4-6 ring PAH ratio | | | 1.20 | 1.09 | 1.18 | 1.10 | 1.37 | 1.06 | 1.24 | 1.10 | 1.14 | 1.09 | 1.22 |



Polyaromatic Hydrocarbon Concentrations (ng/g dry weight basis) EPA 16 PAHs

| PAH Fraction | # PAH | Station : Mass | A/21/ENV | A/22/ENV | A/26/EN | A/27/ENV | A/202/EN | A/204/ENV | A/301/EN | A/302/EN | A303/EN | A/304/EN |
|---------------------------|--------------|---------------------------|-----------------|-----------------|----------------|-----------------|-----------------|------------------|-----------------|-----------------|----------------|-----------------|
| Naphthalene | 1 | 128 | <1 | <1 | 1.2 | 1.3 | <1 | 2.7 | <1 | 1.2 | <1 | <1 |
| C1 Naphthalenes | | 142 | 2.3 | 2.5 | 3.4 | 3.5 | 1.7 | 7.0 | 1.5 | 2.4 | 1.7 | 2.1 |
| C2 Naphthalenes | | 156 | 4.3 | 4.6 | 6.4 | 6.7 | 2.9 | 12.4 | 2.8 | 4.3 | 2.8 | 3.9 |
| C3 Naphthalenes | | 170 | 7.3 | 8.3 | 7.2 | 10.1 | 4.2 | 18.1 | 3.8 | 6.9 | 4.4 | 5.7 |
| C4 Naphthalenes | | 184 | 5.7 | 5.4 | 8.5 | 6.3 | 3.0 | 14.1 | 3.0 | 5.1 | 3.1 | 4.5 |
| Sum Naphthalenes | | | 20 | 21 | 27 | 28 | 12 | 54 | 11 | 20 | 12 | 16 |
| Phenanthrene / Anthracene | 2 | 178 | 2.7 | 2.9 | 4.1 | 3.8 | 1.7 | 8.6 | 1.7 | 3.0 | 1.9 | 2.3 |
| C1 178 | | 192 | 5 | 6 | 8 | 7 | 3 | 15 | 3 | 6 | 3 | 5 |
| C2 178 | | 206 | 5 | 6 | 8 | 8 | 3 | 15 | 3 | 5 | 3 | 5 |
| C3 178 | | 220 | 4 | 4 | 7 | 6 | 3 | 10 | 2 | 4 | 2 | 4 |
| Sum 178 | | | 18 | 19 | 27 | 24 | 10 | 49 | 10 | 18 | 11 | 16 |
| Dibenzthiophene | | 184 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 | <1 |
| C1 Dibenzthiophenes | | 198 | <1 | <1 | 1 | 1 | <1 | 2 | <1 | <1 | <1 | <1 |
| C2 Dibenzthiophenes | | 212 | <1 | <1 | 1 | <1 | <1 | 2 | <1 | <1 | <1 | <1 |
| C3 Dibenzthiophenes | | 226 | <1 | <1 | <1 | <1 | <1 | 1 | <1 | <1 | <1 | <1 |
| Sum Dibenzthiophenes | | | 0 | 0 | 2 | 1 | 0 | 5 | 0 | 0 | 0 | 0 |
| Fluoranthene / pyrene | 2 | 202 | 1 | 1 | 2 | 2 | <1 | 5 | <1 | 1 | <1 | 1 |
| C1 202 | | 216 | 3 | 3 | 4 | 4 | 2 | 7 | 2 | 3 | 2 | 2 |
| C2 202 | | 230 | 4 | 4 | 5 | 5 | 2 | 10 | 2 | 3 | 2 | 3 |
| C3 202 | | 244 | 3 | 4 | 5 | 5 | 1 | 9 | 2 | 3 | 2 | 3 |
| Sum 202 | | | 11 | 13 | 15 | 15 | 5 | 31 | 5 | 11 | 5 | 9 |
| Benzanthracene / chrysene | 2 | 228 | 2 | 2 | 3 | 3 | 1 | 6 | <1 | 2 | 1 | 2 |
| C1 228 | | 242 | 4 | 4 | 4 | 4 | 2 | 8 | 2 | 3 | 2 | 3 |
| C2 228 | | 256 | 3 | 4 | 5 | 5 | 2 | 9 | 2 | 3 | 2 | 3 |
| Sum 228 | | | 9 | 10 | 12 | 11 | 5 | 24 | 3 | 8 | 5 | 7 |
| Benzfluoranthenes / | 3 | 252 | 4 | 4 | 4 | 4 | <1 | 11 | <1 | 3 | 1 | 3 |
| C1 252 | | 266 | 5 | 5 | 7 | 7 | 3 | 10 | 2 | 5 | 3 | 3 |
| C2 252 | | 280 | 5 | 4 | 6 | 6 | 2 | 11 | 2 | 4 | 2 | 3 |
| Sum 252 | | | 14 | 13 | 17 | 17 | 5 | 32 | 4 | 12 | 6 | 9 |
| Aranthranthenes / | 3 | 276 | 2 | 2 | 2 | 2 | <1 | 6 | <1 | 2 | <1 | 1 |
| C1 276 | | 290 | 2 | 2 | 3 | 3 | 1 | 5 | 1 | 2 | 1 | 2 |
| C2 276 | | 304 | <1 | 1 | <1 | 1 | <1 | 2 | <1 | <1 | <1 | <1 |
| Sum 276 | | | 4 | 5 | 6 | 7 | 1 | 13 | 1 | 3 | 1 | 3 |
| Sum of all fractions | | | 75 | 82 | 106 | 104 | 38 | 208 | 35 | 71 | 40 | 60 |
| Sum of NPD fraction | | | 37 | 40 | 56 | 53 | 22 | 108 | 21 | 37 | 23 | 32 |
| NPD / 4-6 ring PAH ratio | | | 0.99 | 0.96 | 1.12 | 1.06 | 1.42 | 1.09 | 1.49 | 1.12 | 1.28 | 1.11 |



Polyaromatic Hydrocarbon Concentrations (ng/g dry weight basis) EPA 16 PAHs

| <i>PAH Fraction</i> | <i># PAH</i> | <i>Station : Mass</i> | A/305/EN | A/306/EN | A/307/EN | A/309/EN | A/3010/EN |
|---------------------------|--------------|---------------------------|----------|----------|----------|----------|-----------|
| Naphthalene | 1 | 128 | <1 | 1.7 | 1.1 | 1.0 | 1.0 |
| C1 Naphthalenes | | 142 | 2.9 | 2.6 | 2.2 | 2.0 | 2.2 |
| C2 Naphthalenes | | 156 | 5.4 | 3.9 | 3.7 | 3.9 | 4.7 |
| C3 Naphthalenes | | 170 | 8.2 | 5.6 | 5.8 | 5.7 | 7.5 |
| C4 Naphthalenes | | 184 | 5.7 | 3.8 | 4.3 | 4.9 | 5.3 |
| Sum Naphthalenes | | | 22 | 18 | 17 | 17 | 21 |
| Phenanthrene / Anthracene | 2 | 178 | 3.5 | 2.2 | 2.5 | 2.3 | 2.9 |
| C1 178 | | 192 | 7 | 4 | 5 | 4 | 6 |
| C2 178 | | 206 | 7 | 4 | 5 | 4 | 6 |
| C3 178 | | 220 | 6 | 3 | 3 | 4 | 4 |
| Sum 178 | | | 23 | 14 | 15 | 14 | 18 |
| Dibenzthiophene | | 184 | <1 | <1 | <1 | <1 | <1 |
| C1 Dibenzthiophenes | | 198 | 1 | <1 | <1 | <1 | <1 |
| C2 Dibenzthiophenes | | 212 | <1 | <1 | <1 | <1 | <1 |
| C3 Dibenzthiophenes | | 226 | <1 | <1 | <1 | <1 | <1 |
| Sum Dibenzthiophenes | | | 1 | 0 | 0 | 0 | 0 |
| Fluoranthene / pyrene | 2 | 202 | 2 | <1 | 1 | 1 | 1 |
| C1 202 | | 216 | 4 | 2 | 2 | 2 | 3 |
| C2 202 | | 230 | 5 | 3 | 3 | 3 | 4 |
| C3 202 | | 244 | 3 | 3 | 3 | 3 | 3 |
| Sum 202 | | | 13 | 8 | 9 | 10 | 11 |
| Benanthracene / chrysene | 2 | 228 | 2 | 1 | 2 | 2 | 2 |
| C1 228 | | 242 | 4 | 3 | 3 | 3 | 3 |
| C2 228 | | 256 | 4 | 3 | 3 | 3 | 4 |
| Sum 228 | | | 11 | 6 | 7 | 8 | 10 |
| Benzfluoranthenes / | 3 | 252 | 4 | 1 | 1 | 3 | 3 |
| C1 252 | | 266 | 6 | 4 | 3 | 4 | 5 |
| C2 252 | | 280 | 5 | 3 | 3 | 3 | 4 |
| Sum 252 | | | 16 | 9 | 8 | 10 | 13 |
| Aranthanthrenes / | 3 | 276 | 2 | 1 | 1 | 1 | 2 |
| C1 276 | | 290 | 3 | 2 | 2 | 2 | 2 |
| C2 276 | | 304 | <1 | <1 | <1 | <1 | <1 |
| Sum 276 | | | 5 | 3 | 3 | 3 | 4 |
| Sum of all fractions | | | 91 | 57 | 59 | 62 | 76 |
| Sum of NPD fraction | | | 46 | 31 | 33 | 32 | 39 |
| NPD / 4-6 ring PAH ratio | | | 1.02 | 1.20 | 1.21 | 1.06 | 1.04 |

APPENDIX VI - SAMPLING LOG SHEETS



| Seabed Sampling (Deck Observations) | | | | | | | | | | | | | |
|-------------------------------------|------------|--------------------------------|-----------------|------------|------------|--------|---------------------------|--|-----------------|-----------------------|--|---------|------------------|
| Job No: | | BSL 1334 | | Operator: | | BSL | | Vessel: | | MV Poseidon | | Client: | Noble Energy Inc |
| Date: | | from: 31/12/13 to: 16/03/14 | | Project: | | FISA | | | | | | | |
| Sample Number | Station | Sampler Used | Water Depth (m) | Time (UTC) | Date | Volume | Sample Name | Comments | Sediment Colour | Sediment Description | Conspicuous Fauna/Comments | | |
| 1 | A/208/ENV | CAM | 1206 | 09:47 | 31/12/2013 | - | - | Time on overlay in local time (UTC -3h). Sonar cable coiled around camera frame, recovered to deck and switched top secondary winch. | - | - | - | | |
| 2 | A/208/ENV | CAM | 1200 | 13:45 | 31/12/2013 | - | - | Re-attempting previous camera location. | - | Fine sand and gravel. | White Branching bryozoan, dog whelk, holothurian, <i>Munida</i> , ophiuroids, Porifera, <i>Flustra</i> , Asteroidea, Anthozoa, pennatulids, annelids and Octocorallia. | | |
| 3 | A/09/ENV | CAM | 1260 | 23:15 | 31/12/2013 | - | - | - | - | Fine sand and gravel. | Pennatulids, cup coral, large isopod (possibly <i>Acanthoserolis schythei</i>), soft coral and large burrows. | | |
| 4 | A/10/ENV | CAM | 1261 | 02:50 | 01/01/2014 | - | - | Failed to locate beacon signal - recovered to deck. | - | - | - | | |
| 5 | A/10/ENV | CAM | 1261 | 03:10 | 01/01/2014 | - | - | Re-attempt of A/10/ENV - voltage leak - recovering - Two severed cables found and replaced (possibly severed by bridle shackles) - soft tow connection on dynema also broken by swell. | - | - | - | | |
| 6 | A/20/ENV | CAM | 1350 | 04:55 | 03/01/2014 | - | - | Failed deployment attempt due to weather/sea-state. | - | - | - | | |
| 7 | A/20/ENV | CAM | 1350 | 07:16 | 03/01/2014 | - | - | Re-attempt of A/20/ENV. | - | - | - | | |
| 8 | A/20/ENV | DVV | 1350 | 11:44 | 03/01/2014 | 35% | FB | N/S on one side and small amount recovered on the other. | - | - | - | | |
| 9 | A/20/ENV | DVV | 1350 | 12:30 | 03/01/2014 | N/S | N/S | Re-attempt of A/20/ENV DVV. | - | - | - | | |
| 10 | A/20/ENV | DVV | 1350 | 14:00 | 03/01/2014 | 70% | FA, PC (Supplementary FB) | Re-attempt of A/20/ENV DVV. | - | - | FB is supplementary fauna left from PC sample - Do not process first, - FA and FC should be proceed. | | |
| 11 | A/203/ENV | CAM | 1321 | 16:17 | 03/01/2014 | - | - | Attempt aborted due to cable twisting. | - | - | - | | |
| 12 | A/203/ENV | CAM | 1321 | 18:30 | 03/01/2014 | - | - | Re-attempt of A/203/ENV (No DVV). | - | - | - | | |
| 13 | A/1015/ENV | CAM | 1362 | 22:40 | 03/01/2014 | - | - | Cable issues, attempt aborted. | - | - | - | | |
| 14 | A/09/ENV | DVV | 1260 | 06:30 | 04/01/2014 | N/S | N/S | Failed attempt - no sample | - | - | - | | |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | | |
|-------------------------------------|-----------|----------|------|-----------|------------|-----|----------|--|--|--|---|--|
| Job No: | BSL 1334 | | | Operator: | BSL | | Vessel: | MV Poseidon | | Client: | Noble Energy Inc | |
| Date: | from: | 31/12/13 | | | | | Project: | FISA | | | | |
| | to: | 16/03/14 | | | | | | recovered. | | | | |
| 15 | A/09/ENV | DVV | 1260 | 07:35 | 04/01/2014 | 70% | FA, PC | Re-attempt - successful FA and PC. | Yellow brown, coarse black dusting on surface. | Fine sand and gravel. | Coarse dusting of black sediment over hard compact fine sand with gravel. | |
| 16 | A/09/ENV | DVV | 1260 | 08:50 | 04/01/2014 | N/S | N/S | FB and FC failed attempt - no sample recovered. | - | - | | |
| 17 | A/09/ENV | DVV | 1260 | 10:45 | 04/01/2014 | N/S | N/S | Failed attempt - no sample recovered. | - | - | | |
| 18 | A/10/ENV | CAM | 1261 | 13:00 | 04/01/2014 | - | - | No beacon detected at 90m, recovered to deck. | - | - | | |
| 19 | A/10/ENV | CAM | 1261 | 13:30 | 04/01/2014 | - | - | Re-attempt - successful. | - | - | | |
| 20 | A/10/ENV | DVV | 1261 | 16:20 | 04/01/2014 | 85% | FA, PC | - | Olive brown with speckled grey (sand). | Fine/medium sand with some small gravel. | Cup coral, ophiuroid, polychaetes, possible Foraminifera | |
| 21 | A/10/ENV | DVV | 1261 | 17:30 | 04/01/2014 | 80% | FB, FC | - | Olive brown | Fine/medium sand with some small gravel. | Bryozoan and tubeworms. | |
| 22 | A/12/ENV | CAM | 1280 | 19:20 | 04/01/2014 | - | - | - | - | - | | |
| 23 | A/12/ENV | DVV | 1280 | 22:00 | 04/01/2014 | 40% | FA, FC | FA and FC samples gained. | Olive brown with speckled grey (Sand) | - | Cup coral, calcareous bryozoans and polychaetes. | |
| 24 | A/12/ENV | DVV | 1280 | 23:00 | 04/01/2014 | N/S | N/S | Re-deployed for PC samples - failed. | - | - | | |
| 25 | A/12/ENV | DVV | 1280 | 00:30 | 05/01/2014 | N/S | N/S | Re-deployed for PC samples - failed. | - | - | | |
| 26 | A/26/ENV | CAM | 1260 | 02:03 | 05/01/2014 | - | - | No beacon detected at 50m, recovered to deck. | - | - | | |
| 27 | A/26/ENV | CAM | 1260 | 03:20 | 05/01/2014 | - | - | Beacon dropping in and out fixes taken when it comes in. | Cream with dark flecks. | Compact medium sand with some coarse sediment and drop stones. | Bryozoan, encrusting sponge and worm tubes. | |
| 28 | A/26/ENV | DVV | 1260 | 05:27 | 05/01/2014 | 70% | FA, FC | Beacon secured to cable 500m from grab to negate dropping out. | Cream with dark flecks. | Compact fine sands with mixed pebbles and cobbles. | | |
| 29 | A/26/ENV | DVV | 1260 | 06:45 | 05/01/2014 | 60% | PC, FB | Beacon replaced on grab - one large cobble in FB - no epifauna so pics then removed. | Cream with dark flecks. | Compact fine sands with mixed pebbles and cobbles. | | |
| 30 | A/207/ENV | CAM | 1250 | 09:40 | 05/01/2014 | - | - | Transect north of original. | - | Compact sands with mixed pebbles and cobbles. Larger dropstones with encrusting, sponges and soft coral. | | |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | | |
|-------------------------------------|------------|----------|------|-----------|------------|-----|----------|---|---------------------------------------|--|--|---|
| Job No: | BSL 1334 | | | Operator: | BSL | | Vessel: | MV Poseidon | | Client: | Noble Energy Inc | |
| Date: | from: | 31/12/13 | | | | | Project: | FISA | | | | |
| | to: | 16/03/14 | | | | | | | | | | |
| 31 | A/1011/ENV | CAM | 1315 | 11:07 | 05/01/2014 | - | - | N/S | - | - | - | - |
| 32 | A/1011/ENV | DVV | 1315 | 14:30 | 05/01/2014 | N/S | N/S | N/S, down on weather. | - | - | - | - |
| 33 | A/304/ENV | CAM | 1250 | 14:40 | 08/01/2014 | - | - | - | - | - | - | - |
| 34 | A/304/ENV | DVV | 1250 | 17:44 | 08/01/2014 | 80% | PC, FC | PC and FC samples gained. | Cream with dark flecks. | Fine to medium sand with occasional fine gravel. | Scaphopoda tusk shell, polychaetes, worm tubes and pycnogonid. | - |
| 35 | A/304/ENV | DVV | 1250 | 18:59 | 08/01/2014 | N/S | N/S | N/S | - | - | - | - |
| 36 | A/304/ENV | DVV | 1250 | 20:03 | 08/01/2014 | 80% | FA/FB | FA and FB samples gained. | Cream with dark flecks. | Fine to medium sand with occasional fine gravel. | Scaphopoda tusk shell, polychaete, worm tubes, urchin, ophiuroid and bivalve shells. | - |
| 37 | A/301/ENV | CAM | 1250 | 23:05 | 08/01/2014 | - | - | Coral garden and concretion. Cable re-term. | - | - | - | - |
| 38 | A/301/ENV | DVV | 1250 | 01:50 | 09/01/2014 | 70% | FC/PC | FC slightly washed out. | Cream with dark flecks. | Darker coarser surface layer then fine to medium compact sand with occasional fine gravel. | Scaphopoda tusk shells, bivalve shells and tube casts. | - |
| 39 | A/301/ENV | DVV | 1250 | 03:30 | 09/01/2014 | - | N/S | Small amount of sand as grab bounced on seabed. | - | - | - | - |
| 40 | A/301/ENV | DVV | 1250 | 04:15 | 09/01/2014 | - | N/S | Small amount of sand as grab bounced on seabed. | - | - | - | - |
| 41 | A/27/ENV | CAM | 1260 | 06:45 | 09/01/2014 | - | - | Cable re-term after drop. | Cream with dark flecks. | Boulders with epifauna, fine and compact sand. | - | - |
| 42 | A/27/ENV | DVV | 1260 | 09:00 | 09/01/2014 | 80% | FA/PC | FA and PC samples gained. | Cream with darker surface layer. | Darker, coarse surface layer over cream fine to med compact sands. | Worm tubes, polychaetes, bivalve shells and gastropods. | - |
| 43 | A/27/ENV | DVV | 1260 | 10:10 | 09/01/2014 | 85% | FB/FC | FC and FB samples gained. | Cream with darker surface layer. | Darker, coarse surface layer over cream fine to med compact sands. | Bryozoan, worm tubes, ophiuroids, polychaetes, bivalve shells, gastropod | - |
| 44 | A/303/ENV | CAM | 1285 | 11:40 | 09/01/2014 | - | - | Flash and winch issues but enough images taken. | - | - | - | - |
| 45 | A/303/ENV | DVV | 1285 | 15:00 | 09/01/2014 | 80% | FA/PC | FA and PC samples gained. | Cream with darker surface layer. | Darker, coarse surface layer over cream fine to med compact sands | Cup coral fragments, worm tubes and polychaetes. | - |
| 46 | A/303/ENV | DVV | 1285 | 16:00 | 09/01/2014 | 45% | FB/FC | FB and FC samples gained. | Mostly dark, some lighter grey. | Dark coarse gravel and rocks then fine to medium sand component. | Worm tubes, polychaetes and gastropods. | - |
| 47 | A/302/ENV | CAM | 1330 | 18:40 | 09/01/2014 | - | - | - | - | - | - | - |
| 48 | A/302/ENV | DVV | 1330 | 22:30 | 09/01/2014 | 80% | FA/FC | FA and FC samples gained. | Olive grey with darker surface layer. | Mostly clay with flecks of dark gravel. | Polychaetes, worm tubes and ophiuroid. | - |
| 49 | A/302/ENV | DVV | 1330 | 23:20 | 09/01/2014 | 80% | PC/NS | PC gained and N/S. | - | - | - | - |
| 50 | A/302/ENV | DVV | 1330 | 23:53 | 09/01/2014 | 30% | FC(2) | One very small washed out | Dark. | Largely dark medium | Ascidian, polychaetes and bivalve | - |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | |
|-------------------------------------|------------|----------|------|-----------|------------|-----|--------------|---|--|--|--|
| Job No: | BSL 1334 | | | Operator: | BSL | | Vessel: | MV Poseidon | | Client: | Noble Energy Inc |
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| | | | | | | | | sample - likely bounced - taken as FC2. | | sands with some large pebbles/gravel. | shell. |
| 51 | A/302/ENV | DVV | 1330 | 00:50 | 10/01/2014 | 60% | FB | - | Dark surface layer, cream below. | Darker, coarse surface layer over cream fine to med compact sands. | Polychaetes, possible echiuran, bivalve and gastropod shells. |
| 52 | A/305/ENV | CAM | 1267 | 04:45 | 12/01/2014 | - | - | No beacon signal at 50m - recovered too deck. | - | - | - |
| 53 | A/305/ENV | CAM | 1267 | 06:04 | 12/01/2014 | - | - | Redeployment. | White cream sands - homogenous. | Darker, coarse surface layer over cream fine to medium compact sands - disturbance on surface. | Cup corals and crustacean. |
| 54 | A/305/ENV | BC | 1267 | 09:26 | 12/01/2014 | 60% | FA/FB/PC/ARC | FA/FB/PC/ Archive core gained. | Dark surface layer, cream below. | Darker, coarse surface layer over cream fine to medium compact sands. | Isopod (FB), polychaetes and tubes. |
| 55 | A/306/ENV | CAM | 1350 | 13:11 | 12/01/2014 | - | - | - | - | - | - |
| 56 | A/306/ENV | HG | 1350 | 15:00 | 12/01/2014 | 70% | FA | FA sample gained. | Dark with light grey clay. | Coarse pebbles and rocks suspended in clay. | Bryozoans, worm tubes, polychaetes, bivalve shells. |
| 57 | A/306/ENV | HG | 1350 | 16:30 | 12/01/2014 | 60% | PC | PC sample gained. | Dark with light grey clay. | Coarse pebbles and rocks suspended in clay. | - |
| 58 | A/306/ENV | HG | 1350 | 17:30 | 12/01/2014 | 70% | FB | FB sample gained. | Dark with light grey clay. | Coarse pebbles and rocks suspended in clay. | Bryozoans, ophiuroid, worm tubes and polychaetes. |
| 59 | A/1013/ENV | CAM | 1350 | 20:12 | 12/01/2014 | - | - | - | - | - | - |
| 60 | A/1013/ENV | HG | 1350 | 22:00 | 12/01/2014 | 75% | FA | FA sample gained. | Freckled black sand with cement coloured <i>Lophelia</i> . | Clay, sand and dead <i>Lophelia</i> . | Dead <i>Lophelia</i> and worm tubes. |
| 61 | A/1013/ENV | HG | 1350 | 23:00 | 12/01/2014 | N/S | N/S | No sample. | - | - | - |
| 62 | A/1013/ENV | HG | 1350 | 23:38 | 12/01/2014 | 75% | PC | PC sample gained. | Freckled black sand with cement coloured <i>Lophelia</i> . | Hard compact sand and dead <i>Lophelia</i> . | Dead <i>Lophelia</i> . |
| 63 | A/1013/ENV | HG | 1350 | 00:55 | 13/01/2014 | - | FB | FB sample gained. | Freckled black sand with cement coloured <i>Lophelia</i> . | Hard compact sand and dead <i>Lophelia</i> . | Dead <i>Lophelia</i> , urchin spine, polychaetes, worm tubes, bivalve shells. |
| 64 | A/1008/ENV | CAM | 1280 | 02:23 | 13/01/2014 | - | - | - | - | - | - |
| 65 | A/1008/ENV | HG | 1280 | 05:54 | 13/01/2014 | 80% | FA | FA sample gained. | Beige sand with black specks. | Hard compact coarse sands with cobbles and dead <i>Lophelia</i> . | Ophiuroids, polychaetes, bivalve shells, dead <i>Lophelia</i> and urchin spines. |
| 66 | A/1008/ENV | DVV | 1280 | 06:43 | 13/01/2014 | 70% | PC/FB | FB/PC sample gained. | Beige sand with | Soft sands with cobbles. | - |



Seabed Sampling (Deck Observations)

| Job No: | BSL 1334 | | | Operator: | BSL | Vessel: | MV Poseidon | Client: | Noble Energy Inc | | |
|---------|------------|----------|------|-----------|------------|----------|--------------|---|---|--|---|
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| 67 | A/1015/ENV | CAM | 1358 | 07:40 | 13/01/2014 | - | - | - | black specks. | - | - |
| 68 | A/1015/ENV | BC | 1358 | 12:00 | 13/01/2014 | - | FA/FB/PC/ARC | FA/FB/PC/ Archive core gained. | Dark surface layer with light grey clay. | Coarse pebbles and rocks suspended in clay. | Tunicata, bivalve shells, tubeworms, polychaetes, ophiuroids and bryozoans. |
| 69 | A/14/ENV | CAM | 1316 | 12:55 | 13/01/2014 | - | - | Camera deployed then retrieved after signal failure. Re-term. | - | - | - |
| 70 | A/14/ENV | CAM | 1316 | 14:05 | 13/01/2014 | - | - | Redeployed. | - | - | - |
| 71 | A/14/ENV | BC | 1316 | 16:05 | 13/01/2014 | 55% | FA/FB/PC/ARC | FA/FB/PC/ Archive core gained. | Dark surface layer with mid-grey clay. | Coarse pebbles and rocks suspended in clay. | Bryozoans, calcified bryozoans, worm tubes, polychaetes, bivalve shells. |
| 72 | A/308/ENV | CAM | 1380 | 17:40 | 13/01/2014 | - | - | - | - | - | - |
| 73 | A/308/ENV | BC | 1380 | 19:53 | 13/01/2014 | N/S | N/S | Box corer damaged by large rock. No sample. | Dark surface layer with mid-grey clay. | Coarse pebbles and rocks suspended in clay. | Bryozoans, calcified bryozoans, worm tubes, polychaetes, bivalve shells. |
| 74 | A/309/ENV | CAM | 1395 | 21:30 | 13/01/2013 | - | - | - | - | - | - |
| 75 | A/309/ENV | BC | 1395 | 23:44 | 13/01/2012 | 30% | FA/FB/PC | Good sample although small. | Dark speckled. | Dark compact medium sand. | Tubeworm casts, tubeworms, gastropod shell, bryozoan. |
| 76 | A/21/ENV | CAM | 1430 | 02:00 | 14/01/2014 | - | - | - | - | - | - |
| 77 | A/21/ENV | BC | 1430 | 04:11 | 14/01/2014 | 30% | FA/FB/PC | Good sample although small. | Dark homogenous. | Dark compact medium sand. | Worm tubes, gastropod shell, bivalve shell. |
| 78 | A/307/ENV | CAM | 1400 | 06:35 | 14/01/2014 | - | - | - | - | - | - |
| 79 | A/307/ENV | BC | 1400 | 08:54 | 14/01/2014 | N/S | N/S | BC failed to trigger at seafloor. | - | - | - |
| 80 | A/307/ENV | BC | 1400 | 10:16 | 14/01/2014 | - | FA/FB/PC | Redeployment - no sieve photo for FB. | - | - | - |
| 81 | A/12/ENV | HG | 1280 | 13:20 | 14/01/2014 | - | PC | - | - | - | - |
| 82 | A/1011/ENV | BC | 1325 | 14:57 | 14/01/2014 | 30% | FA/FB/PC/ARC | - | Dark compact med clay/sand with thin cream surface layer. | Dark compact medium clay/sand with thin cream surface layer. | - |
| 83 | A/301/ENV | HG | 1250 | 16:47 | 14/01/2014 | 40% | FA | - | Light/olive grey sediment. | Broken rock fragments and hard clay substrate with fauna attached. | - |
| 84 | A/301/ENV | HG | 1250 | 17:59 | 14/01/2014 | - | - | HG failed to trigger at seafloor. | - | - | - |
| 85 | A/202/ENV | CAM | 1153 | 22:35 | 14/01/2014 | - | - | - | - | - | - |
| 86 | A/202/ENV | BC | 1153 | 00:50 | 15/01/2014 | - | FA/FB/PC/ARC | - | Light/olive grey sediment with dark surface layer. | Hard compact medium sand with clay lumps. | Polychaetes, worm tubes, bryozoan, sponge, asteroid. |
| 87 | A/201/ENV | CAM | 1120 | 04:15 | 15/01/2014 | - | - | - | Hard compact fine sands with | - | - |



Seabed Sampling (Deck Observations)

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| Date: | from: | 31/12/13 | | | | Project: | FISA | | | | |
| | to: | 16/03/14 | | | | | | | cobbles and boulders. | | |
| 88 | A/106/ENV | CAM | 1302 | 20:55 | 16/01/2014 | - | - | - | - | - | - |
| 89 | A/106/ENV | BC | 1302 | 22:52 | 16/01/2014 | N/S | N/S | Failed to trigger. | - | - | - |
| 90 | A/106/ENV | BC | 1302 | 23:40 | 16/01/2014 | N/S | N/S | Insufficient sample. | - | - | - |
| 91 | A/106/ENV | BC | 1302 | 00:41 | 17/01/2014 | N/S | N/S | Triggered in water column/bounced. | - | - | - |
| 92 | A/204/ENV | CAM | 1302 | 04:00 | 17/01/2014 | - | - | - | - | - | - |
| 93 | A/204/ENV | BC/NISKIN | 1302 | 06:18 | 17/01/2014 | 55% | FA/FB/PC/ARC | Niskin failed to trigger. | Light/olive grey sediment with dark surface layer. | Hard compact medium sand with clay from 10cm down. | Dead <i>Lophelia</i> , polychaetes, worm tubes, anemones and gastropod shells. |
| 94 | A/11/GEO | 6m GC | 1500 | 09:48 | 17/01/2014 | N/S | N/S | Bent barrel. | - | - | - |
| 95 | A/12/GEO | 6m GC | 1340 | 13:14 | 17/01/2014 | N/S | N/S | Bent barrel. | - | - | - |
| 96 | A/09/GEO | CAM | 1070 | 17:25 | 17/01/2014 | - | - | - | - | - | - |
| 97 | A/09/GEO | 3m GC | 1070 | 19:13 | 17/01/2014 | 250cm | 2xHSG, 2xLPH, MPOG, Eh, Archive | - | - | - | - |
| 98 | A/10A/GEO | 3m GC | 1217 | 21:05 | 17/01/2014 | 30cm | Archive | - | - | - | - |
| 99 | A/10B/GEO | 3m GC | 1217 | 21:57 | 17/01/2014 | 30cm | Archive | - | - | - | - |
| 100 | A/21/GEO | 3m GC | 1304 | 23:42 | 17/01/2014 | 22cm | Archive | - | Brown/grey | Wet, stiff clay over hard dried clay/mud. | - |
| 101 | A/20/GEO | 3m GC | 1300 | 01:57 | 18/01/2014 | 102cm | HSG, LPH, MPOG, Eh | - | Dark grey. | Firm fine sands to muds - some clay. | - |
| 102 | A/12B/GEO | 3m GC | 1340 | 04:00 | 18/01/2014 | 62cm | LPH, MPOG, Eh | - | Grey to dark grey. | Firm fine sands to muds. | - |
| 103 | A/12C/GEO | 3m GC | 1340 | 04:59 | 18/01/2014 | 55cm | Archive | - | Grey to dark grey. | Firm fine sands to muds. | - |
| 104 | A/11B/GEO | 3m GC | 1500 | 07:12 | 18/01/2014 | 22cm | Archive | Small sample. | Olive and black speckled. | Hard compact medium sand. | - |
| 105 | A/11C/GEO | 3m GC | 1500 | 08:07 | 18/01/2014 | N/S | N/S | Some trace of gravel, likely bounced. | - | - | - |
| 106 | A/22/GEO | 3m GC | 917 | 12:44 | 18/01/2014 | 145cm | LPH, MPOG, Eh, HSG, GORE | - | Grey to dark grey. | Silty sand. | - |
| 107 | A/23/GEO | 3m GC | 1087 | 15:09 | 18/01/2014 | 220m | LPH, MPOG, Eh, HSG, GORE | - | Grey to dark grey. | - | - |
| 108 | A/24/GEO | 3m GC | 985 | 17:31 | 18/01/2014 | 60cm | MPOG | - | Grey to dark grey. | - | - |
| 109 | A/08/GEO | 3m GC | 1187 | 19:52 | 18/01/2014 | 40cm | Archive | Small sample. | Grey to dark grey. | - | - |
| 110 | A/08B/GEO | 3m GC | 1187 | 20:33 | 18/01/2014 | N/S | N/S | N/S | - | - | - |
| 111 | A/08C/GEO | 3m GC | 1187 | 21:08 | 18/01/2014 | 70cm | MPOG, Archive | Small sample (third attempt). | Mid-grey and black speckled. | - | - |
| 112 | A/14A/GEO | 3m GC | 1206 | 22:48 | 18/01/2014 | 160cm | HSG, LPH, | Large sample with 3m | Olive and black | Medium compact sand | - |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | | |
|-------------------------------------|-----------|----------|------|-----------|------------|-------|------------------------------|--|---|---|------------------|--|
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| | | | | | | | MPOG, Eh, Archive | barrel. | speckled at surface; dark grey/green at bottom. | with some mud at surface; dense, moist, compact sandy mud at bottom. | | |
| 113 | A/14B/GEO | 6m GC | 1206 | 23:35 | 18/01/2014 | 203cm | HSG, LPH, MPOG, Eh, Archive | Re-attempt with 6m barrel. | Olive and black speckled at surface; dark grey at bottom. | Medium compact sand with some mud at surface; dense, firm, compact sandy mud at bottom. | - | |
| 114 | A/07/GEO | 3m GC | 1265 | 11:22 | 19/01/2014 | 90cm | LPH, MPOG, Eh, Archive | No HSG. | Olive and black speckled at surface; dark grey at bottom. | Medium compact sand with some mud at surface; dense, firm, compact sandy mud at bottom. | - | |
| 115 | A/07B/GEO | 3m GC | 1275 | 12:11 | 19/01/2014 | N/S | N/S | N/S (liner cut too short) | - | - | - | |
| 116 | A/13/GEO | 3m GC | 1267 | 14:13 | 19/01/2014 | N/S | N/S | N/S | - | - | - | |
| 117 | A/13B/GEO | 3m GC | 1267 | 14:54 | 19/01/2014 | N/S | N/S | N/S | - | - | - | |
| 118 | A/6/GEO | 3m GC | 1340 | 16:21 | 19/01/2014 | N/S | N/S | N/S | - | - | - | |
| 119 | A/6B/GEO | 3m GC | 1340 | 17:06 | 19/01/2014 | N/S | N/S | N/S | - | - | - | |
| 120 | A/6C/GEO | 3m GC | 1340 | 18:03 | 19/01/2014 | 60cm | MPOG | Archived. | - | - | - | |
| 121 | A/5/GEO | 3m GC | 1335 | 19:54 | 19/01/2014 | 115cm | HSG, LPH, MPOG, Eh, Archive | - | Beige to very dark grey. | Very soft silty fine sand. | - | |
| 122 | A/19/GEO | 3m GC | 1330 | 09:25 | 20/01/2014 | N/S | Archive (bag) | No core, just gravel. Bagged as archive. | - | Medium to coarse gravel - mixed materials. | - | |
| 123 | A/4/GEO | 3m GC | 1265 | 12:10 | 20/01/2014 | 25cm | Archive (bag) | Small sample (archived). | Grey to dark grey silty SAND. | - | - | |
| 124 | A/4B/GEO | 3m GC | 1265 | 12:56 | 20/01/2014 | 32cm | Archive (bag) | Small sample (archived). | Black sand and dark grey silt. | Fine sand. | - | |
| 125 | A/2/GEO | 3m GC | 1334 | 15:29 | 20/01/2014 | 52cm | MPOG and Archive (bag) | Small sample (archived). | Black/grey silty sand. | Fine sand. | - | |
| 126 | A/2B/GEO | 3m GC | 1334 | 16:06 | 20/01/2014 | 39cm | Archive (bag) | Small sample (archived). | Grey to dark grey silty sand. | - | - | |
| 127 | A/3/GEO | 3m GC | 1247 | 17:59 | 20/01/2014 | <5cm | Archive (bag) | Small sample (archived). | Grey to dark grey silty sand. | - | - | |
| 128 | A/3B/GEO | 3m GC | 1247 | 18:33 | 20/01/2014 | 23cm | Archive (bag) | Small sample (archived). | Black/grey silty sand. | Fine sand | - | |
| 129 | A/18/GEO | 3m GC | 1266 | 20:25 | 20/01/2014 | 87cm | LPH, MPOG, Eh, Gore, Archive | - | Grey to dark grey silty sand. | - | - | |
| 130 | A/18B/GEO | 3m GC | 1266 | 20:59 | 20/01/2014 | 90cm | LPH, MPOG, Eh, Gore, Archive | - | Grey to dark grey silty sand. | - | - | |
| 131 | A/17A/GEO | 3m GC | 1170 | 12:34 | 21/01/2014 | 39cm | Archive | - | Olive with black speckles surface - dark | Compact sand surface - spongy silty mud at bottom. | - | |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | |
|-------------------------------------|-----------|----------|------|-----------|------------|-------|---|--------------------------------------|---|---|------------------|
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| | | | | | | | | | grey bottom. | | |
| 132 | A/17B/GEO | 3m GC | 1170 | 01:13 | 21/01/2014 | 275cm | HSG x2, LPH x2, MPOG, Eh x2, GORE, Archive x2 | - | - | - | - |
| 133 | A/17C/GEO | 6m GC | 1170 | 02:01 | 21/01/2014 | 250cm | HSG x2, LPH x2, MPOG, Eh x2, GORE, Archive x2 | - | - | - | - |
| 134 | A/1A/GEO | 3m GC | 1085 | 03:19 | 21/01/2014 | 26cm | Archive | - | Olive with black speckles. | Compact sand. | - |
| 135 | A/1B/GEO | 3m GC | 1085 | 04:07 | 21/01/2014 | 26cm | Archive | - | Olive with black speckles. | Compact sand. | - |
| 136 | A/16A/GEO | 3m GC | 1070 | 05:15 | 21/01/2014 | N/S | Archive bag | Only fine gravel in core. | Dark brown. | Fine gravel - likely cut from concretion - clay based. | - |
| 137 | A/16B/GEO | 3m GC | 1070 | 06:00 | 21/01/2014 | 5cm | Archive bag | Very hard concretion at 3cm depth. - | Olive with black speckles surface - dark cream/yellow concretion below. | Compact sand surface - solid concretion below, likely clay based. | - |
| 138 | A/15A/GEO | 3m GC | 1070 | 07:16 | 21/01/2014 | 49cm | Archive core | - | Olive with black speckles. | Compact sand. | - |
| 139 | A/15B/GEO | 3m GC | 1070 | 07:54 | 21/01/2014 | 34cm | Archive core | - | Olive with black speckles. | Compact sand. | - |
| 140 | A/25/GEO | 3m GC | 820 | 11:46 | 21/01/2014 | 230cm | HSG, LPH, MPOG, Eh, Archive | - | Olive grey to black. | Soft cohesive CLAY with fine sand through to black sand. | - |
| 141 | A/25B/GEO | 3m GC | 820 | 12:21 | 21/01/2014 | 240cm | HSG, LPH, MPOG, Eh, Archive | Claystones present. | Olive grey to black. | Soft cohesive CLAY with fine sand through to black sand. | - |
| 142 | A/27/GEO | CAM | 484 | 18:40 | 21/01/2014 | - | - | Original core site. | - | - | - |
| 143 | A/27B/GEO | CAM | 484 | 19:55 | 21/01/2014 | - | - | Fault site. | - | - | - |
| 144 | A/27/GEO | 3m GC | 502 | 21:21 | 21/01/2014 | 250cm | HSG (a&b), LPH (a&b), MPOG, Eh, Archive | - | Olive grey/green becoming darker grey with core depth. | Soft CLAY with occasional fine sand becoming darker grey with a larger cohesive CLAY component with core depth. | - |
| 145 | A/27B/GEO | 3m GC | 502 | 21:51 | 21/01/2014 | 204cm | HSG, LPH, MPOG, Eh, Archive | - | Olive grey/green becoming darker grey with core depth. | Soft CLAY with occasional fine sand becoming darker grey with a larger cohesive CLAY component with core depth. | - |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | | |
|-------------------------------------|-----------------------|----------|------|-----------|------------|-------|--|-------------|--|---|------------------|--|
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| 146 | A/28/GEO | 3m GC | 345 | 01:22 | 22/01/2014 | 77cm | LPH, MPOG, Eh, Archive | - | Olive grey/green becoming darker grey with core depth. | Very firm compact clayey sands, olive grey small nodules and concretion sediments within. | - | |
| 147 | A/28B/GEO | 3m GC | 345 | 01:52 | 22/01/2014 | 97cm | LPH, MPOG, Eh, Archive | - | Olive grey/green becoming darker grey with core depth. | Very firm compact clayey sands, olive grey small nodules and concretion sediments within. | - | |
| 148 | A/29/GEO | 3m GC | 673 | 04:33 | 22/01/2014 | 270cm | HSG, LPH x2, MPOG, Ehx2, GORE, Archive | - | Olive grey/green becoming darker grey with core depth. | Soft clayey sand with coarser granules. | - | |
| 149 | A/29B/GEO | 6m GC | 673 | 05:06 | 22/01/2014 | 310cm | HSG, LPH x2, MPOG, Ehx2, GORE, Archive | - | Olive grey/green becoming darker grey with core depth. | Soft clayey sand with coarser granules. | - | |
| 150 | A/26/GEO | 6m GC | 520 | 09:33 | 22/01/2014 | 230cm | HSG, LPH x2, MPOG, Ehx2, GORE, Archive | - | Olive grey/green becoming darker grey with core depth. | Soft clayey sand with coarser granules. | - | |
| 151 | A/26B/GEO | 6m GC | 520 | 10:15 | 22/01/2014 | 210cm | HSG, LPH x2, MPOG, Ehx2, GORE, Archive | - | Olive grey/green becoming darker grey with core depth. | Soft clayey sand with coarser granules. | - | |
| 152 | A/23/GEO (revisited) | 3m GC | 1100 | 14:35 | 22/01/2014 | 18cm | Archive | - | - | - | - | |
| 153 | A/23B/GEO (revisited) | 3m GC | 1100 | 15:20 | 22/01/2014 | 16cm | Archive | - | - | - | - | |
| 154 | A/23/Water 1100m | Niskin | 1100 | 16:30 | 22/01/2014 | - | N/S | - | Did not trigger. | - | - | |
| 155 | A/23/Water 1100m | Niskin | 1100 | 17:00 | 22/01/2014 | - | 3x plastic 1L, 1x glass 1L | - | - | - | - | |
| 156 | A/23/Water 500m | Niskin | 1100 | 17:40 | 22/01/2014 | - | 3x plastic 1L, 1x glass 1L | - | - | - | - | |
| 157 | A/23/Water 50m | Niskin | 1100 | 18:00 | 22/01/2014 | - | 3x plastic 1L, 1x glass 1L | - | - | - | - | |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | |
|-------------------------------------|---------------|----------|---------|-----------|------------|----------|----------------------------|---|--|--|---|
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| 158 | A/23/Water 0m | Niskin | 1100 | 18:15 | 22/01/2014 | - | 3x plastic 1L, 1x glass 1L | - | - | - | - |
| 190 | A/18/ENV | CAM | 1415 | 12:40 | 01/02/2014 | - | - | - | - | - | - |
| 191 | A/18/ENV | BC | 1415 | 15:00 | 01/02/2014 | 55% | FA, FB, HC, HM, PSA, TOC | - | Light/olive grey sediment with dark surface layer. | Fine to medium sand with occasional fine gravel. | Ophiuroids and polychaetes. |
| 192 | A/22/ENV | CAM | 1524 | 17:48 | 01/02/2014 | - | - | - | - | - | Pennatulids and urchins. |
| 193 | A/22/ENV | BC | 1522 | 19:53 | 01/02/2014 | 54% | FA, FB, HC, HM, PSA, TOC | - | Light/olive grey sediment with dark surface layer. | Fine to medium sand with fine gravel. | Hard coral, worm tubes, bryozoan, orange encrusting sponge. Possible relic <i>Lophelia</i> . |
| 194 | A/206/ENV | CAM | 1530 | 22:52 | 01/02/2014 | - | - | - | - | - | Stalked crinoid, relic <i>Lophelia</i> , <i>Munida</i> , anemone, hard corals and sea whips. |
| 195 | A/205/ENV | CAM | 1540 | 04:25 | 02/02/2014 | - | - | No flash function - so stills used longer aperture time so some blurriness - screenshots taken additionally - some beacon problems. | - | - | Coarse sediment with shell and relic coral possibly <i>Lophelia</i> , rocky outcrops covered with sponges (05:33 possible <i>Phakellia</i> sp.), hard corals and sea whips. |
| 196 | A/3/ENV | CAM | 1124 | 11:30 | 02/02/2014 | - | - | No flash function - some blurriness in stills, some screen shots additionally, 2 videos as camera rebooted during line. | - | - | Ophiuroids, pennatulids and batoid. |
| 197 | A/3/ENV | BC | 1134 | 13:52 | 02/02/2014 | 20% | FA, FB, HC, HM, PSA, TOC | No depth reading from the beacon. | Light/olive grey sediment with dark surface layer. | Silty SAND. | Isopod, hydroid and polychaetes. |
| 198 | A/3/ENV | Niskin | 1134 | 13:52 | 02/02/2014 | - | HM, NUT, spare, HC. | Seabed, attached to BC. | - | - | - |
| 199 | A/3/ENV | Niskin | Surface | 16:55 | 02/02/2014 | - | HM, NUT, spare, HC. | Wire parted on winch, no further samples could be obtained at this location - further samples to be taken at the end of wreck multibeam line. | - | - | - |
| 200 | A/3/ENV | Niskin | 50 | 01:03 | 03/02/2014 | - | N/S | Niskin bottle failed to trigger. | - | - | - |
| 201 | A/3/ENV | Niskin | 50 | 01:09 | 03/02/2014 | - | N/S | Niskin bottle failed to trigger. | - | - | - |
| 202 | A/3/ENV | Niskin | 50 | 01:54 | 03/02/2014 | - | N/S | Niskin bottle failed to trigger. | - | - | - |



| Seabed Sampling (Deck Observations) | | | | | | | | | | | | |
|-------------------------------------|------------|----------|------|-----------|------------|------|-------------------------------------|--|--|---------------------------------------|--|---|
| Job No: | BSL 1334 | | | Operator: | BSL | | Vessel: | MV Poseidon | | Client: | Noble Energy Inc | |
| Date: | from: | 31/12/13 | | | | | Project: | FISA | | | | |
| | to: | 16/03/14 | | | | | | | | | | |
| 203 | A/3/ENV | Niskin | 50 | 02:34 | 03/02/2014 | - | HM, NUT, spare, HC. | 50m water sample. | | - | - | - |
| 204 | A/3/ENV | Niskin | 400 | 02:52 | 03/02/2014 | - | N/S | Niskin bottle failed to trigger. | | - | - | - |
| 205 | A/3/ENV | Niskin | 400 | 03:25 | 03/02/2014 | - | N/S | Niskin bottle failed to trigger. | | - | - | - |
| 206 | A/3/ENV | Niskin | 400 | 04:02 | 03/02/2014 | - | HM, NUT, spare, HC. | 400m water sample (max length on dyneema). | | - | - | - |
| 207 | A/3010/ENV | CAM | 1540 | 13:10 | 05/02/2014 | - | - | - | | - | - | - |
| 208 | A/3010/ENV | BC | 1540 | 15:10 | 05/02/2014 | N/S | N/S | Sample rejected due to washout, small gravel caught between bucket edge and spade. | | - | - | - |
| 209 | A/3010/ENV | BC | 1540 | 16:35 | 05/02/2014 | 50% | FA, FB, HC, HM, PSA, TOC | - | Light/olive grey sediment with dark surface layer. | Fine to medium sand with fine gravel. | Polychaetes and possible Foraminifera. | |
| 302 | A/30/GEO | 3m GC | 1040 | 10:39 | 16/03/2014 | 70cm | Archive C, LPHx2, MPOG x 2, X and Y | - | - | Medium sandy silt to fine sandy silt. | - | |
| 303 | A/30B/GEO | 3m GC | 1040 | 11:31 | 16/03/2014 | 90cm | Archive C, LPHx2, MPOG x 2, X and Y | - | - | Medium sandy silt to fine sandy silt. | - | |

GC = Gravity core (geochemical sample)
 BC = Box corer (environmental sample)
 DVV = double van Veen grab (Environmental sample)
 CAM = seabed camera
 HSG = Headspace gas (geochemical sample)
 LPH = Liquid phase hydrocarbons (geochemical sample)
 MPOG = Microbial prospecting for Oil and Gas (geochemical sample)
 eH = Redox potential (geochemical sample)
 HC = Hydrocarbon sample (Environmental sample)
 HM = Heavy metals (Environmental sample)
 PSA = Particle size analysis (Environmental sample)
 TOC = Total organic carbon (Environmental sample)

APPENDIX VII – MACRO-INVERTEBRATE MATRIX

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | Authority | A3_ENV_FA | A3_ENV_FB | A09_ENV_FA | A09_ENV_FB | A09_ENV_FC | A10_ENV_FA | A10_ENV_FB | A10_ENV_FC | A12_ENV_FA | A12_ENV_FC | A14_ENV_FA | A14_ENV_FB | A18_ENV_FA | A18_ENV_FB |
|----------|---|--------------------------------------|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|
| 1267 | CNIDARIA | | | | | | | | | | | | | | | |
| 128484 | Pennatulidae | Ehrenberg, 1834 | | | | | | | 1 | | | | | | | |
| 1360 | <i>Actinaria</i> | | | | | | | | | | | | | | | |
| 100665 | Edwardsiidae | Andres, 1881 | | | 1 | | | | | | | | 1 | | | 1 |
| 173052 | <i>Flabellum (Flabellum) curvatum</i> | Moseley, 1881 | | | | | | 1 | | | | | | | | |
| 152391 | NEMERTEA | | | | | | | | | | | | | | | |
| 152391 | Nemertea | | 1 | | | 2 | | | | | | | | | | 1 |
| 799 | NEMATODA | | | | | | | | | | | | | | | |
| 799 | Nematoda | | 2 | | 1 | 1 | | | | | | 1 | 1 | | | |
| 1268 | SIPUNCULA | | | | | | | | | | | | | | | |
| 410734 | <i>Nephasoma (Nephasoma) diaphanes</i> | (Gerould, 1913) | | | 1 | | | | | | | | | 3 | 3 | 1 |
| | ECHIURIDA | | | | | | | | | | | | | | | |
| 110352 | <i>Bonellia sp</i> | Rolando, 1821 | | | | | | | | | | | | | | 1 |
| 882 | ANNELIDA | | | | | | | | | | | | | | | |
| 129285 | <i>Euphrosine Sp 1</i> (no recorded NHM)† | Lamarck, 1818 | | | | | | | | | | | | | | |
| 173263 | <i>Euphrosine cirrata magellanica</i> | Ehlers, 1900 | | | | | | | | | | | | | | 1 |
| 939 | Polynoidae | Kinberg, 1856 | | | | | | | | | | | | 2 | | |
| 129487 | <i>Eunoe</i> | Malmgren, 1866 | | | 1 | | | 2 | | | | 1 | | | 1 | |
| 174391 | <i>Polyeunoe laevis</i> | McIntosh, 1885 | 1 | | | | | | | | | | | | | |
| 174468 | <i>Harmothoe magellanica</i> | (McIntosh, 1885) | 1 | | | | | | | | | | | | | |
| 129590 | <i>Leanira</i> | Kinberg, 1856 | | | | | | | | | | | | | | |
| | Sigalionidae (not recorded by NHM)† | | | | | | | | | | | | | | | |
| 129443 | <i>Eteone</i> | Savigny, 1818 | | | | | | | | | | | | | | 1 |
| 129450 | <i>Mystides</i> | Théel, 1879 | | | | | | | | | | | | | | |
| 130118 | <i>Glycera capitata</i> | Ørsted, 1843 | | | | | 1 | 2 | | | | | | 1 | | |
| 948 | Syllidae | Grube, 1850 | 1 | | | 1 | | | | | | | 1 | 1 | | |
| 129677 | <i>Sphaerosyllis</i> | Claparède, 1863 | | | | | | | | | | | | | | |
| 340445 | <i>Gymnonereis fauveli</i> | (Hartmann-Schröder, 1962) | 2 | 7 | | 2 | | | | 2 | | | 1 | 3 | 4 | 1 |
| 173522 | <i>Lumbrineris magalhaensis</i> | Kinberg, 1865 | | | | 2 | 1 | | | | | | | | | |
| 129280 | <i>Lysidice</i> | Lamarck, 1818 | | | | | | | | | | 1 | | | | |
| 129366 | <i>Aglaophamus (posteriobranchiatus)</i> NHM) | Kinberg, 1865 | | | 1 | | | | | 1 | | | | | | 1 |
| | <i>Aglaophanus sp I</i> | | | | | | | | | | | | | | | 1 |
| 129370 | <i>Nephtys</i> | Cuvier, 1817 | | | | | | | | | | | 1 | | | |
| 196680 | <i>Rhampobrachium (Spinigerium) ehlersi</i> | Monro, 1930 | | | | | | 4 | 8 | 4 | | | | 11 | 15 | 6 |
| 966 | Eunicidae | Berthold, 1827 | | 1 | 1 | 9 | 5 | | | | | | | | | 2 |
| 129278 | <i>Eunice</i> | Cuvier, 1817 | | | | | | | | | | | | | | |
| 130467 | <i>Nothria conchylega</i> | (Sars, 1835) | | | | | | 1 | | | | | | 1 | | 1 |
| 129425 | Scoloplos | Blainville, 1828 | | | | | | | | | | | | | | |
| 129430 | Aricidea | Webster, 1879 | 2 | 1 | | | 1 | | | | | | | 1 | 2 | |
| 326595 | Aricidea (<i>Allia</i>) <i>oculata</i> | Hartmann-Schröder & Rosenfeldt, 1990 | | | | | | 1 | | | | | | | | |
| 525485 | Aricidea (<i>Acmira</i>) <i>strelzovi</i> | Hartmann-Schröder & Rosenfeldt, 1990 | | | | | | | 1 | 1 | | | 1 | | | |
| 903 | Paraonidae | Cerruti, 1909 | | 1 | 1 | 2 | | | | | | | | | 1 | |
| 129198 | <i>Apistobrachus sp I</i> | Levinsen, 1883 | | 1 | 2 | 3 | 1 | | 2 | 1 | | | | 2 | 1 | 1 |
| 129198 | <i>Apistobrachus sp II</i> | Levinsen, 1883 | | | | | | | | | | | | | | |
| 913 | Spionidae | Grube, 1850 | | | | | | | | | | | | | | |
| 129620 | <i>Prionospio</i> | Malmgren, 1867 | | | 1 | | | | 1 | | | | | | | |
| 129623 | <i>Scolecopsis</i> | Blainville, 1828 | | | | | | | | | | | | | | |
| 129613 | <i>Laonice (Laonice sp2)</i> NHM) | Malmgren, 1867 | | | | | | | | | | | | | | 1 |
| 129291 | <i>Flabelligera</i> | Sars, 1829 | | | | | | | | | | | | | | |
| 129289 | <i>Brada sp</i> | Stimpson, 1854 | | | | | | | | | | | | | | |
| 923 | Maldanidae | Malmgren, 1867 | | 1 | | 1 | | | 3 | 1 | | 1 | 1 | | 2 | |
| 173557 | <i>Asychis amphiglypta</i> | (Ehlers, 1897) | | | | | | | | 1 | | | | | | 1 |
| 130305 | <i>Maldane sarsi</i> | Malmgren, 1865 | | | | | | | | | | | | | | |
| 130331 | <i>Rhodine loveni</i> | Malmgren, 1865 | | | | | | | | | | | | | | 2 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | Authority | A3_ENV_FA | A3_ENV_FB | A09_ENV_FA | A09_ENV_FB | A09_ENV_FC | A10_ENV_FA | A10_ENV_FB | A10_ENV_FC | A12_ENV_FA | A12_ENV_FC | A14_ENV_FA | A14_ENV_FB | A18_ENV_FA | A18_ENV_FB |
|-----------------------|---|--|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|
| 129924 | Spiochaetopterus typicus | M. Sars, 1856 | 1 | | 2 | | | 1 | | | | | | | 2 | |
| | Chaetozone sp 8 † (NHM) | | | | | | | | | | | | | | | |
| 129243 | Cirratulus Sp2 | Lamarck, 1818 | | | | | 1 | | | | | | | | | 3 |
| 129245 | Pseudoscalibregma bransfeldiat | Hartman, 1936 | | | | | | | | | | | | | | 2 |
| 129241 | Caulleriella | Chamberlin, 1919 | | 1 | 1 | 2 | 2 | | 4 | 1 | | | 1 | 2 | | 1 |
| 129955 | Chaetozone setosa (Chaetozone sp 5 NHM) | Malmgren, 1867 | | 1 | | 1 | | | | | | | | 1 | | |
| 173159 | Tharyx fusiformis | Monro, 1939 | | | | | | | | | | | | 1 | | |
| 129249 | Tharyx | Webster & Benedict, 1887 | | | | | | | | | | | | | | |
| 129898 | Notomastus latericeus | Sars, 1851 | | | | | | | | 1 | | | | | | |
| 130503 | Ophelina cylindricaudata | (Hansen, 1879) | | | | | | | | | | | | | | 1 |
| 129413 | Ophelia | Savigny, 1822 | 1 | | | | | | | 1 | | | | | | |
| 130980 | Scalibregma inflatum | Rathke, 1843 | | | 1 | | | | | | | | | | | 1 |
| 152252 | Ampharetinae | Malmgren, 1866 | | | | | | | | | | | | | | |
| 129177 | Samytha | Malmgren, 1866 | | | | | | 1 | | | | | | | | 1 |
| 129153 | Amage | Malmgren, 1866 | | 1 | | | | | | | | | | | | 1 |
| 129804 | Melinna cristata (complex) | (M. Sars, 1851) | | | 3 | | | 3 | 3 | 3 | | | | 1 | 1 | 1 |
| 129708 | Pista | Malmgren, 1866 | 1 | | | | | | | | | | | | | |
| 131573 | Terebellides stroemii | Sars, 1835 | 1 | | | 1 | 1 | | | 1 | | | | 2 | | 1 |
| 982 | Terebellidae | Johnston, 1846 | | | | 1 | | | | 1 | | | | | | |
| 174568 | Perkinsiana antarctica | (Kinberg, 1867) | | | | | | | | | | | | | | |
| 985 | Sabellidae | Latreille, 1825 | | 2 | | | | 1 | 1 | | | | | 2 | | |
| 154918 | Fabriciidae | Rioja, 1923 | | | | | | | | | 1 | | | | | 1 |
| 129530 | Fabriciola | Friedrich, 1939 | | | | | | | | 1 | | | | | | |
| 129533 | Jasmineira | Langerhans, 1880 | 1 | | | | | | | | | | | | | |
| 129525 | Chone | Krøyer, 1856 | | 1 | | | | | | | | | | | | |
| 174564 | Euchone pallida (Euchone sp1 NHM) | Ehlers, 1908 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | | 1 | | | | 1 |
| 2036 | Oligochaeta | | | 1 | | | | | | 2 | | | | | 2 | |
| 988 | Serpulidae | Rafinesque, 1815 | | | | 3 | 1 | | | | | | | | | |
| 147018 | Microrbinia | Hartman, 1965 | 2 | | | | | | | | | | | | | |
| 244666 | Phylo felix | Kinberg, 1866 | | | | | | | | | | | 1 | | 2 | |
| 734535 | Phylo minima | (Hartmann-Schröder & Rosenfeldt, 1990) | 2 | | | | | | | | | | | | | |
| 146949 | Galathowenia | Kirkegaard, 1959 | 1 | | | | | | | 1 | | | | 1 | | |
| 1066 CRUSTACEA | | | | | | | | | | | | | | | | |
| 106115 | Scalpellum | Leach, 1818 | | | | 2 | | | | | | | | | | |
| 106140 | Verruca | Schumacher, 1817 | | | | | | | | | | | | | | |
| 106137 | Altiverruca | Pilsbry, 1916 | | | | | | | | | | | | | | |
| 1078 | Ostracoda | Latreille, 1802 | | 1 | | 1 | | | | | | | | | | |
| 1080 | Copepoda | | | | 1 | | | 1 | | | 1 | 1 | | 1 | | |
| 101436 | Phtisica | Slabber, 1769 | | | | | | | | | | | | | | |
| 1135 | Amphipoda | Latreille, 1816 | 2 | | | 2 | | | | | | | | 1 | | |
| 101409 | Stenothoidae | Boeck, 1871 | | | | | | | | | | | | | | |
| 101409 | Stenothoidae | Boeck, 1871 | | | | | | | | | | | | | | |
| 101789 | Urothoe | Dana, 1852 | | 5 | 1 | | | | 1 | 1 | | | 1 | 1 | | 1 |
| 101403 | Phoxocephalidae | Sars, 1891 | | | 1 | | | | | | | | 1 | | | |
| 101403 | Phoxocephalidae | Sars, 1891 | | 1 | | | | | 2 | | | | 1 | 2 | 1 | |
| 101723 | Phoxocephalus | Stebbing, 1888 | | | | | | | | 1 | | | | | | |
| 101675 | Maera | Leach, 1814 | | | 1 | | | | | | | | | | | |
| 101393 | Leucothoidae | Dana, 1852 | | | | | | | | | | | | | | |
| 176788 | Lysianassoidea | Dana, 1849 | 2 | | | | | | | | | | | 1 | | |
| 176788 | Lysianassoidea | Dana, 1849 | | 12 | | | | | | | | | | 1 | | |
| 101445 | Ampelisca | Krøyer, 1842 | 1 | 2 | 5 | 4 | 2 | 2 | 3 | 2 | 1 | | 2 | 4 | | |
| 101445 | Ampelisca | Krøyer, 1842 | 4 | | | | | | | | | | | | | |
| 101445 | Ampelisca | Krøyer, 1842 | | | | | | | | | | | | | | |
| 101445 | Ampelisca | Krøyer, 1842 | | | | | | | | | | | | | 1 | 2 |
| 101365 | Amphilochidae | Boeck, 1871 | | | | 1 | | | | | | | | | | 1 |
| 101410 | Synopiidae | Dana, 1853 | 3 | | | | | | | | | | | | | |
| 101563 | Photis | Krøyer, 1842 | | | | | | 1 | | 1 | | | | 2 | | |
| 101470 | Leptocheirus | Zaddach, 1884 | | | | 2 | | | | | | | | | | |
| 101519 | Eusirus | Krøyer, 1845 | | | | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | Authority | A3_ENV_FA | A3_ENV_FB | A09_ENV_FA | A09_ENV_FB | A09_ENV_FC | A10_ENV_FA | A10_ENV_FB | A10_ENV_FC | A12_ENV_FA | A12_ENV_FC | A14_ENV_FA | A14_ENV_FB | A18_ENV_FA | A18_ENV_FB |
|--------------------|--------------------------|--------------------|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|
| 101734 | Dulichia | Kroyer, 1845 | | | | | | | | | | | | | | |
| 118437 | Gnathia | Leach, 1814 | | | | | | | | | | | | | | |
| 257124 | Caecognathia c.f.polaris | (Hodgson, 1902) | | | | | | | | | | | | | | |
| 118354 | Ilyarachna | Sars, 1870 | | | | | | | | | | | | | | |
| 256200 | Munna argentiniae | Menzies, 1962 | | | | | | | | | | | | | | |
| 118371 | Macrostylis | G.O. Sars, 1864 | | | | | | | | | | | | | | |
| 118371 | Macrostylis II | G.O. Sars, 1864 | | | | | | | | | | | | | | |
| 118351 | Haploniscus | Richardson, 1908 | | | | | | | | | | | | | | |
| 118358 | Ischnomesus | Richardson, 1908 | | | | 1 | | | | | | | | | | |
| 118356 | Haplomesus | Richardson, 1908 | | | | | | | | | | | | | | 1 |
| n/a | Stylomesus elegans | Menzies, 1962 | | | | | | | | | | | | | | |
| 174952 | Stylomesus | Wolff, 1956 | | | | | 1 | | | | | | | | | |
| 118319 | Desmosoma | G.O. Sars, 1864 | | 2 | | 2 | | | | | | | | | 1 | |
| 118323 | Eugerdia | Meinert, 1890 | | | | 1 | | | | | | | | | | |
| 118360 | Janirella | Bonnier, 1896 | | | | | | | | | | | | | | |
| 248358 | Abyssianira | Menzies, 1956 | | | | | | | | | | | | | | |
| 118399 | Cirolana | Leach, 1818 | 1 | 1 | 1 | | | | | | | | | | | |
| 118399 | Cirolana | Leach, 1818 | | | | | | | | | | | | | | |
| 213013 | Pseudanthura lateralis | Richardson, 1911 | | 1 | 1 | | | | | 2 | | | | | | |
| 255448 | Haliophasma antarctica | (Kensley, 1982) | 1 | | | | | 1 | | | | | | | | |
| 174514 | Antarcturus | zur Strassen, 1902 | | | | | 1 | | 1 | | | | 13 | 1 | | |
| 136164 | Tanaidae | Dana, 1849 | 1 | | | | | | | | | | | | | |
| 136164 | Tanaidae | Dana, 1849 | | | | | | | | | | | | | | |
| 136222 | Agathotanaia | Hansen, 1913 | 2 | | | 1 | | | | | | | | | | 1 |
| 136229 | Leptognathia | Sars, 1882 | 3 | 2 | 1 | 3 | | | | 1 | | 1 | | | 1 | 4 |
| 136256 | Typhlotanaia | Sars, 1882 | | | | | | | | | | | 2 | | | |
| 136185 | Apseudes | Leach, 1813 | | | | | | | | | | | | | | 1 |
| 136200 | Pseudosphyrapus sp I | Gutu, 1980 | | | | | | | | | | | | | | |
| 136200 | Pseudosphyrapus sp II | Gutu, 1980 | | | | | | | | | | | | | | |
| 182287 | Eudorella gracilior | Zimmer, 1907 | | | | | | | | | | | | | | |
| 110414 | Leucon | Kroyer, 1846 | 1 | | | | | | | | | | | | | |
| 110398 | Diastylis | Say, 1818 | 1 | 1 | | | | | 1 | | | | | | | 1 |
| 110398 | Diastylis sp II | Say, 1818 | | | | | | | | | | | | | | 1 |
| 110415 | Campylaspis | G.O. Sars, 1865 | | | | | | | | | | | | | | |
| 110415 | Campylaspis spII | G.O. Sars, 1865 | | | | | | | | | | | | | | |
| 110404 | Makrokylindrus | Stebbing, 1912 | 1 | | | | | | | | | | | | | |
| 51 MOLLUSCA | | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | |
| 138249 | "Neomenia " | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | |
| 343675 | Neopilina Lemche, 1957 | Lemche, 1957 | | | | | | | | | | | | | | |
| 104 | Scaphopoda | Bronn, 1862 | | | | | | | | | | | | | | |
| 138024 | Cadulus | Philippi, 1844 | | 1 | | | | | | | | | | | | |
| 138013 | Fissurella | Bruguière, 1789 | | | | | | | | | | | | | | |
| 138508 | Skenea | Fleming, 1825 | | | | | | | | | 1 | | | | | |
| 138508 | Skenea | Fleming, 1825 | | | | | | | | | | | | | | |
| 137970 | Eulima | Risso, 1826 | | | | | | | | | | | | | | |
| 147109 | Polinices | Montfort, 1810 | | | | | | | | | | | | | | |
| 147109 | Polinices | Montfort, 1810 | | | | | | | | | | | | | | |
| 390535 | Fusitriton | Cossmann, 1903 | | | 1 | | | | | 1 | | | | | | |
| 390535 | Fusitriton | Cossmann, 1903 | 1 | 1 | | | | | | | | | | | | |
| 138432 | Retusa | T. Brown, 1827 | | | | | | | | | | | | | | |
| 138217 | Dacrydium | Torell, 1859 | | | | | | | | | | | | | | |
| 138259 | Nuculana | Link, 1807 | | | | | | | | | | | | | | |
| 138672 | Yoldia | Möller, 1842 | | | | | | | | | | | | | | 1 |
| 138132 | Limopsis | Sassi, 1827 | | | | | | | | | | | | 2 | | |
| 138323 | Pecten | O. F. Müller, 1776 | | | | 1 | | | | | | | | | | |
| 147153 | Similipecten | Winckworth, 1932 | | | | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | Authority | A3_ENV_FA | A3_ENV_FB | A09_ENV_FA | A09_ENV_FB | A09_ENV_FC | A10_ENV_FA | A10_ENV_FB | A10_ENV_FC | A12_ENV_FA | A12_ENV_FC | A14_ENV_FA | A14_ENV_FB | A18_ENV_FA | A18_ENV_FB |
|----------|----------------------------|-----------------------------------|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|
| 197227 | Limatula hodgsoni | (E. A. Smith, 1907) | | 1 | | | | 1 | | | | | | | | |
| 505622 | Limea pygmaea | Philippi, 1845) | | | | | | | | | | | | | | |
| 138186 | Mysella | Angas, 1877 | | | | | | | | | | | | | | |
| 138654 | Lyonsiella sp | G.O. Sars, 1872 | | | | | | | | | | | | | | |
| 197467 | Thracia meridionalis | E. A. Smith, 1885 | | | 1 | | | | | | | | | | | |
| 137858 | Cuspidaria | Nardo, 1840 | 1 | 1 | | | 1 | | | | | | | | | 1 |
| 137863 | Tropidomya | Dall & E. A. Smith, 1886 | | | | | | | | | | | | | | |
| 152413 | Axinulus | Verrill & Bush, 1898 | | | | | | | | | 1 | | | | 2 | |
| 138552 | Thyasira | Lamarck, 1818 | | | | 2 | | | | | | | | | 1 | |
| 138552 | Thyasira II | Lamarck, 1818 | | | | | | 2 | | | | | | | 2 | |
| 1806 | ECHINODERMATA | Bruguieres, 1791 [ex Klein, 1734] | | | | | | | | | | | | | | |
| 172707 | Anseropoda antarctica † | Fisher, 1940 | | | | | | | | | | | | | | 1 |
| 172769 | Acodontaster elongatus | (Sladen, 1889) | | | | | | | | | | | | | | |
| 123567 | Ophiophycis aff mirabilis† | Koehler, 1901 | | | | | | | | | | | | | | |
| 173102 | Amphiura belgicae | Koehler, 1900 | | 1 | | | | 1 | | | | | | | | |
| 123206 | Amphiuridae | Ljungman, 1867 | 1 | | | | | | | | | | | | | |
| 123200 | Ophiuridae | Müller & Troschel, 1840 | | | | | | | | | | | | | | |
| 236013 | Ophiozonella falklandica | Mortensen, 1936 | | | | | | | 1 | | | | | | 1 | |
| 160790 | Brisaster antarcticus † | (Doderlein, 1906) | | | | | | | | | | | | | | |
| 123494 | aff Thyone | Jaeger, 1833 | | | | | | | | | | | | | | |
| 123473 | Echinocucumis sp | M. Sars, 1859 | | | | | | | | | | | | | | |
| 146121 | Psolus | Oken, 1815 | | | | | | | | | | | | | | |
| | Psolus spp | | | | | | | | | | | | | | | |
| 123182 | Synaptidae | Burmeister, 1837 | | | | | | | | | | | | | | |
| 123441 | Myriotrochus | Steenstrup, 1851 | | | | | | | | | | | | | | |
| 1803 | BRACHIOPODA | Duméril, 1805 | | | | | | | | | | | | | | |
| 1803 | Brachiopoda | Duméril, 1805 | | | | | | | | | | | | | | |
| 1803 | Brachiopoda II | Duméril, 1805 | | | | | | | | | | | | | | |
| 183339 | Pelagodiscus | atlanticus (King, 1868) | | | | | | | | | | | | | | |
| | CHAETOGNATHA | | | | | | | | | | | | | | | |
| 105413 | Spadella | Langerhans, 1880 | | | | | | | | | | | | | | |
| | S | | 33 | 28 | 25 | 29 | 12 | 17 | 21 | 21 | 17 | 8 | 22 | 36 | 20 | 28 |
| | N | | 49 | 53 | 34 | 56 | 18 | 25 | 40 | 30 | 17 | 8 | 51 | 72 | 30 | 35 |
| | d | | 8.222 | 6.801 | 6.806 | 6.956 | 3.806 | 4.971 | 5.422 | 5.88 | 5.647 | 3.366 | 5.341 | 8.184 | 5.586 | 7.594 |
| | J' | | 0.9694 | 0.8686 | 0.9539 | 0.9352 | 0.9213 | 0.9526 | 0.9142 | 0.9596 | 1 | 1 | 0.8283 | 0.905 | 0.927 | 0.9719 |
| | H'(log2) | | 4.89 | 4.175 | 4.43 | 4.543 | 3.303 | 3.894 | 4.015 | 4.215 | 4.087 | 3 | 3.694 | 4.679 | 4.006 | 4.672 |
| | 1-Lambda' | | 0.9821 | 0.9267 | 0.9715 | 0.9604 | 0.9216 | 0.96 | 0.941 | 0.9701 | 1 | 1 | 0.8894 | 0.9476 | 0.9494 | 0.9832 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A20_ENV_FA | A20_ENV_FB | A20_ENV_FC | A21_ENV_FA | A21_ENV_FB | A22_ENV_FA | A22_ENV_FB | A26_ENV_FA | A26_ENV_FB | A26_ENV_FC | A27_ENV_FA | A27_ENV_FB | A27_ENV_FC | A202_ENV_FA | A202_ENV_FB | A204_ENV_FA |
|----------|---|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|-------------|-------------|-------------|
| 1267 | CNIDARIA | | | | | | | | | | | | | | | | |
| 128484 | Pennatulidae | | | | | | 1 | | 1 | | 1 | | | 1 | | | 14 |
| 1360 | Actinaria | | | | | | | | | | | | | | | 1 | |
| 100665 | Edwardsiidae | | | | | | | | 1 | | | | | 1 | | | |
| 173052 | <i>Flabellum (Flabellum) curvatum</i> | | | | | | | | | | | | | | | | |
| 152391 | NEMERTEA | | | | | | | | | | | | | | | | |
| 152391 | Nemertea | 1 | | | 2 | | | | 1 | 1 | | | | 2 | | 1 | 1 |
| 799 | NEMATODA | | | | | | | | | | | | | | | | |
| 799 | Nematoda | | | | | | | | 3 | | | | 1 | | 1 | | 3 |
| 1268 | SIPUNCULA | | | | | | | | | | | | | | | | |
| 410734 | <i>Nephasoma (Nephasoma) diaphanes</i> | | | | 4 | 2 | 1 | 1 | | 3 | | | 1 | | | 1 | 2 |
| | ECHIURIDA | | | | | | | | | | | | | | | | |
| 110352 | <i>Bonellia sp</i> | | | | | | | | | | | | | | | | |
| 882 | ANNELIDA | | | | | | | | | | | | | | | | |
| 129285 | <i>Euphrosine Sp 1 (no recorded NHM)†</i> | | | | | | | | 2 | | | | | | | | |
| 173263 | <i>Euphrosine cirrata magellanica</i> | | | | | | | | 1 | 1 | | | | | | | |
| 939 | Polynoidae | | | | | | | | | | | | | | | | |
| 129487 | <i>Eunoe</i> | | | | 2 | | 1 | | 1 | 1 | 2 | | | | | | 2 |
| 174391 | <i>Polyeunoe laevis</i> | | | | | | | | | | | | | | | | |
| 174468 | <i>Harmothoe magellanica</i> | | | | | | | | | | | | | | | | |
| 129590 | <i>Leanira</i> | | | | | | | | | | | | | 1 | | | |
| | Sigalionidae (not recorded by NHM)† | | | | | | | | | | | | | 1 | | | |
| 129443 | <i>Eteone</i> | | | | | | | | | 2 | | | | | | 1 | 2 |
| 129450 | <i>Mystides</i> | | | | | | | | | | | 1 | | | | | |
| 130118 | <i>Glycera capitata</i> | | | | | | | | | | | | | | | | |
| 948 | Syllidae | 1 | | | | | | | 2 | 1 | | | | 1 | | | |
| 129677 | <i>Sphaerosyllis</i> | | | | | | | | | | | 2 | | | | | |
| 340445 | <i>Gymnoreris fauveli</i> | | | | | | | | 3 | | | | | | | | |
| 173522 | <i>Lumbrineris magalhaensis</i> | 1 | | | 1 | 4 | 1 | | | | | 1 | | 1 | | | 1 |
| 129280 | <i>Lysidice</i> | | | | | | | | | | | | | | | | |
| 129366 | <i>Aglaophamus (posteriobranchiatus) NHM)</i> | | | | | | | | | | | | | | | | |
| | <i>Aglaophanus sp I</i> | | | | | | | | | | | | | | | | |
| 129370 | <i>Nephtys</i> | | | | | | | | | | | 1 | | 1 | | | |
| 196680 | <i>Rhynchobranchium (Spinigerium) ehlersi</i> | 1 | 4 | | 18 | 3 | 3 | 2 | 6 | 1 | 2 | 7 | 9 | 7 | 4 | 4 | |
| 966 | Eunicidae | | | | | | | | | | | | | | | | 1 |
| 129278 | <i>Eunice</i> | | | | | | | | | | | | | | | | |
| 130467 | <i>Nothria conchylega</i> | | | | | | | | 1 | | | | | | | | |
| 129425 | <i>Scoloplos</i> | | | | | | | | | | | 1 | | | | | |
| 129430 | Aricidea | | | | | | | | | | | | | | | | |
| 326595 | Aricidea (<i>Allia</i>) <i>oculata</i> | | | | | | | | | | | | | | | | |
| 525485 | Aricidea (<i>Acmira</i>) <i>strelzovi</i> | | | | | | | | | | | | | | | | |
| 903 | Paraonidae | | | | | | | | | | | | | | | | |
| 129198 | <i>Apistobranchus sp I</i> | | | | | | | | | | | | | | | 4 | 4 |
| 129198 | <i>Apistobranchus sp II</i> | | | | | | | | | | | | | | | | |
| 913 | Spionidae | | | | | | | | | | | | | | | | |
| 129620 | <i>Prionospio</i> | | | | | | | | | | | | | | | | |
| 129623 | <i>Scoletepis</i> | | | | | | | | | | | | | | | | |
| 129613 | <i>Laonice (Laonice sp2) NHM)</i> | | | | | | | | | | | | | | | | |
| 129291 | <i>Flabelligera</i> | | | | | | | | | | | | | | | | |
| 129289 | <i>Brada sp</i> | | | | | | | | | | | | | | | | |
| 923 | Maldanidae | | | | | | | | 1 | | | | | | | | 3 |
| 173557 | <i>Asychis amphiglypta</i> | | | | | | 1 | 1 | 1 | 1 | | | 1 | | | | |
| 130305 | <i>Maldane sarsi</i> | | | | | | | 2 | 2 | 1 | | | | | | | |
| 130331 | <i>Rhodine loveni</i> | | | | | | | | | | | 2 | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A20_ENV_FA | A20_ENV_FB | A20_ENV_FC | A21_ENV_FA | A21_ENV_FB | A22_ENV_FA | A22_ENV_FB | A26_ENV_FA | A26_ENV_FB | A26_ENV_FC | A27_ENV_FA | A27_ENV_FB | A27_ENV_FC | A202_ENV_FA | A202_ENV_FB | A204_ENV_FA | |
|-----------------------|---|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|-------------|-------------|-------------|----|
| 129924 | Spiochaetopterus typicus | 1 | 1 | | | | | | 1 | | | | 1 | 1 | 1 | 1 | 1 | |
| | Chaetozone sp 8 † (NHM) | | | | | | | | | | | | | | | | 1 | |
| 129243 | Cirratulus Sp2 | | | | | | | | | | | | | | | | 1 | |
| 129245 | Pseudoscalibregma bransfeldiat | | | | | | | | 1 | | | | | | | 1 | | |
| 129241 | Caulierella | | | | 1 | | 2 | 4 | | | 1 | | | | 2 | 1 | 1 | |
| 129955 | Chaetozone setosa (Chaetozone sp 5 NHM) | | 1 | | 1 | | | | 1 | | | | 3 | 2 | 2 | 1 | | |
| 173159 | Tharyx fusiformis | | | | | | | | | | | 1 | | | | | | |
| 129249 | Tharyx | | | | | | | | | | | 2 | | | | | | |
| 129898 | Notomastus latericeus | | | | 1 | | | | | 1 | | 1 | 1 | | | 1 | | |
| 130503 | Ophelina cylindricaudata | | | | | | | | | | | | | | 1 | | | |
| 129413 | Ophelia | | | | | | | | | | | | | | | | | |
| 130980 | Scalibregma inflatum | | | | | | | | | | | | | | | | | |
| 152252 | Ampharetinae | | | | | | | | | | | 1 | | | | | | |
| 129177 | Samytha | | | | | | | | | | | 1 | | | | | | |
| 129153 | Amage | | | | | | | | | | | | | | | | | |
| 129804 | Melinna cristata (complex) | | | | | | | | | | | | | | | 1 | | |
| 129708 | Pista | | | | | | | | | | | 1 | | | | | | |
| 131573 | Terebellides stroemii | | | | | | | | 1 | | | | 1 | | | | | |
| 982 | Terebellidae | | | | | | | | | 1 | | | | | | | | |
| 174568 | Perkinsiana antarctica | | | | | | | | | | | | | | 3 | | | |
| 985 | Sabellidae | | | | | | | 1 | | | | | | | | | | |
| 154918 | Fabriciidae | | | | | | | | 1 | | | | | | 2 | 1 | | |
| 129530 | Fabriciola | | | | | | | | | | | 1 | | | | | | |
| 129533 | Jasmineira | | | | | | | | | | | | | | | | | |
| 129525 | Chone | | | | | | | | | | | | | | | | | |
| 174564 | Euchone pallida (Euchone sp1 NHM) | | 1 | | | 1 | | | | | | | | | 1 | 1 | 2 | |
| 2036 | Oligochaeta | | | | | | | | | | | 4 | | 2 | | 1 | | |
| 988 | Serpulidae | | | | | | | | | | | | | | | 1 | | |
| 147018 | Microrbinia | | | | | | | | | | | | | | | | | |
| 244666 | Phylo felix | | | | | | | | | | | | | 1 | | 1 | | |
| 734535 | Phylo minima | | | | | | | | | | | | | | | | | |
| 146949 | Galathowenia | | | | | | | 1 | 1 | | | | | 2 | | | | |
| 1066 CRUSTACEA | | | | | | | | | | | | | | | | | | |
| 106115 | Scalpellum | | | | | | | | | | | | | | | | | 14 |
| 106140 | Verruca | | | | | | | | 1 | | | | | | | | | 2 |
| 106137 | Altiverruca | | | | | | | | | 1 | | | | | | | | 2 |
| 1078 | Ostracoda | | | | | | | | | | | | | | 1 | | | 2 |
| 1080 | Copepoda | 1 | 13 | 2 | 1 | | | | | | | | 1 | | 13 | 1 | | 2 |
| 101436 | Phtisica | | | | 1 | | | | | | | | | | | | | 2 |
| 1135 | Amphipoda | | | | | | | | | | | 1 | | 1 | | | | 3 |
| 101409 | Stenothoidae | | | | | | | | | | | 2 | | | | | | 2 |
| 101789 | Urothoe | | | | 3 | 2 | 1 | | | | | 2 | 2 | | 3 | 4 | | 2 |
| 101403 | Phoxocephalidae | | | | | | | | | | | | | | | | | 2 |
| 101403 | Phoxocephalidae | | | | 2 | 3 | | | 1 | | | 1 | 1 | | 1 | | | 2 |
| 101723 | Phoxocephalus | | | | | | | | | | | | | | | | | 2 |
| 101675 | Maera | | | | 1 | | 3 | 4 | | 1 | | | | | 2 | | | 2 |
| 101393 | Leucothoidae | | | | | | | | | | | | | | | 2 | | 1 |
| 176788 | Lysianassoidea | | | | | | | | | | | 1 | | | 1 | | | 1 |
| 176788 | Lysianassoidea | | | | | | | | | | | | | | | | | 1 |
| 101445 | Ampelisca | | | | | 2 | | | 1 | | | 2 | 1 | | 3 | | | 1 |
| 101445 | Ampelisca | | | | | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | | | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | 1 | | | | | | | | | | | | | | |
| 101365 | Amphiloichidae | | | | | | | | | | | 1 | 1 | | | | | |
| 101410 | Synopiidae | | | | | | | | | | | | | | | | | |
| 101563 | Photis | | | | | 2 | 3 | | 1 | | | | 1 | | | | | 2 |
| 101470 | Leptocheirus | | | | | | | | | | | | | | 1 | 4 | | |
| 101519 | Eusirus | | | | 1 | | | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A20_ENV_FA | A20_ENV_FB | A20_ENV_FC | A21_ENV_FA | A21_ENV_FB | A22_ENV_FA | A22_ENV_FB | A26_ENV_FA | A26_ENV_FB | A26_ENV_FC | A27_ENV_FA | A27_ENV_FB | A27_ENV_FC | A202_ENV_FA | A202_ENV_FB | A204_ENV_FA |
|--------------------|--------------------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|-------------|-------------|-------------|
| 101734 | Dulichia | | | | 3 | | | | | | | | | | 1 | | |
| 118437 | Gnathia | | | | | | | | | | | | 1 | | | | |
| 257124 | Caecognathia c.f.polaris | | | | | | | | | | | | 1 | | | | 1 |
| 118354 | Ilyarachna | | | | | | | | | | | | 1 | | | | 1 |
| 256200 | Munna argentinae | | | | | | | | | | | | | | | | |
| 118371 | Macrostylis | | | | | | | | | | | | | | | | 1 |
| 118371 | Macrostylis II | | | | | | | | | | | | | | | | 1 |
| 118351 | Haploniscus | | | | | | | | | | | | | | 1 | | 1 |
| 118358 | Ischnomesus | | | | | | 2 | | | | | | | | | | |
| 118356 | Haplomesus | | | | | | | | | | | | | | | | |
| n/a | Stylomesus elegans | | | | | 1 | | | | | | | | | | | |
| 174952 | Stylomesus | | | | | | | | | | | | | | | | 4 |
| 118319 | Desmosoma | | | | 2 | | | | | 1 | | | | | | 2 | 1 |
| 118323 | Eugerdia | | | | | | | | | | | | | | | | |
| 118360 | Janirella | | | | | | | | | | | | | | | | 1 |
| 248358 | Abyssianira | | | | 1 | | | | | | | | | | | | |
| 118399 | Cirolana | | | | | | | | | 1 | | | | | | | |
| 118399 | Cirolana | | | | | | | | | | | | | | | | |
| 213013 | Pseudanthura lateralis | | | | | | | | 1 | | | | | 1 | | | |
| 255448 | Haliophasma antarctica | | | | | | | | | | | 1 | | | 1 | | |
| 174514 | Antarcturus | | | | | | 1 | | | | | | | | | | |
| 136164 | Tanaidae | | | | | | | | | | | | | | | | 1 |
| 136164 | Tanaidae | | | | | | | | | | | | | | | | |
| 136222 | Agathotanaeis | | | | | | | | | | | | | | | | 1 |
| 136229 | Leptognathia | | | | 5 | 1 | 1 | 2 | 1 | | | | | | | 2 | 1 |
| 136256 | Typhlotanaeis | | | | | | | | | | | | | | | | |
| 136185 | Apseudes | | | | | | | | | | | | | | | | |
| 136200 | Pseudosphyrapus sp I | | | | | | | | | | | | 1 | | | 1 | |
| 136200 | Pseudosphyrapus sp II | | | | | | | | | | | | | | | | |
| 182287 | Eudorella gracillior | | | | | | | | | | | | | | | | |
| 110414 | Leucon | | | | | | | | | | | | | | | | 1 |
| 110398 | Diastylis | | | | 3 | 2 | | | | | | 1 | 1 | | | 2 | 3 |
| 110398 | Diastylis sp II | | | | | | | | | | | | | | | | 3 |
| 110415 | Campylaspis | | | | 1 | | | | | | | | | | | | |
| 110415 | Campylaspis spII | | | | | | | | | | | | | | | | 1 |
| 110404 | Makrokylindrus | | | | | | | | | | | | | | | | |
| 51 MOLLUSCA | | | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | | 1 |
| 138249 | "Neomenia " | | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | | |
| 343675 | Neopilina Lemche, 1957 | | | | 1 | | | | | | | | | | | | |
| 104 | Scaphopoda | | | | | | | | | | | | | | | | |
| 138024 | Cadulus | | | | | | | | | | | | | | | | 1 |
| 138013 | Fissurella | | | | | | | | | | | | | | | | |
| 138508 | Skenea | | | | | | | | | | | | | | | | |
| 138508 | Skenea | | | | | | | | | | | | | | | | |
| 137970 | Eulima | | | | | | | | | | | | | | | | |
| 147109 | Polinices | | | | | | | | | | | | | | | | |
| 147109 | Polinices | | | | | | | | | | | | | | | | |
| 390535 | Fusitriton | | | | | | | | | | | | 1 | | | | |
| 390535 | Fusitriton | | | | | | | | | | | | | | | | |
| 138432 | Retusa | | | | | | | | | | | | | | | | |
| 138217 | Dacrydium | | | | 1 | | | | 1 | | 1 | | | | 1 | 1 | |
| 138259 | Nuculana | | | | | | | | | | | | | | | | |
| 138672 | Yoldia | | | | | | | | 1 | | | | | | | | 1 |
| 138132 | Limopsis | | | | | | | | | | | | | | | | |
| 138323 | Pecten | | | | | | | | | | | | | | | | |
| 147153 | Similipecten | | | | | | | | | | | | | | | | 1 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A20_ENV_FA | A20_ENV_FB | A20_ENV_FC | A21_ENV_FA | A21_ENV_FB | A22_ENV_FA | A22_ENV_FB | A26_ENV_FA | A26_ENV_FB | A26_ENV_FC | A27_ENV_FA | A27_ENV_FB | A27_ENV_FC | A202_ENV_FA | A202_ENV_FB | A204_ENV_FA |
|----------|----------------------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|-------------|-------------|-------------|
| 197227 | Limatula hodgsoni | | | | | | | | | | | | | | | | |
| 505622 | Limea pygmaea | | | | | | | | | | | | | | | | |
| 138186 | Mysella | | | | 2 | 1 | | | | | | | | | | | |
| 138654 | Lyonsiella sp | | | | | | | | | | | | | | 1 | | |
| 197467 | Thracia meridionalis | | | | | | | | 1 | | | | | | | | |
| 137858 | Cuspidaria | | | | 1 | | | | | | | 1 | 1 | 1 | | | |
| 137863 | Tropidomya | | | | | | | | | | | | | | | | |
| 152413 | Axinulus | | | | | | | | | | | 1 | | | | | |
| 138552 | Thyasira | | | | 1 | | | | 1 | 1 | | 1 | 1 | | | | |
| 138552 | Thyasira II | | | | | 3 | | 1 | | 2 | | | | | | 4 | |
| 1806 | ECHINODERMATA | | | | | | | | | | | | | | | | |
| 172707 | Anseropoda antarctica † | | | | | | | | | | | | | | | | |
| 172769 | Acodontaster elongatus | | | | | | | | | | | | | | | 1 | |
| 123567 | Ophiophycis aff mirabilis† | | | | | | | | | | | | | | | | 1 |
| 173102 | Amphiura belgicae | | | | 1 | | | | 1 | | | | | | | | 2 |
| 123206 | Amphiuridae | | | | | | | | | | | | | | | | |
| 123200 | Ophiuridae | | | | | | | | | | | | | | | | |
| 236013 | Ophiozonella falklandica | | | | | | | | | 1 | | | 1 | | | | |
| 160790 | Brisaster antarcticus † | | | | | | 1 | | | | | | | | | | |
| 123494 | aff Thyone | | | | | | | | | | | | | | | | |
| 123473 | Echinocucumis sp | | | | | | | | | 1 | | | | | | | |
| 146121 | Psolus | | | | 1 | | | | | | | | | | | | |
| | Psolus spII | | | | | | | | | | | | | | | | |
| 123182 | Synaptidae | | | | 1 | | | | | | | | | 1 | | | |
| 123441 | Myriotrochus | | 1 | | | | | | | 1 | | | | | | 1 | |
| 1803 | BRACHIOPODA | | | | | | | | | | | | | | | | |
| 1803 | Brachiopoda | | | | 2 | | | | | | | | | | | | |
| 1803 | Brachiopoda II | | | | | | | | | | | | | | | | |
| 183339 | Pelagodiscus | | | | | | | | | | | | | | | | |
| | CHAETOGNATHA | | | | | | | | | | | | | | | | |
| 105413 | Spadella | | 1 | | | | | | | | | | | | | | |
| | | 6 | 8 | 1 | 29 | 17 | 14 | 16 | 32 | 21 | 8 | 27 | 28 | 17 | 28 | 34 | 43 |
| | | 6 | 23 | 2 | 64 | 32 | 22 | 25 | 48 | 25 | 10 | 41 | 41 | 26 | 61 | 56 | 94 |
| | | 2.791 | 2.233 | 0 | 6.733 | 4.617 | 4.206 | 4.66 | 8.008 | 6.213 | 3.04 | 7.001 | 7.271 | 4.911 | 6.568 | 8.198 | 9.244 |
| | | 1 | 0.6947 | **** | 0.8526 | 0.9613 | 0.9532 | 0.941 | 0.9513 | 0.9775 | 0.974 | 0.9336 | 0.9151 | 0.9086 | 0.8863 | 0.9449 | 0.8896 |
| | | 2.585 | 2.084 | 0 | 4.142 | 3.929 | 3.629 | 3.764 | 4.756 | 4.294 | 2.922 | 4.439 | 4.399 | 3.714 | 4.261 | 4.807 | 4.827 |
| | | 1 | 0.668 | 0 | 0.9092 | 0.9577 | 0.9524 | 0.95 | 0.9743 | 0.9833 | 0.9556 | 0.961 | 0.9488 | 0.9262 | 0.9355 | 0.974 | 0.9508 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A204_ENV_FB | A301_ENV_FA | A301_ENV_FC | A302_ENV_FA | A302_ENV_FB | A302_ENV_FC | A303_ENV_FA | A303_ENV_FB | A303_ENV_FC | A304_ENV_FA | A304_ENV_FC | A305_ENV_FA | A305_ENV_FB | A306_ENV_FA | A306_ENV_FB |
|----------|--|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| 1267 | CNIDARIA | | | | | | | | | | | | | | | |
| 128484 | <i>Pennatulidae</i> | 1 | | | | | | | | | 1 | | | | | |
| 1360 | <i>Actinaria</i> | 14 | 1 | | | | | | | | | | | 1 | | |
| 100665 | <i>Edwardsiidae</i> | | | | | | | | | | 2 | 1 | | | | |
| 173052 | <i>Flabellum (Flabellum) curvatum</i> | | 1 | | | | | | | | | | | | | 1 |
| 152391 | NEMERTEA | | | | | | | | | | | | | | | |
| 152391 | <i>Nemertea</i> | | | | | | | | | | | | 1 | | 4 | |
| 799 | NEMATODA | | | | | | | | | | | | | | | |
| 799 | <i>Nematoda</i> | 2 | | | | 1 | | | | | | | | | | 1 |
| 1268 | SIPUNCULA | | | | | | | | | | | | | | | |
| 410734 | <i>Nephasoma (Nephasoma) diaphanes</i> | 2 | | | 1 | | | 2 | | | 3 | | 1 | 2 | | 3 |
| | ECHIURIDA | | | | | | | | | | | | | | | |
| 110352 | <i>Bonellia sp</i> | | | | | | | | | | | | | | | 1 |
| 882 | ANNELIDA | | | | | | | | | | | | | | | |
| 129285 | <i>Euphrosine Sp 1 (no recorded NHM)†</i> | | | | | | | 1 | | | | | | | | |
| 173263 | <i>Euphrosine cirrata magellanica</i> | | | | | | | | | | | | | | | |
| 939 | <i>Polynoidae</i> | | | | | | | | | | | | | | | |
| 129487 | <i>Eunoe</i> | | 2 | | | | | | | | 1 | | | | | |
| 174391 | <i>Polyeunoe laevis</i> | | | | | | | | | | | | | | | |
| 174468 | <i>Harmothoe magellanica</i> | | | | | | | | | | | | | | | |
| 129590 | <i>Leanira</i> | | | | | | | | | | | | | | | |
| | <i>Sigalionidae (not recorded by NHM)†</i> | | | | | | | | | | | | | | | |
| 129443 | <i>Eteone</i> | | | | | | | 1 | | | | | | | | |
| 129450 | <i>Mystides</i> | | | | | | | | | | | | | | | |
| 130118 | <i>Glycera capitata</i> | | | | | | | | | | 2 | | 1 | 2 | | |
| 948 | <i>Syllidae</i> | 1 | 1 | | 2 | 1 | | | | | 1 | | | 1 | | |
| 129677 | <i>Sphaerosyllis</i> | | | | | | | | | | | | | | | |
| 340445 | <i>Gymnoreris fauveli</i> | | | | 2 | | | 1 | | | 2 | | 3 | 1 | 4 | 2 |
| 173522 | <i>Lumbrineris magalhaensis</i> | | | | | 2 | | | | | | 1 | | 2 | | |
| 129280 | <i>Lysidice</i> | | | | | | | | | | | 1 | | | | |
| 129366 | <i>Aglaophamus (posteriobranchiatus) NHM)</i> <i>Aglaophanus sp I</i> | | | | 1 | | | | | | | | | | | |
| 129370 | <i>Nephtys</i> | | | | | | | | | | | | 1 | | | |
| 196680 | <i>Rhynchobranchium (Spinigerium) ehlersi</i> | 1 | | 6 | 11 | 2 | 2 | 1 | 1 | 1 | 8 | 3 | 9 | 4 | 7 | 8 |
| 966 | <i>Eunicidae</i> | | | | | | | | | | | | | | | |
| 129278 | <i>Eunice</i> | | | | | | | | | | | | | | | |
| 130467 | <i>Nothria conchylega</i> | | | | | | | | | | | | | | | |
| 129425 | <i>Scoloplos</i> | | | 1 | | | | | | | | | | 2 | 1 | |
| 129430 | <i>Aricidea</i> | | | | | | | | | | | | | 2 | | |
| 326595 | <i>Aricidea (Allia) oculata</i> | | | | | | | | | | | | | | | |
| 525485 | <i>Aricidea (Acmira) strelzovi</i> | | | | | | | | | | | | | | | |
| 903 | <i>Paraonidae</i> | | | | | | | | | | 1 | | | | | |
| 129198 | <i>Apistobranchus sp I</i> | | | | | | | | | | 1 | 1 | | | | |
| 129198 | <i>Apistobranchus sp II</i> | | | | | | | | | | 2 | 1 | 3 | | 1 | 2 |
| 913 | <i>Spionidae</i> | | | | | | | | | | | | | 2 | | |
| 129620 | <i>Prionospio</i> | | | | | | | | | | | | | | | |
| 129623 | <i>Scoletepis</i> | | | | | | | | | | | | | 1 | | |
| 129613 | <i>Laonice (Laonice) sp2 NHM)</i> | | | | | | | | | | | | | | | 2 |
| 129291 | <i>Flabelligera</i> | | | | | | | | | | | | | | | |
| 129289 | <i>Brada sp</i> | | | | | | | | | | | | | | | |
| 923 | <i>Maldanidae</i> | | | 1 | 3 | 1 | 4 | | | | 2 | 3 | 2 | 1 | 2 | |
| 173557 | <i>Asychis amphiglypta</i> | | | | | 1 | | | | | | | | | | 3 |
| 130305 | <i>Maldane sarsi</i> | | | | | | | | | | | | | | | |
| 130331 | <i>Rhodine loveni</i> | | | | | | | | | | | | | | 1 | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A204_ENV_FB | A301_ENV_FA | A301_ENV_FC | A302_ENV_FA | A302_ENV_FB | A302_ENV_FC | A303_ENV_FA | A303_ENV_FB | A303_ENV_FC | A304_ENV_FA | A304_ENV_FC | A305_ENV_FA | A305_ENV_FB | A306_ENV_FA | A306_ENV_FB |
|----------|---|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| 129924 | Spiochaetopterus typicus | | | | | | | 1 | 1 | | 2 | 2 | | 1 | 3 | 2 |
| | Chaetozone sp 8 † (NHM) | | | | | | | | | | | | | | | |
| 129243 | Cirratulus Sp2 | | | | | | | | | | | | 1 | | | |
| 129245 | Pseudoscalibregma bransfeldiat | 1 | | | | | | | | | | | 2 | | 1 | 1 |
| 129241 | Cauilleriella | | 1 | | | | | | | | 2 | 2 | | | 1 | 1 |
| 129955 | Chaetozone setosa (Chaetozone sp 5 NHM) | | | | 2 | | | | | | 1 | 1 | | | 1 | |
| 173159 | Tharyx fusiformis | | | | | | | | | | | | | | | |
| 129249 | Tharyx | | | | | | | | | | | | | | | |
| 129898 | Notomastus latericeus | | | | | | | | | 1 | | | 1 | | 1 | |
| 130503 | Ophelina cylindricaudata | | | | | | | | | | | | | | | |
| 129413 | Ophelia | | | | | | | | | | | | | | | |
| 130980 | Scalibregma inflatum | | | | | | | | | | | 1 | | 1 | | |
| 152252 | Ampharetinae | | | | | | | | | | | | | | | |
| 129177 | Samytha | | | | | | | | | | | | | | | |
| 129153 | Amage | | | | 1 | | | | | | | | | | | |
| 129804 | Melinna cristata (complex) | | | | | | | | 2 | | 1 | 2 | 2 | | 1 | 2 |
| 129708 | Pista | | | | | | | | | | | | | | | |
| 131573 | Terebellides stroemii | | | | | | | | | | 1 | 3 | | 2 | 1 | 1 |
| 982 | Terebellidae | | | | | 1 | | | | | | | | | 1 | |
| 174568 | Perkinsiana antarctica | | | | | | | | | | | | 1 | | 1 | |
| 985 | Sabellidae | | | | 3 | | 1 | | | | | 1 | | 1 | 1 | |
| 154918 | Fabriciidae | | | | | | | | | 1 | | | | | | |
| 129530 | Fabriciola | | | | | | | | | | 2 | | | | | |
| 129533 | Jasmineira | | | | | | 1 | | | | | | | | | |
| 129525 | Chone | | | | | | | | | | | | | | | |
| 174564 | Euchone pallida (Euchone sp1 NHM) | | | | | 2 | | | 2 | | 3 | | | 1 | | 2 |
| 2036 | Oligochaeta | 2 | | | | | | 1 | | | 1 | | | | | |
| 988 | Serpulidae | | | | | | | | | | | | 2 | | | |
| 147018 | Microbinia | | | | | | | | | | | | 1 | | | |
| 244666 | Phylo felix | | | | | | | | | | | | | | 1 | |
| 734535 | Phylo minima | | | | | | | | | | | | | | | |
| 146949 | Galathowenia | 1 | | | | | | | | | | | | 1 | | |
| 1066 | CRUSTACEA | | | | | | | | | | | | | | | |
| 106115 | Scalpellum | | | | | | | | 1 | | | | | | 1 | |
| 106140 | Verruca | | | | | | | | | | | | | | | |
| 106137 | Altiverruca | | | | | | | | | | | | | | | |
| 1078 | Ostracoda | 1 | 1 | | | | | | | | 1 | 1 | 3 | | 18 | |
| 1080 | Copepoda | 1 | | | 6 | 4 | 30 | | 2 | 2 | | | 2 | | | |
| 101436 | Phtisica | | | | | | | | | | | | | | | 1 |
| 1135 | Amphipoda | 1 | 1 | | | | | | 1 | | | | | | | |
| 101409 | Stenothoidae | | | | | | | | | | | | | | | |
| 101409 | Stenothoidae | | | | | | | | | | | | | | | |
| 101789 | Urothoe | | | | 1 | | 1 | | | 1 | 4 | 1 | 3 | 4 | 3 | 2 |
| 101403 | Phoxocephalidae | | | | | | | | | | | | | | | |
| 101403 | Phoxocephalidae | | | | | 1 | | | | | 2 | | 1 | | 3 | |
| 101723 | Phoxocephalus | | | | | | 1 | | | | | | | | | |
| 101675 | Maera | 3 | | | | | | | | | | | | | 1 | |
| 101393 | Leucothoidae | | | | | | | | | | 1 | | | | | |
| 176788 | Lysianassoidea | | 1 | | | | | | 1 | | | | | | | |
| 176788 | Lysianassoidea | | | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | | | 4 | 1 | | 2 | | 2 | 1 | | 1 | | |
| 101445 | Ampelisca | | | | | | 1 | | | | | 2 | | | | |
| 101445 | Ampelisca | | | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | | | | | | | | | | | | | |
| 101365 | Amphilochildae | | | | | | 2 | 1 | | 1 | 2 | 1 | | | | |
| 101410 | Synopiidae | | | | | | | | | | | | | | | |
| 101563 | Photis | | | | 1 | 2 | | | | | | | | | | 1 |
| 101470 | Leptocheirus | | | | | | | | | | | | | | | |
| 101519 | Eusirus | | | | 1 | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A204_ENV_FB | A301_ENV_FA | A301_ENV_FC | A302_ENV_FA | A302_ENV_FB | A302_ENV_FC | A303_ENV_FA | A303_ENV_FB | A303_ENV_FC | A304_ENV_FA | A304_ENV_FC | A305_ENV_FA | A305_ENV_FB | A306_ENV_FA | A306_ENV_FB |
|--------------------|--------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| 101734 | Dulichia | | | | | | | | | | | | | | | |
| 118437 | Gnathia | | | | | | | | | | | | | | | |
| 257124 | Caecognathia c.f.polaris | | | | | | | | | | | | | | | |
| 118354 | Ilyarachna | | | | | | | | | | | | | | | |
| 256200 | Munna argentinae | | | | | | | | 1 | | | | | | | |
| 118371 | Macrostylis | | | | | | | | | | 1 | | | | | |
| 118371 | Macrostylis II | 2 | | | | | | | 2 | | | | | | | |
| 118351 | Haploniscus | | | | | | | | | | | | | | | |
| 118358 | Ischnomesus | | | | | | | | | | | | | | | |
| 118356 | Haplomesus | | | | | | | | | | | | | | | |
| n/a | Stylomesus elegans | | | | | | | | | | | | | | | |
| 174952 | Stylomesus | | | | | | | | | | | | | | | |
| 118319 | Desmosoma | 1 | | | 1 | 2 | | | | 2 | | | | | | |
| 118323 | Eugerdia | | | | | | | | | | | | | | | |
| 118360 | Janirella | | | | | | | | | | | | | | | |
| 248358 | Abysianira | | | | | | | | | | | | | | | |
| 118399 | Cirolana | | | | | | | | | | | | 1 | | | |
| 118399 | Cirolana | | | | | | | | 2 | | | | | | | |
| 213013 | Pseudanthura lateralis | | | | | | | | | | | | | 2 | | 3 |
| 255448 | Haliopasma antarctica | | | | | | | | | | | | | 2 | | |
| 174514 | Antarcturus | | | | | | | | 1 | | | 1 | | | | |
| 136164 | Tanaidae | | | | | | | | | | | | | | | |
| 136164 | Tanaidae | | | | | | | | | | | | | | | |
| 136222 | Agathotanales | | | | | | | | | | | | | | | |
| 136229 | Leptognathia | | | | | | | | | 2 | | | 1 | | | 1 |
| 136256 | Typhlotanales | | | | | | | | | | | | | | | |
| 136185 | Apeudes | | | | | | | | | | | | 1 | | | |
| 136200 | Pseudosphyrapus sp I | | | | | | | | | | | | | | | |
| 136200 | Pseudosphyrapus sp II | | | | | | | | | | | | | | | |
| 182287 | Eudorella gracilior | | | | | | | | | | | | | 1 | | |
| 110414 | Leucon | | | | | | | | | | | | 1 | | | |
| 110398 | Diastylis | | | | 1 | | | | 1 | | | 1 | | | | |
| 110398 | Diastylis sp II | | | | | | | | | | | | | | | |
| 110415 | Campylaspis | | | | | | | | | | | | | | | |
| 110415 | Campylaspis spII | | | | | | | | | | | | | | | |
| 110404 | Makrokylindrus | | | | | | | | | | | | | | | |
| 51 MOLLUSCA | | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | |
| 138249 | "Neomenia " | | | 1 | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | | |
| 343675 | Neopilina Lemche, 1957 | | | | | | | | | | | | | | | |
| 104 | Scaphopoda | | | | | | | | | | | | | | | |
| 138024 | Cadulus | | | | | | | | | | | | | | | |
| 138013 | Fissurella | | | | | | | | | | | | | | | |
| 138508 | Skenea | | | | | | | | | | | | | | | |
| 138508 | Skenea | | | | | | | | | | | | | | | |
| 137970 | Eulima | | | | | | | | | | | | | | | |
| 147109 | Polinices | 1 | | | | | | | | | | | | | | |
| 147109 | Polinices | | | | | | | | | | | | | | | |
| 390535 | Fusitriton | | | | | | | | | | | | | | | |
| 390535 | Fusitriton | | | | | | | | | | | | | | | |
| 138432 | Retusa | | | | | | | | | | | | | | | |
| 138217 | Dacrydium | 1 | 3 | | | | | | | 1 | | 2 | | | | |
| 138259 | Nuculana | | | | | | | | | | | | | | | |
| 138672 | Yoldia | | | | | | | | | | | 1 | | | | |
| 138132 | Limopsis | | 1 | | | | | | | | | | | | | |
| 138323 | Pecten | | | | | | | | | | | | | | | |
| 147153 | Similipecten | | | | | | | | 2 | 1 | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A204_ENV_FB | A301_ENV_FA | A301_ENV_FC | A302_ENV_FA | A302_ENV_FB | A302_ENV_FC | A303_ENV_FA | A303_ENV_FB | A303_ENV_FC | A304_ENV_FA | A304_ENV_FC | A305_ENV_FA | A305_ENV_FB | A306_ENV_FA | A306_ENV_FB |
|----------|----------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| 197227 | Limatula hodgsoni | | | | | | | | | | | | | | | |
| 505622 | Limea pygmaea | 1 | | | | | | | | | | | | | | |
| 138186 | Mysella | | | | | | | 3 | | | | | | | 1 | |
| 138654 | Lyonsiella sp | | | | | | | | | | | | | | | |
| 197467 | Thracia meridionalis | | | | | | | | | | | | | | | |
| 137858 | Cuspidaria | | | | | | | | | | | | | | | |
| 137863 | Tropidomya | | | | | | | | | | | | | | | |
| 152413 | Axinulus | | | | | | | | | | | | | | | |
| 138552 | Thyasira | | | | | | | | | | | | 2 | | | |
| 138552 | Thyasira II | | | | 1 | | | | | | 2 | 2 | | 2 | | |
| 1806 | ECHINODERMATA | | | | | | | | | | | | | | | |
| 172707 | Anseropoda antarctica † | | | | | | | | | | | | | | | |
| 172769 | Acodontaster elongatus | | | | | | | | | | | | | | | |
| 123567 | Ophiophycis aff mirabilis† | | | | | | | | | | | | | | | |
| 173102 | Amphiura belgicae | | 1 | | | | | | | | | | | | | 2 |
| 123206 | Amphiuridae | | | | | | | | | | | | | | | |
| 123200 | Ophiuridae | | | | | | | | | | | | | | | |
| 236013 | Ophiozonella falklandica | | 1 | | | | | | | | | | | | | |
| 160790 | Brisaster antarcticus † | | | | | | | | | | 1 | | | | | |
| 123494 | aff Thyone | | | | | | | | | | | | | | | |
| 123473 | Echinocucumis sp | | | | | | | | | | 1 | | | | | |
| 146121 | Psolus | | 1 | | | | | | | | | | | | | |
| | Psolus spII | | | | | | 1 | | | | | | | | | |
| 123182 | Synaptidae | | | | | | | | | | | | | | | |
| 123441 | Myriotrochus | | | | | | | | | | | | | | | |
| 1803 | BRACHIOPODA | | | | | | | | | | | | | | | |
| 1803 | Brachiopoda | | 2 | | | | | | | | | | | | | |
| 1803 | Brachiopoda II | | | | | | | | | | | | | | | |
| 183339 | Pelagodiscus | | | | | | 1 | | | | | | | | | 1 |
| | CHAETOGNATHA | | | | | | | | | | | | | | | |
| 105413 | Spadella | | | | | | | | | | | | | | | |
| | | 18 | 14 | 4 | 16 | 20 | 18 | 5 | 26 | 6 | 35 | 24 | 24 | 23 | 24 | 20 |
| | | 37 | 18 | 9 | 38 | 34 | 52 | 6 | 37 | 7 | 62 | 36 | 46 | 38 | 61 | 40 |
| | | 4.708 | 4.498 | 1.365 | 4.124 | 5.388 | 4.302 | 2.232 | 6.923 | 2.569 | 8.238 | 6.418 | 6.007 | 6.048 | 5.595 | 5.151 |
| | | 0.8211 | 0.9675 | 0.7233 | 0.8576 | 0.9594 | 0.6328 | 0.9697 | 0.9731 | 0.9755 | 0.9485 | 0.9685 | 0.9224 | 0.9623 | 0.8324 | 0.9291 |
| | | 3.424 | 3.684 | 1.447 | 3.43 | 4.146 | 2.639 | 2.252 | 4.574 | 2.522 | 4.865 | 4.44 | 4.229 | 4.353 | 3.816 | 4.015 |
| | | 0.8529 | 0.9673 | 0.5833 | 0.8876 | 0.9643 | 0.6659 | 0.9333 | 0.9805 | 0.9524 | 0.972 | 0.9762 | 0.9478 | 0.9701 | 0.8923 | 0.9436 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A307_ENV_FA | A307_ENV_FB | A309_ENV_FA | A309_ENV_FB | A1008_ENV_FA | A1008_ENV_FB | A1011_ENV_FA | A1011_ENV_FB | A1013_ENV_FA | A1013_ENV_FB | A1015_ENV_FA | A1015_ENV_FB | A3010_ENV_FA | A3010_ENV_FB |
|----------|---|-------------|-------------|-------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| 1267 | CNIDARIA | | | | | | | | | | | | | | |
| 128484 | Pennatulidae | | | | | 1 | | | | 1 | | 1 | | 1 | 2 |
| 1360 | <i>Actinaria</i> | | 1 | | | | 1 | | | | | | | | |
| 100665 | Edwardsiidae | | | | | | | | | | | | | | |
| 173052 | <i>Flabellum (Flabellum) curvatum</i> | | | | | | | | | | | | | | |
| 152391 | NEMERTEA | | | | | | | | | | | | | | |
| 152391 | Nemertea | 1 | | 1 | | | 1 | 1 | | 1 | | | 1 | | |
| 799 | NEMATODA | | | | | | | | | | | | | | |
| 799 | Nematoda | | 1 | | | | 1 | | 1 | 1 | | 1 | | | 3 |
| 1268 | SIPUNCULA | | | | | | | | | | | | | | |
| 410734 | <i>Nephasoma (Nephasoma) diaphanes</i> | | 2 | 1 | 1 | | 2 | 1 | | 3 | 1 | | 3 | | 3 |
| | ECHILURIDA | | | | | | | | | | | | | | |
| 110352 | <i>Bonellia sp</i> | | | | | | | | | | | | | | |
| 882 | ANNELIDA | | | | | | | | | | | | | | |
| 129285 | <i>Euphrosine Sp 1 (no recorded NHM)†</i> | | | | | | | | | | | 2 | | | |
| 173263 | <i>Euphrosine cirrata magellanica</i> | | | | | 1 | | | | | | | | | |
| 939 | Polynoidae | | | | | | | | | | | | | | |
| 129487 | <i>Eunoe</i> | | | | | 1 | 1 | | | 2 | | 1 | | | 1 |
| 174391 | <i>Polyeunoe laevis</i> | | | | | | | | | | | | | | |
| 174468 | <i>Harmothoe magellanica</i> | | | | | | | | | | | | | | |
| 129590 | <i>Leanira</i> | | | | | | | | | | | | | | |
| | Sigalionidae (not recorded by NHM)† | | | | | | | | | | | | | | |
| 129443 | <i>Eteone</i> | | 1 | | | | | | | | | | 2 | | |
| 129450 | <i>Mystides</i> | | | | | | | | | | | | | | |
| 130118 | <i>Glycera capitata</i> | | | | | | | | | | | | | 1 | 1 |
| 948 | Syllidae | | 3 | | | 14 | 3 | | | | | | 4 | 1 | |
| 129677 | <i>Sphaerosyllis</i> | | | | | | | | | 2 | | | 2 | | |
| 340445 | <i>Gymnonereis fauveli</i> | 1 | | | | | | 4 | 1 | | | 1 | 2 | 2 | |
| 173522 | <i>Lumbrineris magalhaensis</i> | | | | | 1 | | | | | | 1 | | 1 | |
| 129280 | <i>Lysidice</i> | | | | | | | | | | | | | | |
| 129366 | <i>Aglaophamus (posteriobranchiatus) NHM</i> | | | | | | | | | | | | 2 | | |
| | <i>Aglaophanus sp I</i> | | | | | | | | | | | | | | |
| 129370 | <i>Nephtys</i> | | | 1 | | | 1 | | | | | | | | |
| 196680 | <i>Rhynchobranchium (Spinigerium) ehlersi</i> | | 2 | 4 | 2 | 4 | | 4 | 4 | 4 | 2 | 3 | 4 | 5 | 7 |
| 966 | Eunicidae | | | | | | | | | | | | | | |
| 129278 | <i>Eunice</i> | | | | | | | | | | | | | | |
| 130467 | <i>Nothria conchylega</i> | | | | | | 2 | | 1 | | | 1 | | | 1 |
| 129425 | <i>Scoloplos</i> | | | | | | | | | | | | | | |
| 129430 | Aricidea | | | | | | | | | | | | | | |
| 326595 | Aricidea (<i>Allia</i>) <i>oculata</i> | | | | | | | | | | | | | | |
| 525485 | Aricidea (<i>Acmira</i>) <i>strelzovi</i> | | | | | | | 1 | | | | | | | 1 |
| 903 | Paraonidae | 1 | | | | | 1 | | 1 | | | | | | |
| 129198 | <i>Apistobranchus sp I</i> | 2 | 2 | | 1 | 2 | 3 | 2 | 3 | | | | | 1 | 2 |
| 129198 | <i>Apistobranchus sp II</i> | | | | | | | | | | | | | | |
| 913 | Spionidae | | | | | | | | | | | | | | |
| 129620 | <i>Prionospio</i> | | | | | | | | 1 | | | | | | |
| 129623 | <i>Scolecopsis</i> | | | | | | | | | | | | | | |
| 129613 | <i>Laonice (Laonice sp2) NHM</i> | | | | | | | 1 | | | | | | 2 | 1 |
| 129291 | <i>Flabelligera</i> | | | | | | | | | | | | 1 | | |
| 129289 | <i>Brada sp</i> | | | | | | | | | | | 1 | | | |
| 923 | Maldanidae | | | 1 | | | | | | | | | 1 | 1 | 1 |
| 173557 | <i>Asychis amphiglypta</i> | | | | | | | 1 | | | | | | | |
| 130305 | <i>Maldane sarai</i> | | | | | | | | | | | | | | |
| 130331 | <i>Rhodine loveni</i> | | | | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A307_ENV_FA | A307_ENV_FB | A309_ENV_FA | A309_ENV_FB | A1008_ENV_FA | A1008_ENV_FB | A1011_ENV_FA | A1011_ENV_FB | A1013_ENV_FA | A1013_ENV_FB | A1015_ENV_FA | A1015_ENV_FB | A3010_ENV_FA | A3010_ENV_FB |
|----------|---|-------------|-------------|-------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| 129924 | Spiochaetopterus typicus | 1 | | 1 | | | | | | 2 | | 1 | 1 | | |
| | Chaetozone sp 8 † (NHM) | | | | | | | | | | | | | | |
| 129243 | Cirratulus Sp2 | | | | | | | | | | | | | | |
| 129245 | Pseudoscalibregma bransfeldiat | | | | | | | | | | | 1 | | 1 | |
| 129241 | Caulleriella | 1 | | 2 | | | 1 | | | | | 4 | | | 2 |
| 129955 | Chaetozone setosa (Chaetozone sp 5 NHM) | | | | | | | | | | | | | | |
| 173159 | Tharyx fusiformis | | | | | | | | | | | | | | |
| 129249 | Tharyx | | | | | | | | | | | | | | |
| 129898 | Notomastus latericeus | | | | | 1 | | 1 | | | | | | | |
| 130503 | Ophelina cylindricaudata | | 1 | | | | | | | | | | | | |
| 129413 | Ophelia | | | | | | | | | | | | | | |
| 130980 | Scalibregma inflatum | | | | | | | | | | | | | | |
| 152252 | Ampharetinae | | | | | | | | | | | | | | |
| 129177 | Samytha | | | | | | | | | | | | 1 | | |
| 129153 | Amage | | | | | | | 1 | | | | | | | |
| 129804 | Melinna cristata (complex) | | | | | 3 | | | 2 | | | | 1 | | |
| 129708 | Pista | | | | | | | | | | | | | | |
| 131573 | Terebellides stroemii | 1 | | | | | | | | | | | | | |
| 982 | Terebellidae | 1 | | | | | 1 | | | | | | 1 | | |
| 174568 | Perkinsiana antarctica | | | | | | | | | | | | | | |
| 985 | Sabellidae | | | 1 | | | | | | 1 | | | 1 | 1 | 2 |
| 154918 | Fabriciidae | | 2 | | | | | 1 | | | | | | | |
| 129530 | Fabriciola | | | | | | | | | | 1 | | | | |
| 129533 | Jasminelira | | | | | | | | | | | | | | |
| 129525 | Chone | | | | | | 1 | | | | | | | | |
| 174564 | Euchone pallida (Euchone sp1 NHM) | | 1 | 1 | 1 | 3 | | | | | 2 | 1 | | 1 | |
| 2036 | Oligochaeta | | 1 | | | | 1 | 3 | | 1 | | | 2 | | 1 |
| 988 | Serpulidae | | | | | | | | | | | | | | |
| 147018 | Microrbinia | | | | | | | | | | | | | | |
| 244666 | Phylo felix | | | 2 | | | 1 | 1 | | 1 | | | | | |
| 734535 | Phylo minima | | | | | | | | | | | | | | |
| 146949 | Galathowenia | | 1 | 1 | | 1 | | | | | | | | | |
| 1066 | CRUSTACEA | | | | | | | | | | | | | | |
| 106115 | Scalpellum | | | | | | | | | | | | 1 | | |
| 106140 | Verruca | | | | | | | | | | | | | | |
| 106137 | Altiverruca | | | | | | | | | | | | | | |
| 1078 | Ostracoda | 1 | | | | | | | | | | 6 | 1 | 1 | 1 |
| 1080 | Copepoda | | | 2 | 7 | | | 3 | | 1 | 5 | 1 | 4 | | |
| 101436 | Phtisica | | 7 | | | 1 | 1 | | | | | | | | 1 |
| 1135 | Amphipoda | | | | | | | | | | | | | | |
| 101409 | Stenothoidae | | | | | | | | 1 | | | | | | |
| 101409 | Stenothoidae | | | | | | | | | | | | | | |
| 101789 | Urothoe | | 1 | 1 | | | | 5 | 1 | | | | | 2 | |
| 101403 | Phoxocephalidae | | | | | | | | | | | | | | |
| 101403 | Phoxocephalidae | | 2 | | | | | | 1 | 1 | | | | | |
| 101723 | Phoxocephalus | | | | | | | | | | | | | | |
| 101675 | Maera | 1 | | | | | | | | | 1 | 2 | | 2 | |
| 101393 | Leucothoidae | | | | | | | | | | | | | | |
| 176788 | Lysianassoidea | | 2 | | | | | | 1 | | | | 3 | | 2 |
| 176788 | Lysianassoidea | | | | | | | | | | | | | | |
| 101445 | Ampelisca | | 3 | | | 1 | 2 | | 2 | | | | | | 2 |
| 101445 | Ampelisca | | | | | | | 2 | | | | | | | |
| 101445 | Ampelisca | | | | | | | 1 | | | | | | | |
| 101445 | Ampelisca | | | | | | | | | | | | | | |
| 101365 | Amphiloichidae | | | | | | 1 | 2 | | | | | 2 | | |
| 101410 | Synopiidae | | | | | | | | | | | | | | |
| 101563 | Photis | | | | | | | | | | | | | | |
| 101470 | Leptocheirus | | | | | | | | | | | | | | |
| 101519 | Eusirus | | | | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A307_ENV_FA | A307_ENV_FB | A309_ENV_FA | A309_ENV_FB | A1008_ENV_FA | A1008_ENV_FB | A1011_ENV_FA | A1011_ENV_FB | A1013_ENV_FA | A1013_ENV_FB | A1015_ENV_FA | A1015_ENV_FB | A3010_ENV_FA | A3010_ENV_FB |
|----------|--------------------------|-------------|-------------|-------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| 101734 | Dulichia | | | | | | 3 | | | | | | | | 3 |
| 118437 | Gnathia | | | | | | | | | | | | | | |
| 257124 | Caecognathia c.f.polaris | | | | | | | | | | | | | | |
| 118354 | Ilyarachna | | | | | | | | | | | | | | |
| 256200 | Munna argentinae | | | | | | | | | | | | | | |
| 118371 | Macrostylis | | | | | | | | | | | | | | |
| 118371 | Macrostylis II | | | | | | | | | | | | | | |
| 118351 | Haplomiscus | | | | | | | | | 1 | | 1 | | | |
| 118358 | Ischnomesus | | | | | | 1 | | | | | | | | |
| 118356 | Haplomesus | | | | | | | | | | | | | | |
| n/a | Stylomesus elegans | | | | | | | | | | | | | | |
| 174952 | Stylomesus | | | | | | | | | | | 1 | | | |
| 118319 | Desmosoma | | | | | | | | | | | | 1 | 1 | 1 |
| 118323 | Eugerda | | | | | | | | | | | | | | |
| 118360 | Janirella | | | | | | | | | | | | | | |
| 248358 | Abyssianira | | | | | | | | | | | | | | |
| 118399 | Cirolana | | | | | | | | | | | | | | |
| 118399 | Cirolana | | | | | | | | | | | | | | |
| 213013 | Pseudanthura lateralis | | | | 1 | | | | | | | | | | |
| 255448 | Haliophasma antarctica | | | | | | | | | | | | | | |
| 174514 | Antarcturus | | | | | | 2 | | | 1 | | 1 | | | |
| 136164 | Tanaidae | | | | | | | | | | | | | | |
| 136164 | Tanaidae | | | | | | | 2 | | | | | | | |
| 136222 | Agathotanaia | | | | | | | 2 | | | | | | | |
| 136229 | Leptognathia | | 1 | | | 1 | 3 | 2 | 1 | | | | | 2 | 1 |
| 136256 | Typhlotanaia | | | | | | | | | | | | | | 1 |
| 136185 | Apseudes | | | | | | 1 | | | | | | 1 | | |
| 136200 | Pseudosphyrapus sp I | | 2 | | | | | | | | | | | | |
| 136200 | Pseudosphyrapus sp II | | | | | | | | | 1 | | | | | |
| 182287 | Eudorella gracillior | | | | | | | | | | | | | | |
| 110414 | Leucon | | 1 | | | | | | | | | | | | |
| 110398 | Diastylis | | | | | | 1 | | | | | 1 | | | |
| 110398 | Diastylis sp II | | 1 | | | | | | | | | | | | |
| 110415 | Campylaspis | | | | | | | | 1 | | | | | | |
| 110415 | Campylaspis spII | | | | | | | | | | | | | | |
| 110404 | Makrokyllindrus | | | | | | | | | | | | | | |
| 51 | MOLLUSCA | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | 1 | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | |
| 138249 | "Neomenia " | | 1 | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | | |
| 343675 | Neopilina Lemche, 1957 | | | | | | 1 | | | | | | | | |
| 104 | Scaphopoda | | | | | | | | | | | | | | |
| 138024 | Cadulus | | | | | | | | | | | | | | |
| 138013 | Fissurella | | | | | | | | | | | | 1 | | |
| 138508 | Skenea | | | | | | | | | | | | 1 | | |
| 138508 | Skenea | | | | | | | | | | | 1 | | | |
| 137970 | Eulima | | | | | | | | | | | | | | |
| 147109 | Pollinices | | | | | | | | | | | | | | |
| 147109 | Pollinices | | | | | | | | | | | | | | |
| 390535 | Fusitriton | | | | | | 1 | | | | | | 1 | | |
| 390535 | Fusitriton | | | | | | | | | | | | | | |
| 138432 | Retusa | | 1 | | | | | | | | | | | | |
| 138217 | Dacrydium | | | | | | | | | | | | | 1 | |
| 138259 | Nuculana | | | | | | | | | | | 1 | | | |
| 138672 | Yoldia | | | | | | | | | | | | | | 1 |
| 138132 | Limopsis | | | | | | | | | 1 | | 1 | | | |
| 138323 | Pecten | | | | | | | | | | | | | 1 | |
| 147153 | Similipecten | | | | | | | | | | | | | | 1 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | A307_ENV_FA | A307_ENV_FB | A309_ENV_FA | A309_ENV_FB | A1008_ENV_FA | A1008_ENV_FB | A1011_ENV_FA | A1011_ENV_FB | A1013_ENV_FA | A1013_ENV_FB | A1015_ENV_FA | A1015_ENV_FB | A3010_ENV_FA | A3010_ENV_FB |
|----------|----------------------------|-------------|-------------|-------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| 197227 | Limatula hodgsoni | | | | | 1 | | | | | | | | | |
| 505622 | Limea pygmaea | | | | | | | | | | | | | 1 | |
| 138186 | Mysella | | | | | | | | | | | | | 1 | |
| 138654 | Lyonsiella sp | | | | | | | | | | | | | | |
| 197467 | Thracia meridionalis | | | | | | | | | | | 1 | | | |
| 137858 | Cuspidaria | | | | | 2 | | | | 1 | | | | | |
| 137863 | Tropidomya | | | | | | | | | | | | | | |
| 152413 | Axinulus | | | | | | | 1 | | | | | | | |
| 138552 | Thyasira | | | | | | | 2 | | | | | | | |
| 138552 | Thyasira II | | | | | | | | 2 | | | | | 2 | 2 |
| 1806 | ECHINODERMATA | | | | | | | | | | | | | | |
| 172707 | Anseropoda antarctica † | | | | | | | 1 | | | | | | | |
| 172769 | Acodontaster elongatus | | | | | | | | | | | | | | |
| 123567 | Ophiophycis aff mirabilis† | | | | | | | | | | | | | | |
| 173102 | Amphiura belgicae | | | | | 2 | 1 | | | | | 2 | 2 | | |
| 123206 | Amphiuridae | | | | | | | | | | | | | | |
| 123200 | Ophiuridae | | | | | | | | | | | | | | |
| 236013 | Ophiozonella falklandica | | | | | 1 | | | | 1 | | | | | |
| 160790 | Brisaster antarcticus † | | | | | | | | 1 | | | | | | |
| 123494 | aff Thyone | | | | | | | | | | | | | | |
| 123473 | Echinocucumis sp | | | | | | | | | | | | | | 1 |
| 146121 | Psolus | | | | | | | | | | | 1 | | | |
| | Psolus spp | | | | | | | | | | | | | | |
| 123182 | Synaptidae | | 3 | | | | | | | | | | 3 | | |
| 123441 | Myriotrochus | | | | | | | | | | | | | | |
| 1803 | BRACHIOPODA | | | | | | | | | | | | | | |
| 1803 | Brachiopoda | | | | | | | | | | | | | | |
| 1803 | Brachiopoda II | | 1 | | | | | | | | | 1 | 1 | | |
| 183339 | Pelagodiscus | | | | | | | | | | | | | | |
| | CHAETOGNATHA | | | | | | | | | | | | | | |
| 105413 | Spadella | | | | | | | 1 | | | | | | | |
| | | 10 | 25 | 14 | 6 | 16 | 35 | 24 | 15 | 17 | 10 | 29 | 30 | 25 | 23 |
| | | 11 | 44 | 20 | 13 | 36 | 53 | 45 | 22 | 24 | 17 | 42 | 52 | 40 | 38 |
| | | 3.753 | 6.342 | 4.34 | 1.949 | 4.186 | 8.564 | 6.042 | 4.529 | 5.035 | 3.177 | 7.491 | 7.339 | 6.506 | 6.048 |
| | | 0.9867 | 0.9411 | 0.9513 | 0.7872 | 0.806 | 0.9676 | 0.95 | 0.9465 | 0.9509 | 0.9186 | 0.9421 | 0.9569 | 0.961 | 0.9323 |
| | | 3.278 | 4.37 | 3.622 | 2.035 | 3.224 | 4.963 | 4.356 | 3.698 | 3.887 | 3.052 | 4.576 | 4.695 | 4.463 | 4.217 |
| | | 0.9818 | 0.9609 | 0.9526 | 0.7179 | 0.8381 | 0.9826 | 0.9646 | 0.9524 | 0.9601 | 0.9044 | 0.9686 | 0.9744 | 0.9718 | 0.9545 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A3_ENV | FISA_A09_ENV | FISA_A10_ENV | FISA_A12_ENV | FISA_A14_ENV | FISA_A18_ENV | FISA_A20_ENV | FISA_A21_ENV | FISA_A22_ENV | FISA_A26_ENV | FISA_A27_ENV | FISA_A202_ENV | FISA_A204_ENV |
|----------|--|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|---------------|---------------|
| 1267 | CNIDARIA | | | | | | | | | | | | | |
| 128484 | Pennatulidae | | | | | | | | | | | | | |
| 1360 | <i>Actinaria</i> | | | 1 | | | | | | 1 | 1 | 1 | | 1 |
| 100665 | Edwardsiidae | | 1 | | | 1 | 1 | | 1 | | 1 | 1 | 1 | 28 |
| 173052 | <i>Flabellum (Flabellum) curvatum</i> | | | 1 | 1 | | | | | | | | | |
| 152391 | NEMERTEA | | | | | | | | | | | | | |
| 152391 | Nemertea | 1 | 2 | | | | 1 | 1 | 2 | | 2 | 2 | 1 | 1 |
| 799 | NEMATODA | | | | | | | | | | | | | |
| 799 | Nematoda | 2 | 2 | | 2 | | | | | | 3 | 1 | 1 | 5 |
| 1268 | SIPUNCULA | | | | | | | | | | | | | |
| 410734 | <i>Nephasoma (Nephasoma) diaphanes</i> | | 1 | | | 3 | 4 | | 6 | 2 | 3 | 1 | 1 | 4 |
| | ECHILURIDA | | | | | | | | | | | | | |
| 110352 | <i>Bonellia sp</i> | | | | | | 1 | | | | | | | |
| 882 | ANNELIDA | | | | | | | | | | | | | |
| 129285 | <i>Euphrosine Sp 1</i> (no recorded NHM)† | | | | | | | | | | 2 | | | |
| 173263 | <i>Euphrosine cirrata magellanica</i> | | | | | | 1 | | | | 2 | | | |
| 939 | Polynoidae | | | | | 2 | | | | | | | | |
| 129487 | <i>Eunoe</i> | | 1 | 2 | 1 | | 1 | | 2 | 1 | 4 | | | 2 |
| 174391 | <i>Polyeunoe laevis</i> | 1 | | | | | | | | | | | | |
| 174468 | <i>Harmothoe magellanica</i> | 1 | | | | | | | | | | | | |
| 129590 | <i>Leanira</i> | | | | | | | | | | | 1 | | |
| | Sigalionidae (not recorded by NHM)† | | | | | | | | | | | 1 | | |
| 129443 | <i>Eteone</i> | | | | | | 1 | | | | 2 | | 1 | 2 |
| 129450 | <i>Mystides</i> | | | | | | | | | | | 1 | | |
| 130118 | <i>Glycera capitata</i> | | 1 | 2 | 1 | 1 | | | | | 1 | | 2 | |
| 948 | Syllidae | 1 | 1 | | | 1 | | 1 | | | 2 | 1 | | 1 |
| 129677 | <i>Sphaerosyllis</i> | | | | | | | | | | | 2 | | |
| 340445 | <i>Gymnonereis fauveli</i> | 9 | 2 | 2 | 1 | 7 | 3 | | | | 3 | | 10 | |
| 173522 | <i>Lumbrineris magalhaensis</i> | | 3 | | | | | 1 | 5 | 1 | | 2 | | 1 |
| 129280 | <i>Lysidice</i> | | | | 1 | | | | | | | | | |
| 129366 | <i>Aglaophamus (posteriobranchiatus</i> NHM) | | 1 | 1 | | | 1 | | | | | | | |
| | <i>Aglaophanus sp I</i> | | | | | | 1 | | | | | | | |
| 129370 | Nephtys | | | | | 1 | | | | | | 2 | | |
| 196680 | <i>Rhynchophobranchium (Spinigerium) ehlersi</i> | 1 | 15 | 16 | 1 | 26 | 8 | 5 | 21 | 5 | 9 | 23 | 8 | 1 |
| 966 | Eunicidae | | | | | | | | | | | | | 1 |
| 129278 | Eunice | | | | | | | | | | | 1 | | |
| 130467 | <i>Nothria conchylega</i> | | | 1 | | 1 | 1 | | | | 1 | | | |
| 129425 | Scoloplos | | | | | | | | 1 | | | 1 | | |
| 129430 | Aricidea | 3 | 1 | | | 3 | | | | | | | | |
| 326595 | Aricidea (<i>Allia</i>) <i>oculata</i> | | | 1 | | | | | | | | | | |
| 525485 | Aricidea (<i>Acmira</i>) <i>strelzovi</i> | | | 2 | 1 | 1 | | | | | | 1 | | |
| 903 | Paraonidae | 1 | 3 | | | 1 | | | | | | | | |
| 129198 | <i>Apistobranchus sp I</i> | 3 | 6 | 3 | | 3 | 1 | | | | 1 | | 8 | |
| 129198 | <i>Apistobranchus sp II</i> | | | | | | | | | | | | | |
| 913 | Spionidae | | | | | 1 | | | | | | | | |
| 129620 | <i>Prionospio</i> | | 1 | 1 | | | | | | | | 1 | 1 | |
| 129623 | <i>Scoletepis</i> | | | | 1 | | 1 | | 1 | | | | 1 | |
| 129613 | <i>Laonice (Laonice sp2</i> NHM) | | | | | | | | | | | | | |
| 129291 | <i>Flabelligera</i> | | | | | | | | | | | | | |
| 129289 | <i>Brada sp</i> | | | | | | | | | | 1 | | | 3 |
| 923 | Maldanidae | 1 | 1 | 4 | 2 | 2 | 1 | | | 2 | 2 | 1 | 1 | |
| 173557 | <i>Asychis amphiglypta</i> | | | 1 | 1 | | | | | 2 | 3 | | 1 | |
| 130305 | <i>Maldane sarsi</i> | | | | | | | | | 1 | | | | |
| 130331 | <i>Rhodine loveni</i> | | | | | | 2 | | | | | 2 | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A3_ENV | FISA_A09_ENV | FISA_A10_ENV | FISA_A12_ENV | FISA_A14_ENV | FISA_A18_ENV | FISA_A20_ENV | FISA_A21_ENV | FISA_A22_ENV | FISA_A26_ENV | FISA_A27_ENV | FISA_A202_ENV | FISA_A204_ENV |
|-----------------------|---|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|---------------|---------------|
| 129924 | Spiochaetopterus typicus | 1 | 2 | 1 | | 2 | | 2 | | | 1 | 2 | 2 | 1 |
| | Chaetozone sp 8 † (NHM) | | | | | | | | | | | | | 1 |
| 129243 | Cirratulus Sp2 | | 1 | | | | 3 | | | | | | | 1 |
| 129245 | Pseudoscallibregma bransfeldiat | | | | | 2 | 2 | | | | 1 | | 1 | 1 |
| 129241 | Caulleriella | 1 | 5 | 5 | | 3 | 1 | | 1 | 6 | 1 | 5 | 3 | 1 |
| 129955 | Chaetozone setosa (Chaetozone sp 5 NHM) | 1 | 1 | | | 1 | | 1 | 1 | 1 | 1 | | | |
| 173159 | Tharyx fusiformis | | | | | 1 | 1 | | | | | 1 | | |
| 129249 | Tharyx | | | | | | | | | | | 2 | | |
| 129898 | Notomastus latericeus | | | 1 | | | | | 1 | | 1 | 2 | 1 | |
| 130503 | Ophelina cylindricaudata | | | 1 | | | 1 | | | | | | 1 | |
| 129413 | Ophelia | 1 | | | | | | | | | | | | |
| 130980 | Scalibregma inflatum | | 1 | | | | 1 | | | | | | | |
| 152252 | Ampharetinae | | | | | | | | | | | 1 | | |
| 129177 | Samytha | | | 1 | | | 1 | | | | | 1 | | |
| 129153 | Amage | 1 | | | | | 1 | | | | | | | |
| 129804 | Melinna cristata (complex) | | 3 | 9 | 1 | 1 | 2 | | | | | | 1 | |
| 129708 | Pista | 1 | | | | | | | | | | 1 | | |
| 131573 | Terebellides stroemii | 1 | 2 | 1 | | 2 | 1 | | | | 1 | 1 | | |
| | 982 Terebellidae | | 1 | 1 | | | | | | | 2 | | | |
| 174568 | Perkinsiana antarctica | | | | | | | | | | | | 3 | |
| | 985 Sabellidae | 2 | | 2 | | 2 | | | | 1 | 1 | 2 | | |
| 154918 | Fabriciidae | | | | 1 | 1 | 1 | | | | 1 | | | |
| 129530 | Fabriciola | | | 1 | | | | | | | | 1 | 3 | |
| 129533 | Jasmineira | 1 | | | | | | | | | | | | |
| 129525 | Chone | 1 | | | | | | | | | | | | |
| 174564 | Euchone pallida (Euchone sp1 NHM) | 2 | 3 | 3 | 1 | | 1 | 1 | 1 | | | 2 | 2 | 2 |
| | 2036 Oligochaeta | 1 | 4 | 2 | | 2 | | | | | | 4 | 1 | 2 |
| | 988 Serpulidae | | | | | | | | | | | | | |
| 147018 | Microrbinia | 2 | | | | | | | | | | | | |
| 244666 | Phylo felix | | | | 1 | 3 | | | | | | 1 | 1 | |
| 734535 | Phylo minima | 2 | | | | | | | | | | | | |
| 146949 | Galathowenia | 1 | | 1 | 1 | 1 | | | | 1 | 1 | 2 | | 1 |
| 1066 CRUSTACEA | | | | | | | | | | | | | | |
| 106115 | Scalpellum | | 2 | | | | | | | | | | | |
| 106140 | Verruca | | | | | | | | | 1 | 1 | | | |
| 106137 | Altiverruca | | | | | | | | | | | | | 14 |
| | 1078 Ostracoda | 1 | 1 | | | 2 | | 16 | 1 | | 3 | 1 | 1 | 3 |
| | 1080 Copepoda | 1 | 1 | 1 | 2 | 1 | | | | | 2 | 1 | 14 | 3 |
| 101436 | Phtisica | | | | | | | | 1 | | | | | 2 |
| | 1135 Amphipoda | 2 | 2 | | | 1 | | | | | 1 | 2 | 2 | 4 |
| 101409 | Stenothoidae | | | | | | | | | | 2 | | | |
| 101409 | Stenothoidae | | | | | | | | 3 | | | | | 2 |
| 101789 | Urothoe | 5 | 1 | 2 | | 2 | 1 | | 2 | 1 | | 4 | 7 | |
| 101403 | Phoxocephalidae | | 1 | | | 1 | | | | | | | | |
| 101403 | Phoxocephalidae | 1 | | 2 | | 4 | 1 | | 5 | | 1 | 2 | 1 | 2 |
| 101723 | Phoxocephalus | | | 1 | | | | | | | | | | |
| 101675 | Maera | | 1 | | | | | | 1 | 7 | 1 | | 2 | 5 |
| 101393 | Leucothoidae | | | | | 1 | | | | | | | 2 | |
| 176788 | Lysianassoidea | 2 | | | | | | | | | | 1 | 1 | 1 |
| 176788 | Lysianassoidea | 12 | | | | 1 | | | | | | | | |
| 101445 | Ampelisca | 3 | 11 | 7 | 1 | 6 | | | 2 | 1 | 1 | 3 | 3 | 1 |
| 101445 | Ampelisca | 4 | | | | | | | | | | | | |
| 101445 | Ampelisca | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | | | | 3 | 1 | | | | | | |
| 101365 | Amphilocheidae | 3 | 1 | | 1 | | 1 | | | | | 2 | | |
| 101410 | Synopiidae | | | | | | | | | | | | | |
| 101563 | Photis | | | 2 | | 2 | | | 2 | 3 | 1 | 1 | | 2 |
| 101470 | Leptocheirus | | 2 | | | 1 | | | | | | | 5 | |
| 101519 | Eusirus | | | | | | | | 1 | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A3_ENV | FISA_A09_ENV | FISA_A10_ENV | FISA_A12_ENV | FISA_A14_ENV | FISA_A18_ENV | FISA_A20_ENV | FISA_A21_ENV | FISA_A22_ENV | FISA_A26_ENV | FISA_A27_ENV | FISA_A202_ENV | FISA_A204_ENV |
|--------------------|--------------------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|---------------|---------------|
| 101734 | Dulichia | | | | | | | | 3 | | | | 1 | |
| 118437 | Gnathia | | | | | | | | | | | 1 | | |
| 257124 | Caecognathia c.f.polaris | | | | | | | | | | | | | 1 |
| 118354 | Ilyarachna | | | | | | | | | | | 1 | | 1 |
| 256200 | Munna argentinae | | | | | | | | | | | | | |
| 118371 | Macrostylis | | | | | | | | | | | | | 1 |
| 118371 | Macrostylis II | | | | | | | | | | | | | 3 |
| 118351 | Haploniscus | | | | | | | | | | | | 1 | 1 |
| 118358 | Ischnomesus | | 1 | | | | | | | 2 | | | | |
| 118356 | Haplomesus | | | | | | 1 | | | | | | | |
| n/a | Stylomesus elegans | | | | | | | | 1 | | | | | |
| 174952 | Stylomesus | | 1 | | | | | | | | | | | 4 |
| 118319 | Desmosoma | 2 | 2 | | | | 1 | | | | 1 | | 2 | 2 |
| 118323 | Eugerdia | | 1 | | | | | | | | | | | |
| 118360 | Janirella | | | | | | | | | | | | | 1 |
| 248358 | Abysstanira | | | | | | | | 1 | | | | | |
| 118399 | Cirolana | 2 | 1 | | | | | | | | 1 | | | |
| 118399 | Cirolana | | | | | | | | | | | | | |
| 213013 | Pseudanthura lateralis | 1 | 1 | 2 | | | | | | | 1 | 1 | | |
| 255448 | Haliophasma antarctica | 1 | | 1 | | | | | | 1 | | 1 | | |
| 174514 | Antarcturus | | 1 | 1 | | | 14 | | | 1 | | | | |
| 136164 | Tanaidae | 1 | | | | | | | | | | | | 1 |
| 136164 | Tanaidae | | | | | | | | | | | | | |
| 136222 | Agathotanales | 2 | 1 | | | | 1 | | | | | | 1 | |
| 136229 | Leptognathia | 5 | 4 | 1 | 1 | | 5 | | 6 | 3 | 1 | | 2 | 1 |
| 136256 | Typhlotanales | | | | | | 2 | | | | | | | |
| 136185 | Apeudes | | | | | | 1 | | | 1 | | | | |
| 136200 | Pseudosphyrapus sp I | | | | | | | | | | | 1 | 1 | |
| 136200 | Pseudosphyrapus sp II | | | | | | | | | | | | | |
| 182287 | Eudorella gracilior | | | | | | | | | | | | | |
| 110414 | Leucon | 1 | | | | | | | | | | | 1 | |
| 110398 | Diastylis | 2 | | 1 | | | 1 | | 5 | | | 2 | 2 | 3 |
| 110398 | Diastylis sp II | | | | | | 1 | | | | | | 1 | 3 |
| 110415 | Campylaspis | | | | | | | | 1 | | | | | |
| 110415 | Campylaspis spII | | | | | | | | | | | | | 1 |
| 110404 | Makrokylinidrus | 1 | | | | | | | | | | | | |
| 51 MOLLUSCA | | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | 1 |
| 138249 | "Neomenia" | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | 1 | | | | |
| 343675 | Neopilina Lemche, 1957 | | | | | | | | 1 | | | | | |
| 104 | Scaphopoda | | | | | | | | | | | | 1 | |
| 138024 | Cadulus | 1 | | | | | | | | | | | | 1 |
| 138013 | Fisurella | | | | | | | | | | | | | |
| 138508 | Skenea | | | | 1 | | | | | | | | | |
| 138508 | Skenea | | | | | | | | | | | | | |
| 137970 | Eulima | | | | | | 1 | | | | | | | 1 |
| 147109 | Polinices | | | | | | | | | | | | | |
| 147109 | Polinices | | | | | | | | | | | | | |
| 390535 | Fusitriton | | 1 | 1 | | | 1 | | | | | 1 | | |
| 390535 | Fusitriton | 2 | | | | | | | | | | | | |
| 138432 | Retusa | | | | | | | | | | | | | |
| 138217 | Dacrydium | | | | | | | | 1 | | 2 | | 2 | 1 |
| 138259 | Nuculana | | | | | | 1 | | | | | | | 1 |
| 138672 | Yoldia | | | | | | 2 | | | | 1 | | | |
| 138132 | Limopsis | | 1 | | | | | | | | | | | |
| 138323 | Pecten | | | | | | | | | | | | | |
| 147153 | Similipecten | | | | | | | | | | | 1 | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A3_ENV | FISA_A09_ENV | FISA_A10_ENV | FISA_A12_ENV | FISA_A14_ENV | FISA_A18_ENV | FISA_A20_ENV | FISA_A21_ENV | FISA_A22_ENV | FISA_A26_ENV | FISA_A27_ENV | FISA_A202_ENV | FISA_A204_ENV |
|---------------------------|----------------------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|---------------|---------------|
| 197227 | Limatula hodgsoni | 1 | | 1 | | | | | | | | | | 1 |
| 505622 | Limnaea pygmaea | | | | | | | | 3 | | | | 1 | |
| 138186 | Mysella | | | | | | | | | | | | | |
| 138654 | Lyonsiella sp | | | | | | | | | | | | 1 | |
| 197467 | Thracia meridionalis | | 1 | | | | | | | | 1 | | | |
| 137858 | Cuspidaria | 2 | | 1 | | | | | 1 | | | 3 | | |
| 137863 | Tropidomya | | | | | | 1 | | | | | | | |
| 152413 | Axinulus | | | | 1 | 2 | | | | | | 1 | | |
| 138552 | Thyasira | | 2 | | | 1 | | | 1 | | 2 | 2 | | |
| 138552 | Thyasira II | | 2 | 2 | | 2 | | | 3 | 1 | 2 | | 4 | |
| 1806 ECHINODERMATA | | | | | | | | | | | | | | |
| 172707 | Anseropoda antarctica † | | | | | | 1 | | | | | | | |
| 172769 | Acodontaster elongatus | | | | | | | | | | | | 1 | |
| 123567 | Ophiophycis aff mirabilis† | | | | | | | | | | | | | 1 |
| 173102 | Amphiura belgicae | 1 | | 1 | | | | | 1 | | | | | 2 |
| 123206 | Amphiuridae | 1 | | | | | | | | | | | | |
| 123200 | Ophiuridae | | | | | | | | | | | | | |
| 236013 | Ophiozonella falklandica | | | 1 | | 1 | | | | | 1 | 1 | | |
| 160790 | Brisaster antarcticus † | | | | | | | | 1 | | | | | |
| 123494 | aff Thyone | | | | | | | | | | | | | |
| 123473 | Echinocucumis sp | | | | | | | | 1 | | 1 | | | |
| 146121 | Psolus | | | | | | | | | | | | | |
| | Psolus spp | | | | | | | | | | | | | |
| 123182 | Synaptidae | | | | | | | | 1 | | | 1 | | |
| 123441 | Myriotrochus | | | | | | | 1 | | | 1 | | 1 | |
| 1803 BRACHIOPODA | | | | | | | | | | | | | | |
| 1803 | Brachiopoda | | | | | | | | 2 | | | | | |
| 1803 | Brachiopoda II | | | | | | | | | | | | | |
| 183339 | Pelagodiscus | | | | | | | | | | | | | |
| CHAETOGNATHA | | | | | | | | | | | | | | |
| 105413 | Spadella | | | | | | | 1 | | | | | | |
| | | 51 | 50 | 44 | 22 | 47 | 42 | 11 | 39 | 24 | 50 | 56 | 50 | 52 |
| | | 102 | 108 | 95 | 25 | 123 | 65 | 31 | 96 | 47 | 83 | 108 | 117 | 131 |
| | | 10.81 | 10.47 | 9.443 | 6.524 | 9.559 | 9.822 | 2.912 | 8.325 | 5.974 | 11.09 | 11.75 | 10.29 | 10.46 |
| | | 0.9209 | 0.9002 | 0.8885 | 0.9875 | 0.854 | 0.9346 | 0.7084 | 0.8692 | 0.9132 | 0.9496 | 0.8871 | 0.8951 | 0.854 |
| | | 5.224 | 5.081 | 4.851 | 4.404 | 4.744 | 5.04 | 2.451 | 4.594 | 4.187 | 5.359 | 5.152 | 5.052 | 4.868 |
| | | 0.9703 | 0.9619 | 0.953 | 0.99 | 0.9351 | 0.9731 | 0.7183 | 0.9366 | 0.9482 | 0.9797 | 0.9486 | 0.963 | 0.9372 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A301_ENV | FISA_A302_ENV | FISA_A303_ENV | FISA_A304_ENV | FISA_A305_ENV | FISA_A306_ENV | FISA_A307_ENV | FISA_A309_ENV | FISA_A1008_ENV | FISA_A1011_ENV | FISA_A1013_ENV | FISA_A1015_ENV | FISA_A3010_ENV |
|----------|---|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|----------------|----------------|----------------|----------------|----------------|
| 1267 | CNIDARIA | | | | | | | | | | | | | |
| 128484 | Pennatulidae | | | | 1 | | | | | 1 | | 1 | | |
| 1360 | Actinaria | 1 | | | | | | 1 | | | | | 1 | 1 |
| 100665 | Edwardsiidae | | | | 3 | 1 | | | | 1 | | | | 2 |
| 173052 | <i>Flabellum (Flabellum) curvatum</i> | 1 | | | | | 1 | | | | | | | |
| 152391 | NEMERTEA | | | | | | | | | | | | | |
| 152391 | Nemertea | | | | | 1 | 4 | 1 | 1 | 1 | 1 | 1 | 1 | |
| 799 | NEMATODA | | | | | | | | | | | | | |
| 799 | Nematoda | | 1 | | | | 1 | 1 | | 1 | 1 | 1 | 1 | 3 |
| 1268 | SIPUNCULA | | | | | | | | | | | | | |
| 410734 | Nephasoma (<i>Nephasoma</i>) diaphanes | | 1 | 2 | 3 | 3 | 3 | 2 | 2 | 2 | 1 | 4 | 3 | 3 |
| | ECHIURIDA | | | | | | | | | | | | | |
| 110352 | <i>Bonellia</i> sp | | | | | | 1 | | | | | | | |
| 882 | ANNELIDA | | | | | | | | | | | | | |
| 129285 | <i>Euphrosine</i> Sp 1 (no recorded NHM)† | | | 1 | | | | | | | | | 2 | |
| 173263 | <i>Euphrosine cirrata magellanica</i> | | | | | | | | | 1 | | | | |
| 939 | Polynoidae | | | | | | | | | | | | | |
| 129487 | <i>Eunoe</i> | 2 | | | 1 | | | | | 2 | | 2 | 1 | 1 |
| 174391 | <i>Polyeunoe laevis</i> | | | | | | | | | | | | | |
| 174468 | <i>Harmothoe magellanica</i> | | | | | | | | | | | | | |
| 129590 | <i>Leanira</i> | | | | | | | | | | | | | |
| | Sigalionidae (not recorded by NHM)† | | | | | | | | | | | | | |
| 129443 | <i>Eteone</i> | | | 1 | | | | 1 | | | | | 2 | |
| 129450 | <i>Mystides</i> | | | | | | | | | | | | | |
| 130118 | <i>Glycera capitata</i> | | | | 2 | 3 | | | | | | | | 2 |
| 948 | Syllidae | 1 | 3 | | 1 | 1 | 1 | 3 | | 17 | | | 4 | 1 |
| 129677 | <i>Sphaerosyllis</i> | | | | | | | | | | | 2 | 2 | |
| 340445 | <i>Gymnonereis fauveli</i> | | 2 | 1 | 2 | 4 | 6 | 1 | | | 5 | | 3 | 2 |
| 173522 | <i>Lumbrineris magalhaensis</i> | | 2 | | 1 | 2 | | | | 1 | | | 1 | 1 |
| 129280 | <i>Lysidice</i> | | | | 1 | | | | | | | | | |
| 129366 | <i>Aglaophamus (posteriobranchiatus</i> NHM) | | 1 | | | | | | | | | | 2 | |
| | <i>Aglaophanus</i> sp I | | | | | | | | | | | | | |
| 129370 | <i>Nephtys</i> | | | | | 1 | | | 1 | 1 | | | | |
| 196680 | <i>Rhynchobranchium (Spinigerium) ehlersi</i> | 6 | 15 | 3 | 11 | 13 | 15 | 2 | 6 | 4 | 8 | 6 | 7 | 12 |
| 966 | Eunicidae | | | | | | | | | | | | | |
| 129278 | <i>Eunice</i> | | | | | | | | | | | | | |
| 130467 | <i>Nothria conchylega</i> | | 1 | 1 | | | | | | 2 | | | 1 | 1 |
| 129425 | <i>Scoloplos</i> | 1 | | | 1 | 2 | 1 | | | | | | | |
| 129430 | Aricidea | | | 1 | | | | | | | | | | |
| 326595 | Aricidea (<i>Allia</i>) oculata | | | | | | | | | | | | | |
| 525485 | Aricidea (<i>Acmira</i>) strelzovi | | | | | 1 | | | | | | | | 1 |
| 903 | Paraonidae | | 1 | 3 | 2 | | | 1 | | 1 | | | | |
| 129198 | <i>Apistobranchus</i> sp I | | 1 | 3 | 3 | 3 | 3 | 4 | 1 | 5 | 5 | | | 3 |
| 129198 | <i>Apistobranchus</i> sp II | | | | | 2 | | | | | | | | |
| 913 | Spionidae | | | | | | | | | | | | | |
| 129620 | <i>Prionospio</i> | | | | 1 | | | | | | 1 | | | |
| 129623 | <i>Scolecipis</i> | | | | | | | | | | | | | |
| 129613 | <i>Laonice (Laonice</i> sp2 NHM) | | | 1 | | | | 2 | | | 1 | | | 3 |
| 129291 | <i>Flabelligera</i> | | | | | | | | | | | | 1 | |
| 129289 | <i>Brada</i> sp | | | | | | | | | | | | 1 | |
| 923 | Maldanidae | 1 | 8 | | 5 | 3 | 2 | | 1 | | | | 1 | 2 |
| 173557 | <i>Asychis amphiglypta</i> | | 1 | 1 | | | 3 | | | | 1 | | | |
| 130305 | <i>Maldane sarsi</i> | | | | | | | | | | | | | |
| 130331 | <i>Rhodine loveni</i> | | | | | | 1 | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A301_ENV | FISA_A302_ENV | FISA_A303_ENV | FISA_A304_ENV | FISA_A305_ENV | FISA_A306_ENV | FISA_A307_ENV | FISA_A309_ENV | FISA_A1008_ENV | FISA_A1011_ENV | FISA_A1013_ENV | FISA_A1015_ENV | FISA_A3010_ENV |
|----------|---|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|----------------|----------------|----------------|----------------|----------------|
| 129924 | Spiochaetopterus typicus | | | 2 | 4 | 1 | 5 | 1 | 1 | | | 2 | 2 | |
| | Chaetozone sp 8 † (NHM) | | | | | | | | | | | | | |
| 129243 | Cirratulus Sp2 | | | | | 1 | | | | | | | 1 | 1 |
| 129245 | Pseudoscalibregma bransfieldia† | 1 | | | 4 | 2 | 2 | 1 | 2 | 1 | | | 4 | 2 |
| 129241 | Caulerella | | | | | | | | | | | | | |
| 129955 | Chaetozone setosa (Chaetozone sp 5 NHM) | | 2 | | 2 | | 1 | | | | | | | |
| 173159 | Tharyx fusiformis | | | | | | | | | | | | | |
| 129249 | Tharyx | | | | | | | | | | | | | |
| 129898 | Notomastus latericeus | | | 1 | 1 | 1 | 1 | | | 1 | 1 | | | |
| 130503 | Ophelina cylindricaudata | | | | | | | 1 | | | | | | |
| 129413 | Ophelia | | | | | | | | | | | | | |
| 130980 | Scalibregma inflatum | | | | 1 | 1 | | | | | | | | |
| 152252 | Ampharetinae | | | | | | | | | | | | | |
| 129177 | Samytha | | | | | | | | | | | | 1 | |
| 129153 | Amage | | 1 | | | | | | | | | | | |
| 129804 | Melinna cristata (complex) | | | 2 | 3 | 2 | 3 | | | 3 | 2 | 1 | 1 | |
| 129708 | Pista | | | | | | | | | | | | | |
| 131573 | Terebellides stroemii | | | | 4 | 2 | 2 | 1 | | | | | | |
| | 982 Terebellidae | | 1 | | | | 1 | 1 | | 1 | | | 1 | |
| 174568 | Perkinsiana antarctica | | | | | | | | | | | | | |
| | 985 Sabellidae | | 4 | | 1 | 1 | 1 | | 1 | | | 1 | 2 | 2 |
| 154918 | Fabriciidae | | | 1 | 2 | | | 2 | | | | 1 | | |
| 129530 | Fabriciola | | | | | | | | | | 1 | 1 | | |
| 129533 | Jasmineira | | 1 | | | | | | | | | | | |
| 129525 | Chone | | | | | | | | | 1 | | | | |
| 174564 | Euchone pallida (Euchone sp1 NHM) | | 2 | 2 | 3 | 1 | 2 | 1 | 2 | 3 | | 2 | 1 | 1 |
| | 2036 Oligochaeta | | | 1 | 1 | 2 | | 1 | | 1 | 3 | 1 | 2 | 1 |
| | 988 Serpulidae | | | | | 1 | | | | | | | | |
| 147018 | Microrbinia | | | | | | | | | | | | | |
| 244666 | Phylo felix | | | 1 | | 1 | | | 2 | 1 | 1 | 1 | | |
| 734535 | Phylo minima | | | | | | | | | | | | | |
| 146949 | Galathowenia | | | | | 1 | | 1 | 1 | 1 | | | | |
| | 1066 CRUSTACEA | | | | | | | | | | | | | |
| 106115 | Scalpellum | | | 1 | | | 1 | | | | | | 1 | |
| 106140 | Verruca | | | | | | | | | | | | | |
| 106137 | Altiterruca | | | | | | | | | | | | 7 | |
| | 1078 Ostracoda | 1 | | | 2 | 3 | 18 | 1 | | 1 | | | 5 | 2 |
| | 1080 Copepoda | | 40 | 4 | | 2 | | | 9 | | 3 | 6 | | |
| 101436 | Phtisica | | | | | | | | | | | | | |
| | 1135 Amphipoda | 1 | | 1 | | | | 7 | | 2 | | | | 1 |
| 101409 | Stenothoidae | | | | | | | | | | | | | |
| 101409 | Stenothoidae | | | | | | | | | 1 | | | | |
| 101789 | Urothoe | | 2 | 1 | 5 | 7 | 5 | 1 | 1 | | 6 | | 1 | 2 |
| 101403 | Phoxocephalidae | | | | | | | | | | | | | |
| 101403 | Phoxocephalidae | | 1 | | 2 | 1 | 3 | 2 | | 1 | 1 | 1 | | |
| 101723 | Phoxocephalus | | 1 | | | | | | | | | | | |
| 101675 | Maera | | | | | | 1 | 1 | | | | 1 | 2 | 2 |
| 101393 | Leucothoidae | | | | 1 | | | | | | | | | |
| 176788 | Lysianassoidea | 1 | | 1 | | | | 2 | | | | | 3 | 2 |
| 176788 | Lysianassoidea | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | 2 | | | | | | 3 | | | | 2 |
| 101445 | Ampelisca | | 1 | | 2 | 1 | | 3 | | | | | | 2 |
| 101445 | Ampelisca | | | | | | | | | | | | | |
| 101445 | Ampelisca | | | | | | | | | | | | | |
| 101365 | Amphiloichidae | | 3 | 1 | 3 | | | | | 1 | 2 | | 2 | |
| 101410 | Synopiidae | | | 1 | | | | | | | | | | |
| 101563 | Photis | | 3 | | | | | 1 | | | | | | |
| 101470 | Leptocheirus | | | | | | | | | | | | | |
| 101519 | Eusirus | | 1 | | | | | | | | | | | |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A301_ENV | FISA_A302_ENV | FISA_A303_ENV | FISA_A304_ENV | FISA_A305_ENV | FISA_A306_ENV | FISA_A307_ENV | FISA_A309_ENV | FISA_A1008_ENV | FISA_A1011_ENV | FISA_A1013_ENV | FISA_A1015_ENV | FISA_A3010_ENV |
|----------|--------------------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|----------------|----------------|----------------|----------------|----------------|
| 101734 | Dulichia | | | | | | | | | 3 | | | | 3 |
| 118437 | Gnathia | | | | | | | | | | | | | |
| 257124 | Caecognathia c.f.polaris | | | | | | | | | | | | | |
| 118354 | Ilyarachna | | | | | | | | | | | | | |
| 256200 | Munna argentinae | | | 1 | | | | | | | | | | |
| 118371 | Macrostylis | | | | 1 | | | | | | | | | |
| 118371 | Macrostylis II | | | | | | | | | | | | | |
| 118351 | Haploniscus | | 2 | | | | | | | | | 1 | 1 | |
| 118358 | Ischnomesus | | | | | | | | | 1 | | | | |
| 118356 | Haplomesus | | | | | | | | | | | | | |
| n/a | Stylomesus elegans | | | | | | | | | | | | | |
| 174952 | Stylomesus | | | | | | | | | | | | 1 | |
| 118319 | Desmosoma | | 3 | 2 | | | | | | | | | 1 | 2 |
| 118323 | Eugerdia | | | | | | | | | | | | | |
| 118360 | Janirella | | | | | | | | | | | | | |
| 248358 | Abyssianira | | | | | | | | | | | | | |
| 118399 | Cirolana | | | | | | | | | | | | | |
| 118399 | Cirolana | | 2 | | | | | | | | | | | |
| 213013 | Pseudanthura lateralis | | | | | | | | | 1 | | | | |
| 255448 | Hallophasma antarctica | | | | | | | | | | | | | |
| 174514 | Antarcturus | | 1 | | 1 | | | | | 2 | | 1 | 1 | |
| 136164 | Tanaidae | | | | | | | | | | | | | |
| 136164 | Tanaidae | | | | | | | | | | | | | |
| 136222 | Agathotanaia | | | | | | | | | | | | | |
| 136229 | Leptognathia | | | 2 | | | | 1 | | 4 | 2 | | | 3 |
| 136256 | Typhlotanaia | | | | | | | 1 | | | 3 | | | 1 |
| 136185 | Apseudes | | | | | | | | | 1 | | | | |
| 136200 | Pseudosphyrapus sp I | | | | | | | | | | | | | |
| 136200 | Pseudosphyrapus sp II | | | | | | | | | | | | | |
| 182287 | Eudorella gracillor | | | | | | | | | | | 1 | | |
| 110414 | Leucon | | | | | | | | | | | | | |
| 110398 | Diastylis | | 2 | 1 | 1 | | | | | 1 | | | | |
| 110398 | Diastylis sp II | | 1 | | | | | | | | | | | |
| 110415 | Campylaspis | | 1 | | | | | | | | 1 | | | |
| 110415 | Campylaspis spII | | | | | | | | | | | | | |
| 110404 | Makrokyllindrus | | | | | | | | | | | | | |
| 51 | MOLLUSCA | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | |
| 138249 | "Neomenia " | | 1 | | | | | | | | | | | |
| 138249 | "Neomenia" | | | | | | | | | | | | | |
| 343675 | Neopilina Lemche, 1957 | | | | | | | | | | | | | |
| 104 | Scaphopoda | | | | | | | | | | | | | |
| 138024 | Cadulus | | | | | | | | | | | | | |
| 138013 | Fissurella | | | | | | | | | | | | | |
| 138508 | Skenea | | | | | | | | | | | | 1 | |
| 138508 | Skenea | | | | | | | | | | | | 1 | |
| 137970 | Eulima | | | | | | | | | | | | | |
| 147109 | Polinices | | | | | | | | | | | | | |
| 147109 | Polinices | | | | | | | | | | | | | |
| 390535 | Fusitriton | | | | | | | | | | | | 1 | |
| 390535 | Fusitriton | | | | | | | | | | | | | |
| 138432 | Retusa | | | | | | | | | | | | | |
| 138217 | Dacrydium | | 3 | 1 | | 2 | | | | | | | | 1 |
| 138259 | Nuculana | | | | | | | | | | | | 1 | |
| 138672 | Yoldia | | | | | 1 | | | | | | | | 1 |
| 138132 | Limopsis | | 1 | | | | | | | | | | 1 | |
| 138323 | Pecten | | | | | | | | | | | | 1 | 1 |
| 147153 | Similipecten | | | | | | | | | | | | | 1 |

Macroinvertebrate Matrix (Infauna)

FISA

| Aphia ID | Species | FISA_A301_ENV | FISA_A302_ENV | FISA_A303_ENV | FISA_A304_ENV | FISA_A305_ENV | FISA_A306_ENV | FISA_A307_ENV | FISA_A309_ENV | FISA_A1008_ENV | FISA_A1011_ENV | FISA_A1013_ENV | FISA_A1015_ENV | FISA_A3010_ENV |
|---------------------------|----------------------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|----------------|----------------|----------------|----------------|----------------|
| 197227 | Limatula hodgsoni | | | | | | | | | 1 | | | | |
| 505622 | Linea pygmaea | | | | | | | | | | | | | |
| 138186 | Mysella | | | 3 | | | 1 | | | | | | | 1 |
| 138654 | Lyonsiella sp | | | | | | | | | | | | 1 | 1 |
| 197467 | Thracia meridionalis | | | | | | | | | | | | | |
| 137858 | Cuspidaria | | | | | | | | | 2 | | 1 | | |
| 137863 | Tropidomya | | | | | | | | | | | | | |
| 152413 | Axinulus | | | | | | | | | | 1 | | | |
| 138552 | Thyasira | | | | | | 2 | | | | | | | |
| 138552 | Thyasira II | | 1 | | 4 | 2 | | | | 2 | 2 | | | 4 |
| 1806 ECHINODERMATA | | | | | | | | | | | | | | |
| 172707 | Anseropoda antarctica † | | | | | | | | | 1 | | | | |
| 172769 | Acodontaster elongatus | | | | | | | | | | | | | |
| 123567 | Ophiophycis aff mirabilis† | | | | | | | | | | | | | |
| 173102 | Amphiura belgicae | 1 | | | | | 2 | | | | | | 4 | |
| 123206 | Amphiuridae | | | | | | | | | | | | | |
| 123200 | Ophiuridae | | | | | | | | | | | | | |
| 236013 | Ophiozonella falklandica | 1 | | | | | | | | 1 | | 1 | | |
| 160790 | Brisaster antarcticus † | | | | 1 | | | | | | | | | |
| 123494 | aff Thyone | | | | | | 1 | | | | | | | |
| 123473 | Echinocucumis sp | | | | 1 | | | | | | | | | 1 |
| 146121 | Psolus | 1 | | | | | | | | | | | 1 | |
| | Psolus spp | | 1 | | | | | | | | | | | |
| 123182 | Synaptidae | | | | | | | 3 | | | | | 3 | |
| 123441 | Myriotrochus | | | | | | | | | | | | | |
| 1803 BRACHIOPODA | | | | | | | | | | | | | | |
| 1803 | Brachiopoda | 2 | | | | | | | | | | | | |
| 1803 | Brachiopoda II | | | | | | | | | | | | 2 | |
| 183339 | Pelagodiscus | | 1 | | | | 1 | 1 | | | | | | |
| CHAETOGNATHA | | | | | | | | | | | | | | |
| 105413 | Spadella | | | | | | | | | | 1 | | | |
| | | 18 | 39 | 32 | 44 | 41 | 36 | 34 | 16 | 44 | 34 | 24 | 51 | 39 |
| | | 27 | 124 | 50 | 98 | 84 | 101 | 55 | 33 | 89 | 67 | 41 | 94 | 78 |
| | | 5.158 | 7.883 | 7.924 | 9.378 | 9.028 | 7.584 | 8.235 | 4.29 | 9.58 | 7.848 | 6.193 | 11.01 | 8.722 |
| | | 0.9248 | 0.7743 | 0.9647 | 0.9377 | 0.9186 | 0.8648 | 0.9437 | 0.8669 | 0.8988 | 0.9254 | 0.9184 | 0.9418 | 0.9332 |
| | | 3.856 | 4.092 | 4.823 | 5.119 | 4.922 | 4.471 | 4.801 | 3.467 | 4.907 | 4.708 | 4.211 | 5.343 | 4.932 |
| | | 0.943 | 0.8752 | 0.9796 | 0.9726 | 0.9621 | 0.936 | 0.9717 | 0.8958 | 0.9533 | 0.9643 | 0.9512 | 0.979 | 0.9657 |

Epifaunal, Fragmented and Juvenile Species

FISA

| Aphia ID | Species | Authority | A3_ENV_FA | A3_ENV_FB | A09_ENV_FA | A09_ENV_FB | A09_ENV_FC | A10_ENV_FA | A10_ENV_FB | A10_ENV_FC | A12_ENV_FA | A12_ENV_FB | A14_ENV_FA | A14_ENV_FB | A18_ENV_FA | A18_ENV_FB | A20_ENV_FA | A20_ENV_FB | A20_ENV_FC | A21_ENV_FA | A21_ENV_FB | A22_ENV_FA | A22_ENV_FB | |
|-------------------------------|-------------------------------------|-----------------------------------|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|----|
| Epifaunal Species | | | | | | | | | | | | | | | | | | | | | | | | |
| 1267 | CNIDARIA | | | | | | | | | | | | | | | | | | | | | | | |
| 117136 | <i>Lafoes</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | | | | | |
| 117103 | <i>Halicium</i> sp | Oken, 1815 | | | | | | | | | | | | | | | | | | | | | | |
| 116999 | <i>Aglaopheria</i> | Lamouroux, 1812 | | | | | | | | | | | | | | | | | | | | | | |
| 117243 | <i>Lepidopora</i> | Pourtales, 1871 | | | | | | | | | | | | | | | | | | | | | | |
| 174823 | <i>Stylaster densicaulis</i> | Moseley, 1879 | | | | | | | | | | | | | | | | | | | | | | |
| 125287 | <i>Sarcodictyon Forbes</i> | (in Johnston), 1847 | | | | + | | | | | | | + | | + | | | | | | + | | + | |
| 173450 | <i>Callozostiron carlottae</i> | Kükenthal, 1909 | | | | + | | | | | | | | | | | | | | | | | | |
| 409519 | <i>Fannyella (Fannyella) rossii</i> | Gray, 1872 | | | | | | | | | | | | | | | | | | | | | | |
| 1365 | <i>Alcyonacea</i> | | | | | | | | | | | | | | | | | | | | | | | |
| 135220 | <i>Scyphozoa</i> | Milne-Edwards & Haime, 1849 | 16 | 12 | 9 | 13 | 1 | 6 | 9 | 9 | 4 | 2 | | | 2 | 11 | 14 | 12 | 8 | 3 | | 8 | 1 | 22 |
| 135095 | <i>Lophelia</i> | | | | | | | | | | | | | | | | | | | | | | | |
| 1821 CHORDATA | | | | | | | | | | | | | | | | | | | | | | | | |
| 103506 | <i>Eugyra</i> | Alder & Hancock, 1870 | | 1 | | 1 | | | 1 | | | | | | | | | | | | 1 | | | |
| 173279 | <i>Molgula pedunculata</i> | Herdman, 1881 | | | | | | | | | | | | | | | | | | | | | | |
| 579849 GRAPTOLITHOIDEA | | | | | | | | | | | | | | | | | | | | | | | | |
| 137594 | <i>Rhabdopleura</i> | Beklemishev, 1951 Allman, 1869 | | | | | | | | | | | | | | | | | | | | | | |
| 558 PORIFERA | | | | | | | | | | | | | | | | | | | | | | | | |
| 131723 | <i>Sycon</i> | Grant, 1836 | | | | | | | | | | | | | | | | | | | | | | |
| 132110 | <i>Hexacnina</i> | Risso, 1827 | | | | | | | | | | | | | | | | | | | | | | |
| 132110 | <i>Hexacnina</i> | Carter, 1885 | + | | | | | | | | | | | | | | | | | | | | | |
| 131662 | <i>Ancorinidae</i> | Carter, 1885 | + | + | | | | + | + | + | | | | | + | | | | | | | | | |
| 132077 | <i>Tethya</i> | Schmidt, 1870 | | | | | | | | | | | | | | | | | | | | | | |
| 132077 | <i>Tethya</i> | Lamarck, 1815 | | | | | | | | | | | | | | | | | | | | | | |
| 171200 | <i>Gastrophanella</i> | Schmidt, 1879 | | | | | | | | | | | | | | | | | | | | | | |
| 132072 | <i>Suberites</i> | Nardo, 1833 | | | | | | | | | | | | | | | | | | | | | | |
| 132046 | <i>Polymastia</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | | | | | |
| 170677 | aff <i>Radiella</i> sol | Schmidt, 1870 | | | | | | | | | | | | | | | | | | | | | | |
| 131882 | <i>Eurypon</i> | Gray, 1867 | | | | | | | | | | | | | | | | | | | | | | |
| 131882 | <i>Eurypon</i> | Gray, 1867 | | | | 1 | | | | | | | | | | | | | | | | | | |
| 131790 | <i>Halicnemis</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | | | | | |
| 132053 | <i>Tentorium</i> | Vosmaer, 1887 | | | | | | | | | | | | | | | | | | | | | | |
| 131912 | <i>Ulosa</i> | de Laubenfels, 1936 | | | | | | | | | | | | | | | | | | | | | | |
| 131893 | <i>Asbestopuma</i> | Topsent, 1901 | | | | | | | | | | | | | | | | | | | | | | |
| 131893 | <i>Asbestopuma II</i> | Topsent, 1901 | | | | | | | | | | | | | | | | | | | | | | |
| 131893 | <i>Asbestopuma III</i> | Topsent, 1901 | | | | | | | | | | | | | | | | | | | | | | |
| 131936 | <i>Crella</i> | Gray, 1867 | | | | | | | | | | | | | | | | | | | | | | |
| 131906 | <i>Esperiopsis</i> | Carter, 1882 | | | | | | | | | | | | | | | | | | | | | | |
| 131907 | <i>Mycale</i> | Gray, 1867 | | | | | | | | | | | | | | | | | | | | | | |
| 131926 | <i>Inflatella</i> | Schmidt, 1875 | | | | | | | | | | | | | | | | | | | | | | |
| 131905 | <i>Isodictya</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | | | | | |
| 131930 | <i>Lissodendoryx</i> | Topsent, 1892 | | | | | | | | | | | | | | | | | | | | | | |
| 131950 | <i>Hymedesmia I</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | | | | | |
| 131950 | <i>Hymedesmia II</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | | | | | |
| 131969 | <i>Melonanchora</i> | Carter, 1874 | | | | | | | | | | | | | | | | | | | | | | |
| 131641 | <i>Microcionidae</i> | Carter, 1875 | | | | | | | | | | | | | | | | | | | | | | |
| 131786 | <i>Monocrepidium</i> sp | Topsent, 1898 | | | | | | | | | | | | | | | | | | | | | | |
| 131834 | <i>Haliclona</i> | Grant, 1836 | | | | | | | | | | | | | | | | | | | | | | |
| 146142 BRYOZOA | | | | | | | | | | | | | | | | | | | | | | | | |
| 111054 | <i>Tabulipora</i> | Lamarck, 1816 | | | | | | | | | | | | | | | | | | | | | | |
| 111029 | <i>Annectocyma</i> | Hayward & Ryland, 1985 | | | | | | | | | | | | | | | | | | | | | | |
| 111052 | <i>Idmidronea</i> | Canu & Bassler, 1920 | | | | | | | | | | | | | | | | | | | | | | |
| 111041 | <i>Hornera</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | | | | | |
| 111041 | <i>Hornera</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | | | | | |
| 111044 | <i>Disporella</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | | | | | |
| 111048 | <i>Stomatopora</i> | Bronn, 1825 | | | | | | | | | | | | | | | | | | | | | | |
| 468484 | <i>Isosecuriflustra</i> | Liu & Hu, 1991 | | | | | | | | | | | | | | | | | | | | | | |
| 110851 | <i>Callopora</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | | | | | |
| 110849 | <i>Amphiblestrum</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | | | | | |
| 173093 | <i>Notoplites klugei</i> | (Hasenbank, 1932) | | | | | | | | | | | | | | | | | | | | | | |
| 110865 | <i>Notoplites</i> | Harmer, 1923 | | | | | | | | | | | | | | | | | | | | | | |
| 110869 | <i>Cellaria</i> | Ellis & Solander, 1786 | | | | | | | | | | | | | | | | | | | | | | |
| 173311 | <i>Melicerita</i> | Milne Edwards, 1836 | | | | | | | | | | | | | | | | | | | | | | |
| 172987 | <i>Cornucopina</i> | Levinson, 1909 | | | | | | | | | | | | | | | | | | | | | | |
| 110965 | <i>Escharella</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | | | | | |
| 110965 | <i>Escharella</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | | | | | |
| 110972 | <i>Escharina</i> | Milne Edwards, 1836 | | | | | | | | | | | | | | | | | | | | | | |
| 110941 | <i>Fenestrellina</i> | Jullien, 1888 | | | | | | | | | | | | | | | | | | | | | | |
| 110931 | <i>Hippothoa</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | | | | | |
| 110932 | <i>Plesiothoa</i> | Gordon & Hastings, 1979 | | | | | | | | | | | | | | | | | | | | | | |
| 110873 | <i>Cellepora</i> | Linnaeus, 1767 | | | | | | | | | | | | | | | | | | | | | | |
| 173401 | <i>Spigaleos</i> | Hayward, 1992 | | | | | | | | | | | | | | | | | | | | | | |

Epifaunal, Fragmented and Juvenile Species

FISA

| Aphia ID | Species | Authority | A3_ENV_FA | A3_ENV_FB | A09_ENV_FA | A09_ENV_FB | A09_ENV_FC | A10_ENV_FA | A10_ENV_FB | A10_ENV_FC | A12_ENV_FA | A12_ENV_FC | A14_ENV_FA | A14_ENV_FB | A18_ENV_FA | A18_ENV_FB | A20_ENV_FA | A20_ENV_FB | A20_ENV_FC | A21_ENV_FA | A21_ENV_FB | A22_ENV_FA | A22_ENV_FB |
|--|--------------------|----------------------------------|-----------|-----------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|------------|
| Epifaunal Species | | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile and fragmented species | | | | | | | | | | | | | | | | | | | | | | | |
| 1066 | CRUSTACEA | | | | | | | | | | | | | | | | | | | | | | |
| 118394 | <i>Aeگا Frag.</i> | Leach, 1815 | | | | | | | | | | | | | | | | | | | | | |
| 118394 | <i>Aeگا juv</i> | Leach, 1815 | | | | | | | | | | | | | | | | | | | | | |
| 1806 | ECHINODERMATA | Bruguière, 1791 [ex Klein, 1734] | | | | | | | | | | | | | | | | | | | | | |
| 123080 | Asteroides juv | | | | | | | | | | | | | | | | | | | | | | |
| 123083 | Holothurioides juv | | | | | | | | | | | | | | | | | | | | | | |

Epifaunal, Fragmented and Juvenile Species

FISA

| Alpha ID | Species | Authority | A26_ENV_FA | A26_ENV_FB | A26_ENV_FC | A27_ENV_FA | A27_ENV_FB | A27_ENV_FC | A202_ENV_FA | A202_ENV_FB | A204_ENV_FA | A204_ENV_FB | A301_ENV_FA | A301_ENV_FC | A302_ENV_FA | A302_ENV_FB | A302_ENV_FC | A303_ENV_FA | A303_ENV_FB | A303_ENV_FC | A304_ENV_FA | A304_ENV_FC |
|--------------------------|------------------------------|-----------------------------|------------|------------|------------|------------|------------|------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Epifaunal Species | | | | | | | | | | | | | | | | | | | | | | |
| 1267 | CNIDARIA | | | | | | | | | | | | | | | | | | | | | |
| 117136 | Lafoes | Lamouroux, 1821 | | + | | | | | | | | | | | | | | | | | | |
| 111703 | Halicium sp | Oken, 1815 | | | | | | | | | | | | | | | | | | | | |
| 116999 | Aglaophenia | Lamouroux, 1812 | | | | | | | | | | | | | | | | | | | | |
| 117243 | Lepidopora | Pourtales, 1871 | | + | | | | | | | | | | | | | | | | | | |
| 174823 | Stylaster densicaulis | Moseley, 1879 | + | | + | | | | | | | | | | | | | | | | | |
| 125287 | Sarcodictyon Forbes | (in Johnston), 1847 | | | | | | | | | | | | | | | | | | | | |
| 173450 | Callozostrom carlottae | Kükenthal, 1909 | | | | | | | | | | | | | | | | | | | | |
| 409519 | Fannyella (Fannyella) rossii | Gray, 1872 | | + | | | | | | | | | | | | | | | | | | |
| 1365 | Alcyonacea | | | | | | | | | | | | | | | | | | | | | |
| 135220 | Scyphozoa | | 2 | 4 | 2 | 21 | 15 | 2 | 3 | 4 | | | | | | | 20 | 15 | | 11 | 6 | 5 |
| 135095 | Lophelia | Milne-Edwards & Haime, 1849 | + | | | | | | | | + | | | | | | | | | | 3 | 2 |
| 1821 | CHORDATA | | | | | | | | | | | | | | | | | | | | | |
| 103506 | Eugyra | Alder & Hancock, 1870 | | | | 1 | | | | 1 | | | | | | | | | | | | 1 |
| 173279 | Molgula pedunculata | Herdman, 1881 | 1 | | | | | | | | | | | | | | 2 | | | | | |
| 579849 | GRAPTOLITHOIDEA | Beklemishev, 1951 | | | | | | | | | | | | | | | | | | | | |
| 137594 | Rhabdopleura | Allman, 1869 | + | | | | | | | | + | | | | | | | | | | | |
| 558 | PORIFERA | Grant, 1836 | | | | | | | | | | | | | | | | | | | | |
| 131723 | Sycon | Risso, 1827 | | | | | | | | | | | | | | | | | | | | |
| 132110 | Hexactinella | Carter, 1885 | | | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + |
| 132110 | Hexactinella | Carter, 1885 | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + |
| 131662 | Ancorinidae | Schmidt, 1870 | | | | | | | | | | | | | | | | | | | | |
| 132077 | Tethya | Lamarck, 1815 | | | | | | | | | | | | | | | | | | | | |
| 171200 | Gastrophanella | Schmidt, 1879 | | | | | | | | | | | | | | | | | | | | |
| 132072 | Suberites | Nardo, 1833 | | | | | | | | | | | | | | | | | | | | |
| 132046 | Polymastia | Bowerbank, 1864 | | | + | | | | | | | | | | | | | | | | | |
| 170677 | aff Radiella sol | Schmidt, 1870 | + | + | | | | | | | | | | | | | + | | | | | |
| 131882 | Eurypora | Gray, 1867 | | + | + | | | | | | | | | | | | | | | | | |
| 131882 | Eurypora | Gray, 1867 | | | | | | | | | | | | | | | | | | | | |
| 131790 | Halicnemis | Bowerbank, 1864 | | | | | | | | | + | | | | | | | | | | | |
| 132053 | Tentorium | Vosmaer, 1887 | | | | | | | | | | | | | | | | | | | | |
| 131912 | Ulosa | de Laubenfels, 1936 | | | | | | | | | | | | | | | | | | | | |
| 131893 | Asbestopluma | Topsent, 1901 | | | | | | | | | | | | | | | | | | | | |
| 131893 | Asbestopluma II | Topsent, 1901 | | | | | | | | | | | | | | | | | | | | |
| 131893 | Asbestopluma III | Topsent, 1901 | | | | | | | | | | | | | | | | | | | | |
| 131936 | Crella | Gray, 1867 | + | + | + | | | | | | | | | | | | | | | | | |
| 131906 | Esperiopsis | Carter, 1882 | | | | | | | | | | | | | | | | | | | | |
| 131907 | Mycale | Gray, 1867 | + | | | | | | | | | | | | | | | | | | | |
| 131926 | Inflatella | Schmidt, 1875 | | | + | | | | | | | | | | | | | | | | | |
| 131905 | Isodictya | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | | | |
| 131930 | Lissodendoryx | Topsent, 1892 | | | | | | | | | | | | | | | | | | | | |
| 131950 | Hymedesmia I | Bowerbank, 1864 | + | | | | | | | | | | | | | | | | | | | |
| 131950 | Hymedesmia II | Bowerbank, 1864 | | | | | | | | | | | + | | | | | | | | | |
| 131969 | Melonanchora | Carter, 1874 | | | | | | | | | | | | | | | | | | | | |
| 131641 | Microcionidae | Carter, 1875 | | | | | | | | | | | | | | | | | | | | |
| 131786 | Monocrepidium sp | Topsent, 1898 | | | + | | | | | | | | | | | | | | | | | |
| 131834 | Haliclona | Grant, 1836 | | | + | | | | | | | | | | | | | | | | | |
| 146142 | BRYOZOA | | | | | | | | | | | | | | | | | | | | | |
| 111054 | Tubulipora | Lamarck, 1816 | | | | + | | | | | | | | | | | | | | | | |
| 111029 | Annectocyma | Hayward & Ryland, 1985 | | | | | + | | | | | | | | | | | | | | | |
| 111052 | Idmidronea | Canu & Bassler, 1920 | | | | | | | | | | | | | | | | | | | | |
| 111041 | Harnera | Lamouroux, 1821 | | | + | | | | | | | | | | | | | | | | | |
| 111041 | Harnera | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | | | |
| 111044 | Disporella | Gray, 1848 | | | | | | | | | | | | | | | | | | | | |
| 111048 | Stomatopora | Bronn, 1825 | | | | | | | | | | | | | | | | | | | | |
| 468484 | Isoscuriflustra | Liu & Hu, 1991 | | | | | | | | | | | | | | | | | | | | |
| 110851 | Callopora | Gray, 1848 | | | | | | | | | | | | | | | | | | | | |
| 110849 | Amphiblastrum | Gray, 1848 | | | | | | | | | | | | | | | | | | | | |
| 173093 | Notoplites klugei | (Hasenbank, 1932) | | | + | | | | | | | | | | | | | | | | | |
| 110845 | Notoplites | Harmer, 1923 | | | | | | | | | | | | | | | | | | | | |
| 110869 | Cellaria | Ellis & Solander, 1786 | | | | | | | | | | | | | | | | | | | | |
| 173311 | Melicerita | Milne Edwards, 1836 | | | | | | | | | | | | | | | | | | | | |
| 172987 | Cornucopina | Levinsen, 1909 | | | | | | | | | | | | | | | | | | | | |
| 110965 | Escharella | Gray, 1848 | + | | + | | | | | | | | | | | | | | | | | |
| 110965 | Escharella | Gray, 1848 | | | | | | | | | | | | | | | | | | | | |
| 110972 | Escharina | Milne Edwards, 1836 | | | | | | | | | | | | | | | | | | | | |
| 110941 | Fenestrulina | Jullien, 1888 | | | + | | | | | | | | | | | | | | | | | |
| 110931 | Hippothoa | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | | | |
| 110932 | Plesiothoa | Gordon & Hastings, 1979 | | | | | | | | | | | | | | | | | | | | |
| 110873 | Cellepora | Linnaeus, 1767 | | | | | | | | | | | | | | | | | | | | |
| 173401 | Spigaleos | Hayward, 1992 | | | | | | | | | | | | | | | | | | | | |

Epifaunal, Fragmented and Juvenile Species

FISA

| Aphia ID | Species | Authority | A26_ENV_FA | A26_ENV_FB | A26_ENV_FC | A27_ENV_FA | A27_ENV_FB | A27_ENV_FC | A202_ENV_FA | A202_ENV_FB | A204_ENV_FA | A204_ENV_FB | A301_ENV_FA | A301_ENV_FC | A302_ENV_FA | A302_ENV_FB | A302_ENV_FC | A303_ENV_FA | A303_ENV_FB | A303_ENV_FC | A304_ENV_FA | A304_ENV_FC |
|--|--------------------|----------------------------------|------------|------------|------------|------------|------------|------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Epifaunal Species | | | | | | | | | | | | | | | | | | | | | | |
| Juvenile and fragmented species | | | | | | | | | | | | | | | | | | | | | | |
| 1066 | CRUSTACEA | | | | | | | | | | | | | | | | | | | | | |
| 118394 | <i>Aegea Frag.</i> | Leach, 1815 | | | | | | | | | | | | | | | | | | | | |
| 118394 | <i>Aegea juv</i> | Leach, 1815 | | | | | | | | | | | | | | | | | | | | |
| 1806 | ECHINODERMATA | Bruguière, 1791 [ex Klein, 1734] | | | | | | | 1 | | | | | 1 | | | | | | | | |
| 123080 | Asteroides juv | | | | | | | | | | | | | | | | | | | | | |
| 123083 | Holothuridae juv | | | | | | | | | | | | | | | | | | | | | |

Epifaunal, Fragmented and Juvenile Species

FISA

| Aphia ID | Species | Authority | A305_ENV_FA | A305_ENV_FB | A306_ENV_FA | A306_ENV_FB | A307_ENV_FA | A307_ENV_FB | A309_ENV_FA | A309_ENV_FB | A1008_ENV_FA | A1008_ENV_FB | A1011_ENV_FA | A1011_ENV_FB | A1013_ENV_FA | A1013_ENV_FB | A1015_ENV_FA | A1015_ENV_FB | A3010_ENV_FA | A3010_ENV_FB |
|--------------------------|-------------------------------------|-----------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| Epifaunal Species | | | | | | | | | | | | | | | | | | | | |
| 1267 | Cnidaria | | | | | | | | | | | | | | | | | | | |
| 11736 | <i>Laloea</i> | Lamouroux, 1821 | | | | | | | | | + | | | | | | | | | |
| 117103 | <i>Halesium</i> sp | Oken, 1815 | | | | | | | | | | + | | | | | | | | |
| 116999 | <i>Aglaophenia</i> | Lamouroux, 1812 | | | | | | | | | | | | | | | | | | |
| 117243 | <i>Lepidopora</i> | Pourtales, 1871 | | | | | | | | | | | | | | | | | | |
| 174823 | <i>Stylaster densicaulis</i> | Moseley, 1879 | | | + | | | + | | | | | | | | | | | | |
| 125287 | <i>Sarcodictyon Forbes</i> | (in Johnston), 1847 | | | | | | | + | | | | | | | | | | | |
| 173450 | <i>Callazostron carlottae</i> | Kükenthal, 1909 | | | | | | | | | | | | | | | | | | |
| 409519 | <i>Fannyella (Fannyella) rossii</i> | Gray, 1872 | | | | | | | | | | | | | | | | | | |
| 1365 | Alcyonacea | | | | | | | | | | | | | | | | | | | |
| 135220 | <i>Scyphozoa</i> | Milne-Edwards & Haime, 1849 | 2 | 1 | 8 | 11 | 11 | 6 | 12 | 3 | 11 | 1 | 11 | 1 | 1 | | 3 | 6 | 9 | 8 |
| 135095 | <i>Lophelia</i> | | | | | | | | | | | | | | | | | | | |
| 1821 | Chordata | | | | | | | | | | | | | | | | | | | |
| 103506 | <i>Eugyra</i> | Alder & Hancock, 1870 | 7 | | 7 | | | 7 | | | | | | | | | | | | |
| 173279 | <i>Molgula pedunculata</i> | Herdman, 1881 | | | | | | | | | | | | | | | | | | |
| 579849 | GRAPTOLITHOIDEA | Beklemishev, 1951 | | | | | | | | | | | | | | | | | | |
| 137594 | <i>Rhabdopleura</i> | Allman, 1869 | | | + | | | | | | | | | | | | | | | |
| 558 | PORIFERA | Grant, 1836 | | | | | | | | | | | | | | | | | | |
| 131723 | <i>Sycon</i> | Risso, 1827 | | | | | | | + | | | | | | | | | | | |
| 132110 | <i>Hexactinella</i> | Carter, 1885 | | | | | | | + | | | | | | | | | | | |
| 132110 | <i>Hexactinella</i> | Carter, 1885 | | | | | | | + | | | | | | | | | | | |
| 131662 | <i>Ancorinidae</i> | Schmidt, 1870 | + | + | + | + | + | + | + | + | | | + | + | | | | | + | + |
| 132077 | <i>Tethya</i> | Lamarck, 1815 | | | | | | | | | | | | | | | | | | |
| 171200 | <i>Gastrophanella</i> | Schmidt, 1879 | | | | | | | | | | | | | | | | | | |
| 132072 | <i>Suberites</i> | Nardo, 1833 | | | | | | | | | | | | | | | | | | |
| 132046 | <i>Polymastia</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | |
| 170677 | aff <i>Radiella</i> sol | Schmidt, 1870 | | | | | | | | | | | | | | | | | | |
| 131882 | <i>Eurypon</i> | Gray, 1867 | + | + | | | | | | | | | | | | | | | | |
| 131882 | <i>Eurypon</i> | Gray, 1867 | | | | | | | | | | | | | | | | | | |
| 131790 | <i>Halicnemis</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | |
| 132053 | <i>Tentorium</i> | Vosmaer, 1887 | | | | | | | | | | | | | | | | | | |
| 131912 | <i>Ulosa</i> | de Laubenfels, 1936 | | | | | | | | | | | | | | | | | | |
| 131893 | <i>Asbestopluma</i> | Topsent, 1901 | | | | | | | 2 | | 2 | | | | | | | | | |
| 131893 | <i>Asbestopluma II</i> | Topsent, 1901 | | | | | | | 2 | | 1 | | | | | | | | | |
| 131893 | <i>Asbestopluma III</i> | Topsent, 1901 | | | | | | | 3 | | | | | | | | | | | |
| 131936 | <i>Crella</i> | Gray, 1867 | | | | | | | | | | | | | | | | | | |
| 131906 | <i>Esperiopsis</i> | Carter, 1882 | | | | | | | | | | | | | | | | | | |
| 131907 | <i>Mycale</i> | Gray, 1867 | | | | | | | | | | | | | | | | | | |
| 131926 | <i>Inflatilella</i> | Schmidt, 1875 | | | | | | | | | | | | | | | | | | |
| 131905 | <i>Isodictya</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | |
| 131930 | <i>Lissodendoryx</i> | Topsent, 1892 | | | | | | | | | | | | | | | | | | |
| 131950 | <i>Hymedesmia I</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | |
| 131950 | <i>Hymedesmia II</i> | Bowerbank, 1864 | | | | | | | | | | | | | | | | | | |
| 131969 | <i>Melonanchora</i> | Carter, 1874 | | | | | | | | | | | | | | | | | | |
| 131641 | <i>Microcionidae</i> | Carter, 1875 | | | | | | | | | | | | | | | | | | |
| 131786 | <i>Monocrepidium</i> sp | Topsent, 1898 | | | | | | | | | | | | | | | | | | |
| 131834 | <i>Haliclona</i> | Grant, 1836 | | | | | | | | | | | | | | | | | | |
| 146142 | BRYOZOA | | | | | | | | | | | | | | | | | | | |
| 111054 | <i>Tubulipora</i> | Lamarck, 1816 | + | | | | | | | | | | | | | | | | | |
| 111029 | <i>Amnectocyma</i> | Hayward & Ryland, 1985 | | | | | | | | | | | | | | | | | | |
| 111052 | <i>Idmidronea</i> | Canu & Bassler, 1920 | | | | | | | | | | | | | | | | | | |
| 111041 | <i>Hornera</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | |
| 111041 | <i>Hornera</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | |
| 111044 | <i>Disporella</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | |
| 111048 | <i>Stomatopora</i> | Bronn, 1825 | | | | | | | | | | | | | | | | | | |
| 468484 | <i>Isosecuriflustra</i> | Liu & Hu, 1991 | | | | | | | | | | | | | | | | | | |
| 110851 | <i>Callopora</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | |
| 110849 | <i>Amphiblestrum</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | |
| 173093 | <i>Notoplites klugei</i> | (Hasenbank, 1932) | | | | | | | | | | | | | | | | | | |
| 110865 | <i>Notoplites</i> | Harmer, 1923 | | | | | | | | | | | | | | | | | | |
| 110869 | <i>Cellaria</i> | Ellis & Solander, 1786 | | | | | | | | | | | | | | | | | | |
| 173311 | <i>Melicerita</i> | Milne Edwards, 1836 | | | | | | | | | | | | | | | | | | |
| 172987 | <i>Cornucopina</i> | Levinson, 1909 | | | | | | | | | | | | | | | | | | |
| 110965 | <i>Escharella</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | |
| 110965 | <i>Escharella</i> | Gray, 1848 | | | | | | | | | | | | | | | | | | |
| 110972 | <i>Escharina</i> | Milne Edwards, 1836 | | | | | | | | | | | | | | | | | | |
| 110941 | <i>Fenestrulina</i> | Julien, 1888 | | | | | | | | | | | | | | | | | | |
| 110931 | <i>Hippothoa</i> | Lamouroux, 1821 | | | | | | | | | | | | | | | | | | |
| 110932 | <i>Plesiothoa</i> | Gordon & Hastings, 1979 | | | | | | | | | | | | | | | | | | |
| 110873 | <i>Cellepore</i> | Linnaeus, 1767 | | | | | | | | | | | | | | | | | | |
| 173401 | <i>Spilgaleas</i> | Hayward, 1992 | | | | | | | | | | | | | | | | | | |

Epifaunal, Fragmented and Juvenile Species

FISA

| Aphia ID | Species | Authority | A305_ENV_FA | A305_ENV_FB | A306_ENV_FA | A306_ENV_FB | A307_ENV_FA | A307_ENV_FB | A309_ENV_FA | A309_ENV_FB | A1008_ENV_FA | A1008_ENV_FB | A1011_ENV_FA | A1011_ENV_FB | A1013_ENV_FA | A1013_ENV_FB | A1015_ENV_FA | A1015_ENV_FB | A3010_ENV_FA | A3010_ENV_FB |
|--|--------------------|----------------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| Epifaunal Species | | | | | | | | | | | | | | | | | | | | |
| Juvenile and fragmented species | | | | | | | | | | | | | | | | | | | | |
| 1066 | CRUSTACEA | | | | | | | | | | | | | | | | | | | |
| 118394 | <i>Aega Frag.</i> | Leach, 1815 | | 1 | | | | | | | | | | | | | | | | |
| 118394 | <i>Aega juv</i> | Leach, 1815 | | 26 | | | | | | | | | | | | | | | | |
| 1806 | ECHINODERMATA | | | | | | | | | | | | | | | | | | | |
| 123080 | Asterioidea juv | Bruguière, 1791 [ex Klein, 1734] | | | | | | | | | | | | | | | | | | |
| 123083 | Holothurioidea juv | | | | | | 1 | | | | | | | | | | | | | |

APPENDIX VIII – SEABED AND SAMPLE PHOTOGRAPHS

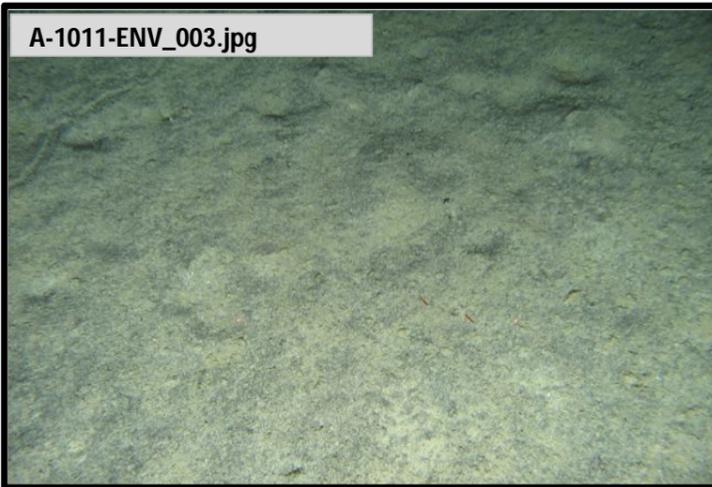


Photo Position: 784828 mE, 4213108 mN

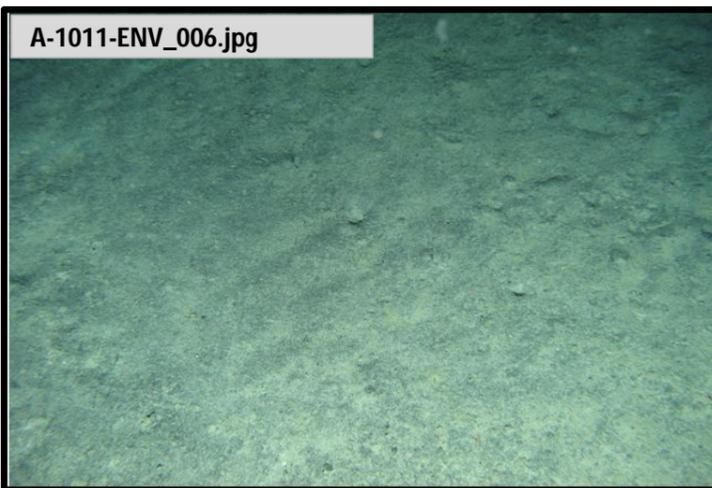


Photo Position: 784826 mE, 4213107 mN

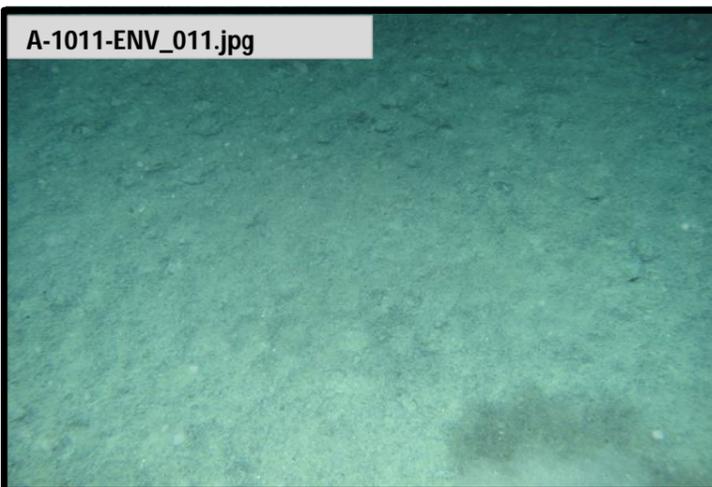


Photo Position: 784800 mE, 4213111 mN

Habitat Summary Information: A/1011/ENV

Survey Area: FISA

No. of Stills: 17 Mins of Video: 28 Track Length: 139m

| | |
|---|---|
| <p>Site Selection Criteria Deepest point of hollow feature</p> | <p>Analogue Interpretation Very slight slope</p> |
| <p>Sediment Description Light grey to black bioturbated slightly silty sand with occasional fine gravels</p> | |
| <p>Conspicuous Fauna Decapod Shrimp sp., Cup Coral: <i>Flabellum</i> sp., Isopod: possibly <i>Acutiserolis neaera</i>,</p> | |

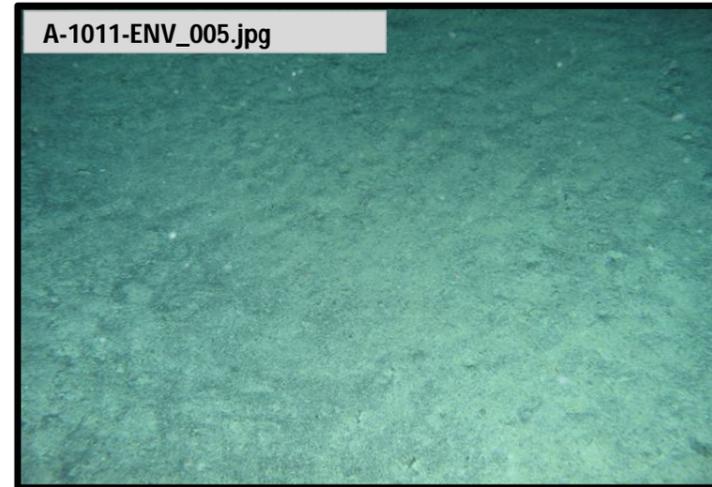


Photo Position: 784824 mE, 4213107 mN

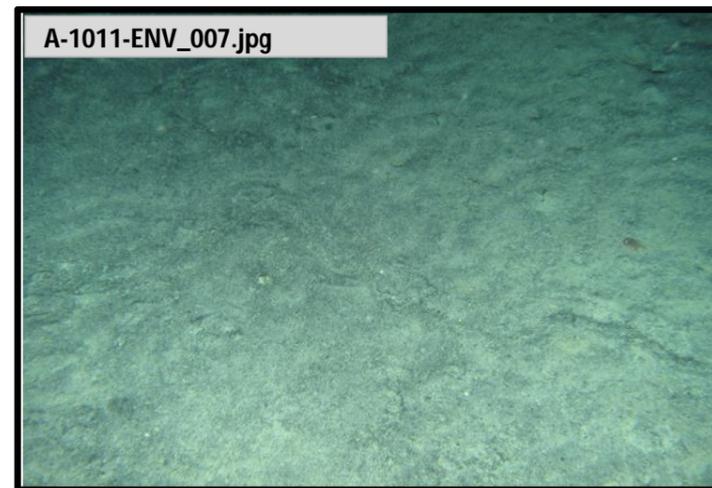


Photo Position: 784823 mE, 4213108 mN

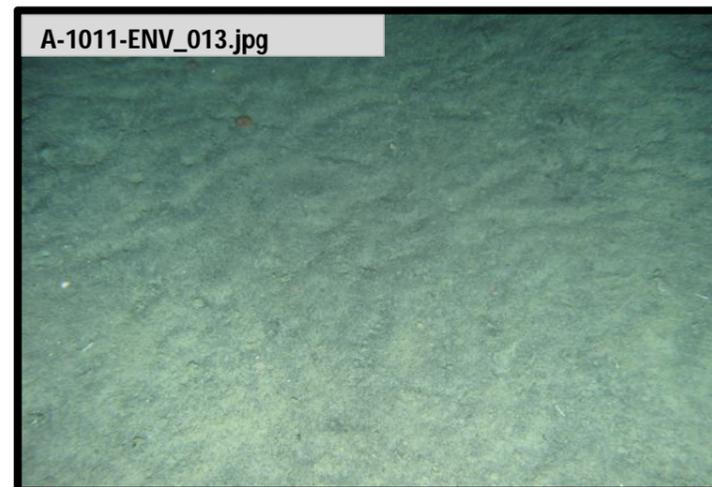
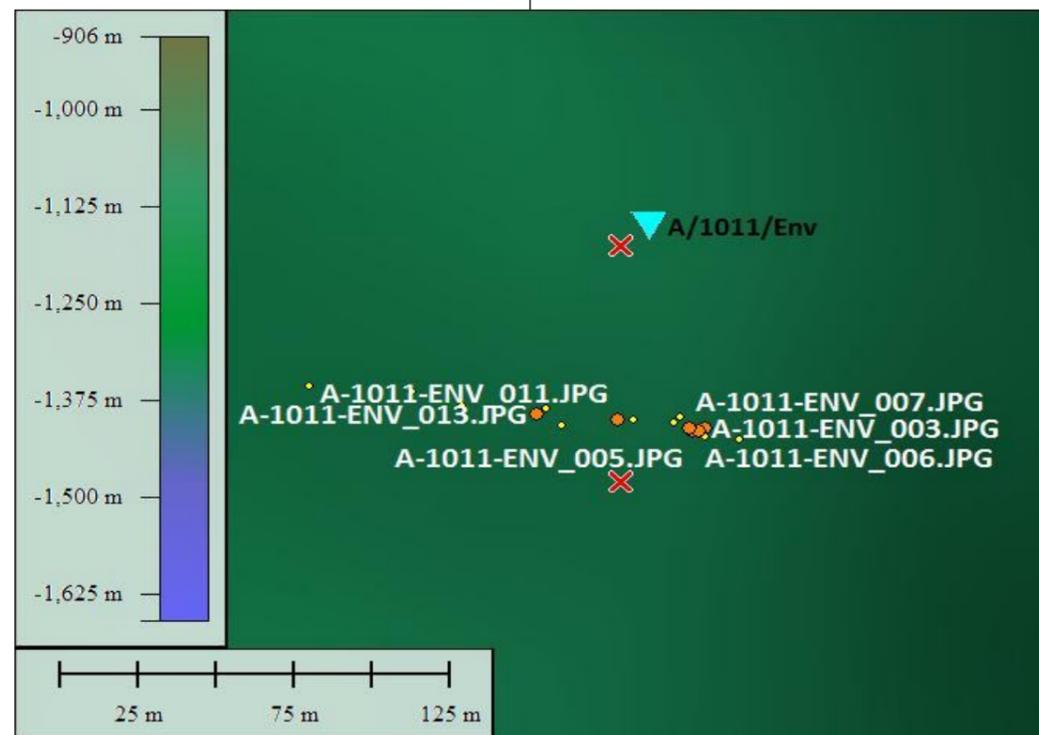


Photo Position: 784774 mE, 4213113 mN



| | |
|---------------------------|---|
| <p>Client </p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
|---------------------------|---|

X Sample Location
● UW Photo
● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

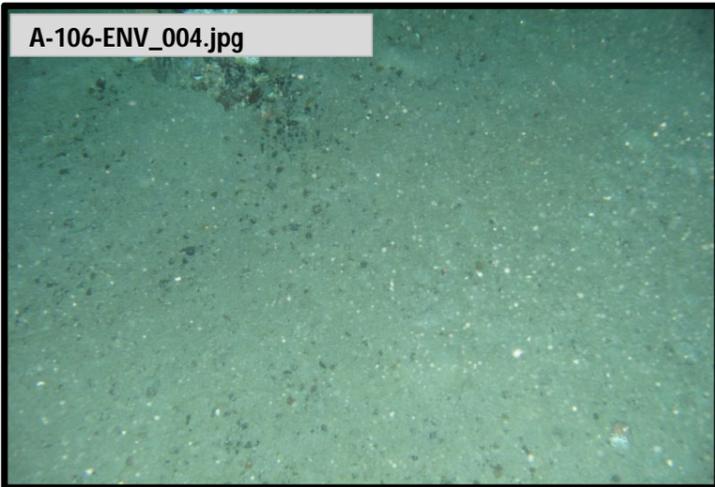


Photo Position: 743498 mE, 4180209 mN

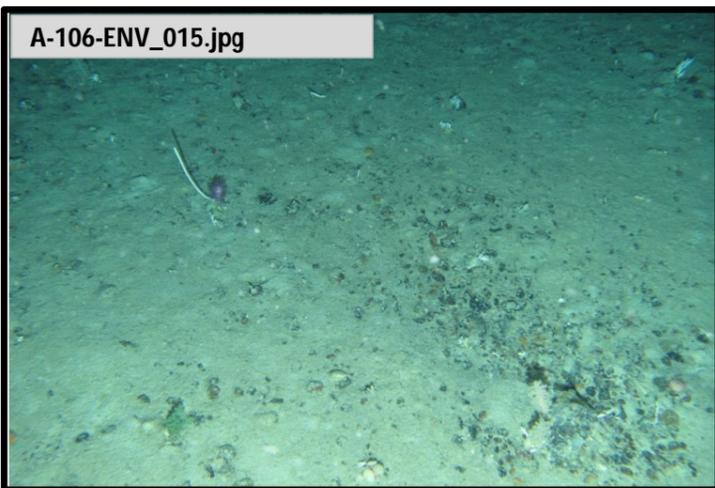


Photo Position: 743149 mE, 4179859 mN

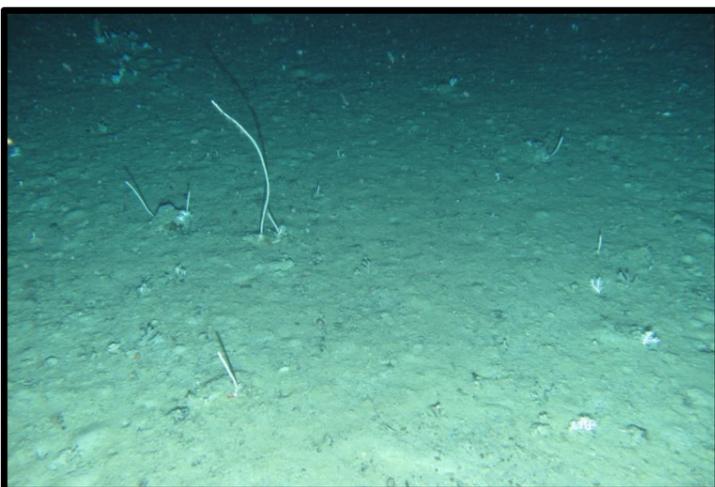


Photo Position: 743055 mE, 4179786 mN

Habitat Summary Information: A/106/ENV

Survey Area: FISA

No. of Stills: 24 Mins of Video: 37 Track Length: 630m

| | |
|---|--|
| <p>Site Selection Criteria Slightly raised linear feature</p> | <p>Analogue Interpretation Slightly raised linear feature</p> |
| <p>Sediment Description Light olive grey and black silty sand with mixed size gravels, pebbles, cobbles and boulders.</p> | |
| <p>Conspicuous Fauna Branched Stone Coral: <i>Stylaster</i> sp., <i>Gorgonian</i> sp., <i>Branched Bryozoa</i> sp., <i>Nephropidae</i> sp., <i>Thymops birsteini</i>, Brittlestar (Ophiuroid), Squat Lobster: <i>Galatheididae</i> sp., <i>Hydroid</i> sp., Sea Urchin: <i>Echinoidea</i> sp., <i>Octocoral</i> sp., Sea Pen: <i>Pennatulacea</i>,</p> | |

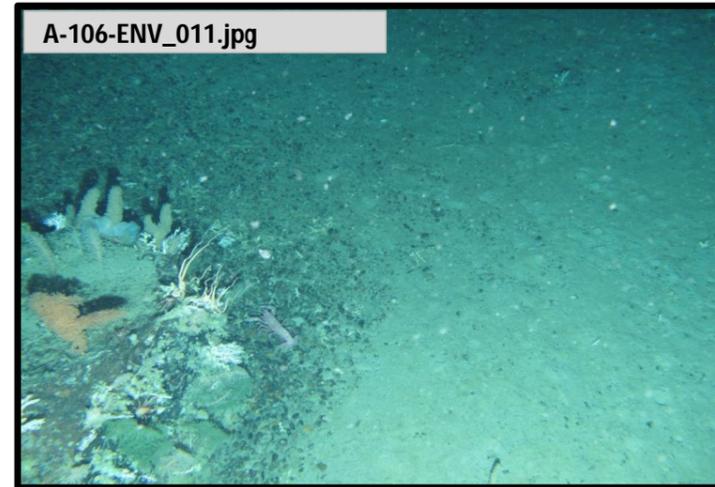


Photo Position: 743221 mE, 4179933 mN

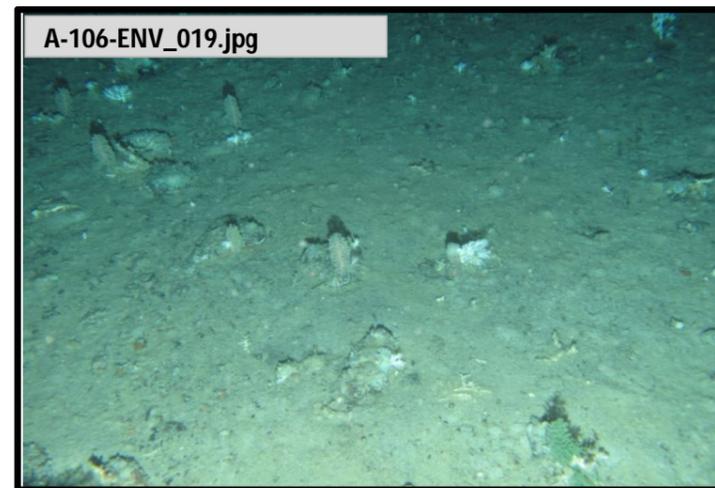


Photo Position: 743101 mE, 4179805 mN

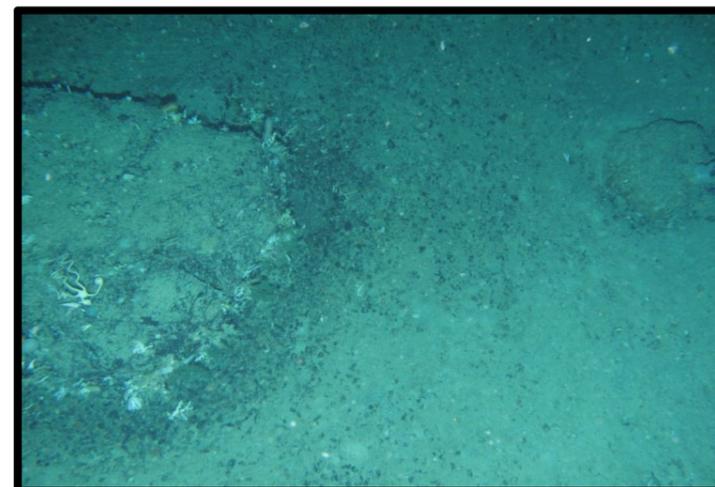
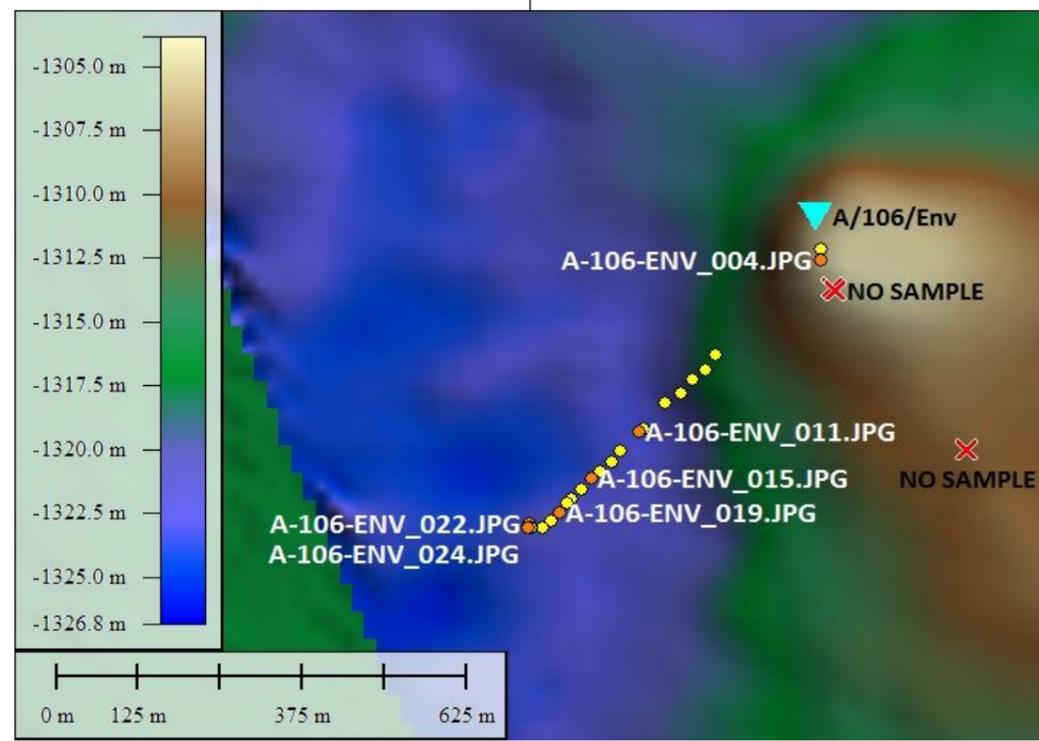


Photo Position: 743053 mE, 4179780 mN



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|---------------------------|---|
| <p>Client </p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
|---------------------------|---|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

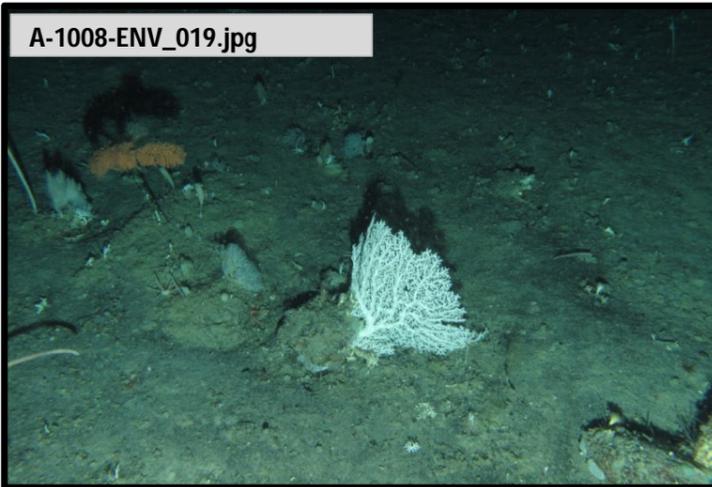


Photo Position: 784327 mE, 4202013 mN

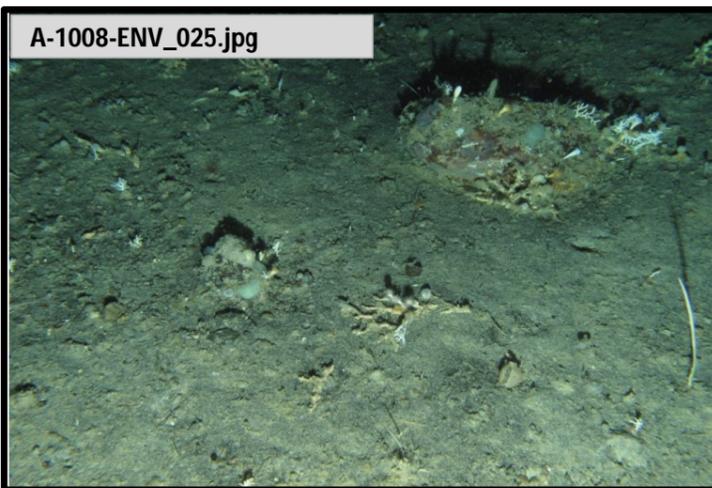


Photo Position: 784324 mE, 4201970 mN



Sediment Example Image

Habitat Summary Information: A/1008/ENV

Survey Area: FISA

No. of Stills: 41 Mins of Video: 40 Track Length: 278m

Site Selection Criteria

Escarpment feature

Analogue Interpretation

Slightly raised linear feature

Sediment Description

Light olive grey and black silty sand with mixed size gravels, pebbles, cobbles and boulders.

Conspicuous Fauna

Branched Stone Coral: *Stylaster* sp., Sea Pen: Pennatulacea, Sponge: Porifera sp., Relic *Lophelia* sp., Sea Lily: Crinoidea sp., Branched Bryozoa sp., Bryozoa sp., Octocoral sp., Gorgonian sp., Pencil Urchin: possibly *Cidaris* sp., Starfish: Asteroidea sp., *Lophelia pertusa*, Brittlestar (Ophiuroid), Hydroid sp.,

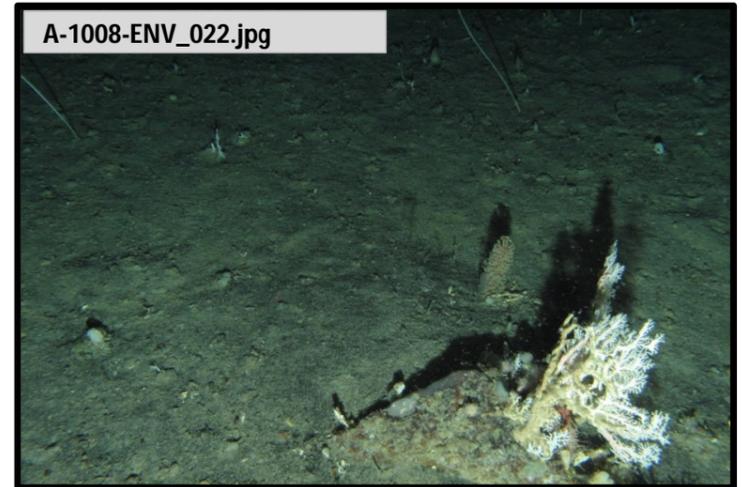


Photo Position: 784326 mE, 4201989 mN

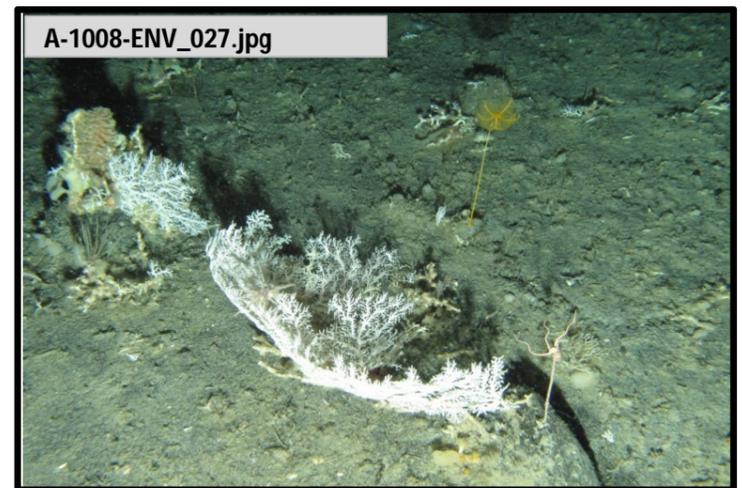
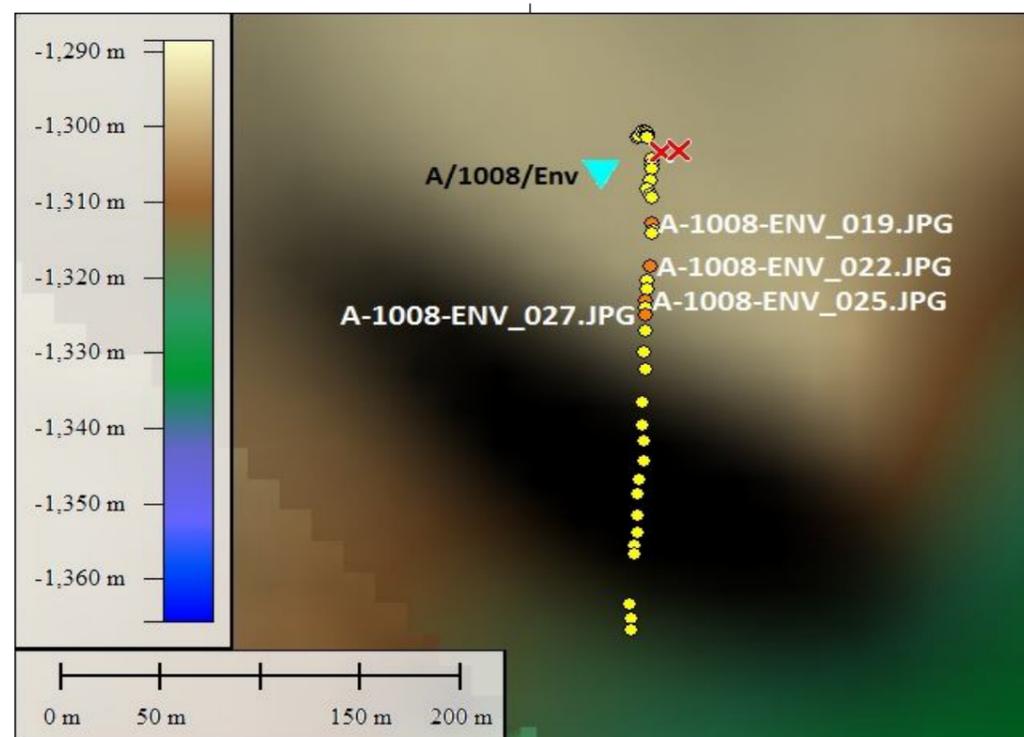


Photo Position: 784324 mE, 4201962 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

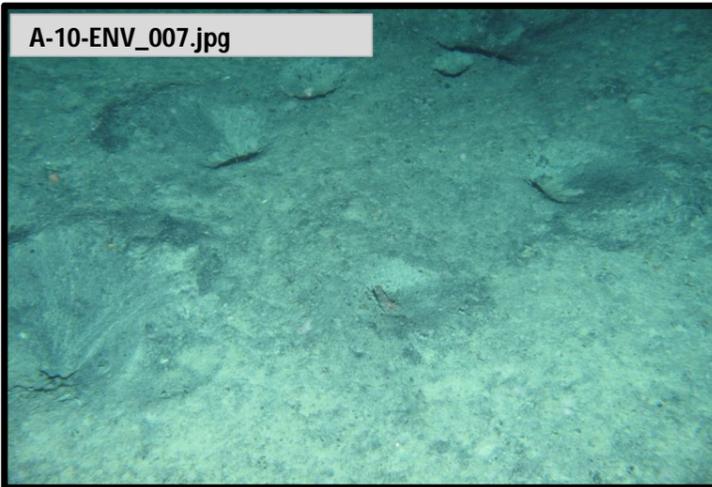


Photo Position: 774447 mE, 4205138 mN

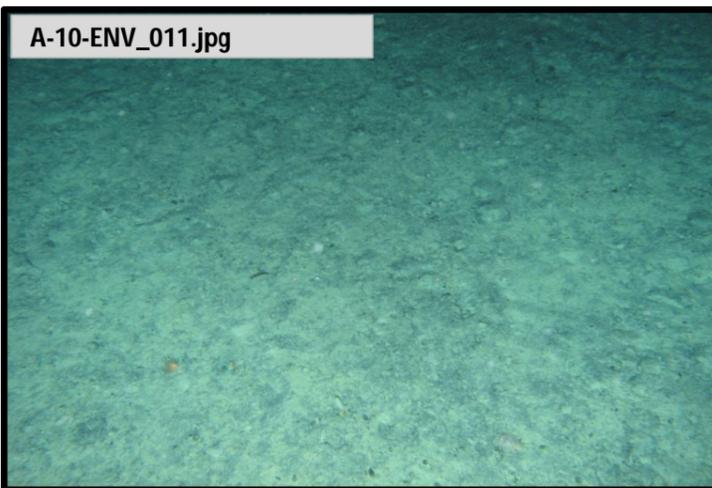


Photo Position: 774435 mE, 4205137 mN



Sediment Example Image

Habitat Summary Information: A/10/ENV

Survey Area: FISA

No. of Stills: 17 Mins of Video: 19 Track Length: 38m

Site Selection Criteria

Medium amplitude flat seabed feature

Analogue Interpretation

Medium amplitude flat seabed feature

Sediment Description

Light grey to black heavily bioturbated silty sand with occasional fine gravel.

Conspicuous Fauna

Sea Pen: Pennatulacea, Brittlestar (Ophiuroid), Decapod Shrimp sp., Batoidea sp., Polychaete sp., Isopod: possibly *Acutiserolis neaera*, Hydroid sp.,

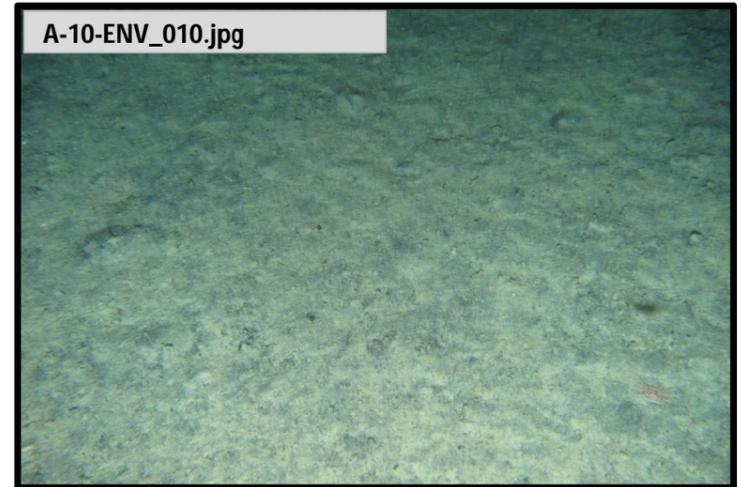


Photo Position: 774439 mE, 4205137 mN

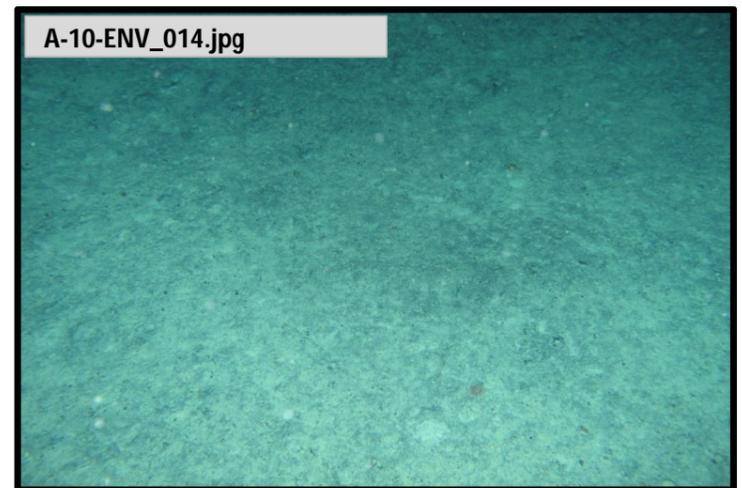
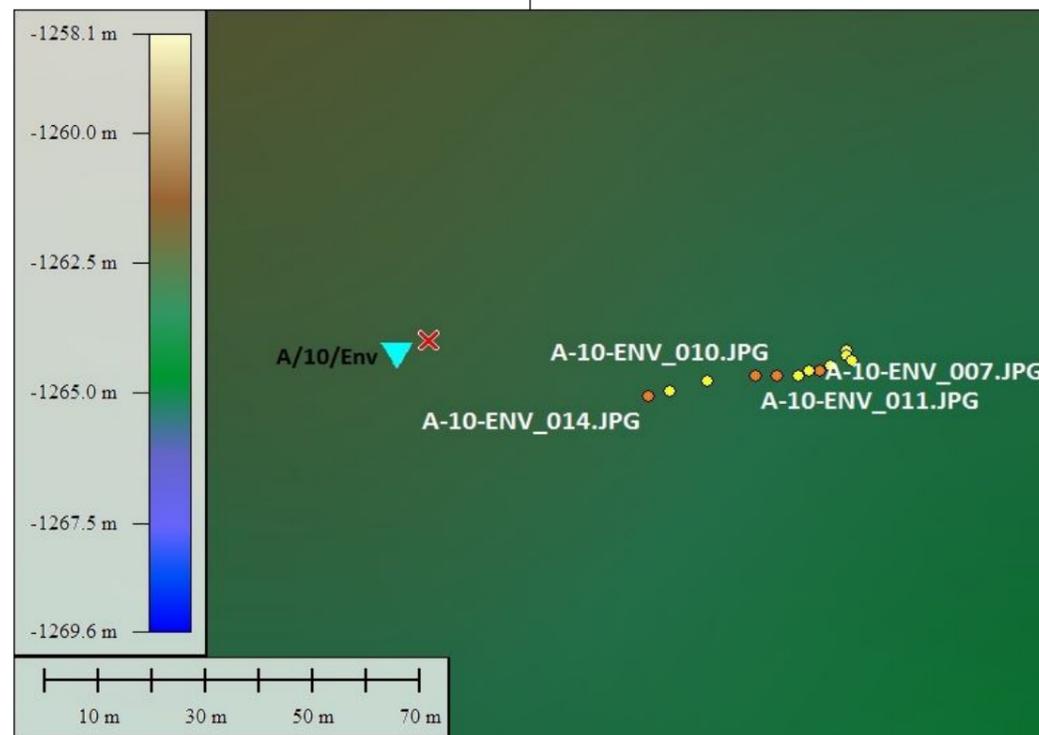


Photo Position: 774415 mE, 4205133 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

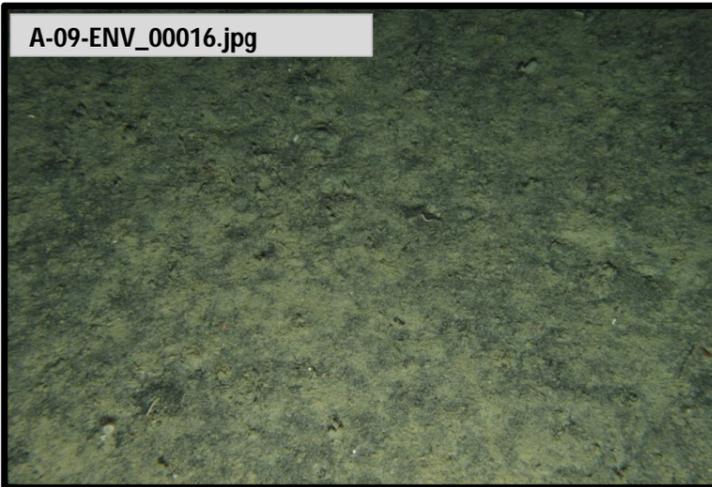


Photo Position: 774028 mE, 4201322 mN

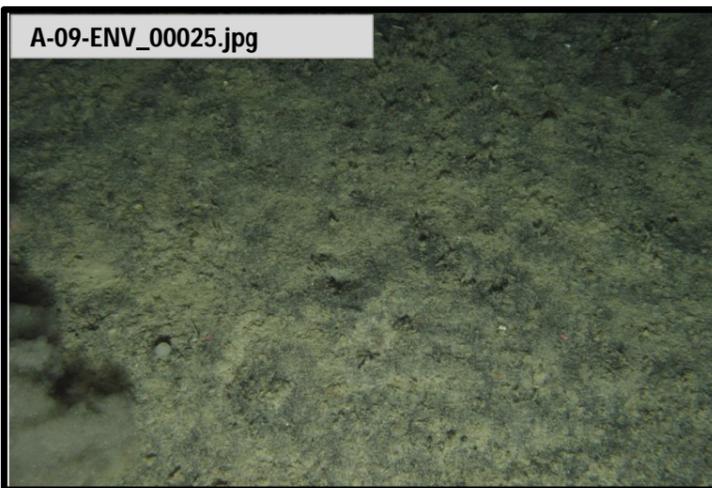
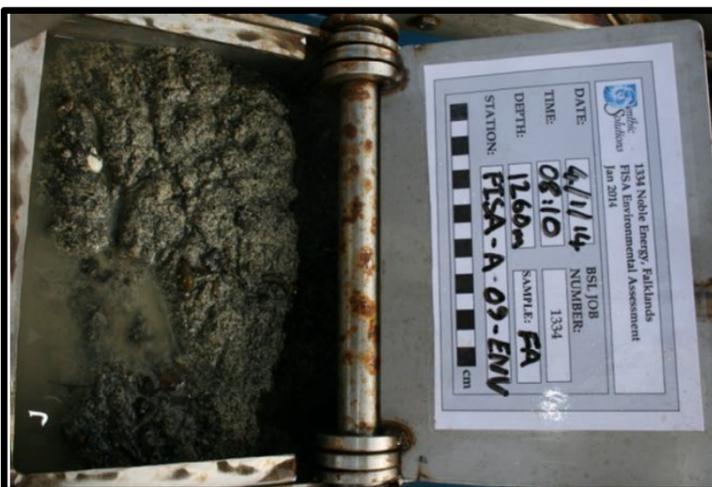


Photo Position: 774042 mE, 4201377 mN



Sediment Example Image

Habitat Summary Information: A/09/ENV

Survey Area: FISA

No. of Stills: 27 Mins of Video: 31 Track Length: 102m

Site Selection Criteria

Medium amplitude flat seabed feature

Analogue Interpretation

Relatively flat seabed

Sediment Description

Light grey to black bioturbated silty sand with occasional fine gravel.

Conspicuous Fauna

Cup Coral: Flabellum sp., Gorgonian sp., Sponge: Porifera sp., Brittlestar (Ophiuroid), Isopod: possibly *Acutiserolis neaera*.

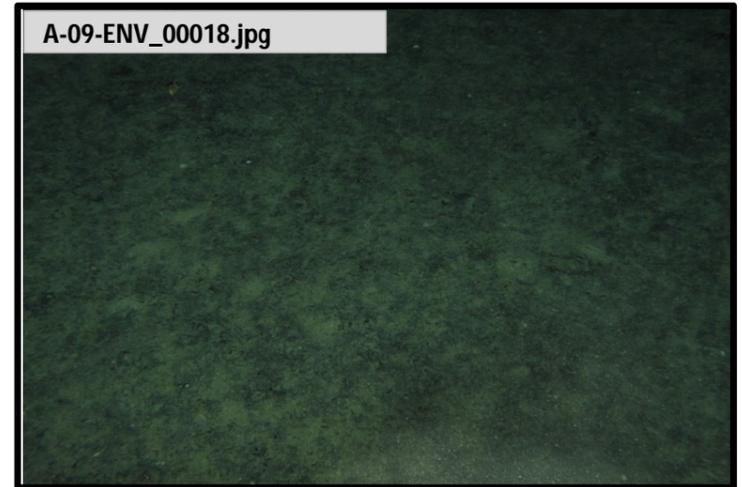


Photo Position: 774032 mE, 4201330 mN

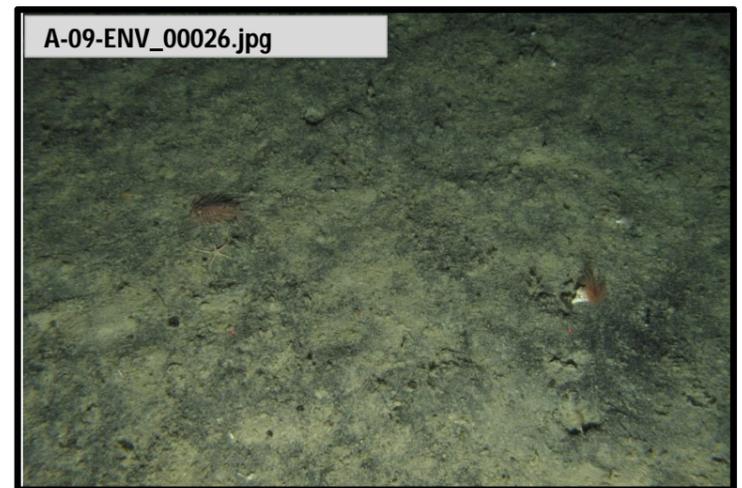
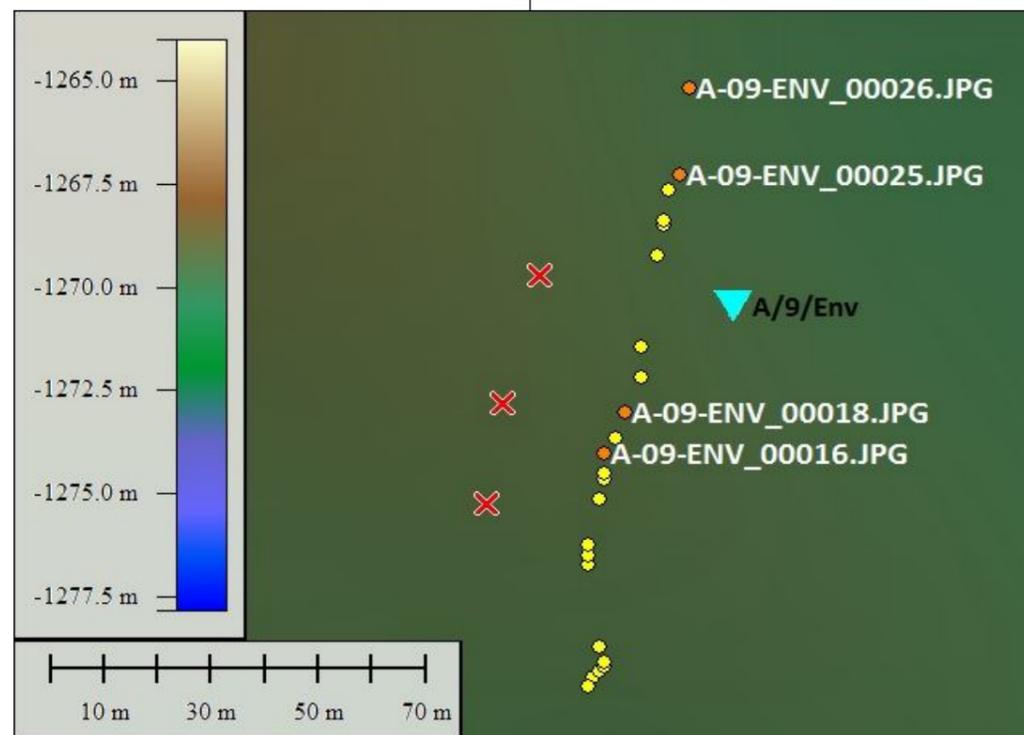


Photo Position: 774044 mE, 4201394 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

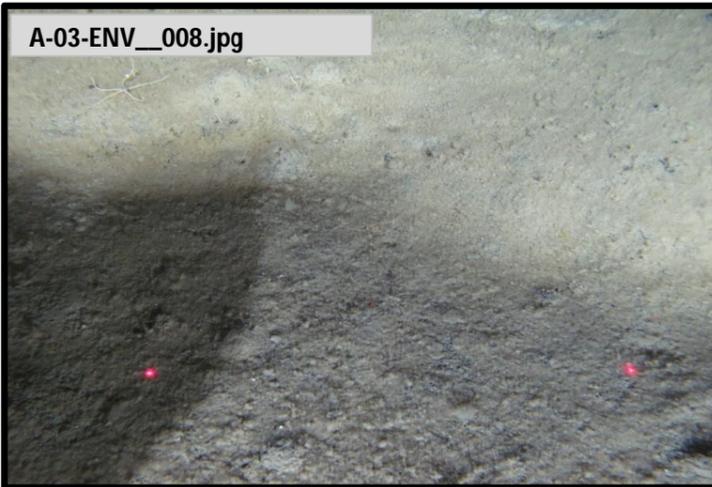


Photo Position: 737024 mE, 4188675 mN

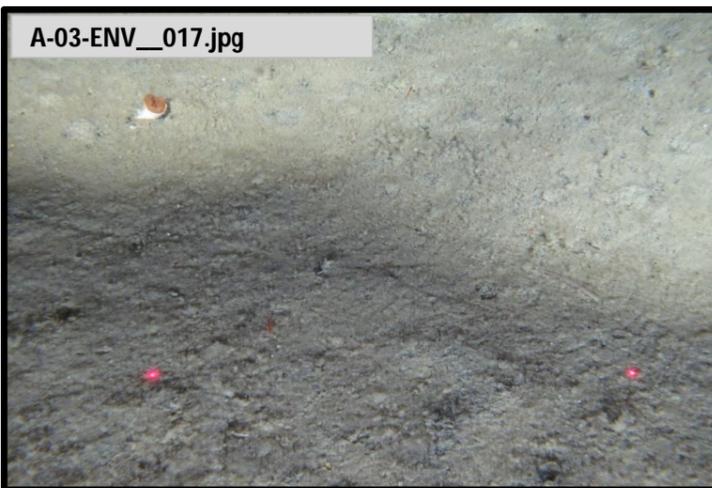


Photo Position: 737018 mE, 4188932 mN



Sediment Example Image

Habitat Summary Information: A/03/ENV

Survey Area: FISA

No. of Stills: 27 Mins of Video: 14 Track Length: 668m

Site Selection Criteria

Regional sample with slight slope

Analogue Interpretation

Very slight slope

Sediment Description

Light olive grey silty sand with a dark surface layer

Conspicuous Fauna

Bryozoa sp., Brittlestar (Ophiuroid), Decapod Shrimp sp., Cup Coral: *Flabellum* sp., Sea Pen: possibly *Anthoptilum grandiflorum*, Sponge: Porifera sp.,

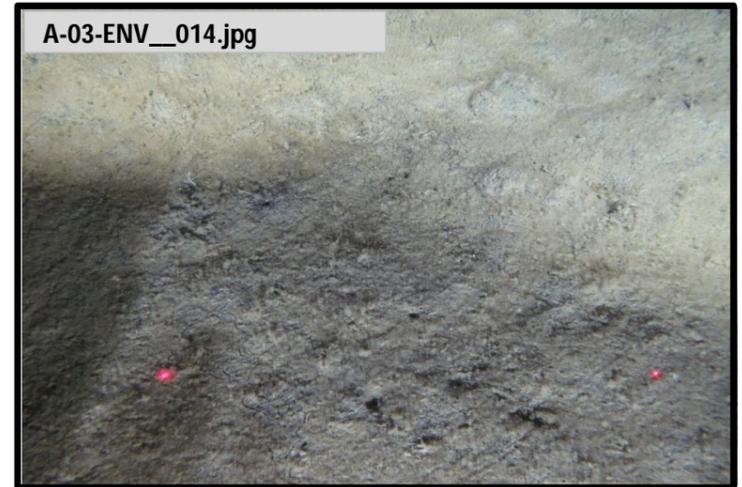


Photo Position: 737018 mE, 4188879 mN

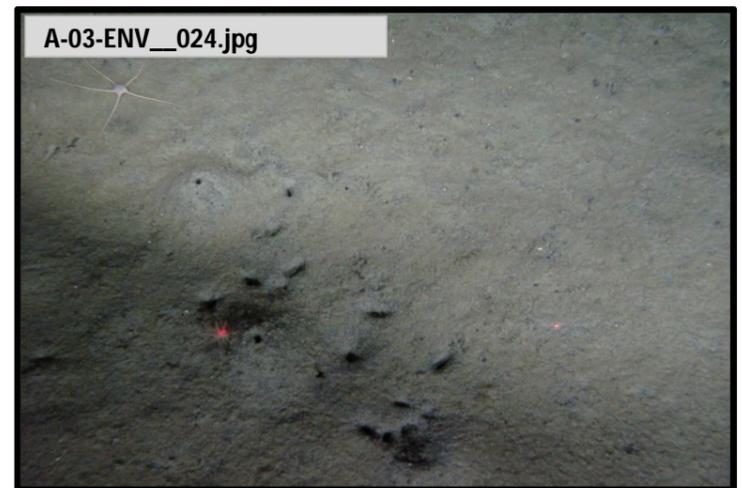
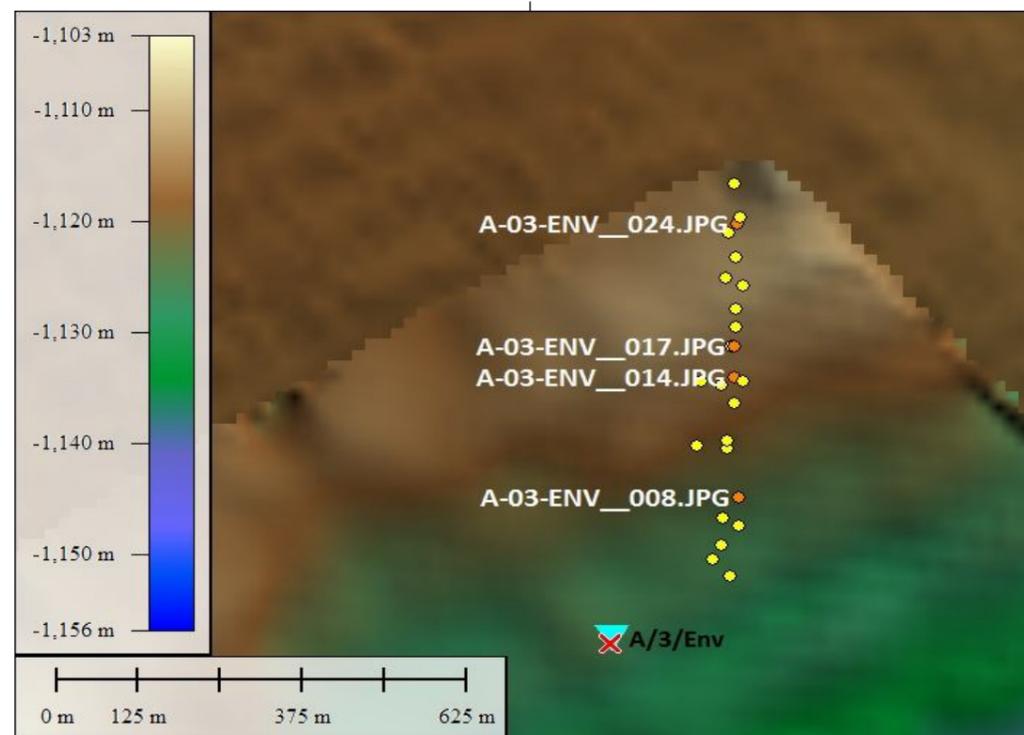


Photo Position: 737022 mE, 4189140 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

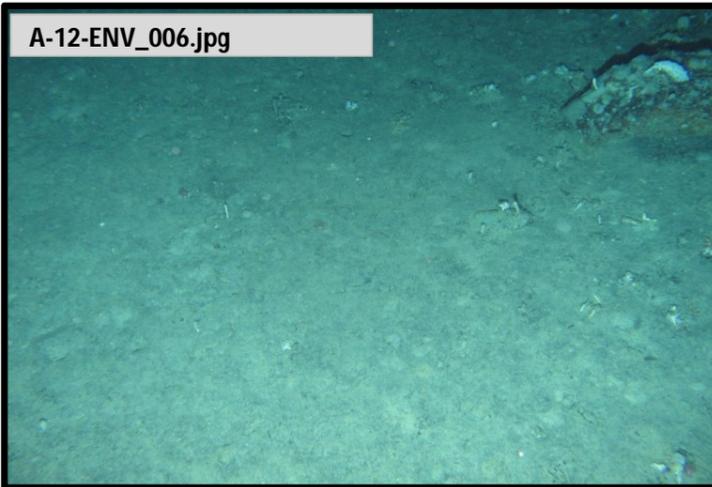


Photo Position: 783879 mE, 4203855 mN

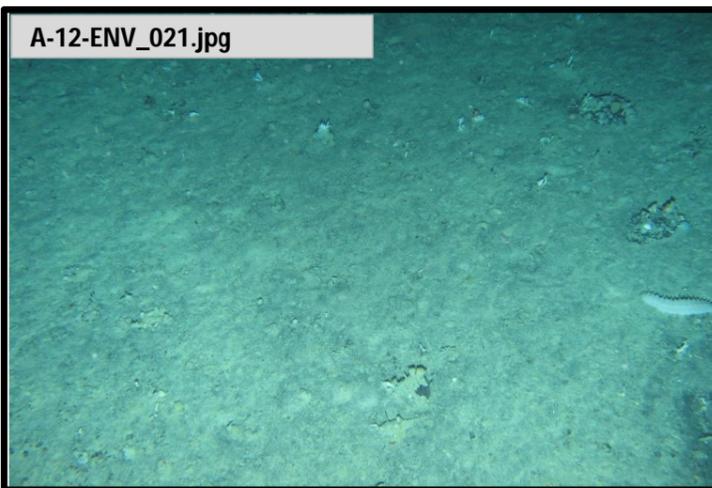


Photo Position: 783799 mE, 4203860 mN



Sediment Example Image

Habitat Summary Information: A/12/ENV

Survey Area: FISA

No. of Stills: 22 Mins of Video: 26 Track Length: 99m

Site Selection Criteria

Regional sample

Analogue Interpretation

Relatively flat seabed

Sediment Description

Light grey to black bioturbated silty sand with occasional fine gravels, pebbles and cobbles

Conspicuous Fauna

Cup Coral: *Flabellum* sp., Polychaete sp., Nephropidae sp. *Thymops birsteini*, Hydroid sp., Holothuroidea sp., Bryozoa sp.,

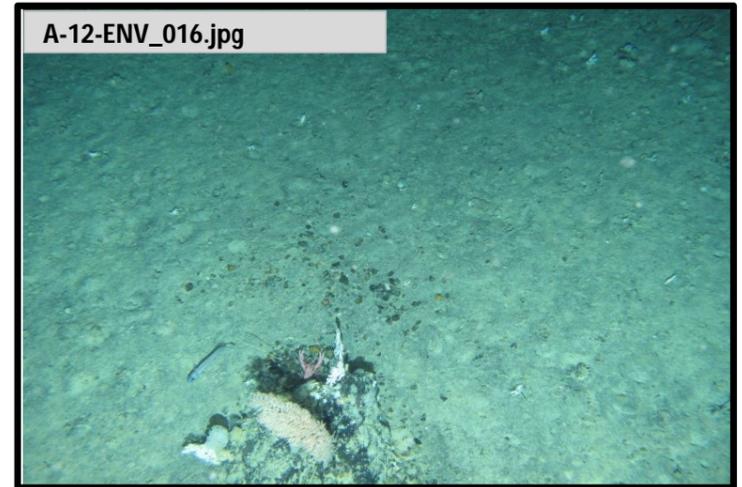


Photo Position: 783838 mE, 4203858 mN

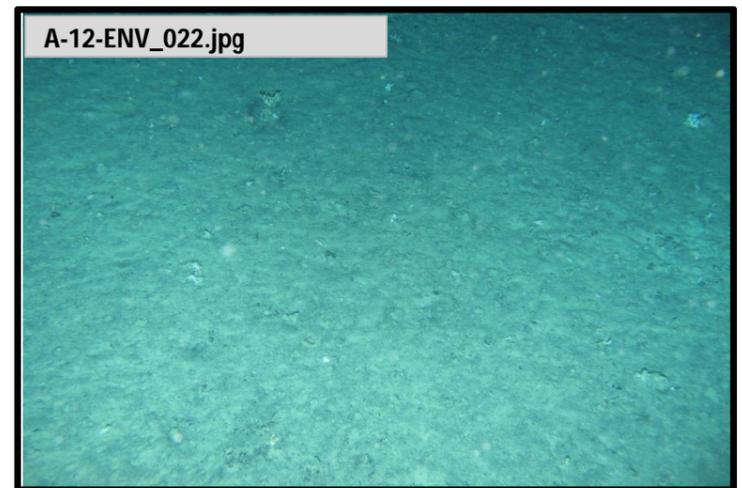
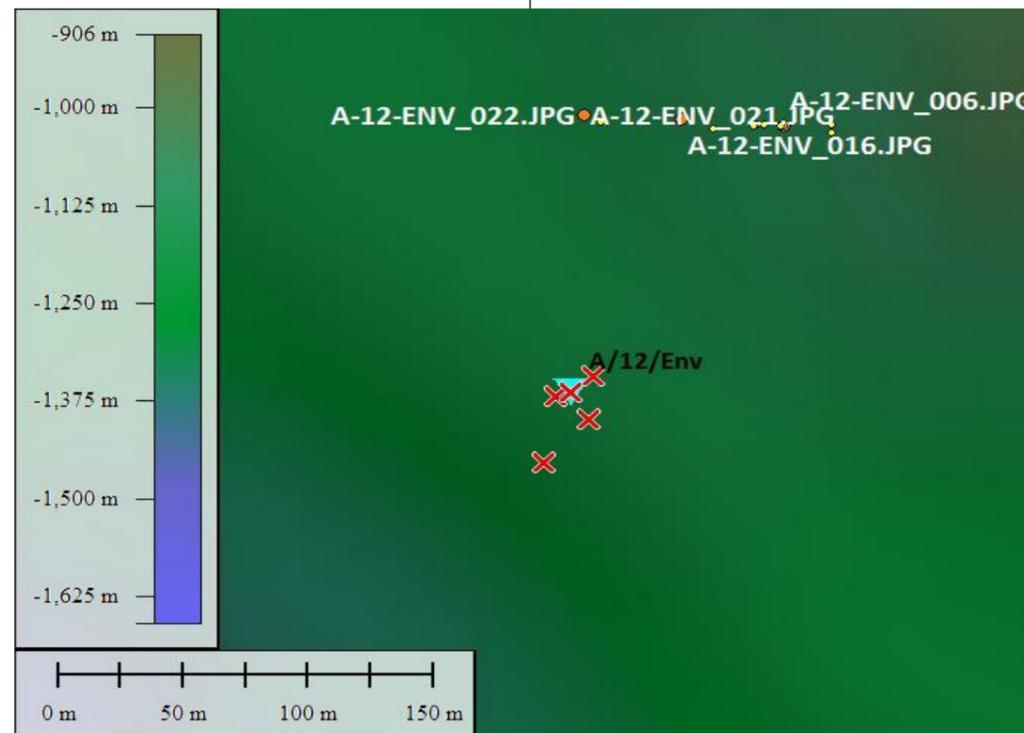
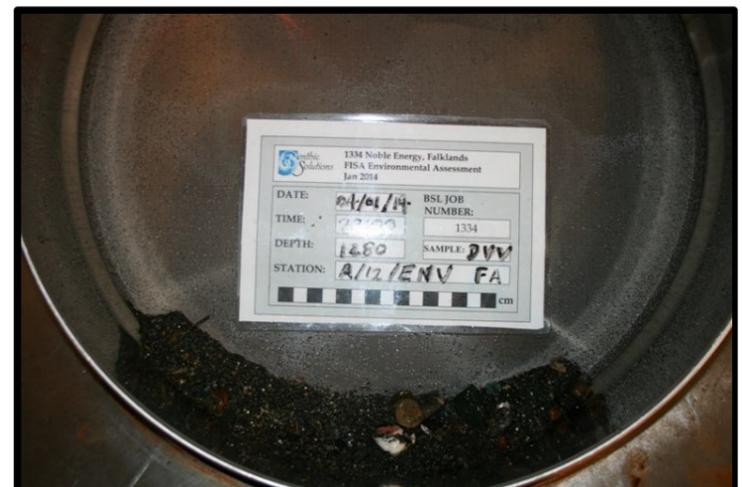


Photo Position: 783806 mE, 4203857 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

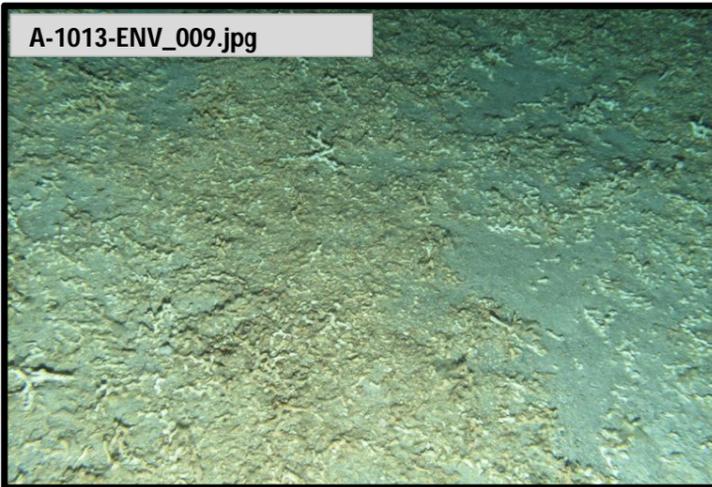


Photo Position: 790314 mE, 4204430 mN

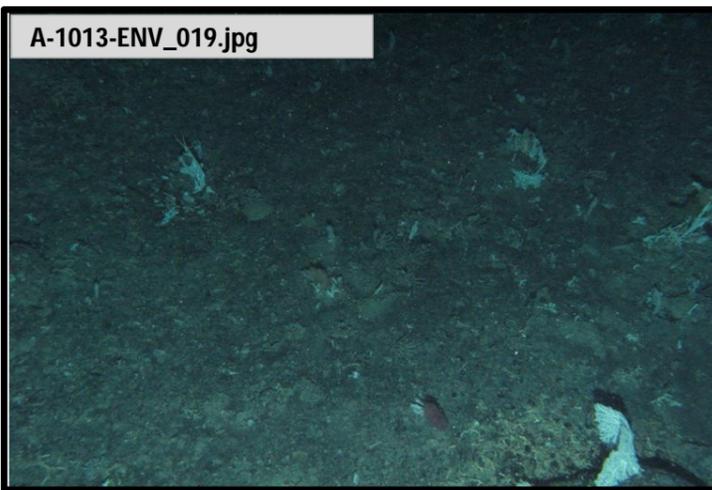


Photo Position: 790314 mE, 4204430 mN



Sediment Example Image

Habitat Summary Information: A/1013/ENV

Survey Area: FISA

No. of Stills: 27 Mins of Video: 31 Track Length: 29m

| | |
|--|--|
| <p>Site Selection Criteria Deepest point of eroded hole</p> | <p>Analogue Interpretation Relatively flat seabed</p> |
| <p>Sediment Description Freckled black sand with cement coloured relic <i>Lophelia</i> sp.</p> | |
| <p>Conspicuous Fauna Tube-building Polychaete, Relic <i>Lophelia</i> sp., Stone Crab: <i>Paralomis</i> sp., Branched Stone Coral: <i>Stylaster</i> sp.,</p> | |

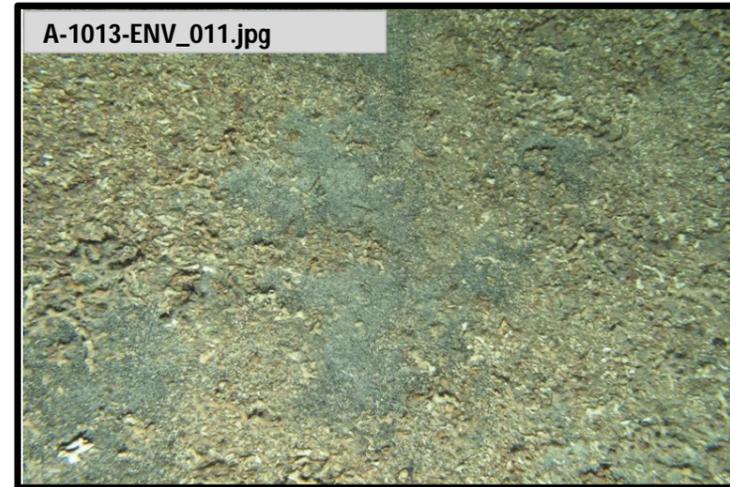


Photo Position: 790314 mE, 4204430 mN

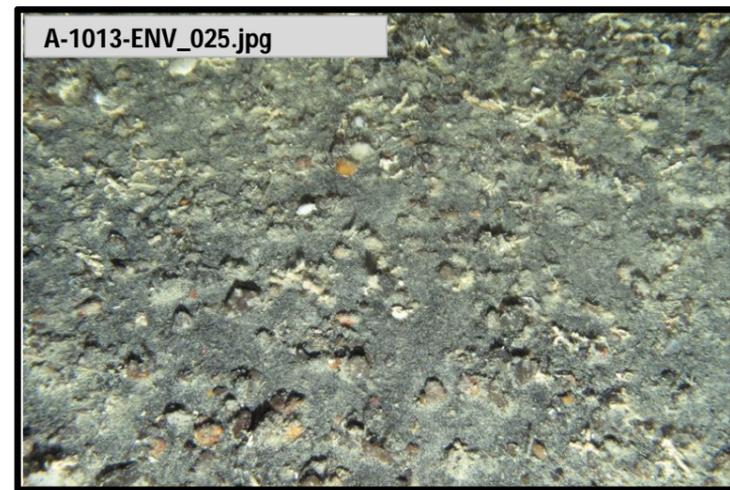
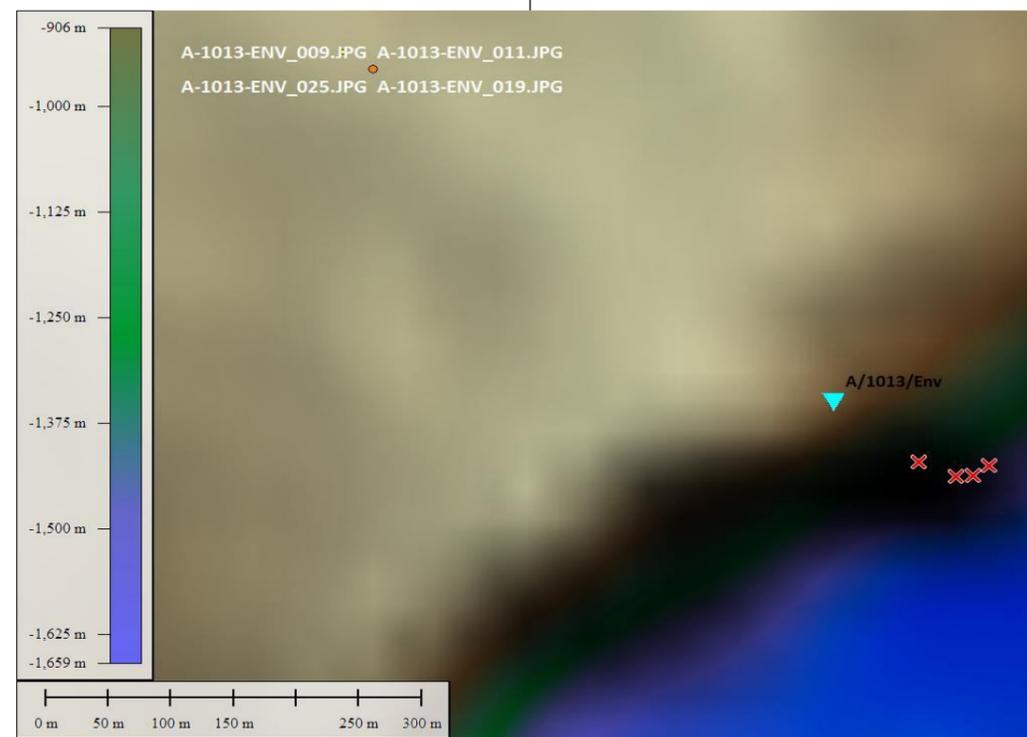


Photo Position: 790314 mE, 4204430 mN



Client
 noble energy

Contractor
 Benthic Solutions
Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

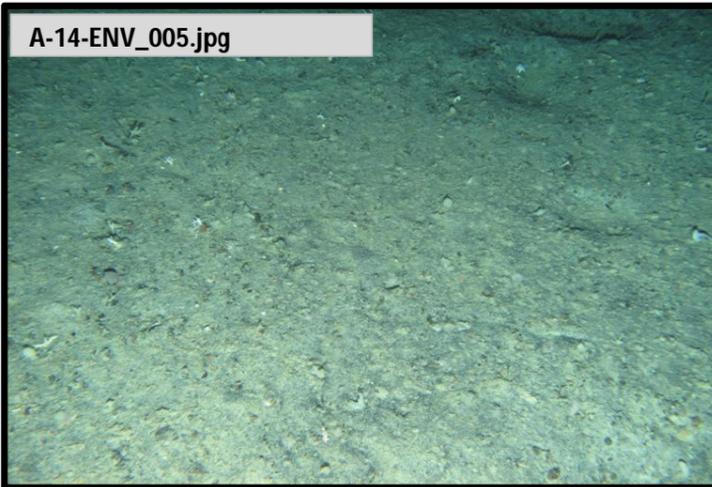


Photo Position: 782524 mE, 4194716 mN

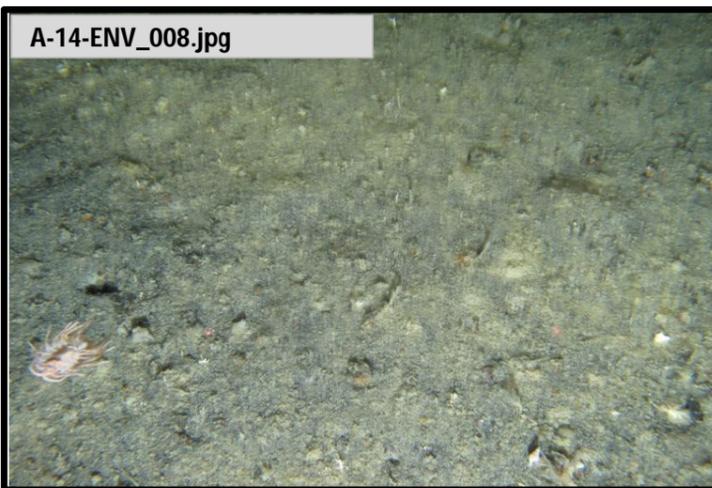


Photo Position: 782549 mE, 4194761 mN



Sediment Example Image

Habitat Summary Information: A/14/ENV

Survey Area: FISA

No. of Stills: 15 Mins of Video: 23 Track Length: 165m

Site Selection Criteria

Edge of regional flat area

Analogue Interpretation

Relatively flat seabed

Sediment Description

Coarse pebbles and rocks suspended in clay

Conspicuous Fauna

Tube-building Polychaete, Polychaete sp., Isopod: possibly *Acutiserolis neaera*, Bryozoa sp.,

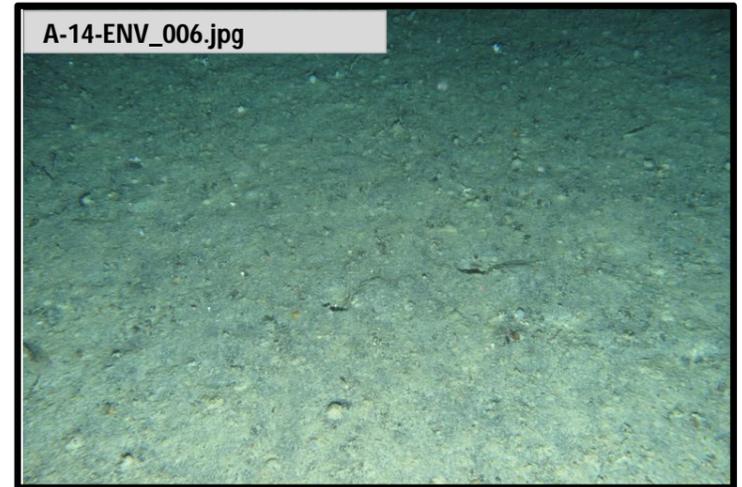


Photo Position: 782528 mE, 4194723 mN

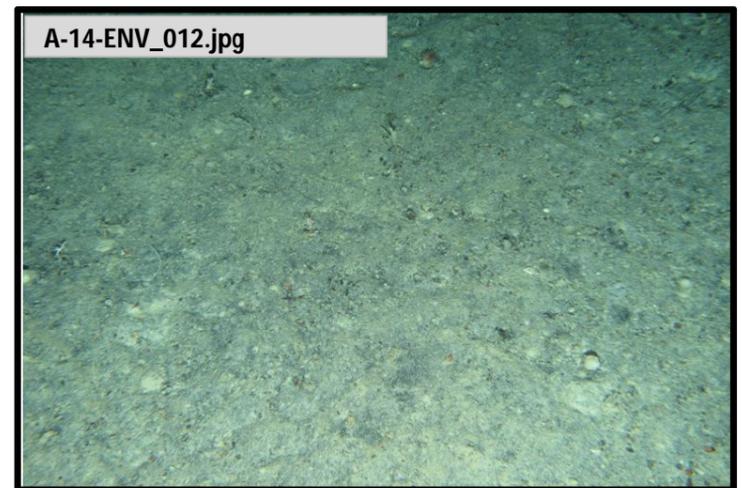
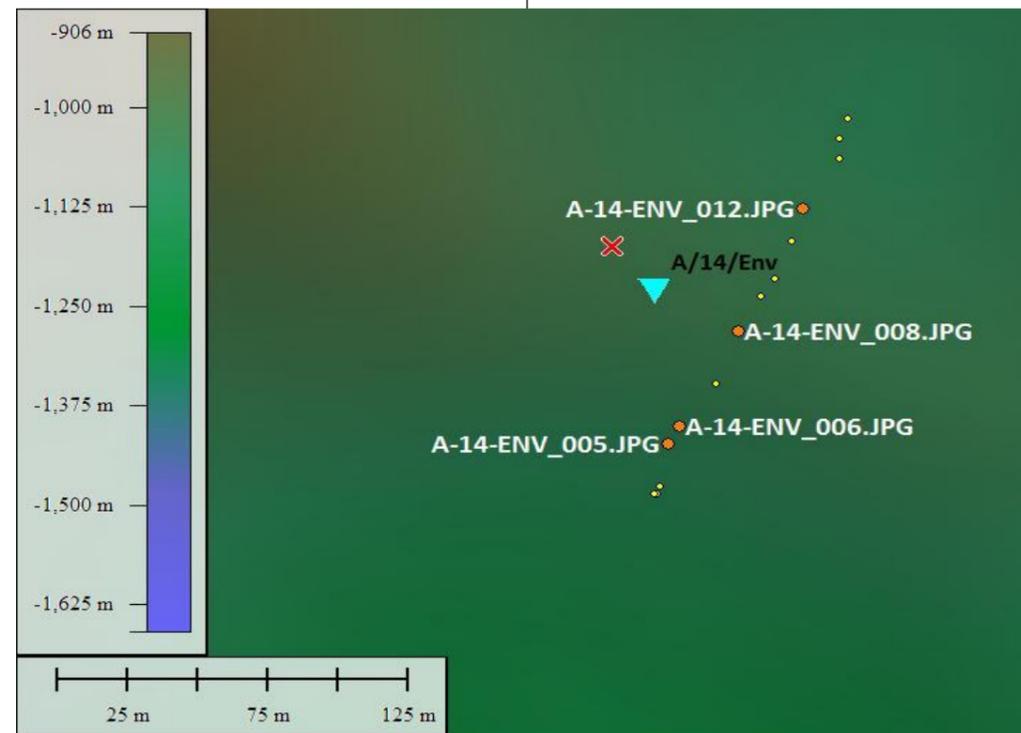


Photo Position: 782572 mE, 4194810 mN



Client
ne noble energy

Contractor
Benthic Solutions
Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

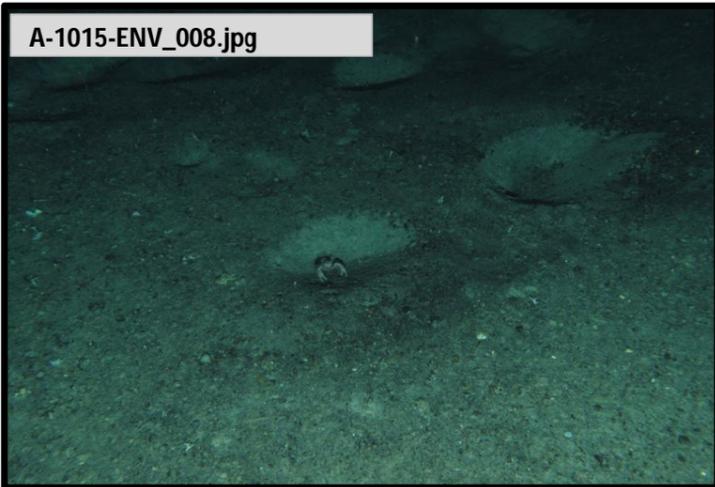


Photo Position: 786584 mE, 4198429 mN

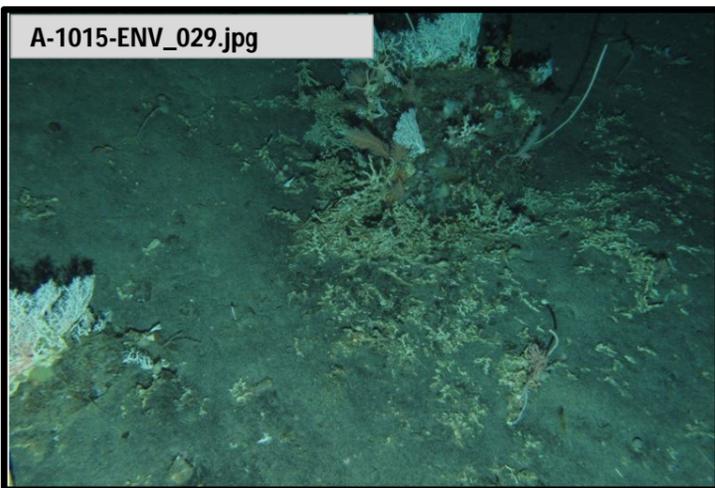


Photo Position: 786407 mE, 4198436 mN



Sediment Example Image

Habitat Summary Information: A/1015/ENV

Survey Area: FISA

No. of Stills: 41

Mins of Video: 41

Track Length: 286m

Site Selection Criteria

Deepest point of hollow feature

Analogue Interpretation

Hollow feature

Sediment Description

Coarse pebbles and rocks suspended in clay

Conspicuous Fauna

Brittlestar (Ophiuroid), Tube-building Polychaete, Polychaete sp., Bryozoa sp., Branched Stone Coral: *Stylaster* sp., Tunicate sp., Relic *Lophelia* sp., Cushion Starfish, Nephropidae sp. *Thymaps birsteini*, Sponge: possibly *Suberites* sp., Branched Sponge: Demospongiae sp.,

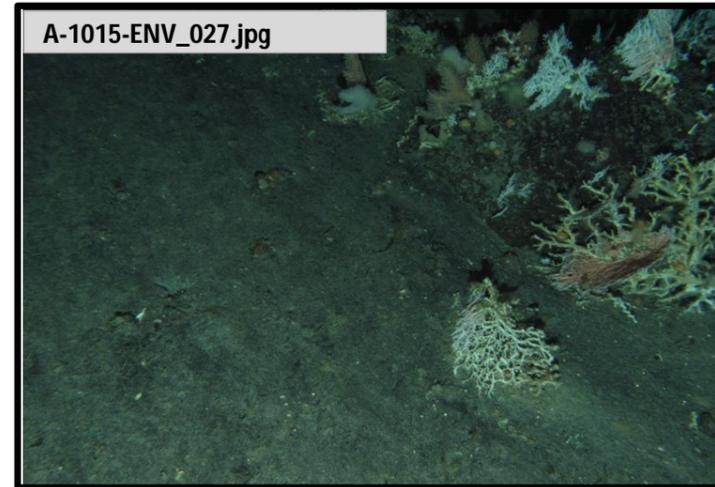


Photo Position: 786420 mE, 4198435 mN

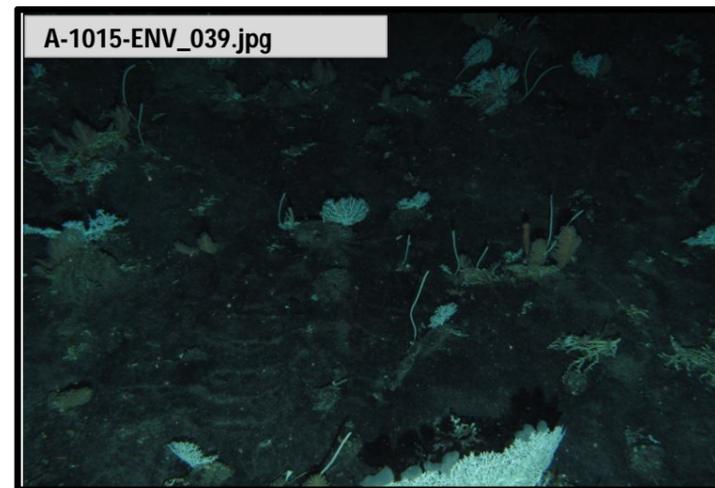
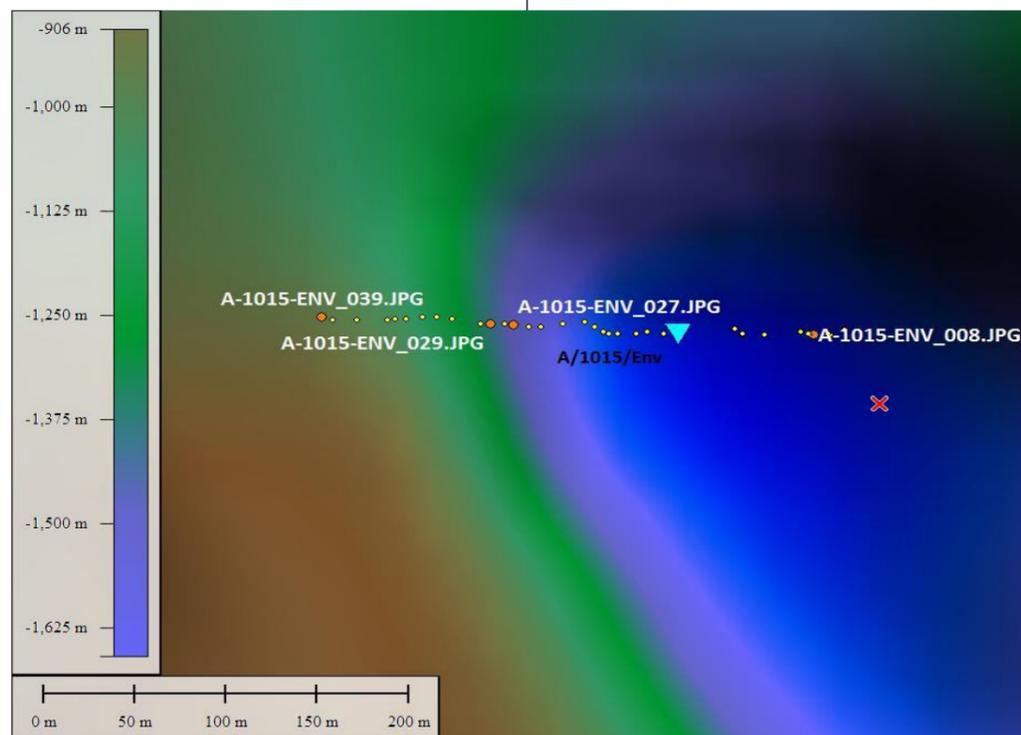


Photo Position: 786315 mE, 4198440 mN



Client
noble energy

Contractor
Benthic Solutions
Benthic Solutions Ltd., Marsh Road,

✗ Sample Location

● UW Photo

● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

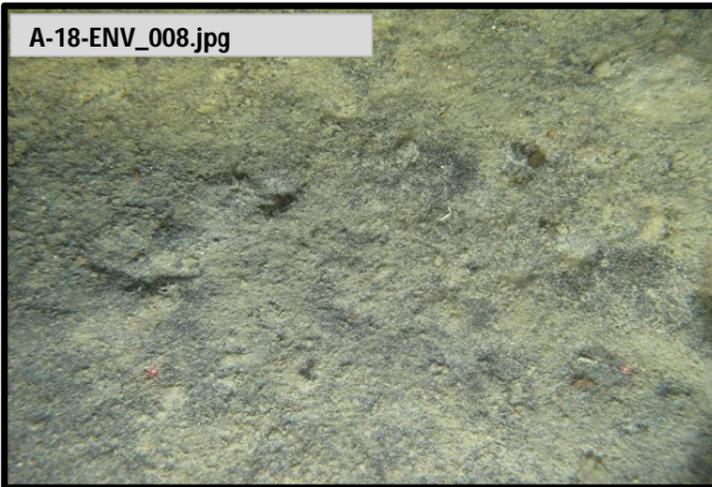


Photo Position: 768048 mE, 4173748 mN

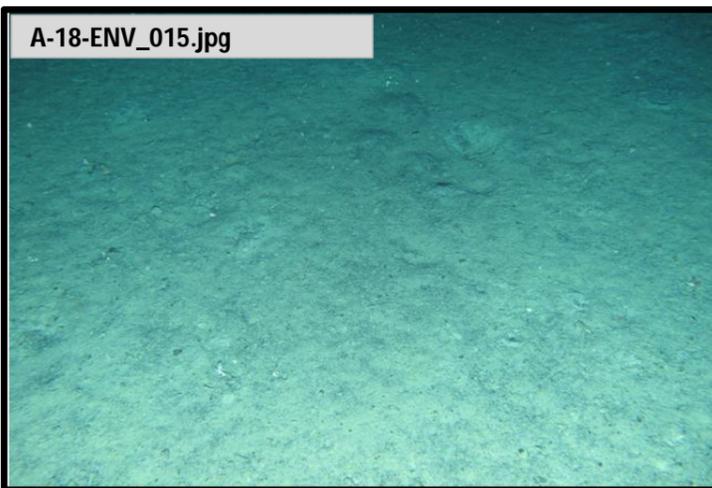


Photo Position: 767936 mE, 4173558 mN



Sediment Example Image

Habitat Summary Information: A/18/ENV

Survey Area: FISA

No. of Stills: 24 Mins of Video: 32 Track Length: 718m

Site Selection Criteria

High amplitude in low relief area

Analogue Interpretation

Small shallow hollow feature

Sediment Description

Homogeneous beige and black silty sand with some gravel, with evidence of bioturbation

Conspicuous Fauna

Stone Crab: *Paralomis* sp., Sponge: (unidentified), Polychaete sp., Cup Coral: *Flabellum* sp.,

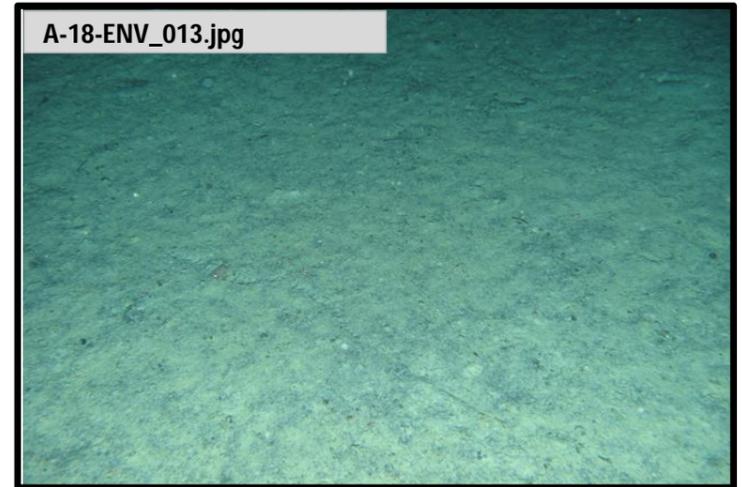


Photo Position: 767954 mE, 4173587 mN

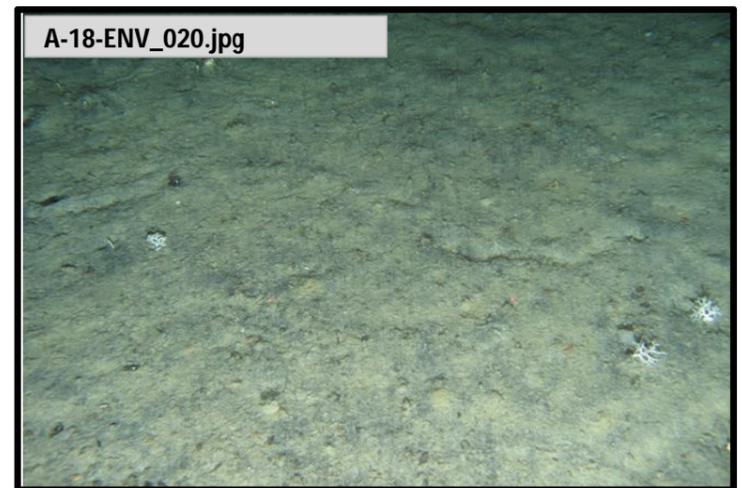
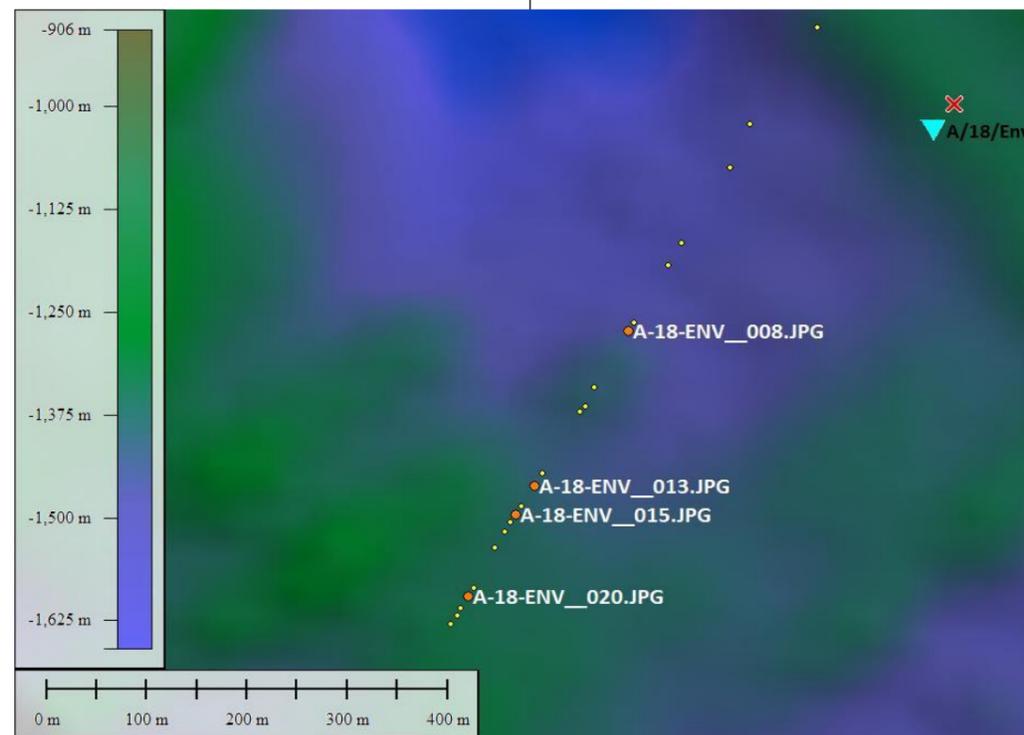
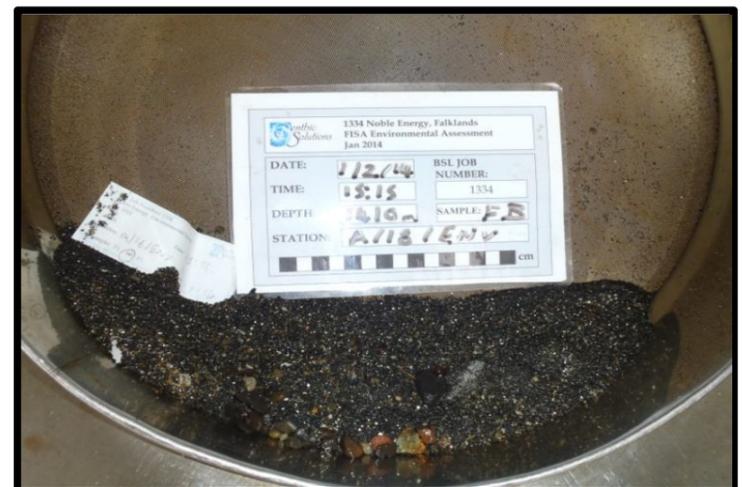


Photo Position: 767888 mE, 4173473 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

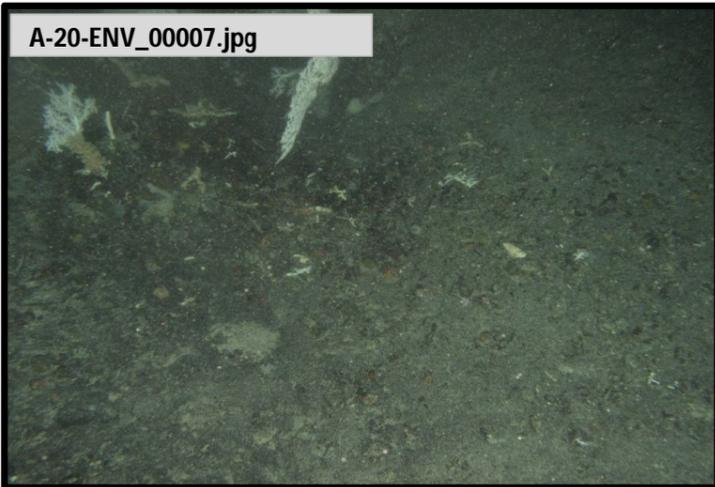


Photo Position: 791007 mE, 4195277 mN

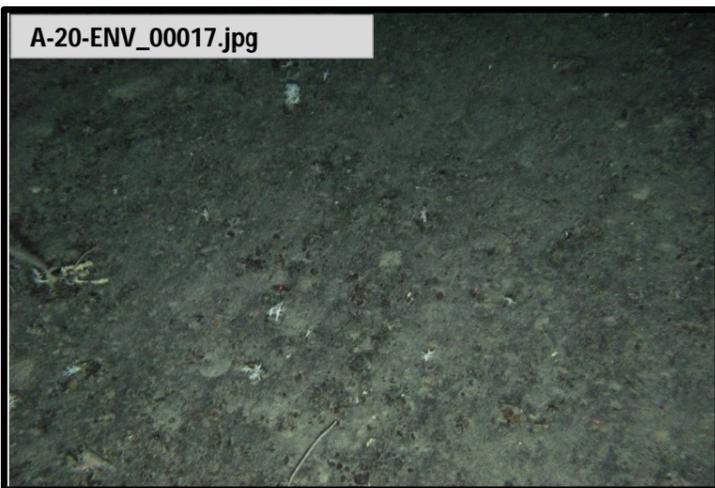


Photo Position: 791042 mE, 4195311 mN



Sediment Example Image

Habitat Summary Information: A/20/ENV

Survey Area: FISA

No. of Stills: 18 Mins of Video: 33 Track Length: 65m

Site Selection Criteria

Regional station

Analogue Interpretation

Relatively flat seabed

Sediment Description

Dark silty sand with gravel

Conspicuous Fauna

Sponge: (unidentified), Sea Pen: Pennatulacea, Brittlestar (Ophiuroid), Branched Stone Coral: *Stylaster sp.*, Hydroid sp.,

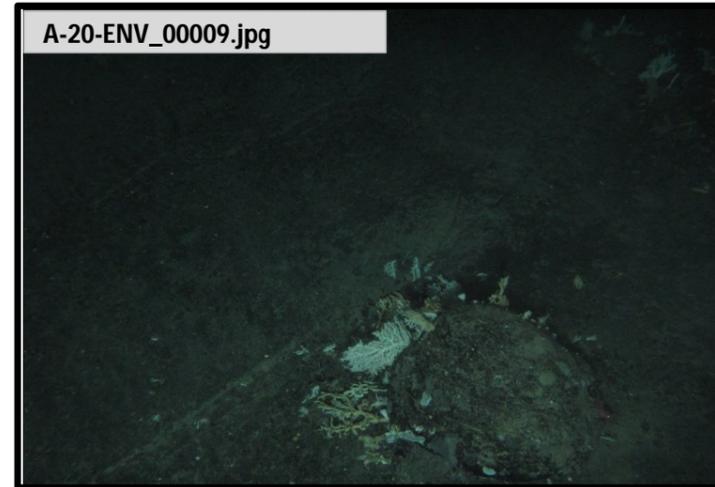


Photo Position: 791009 mE, 4195279 mN

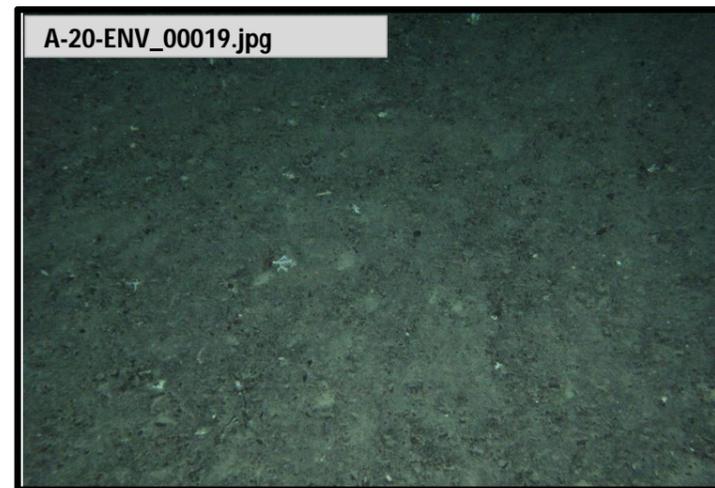


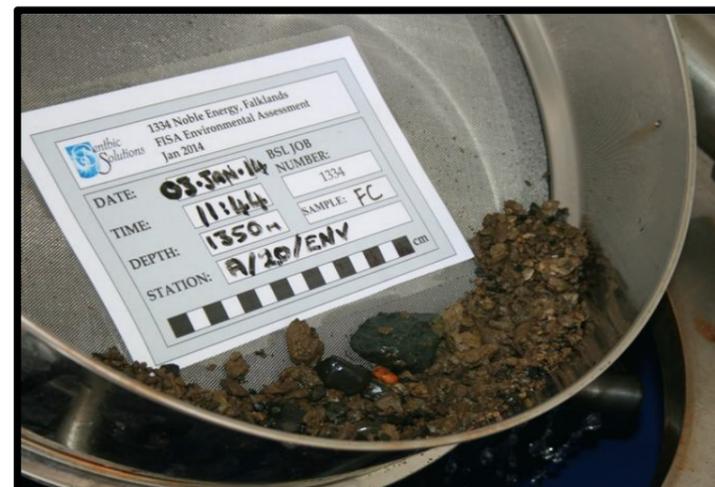
Photo Position: 791055 mE, 4195320 mN



Client **noble energy** Contractor **Benthic Solutions**
 Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West



Sieved Sample Image

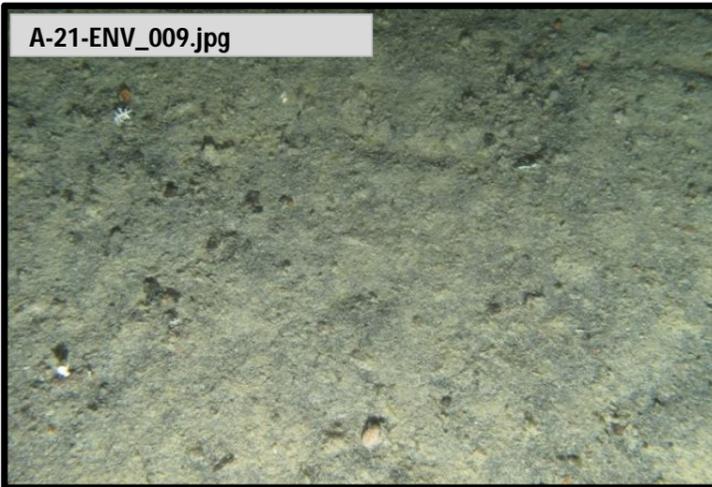


Photo Position: 798264 mE, 4195360 mN

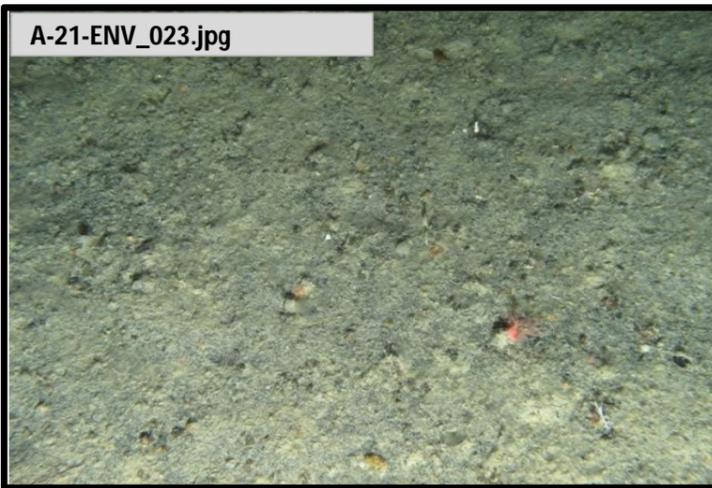


Photo Position: 798204 mE, 4195421 mN



Sediment Example Image

Habitat Summary Information: A/21/ENV

Survey Area: FISA

No. of Stills: 31 Mins of Video: 28 Track Length: 152m

Site Selection Criteria

Regional station

Analogue Interpretation

Relatively flat seabed with slight slope

Sediment Description

Black homogeneous sand

Conspicuous Fauna

Sponge: (unidentified), Tube-building Polychaete, Bryozoa sp.,

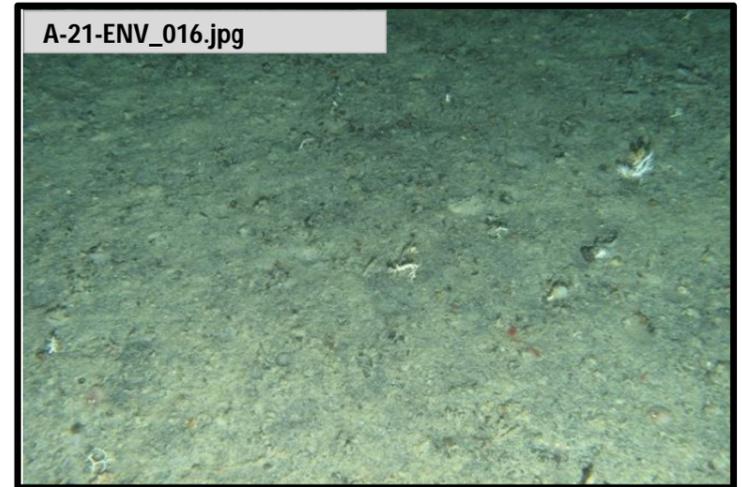


Photo Position: 798235 mE, 4195401 mN

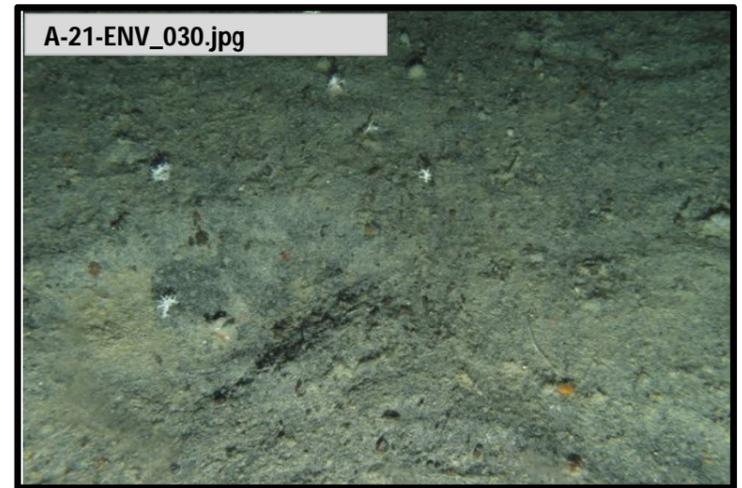
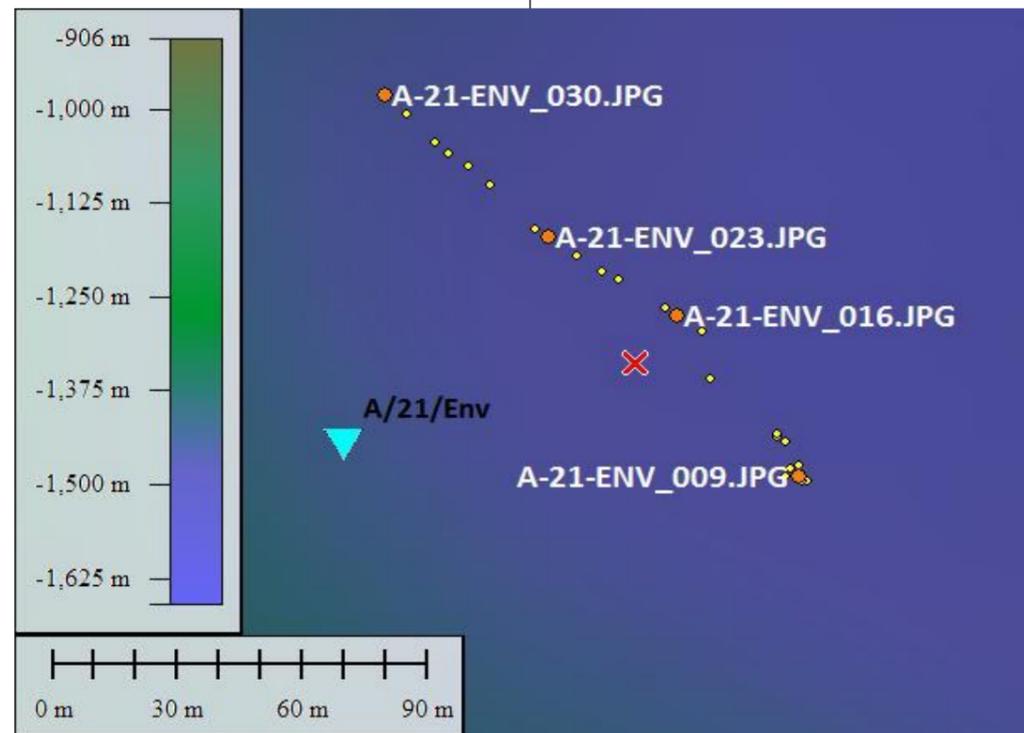


Photo Position: 798165 mE, 4195457 mN



Client
noble energy

Contractor
Benthic Solutions
Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West



Sieved Sample Image

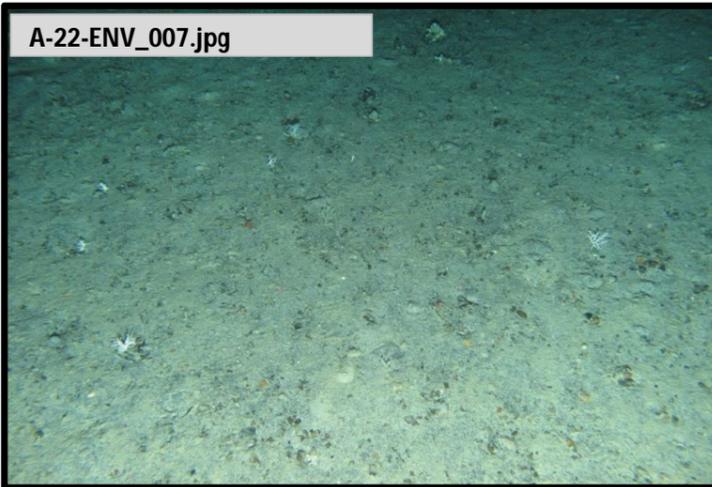


Photo Position: 768038 mE, 4163200 mN

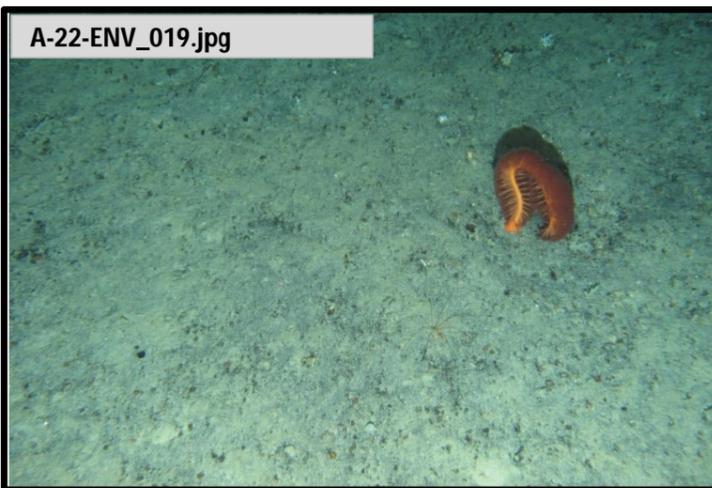


Photo Position: 768056 mE, 4163427 mN



Sediment Example Image

Habitat Summary Information: A/22/ENV

Survey Area: FISA

No. of Stills: 27 Mins of Video: 26 Track Length: 352m

Site Selection Criteria

High amplitude base of slope

Analogue Interpretation

Sloped seabed

Sediment Description

Light/olive grey silty sand with mixed sized gravels

Conspicuous Fauna

Tube-building Polychaete, Bryozoa sp., Sea Pen: possibly *Anthoptilum grandiflorum*, Sea Spider: *Pycnogonida* sp., Isopod: possibly *Acutiserolis neaera*, Sea Pen: Pennatulacea, Relic *Lophelia* sp., Encrusting Sponge, Sea Urchin: Echinoidea sp., Branched Stone Coral: *Stylaster* sp.,



Photo Position: 768032 mE, 4163339 mN

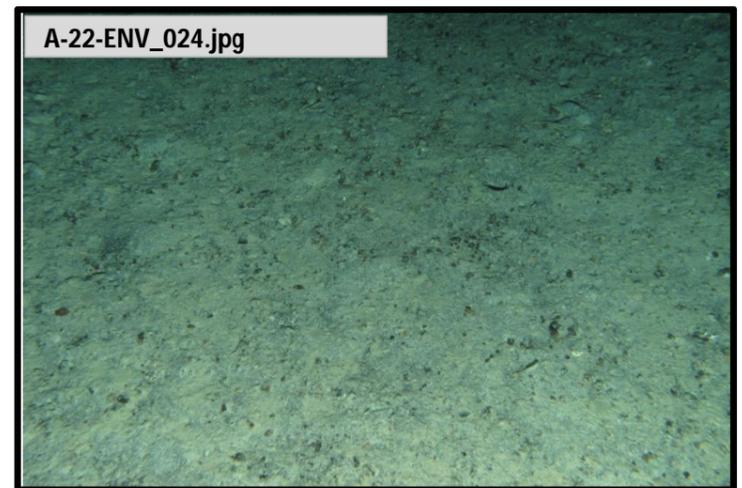
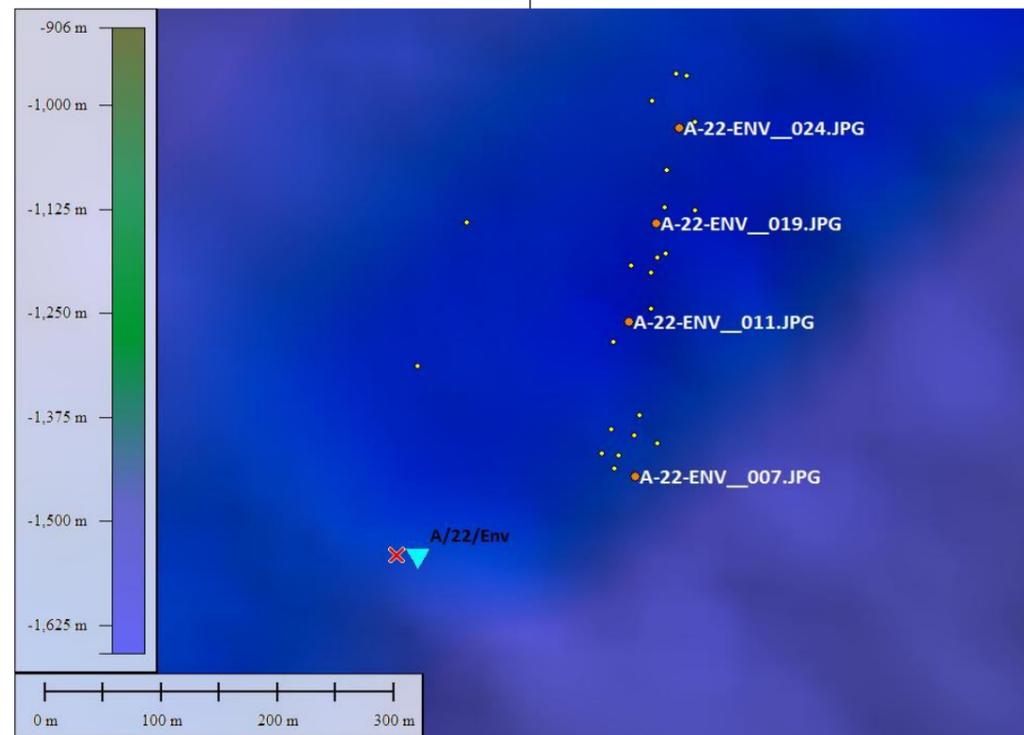


Photo Position: 768076 mE, 4163512 mN



Client **noble energy** Contractor **Benthic Solutions**
 Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

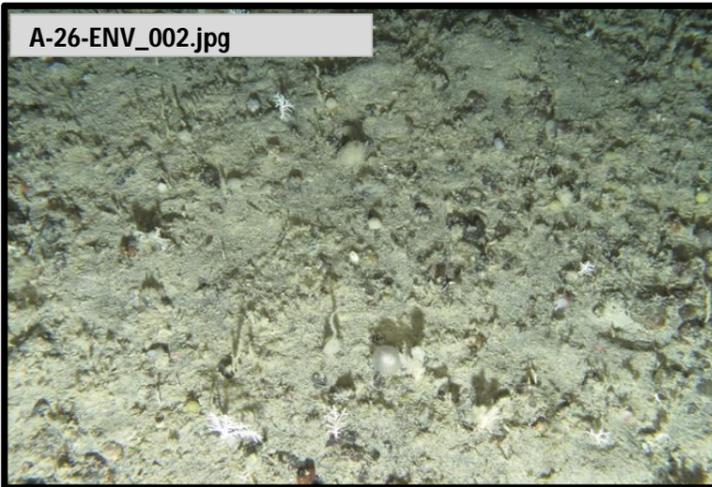


Photo Position: 768038 mE, 4163200 mN

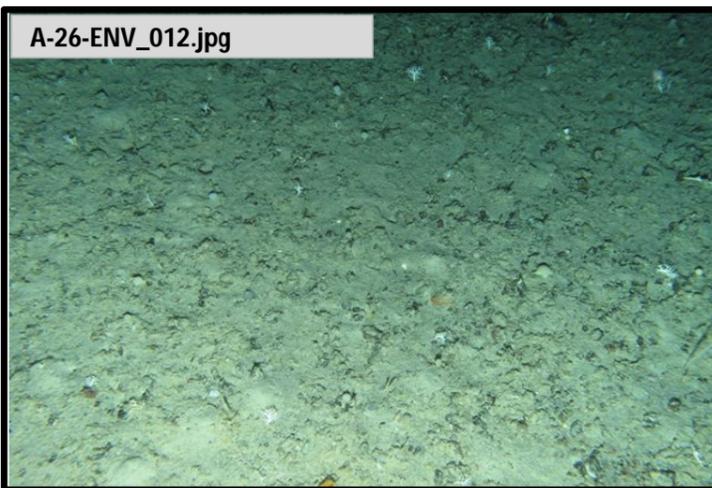


Photo Position: 768056 mE, 4163427 mN



Sediment Example Image

Habitat Summary Information: A/26/ENV

Survey Area: FISA

No. of Stills: 19 Mins of Video: 21 Track Length: 297m

Site Selection Criteria

Close to proposed well location, (Finback-1)

Analogue Interpretation

Slightly sloped seabed

Sediment Description

Compact medium sand with some coarse sediment and stones

Conspicuous Fauna

Tube-building Polychaete, Bryozoa sp., Encrusting Sponge, Isopod: possibly *Acutiserolis neaera*, Branched Stone Coral: *Stylaster* sp., Branched Bryozoa sp., Sea Pen: Pennatulacea,

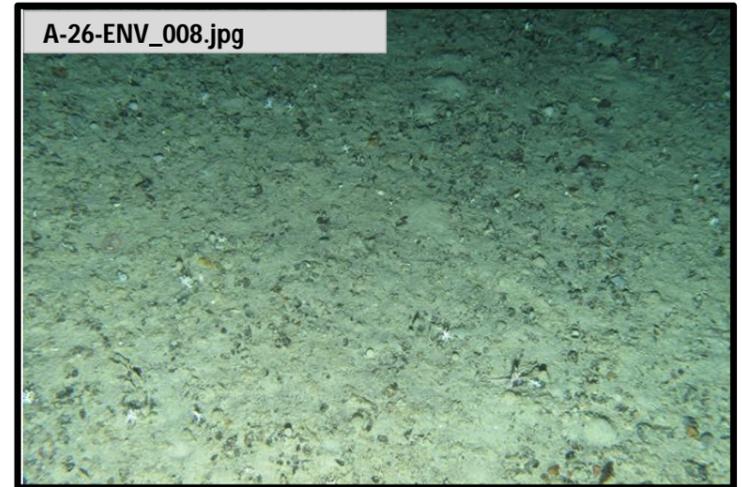


Photo Position: 768032 mE, 4163339 mN

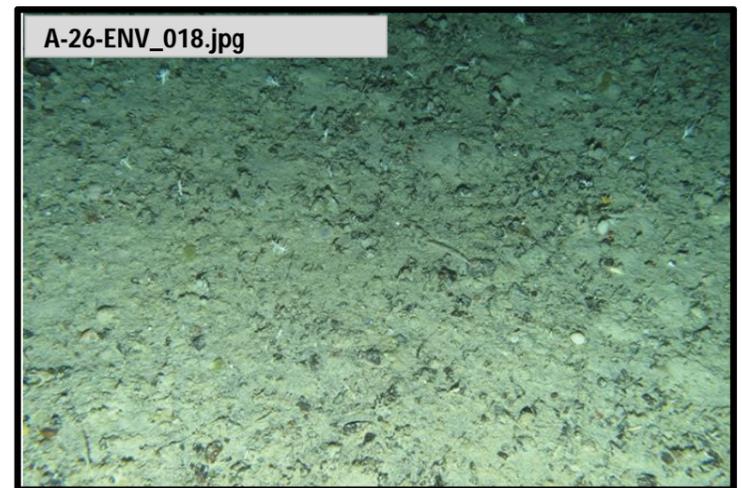
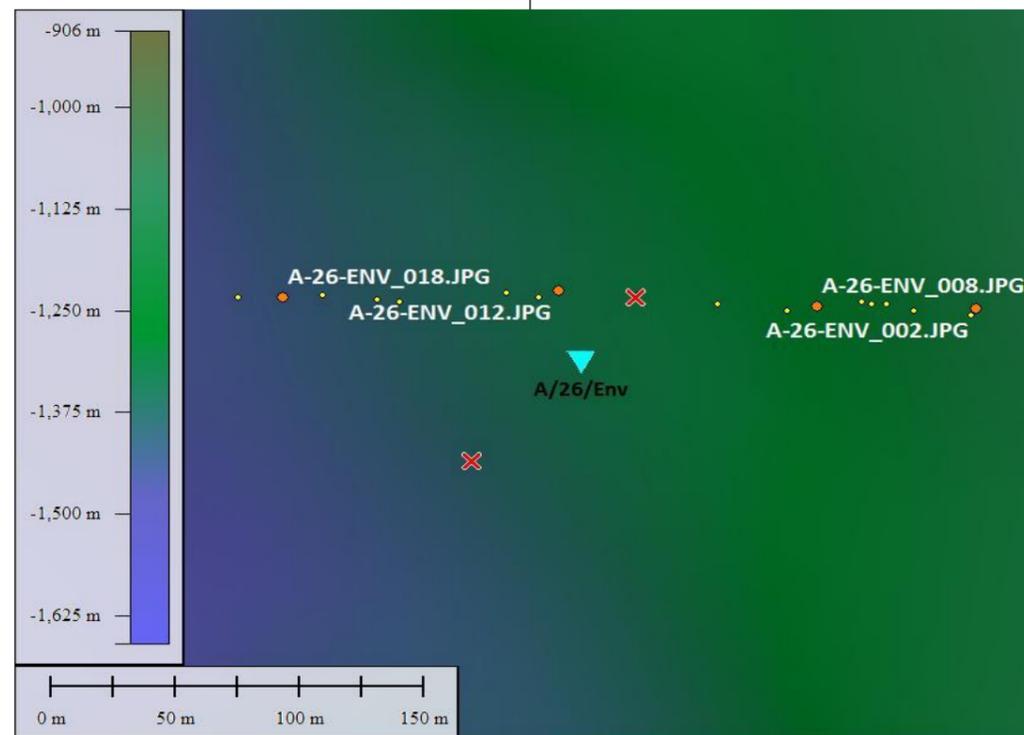


Photo Position: 768076 mE, 4163512 mN



✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West



Photo Position: 792429 mE, 4213811 mN

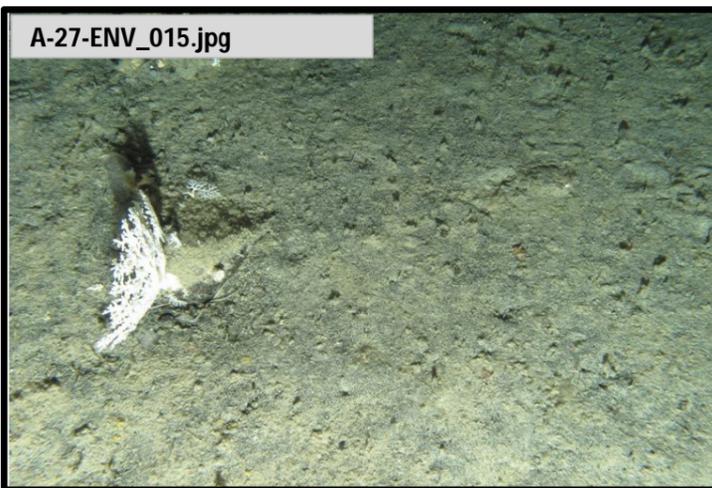


Photo Position: 792427 mE, 4213868 mN



Sediment Example Image

Habitat Summary Information: A/27/ENV

Survey Area: FISA

No. of Stills: 25 Mins of Video: 23 Track Length: 138m

Site Selection Criteria

Close to proposed well location, (Humpback-1)

Analogue Interpretation

Slightly sloped seabed

Sediment Description

Fine and compact beige and black sand with boulders

Conspicuous Fauna

Cup Coral: *Flabellum* sp., Bryozoa sp., Branched Stone Coral: *Stylaster* sp., Holothuroidea sp., Decapod Shrimp sp., Isopod: possibly *Acutiserolis neaera*, Encrusting Sponge, Sea Pen: Pennatulacea, Hydroid sp., Branched Bryozoa sp., Tunicate sp., Nephropidae sp., *Thymops birsteini*, Sea Urchin: Echinoidea sp., Brittlestar (Ophiuroid), Anthozoa: *Cerianthus* sp., Octocoral sp.,

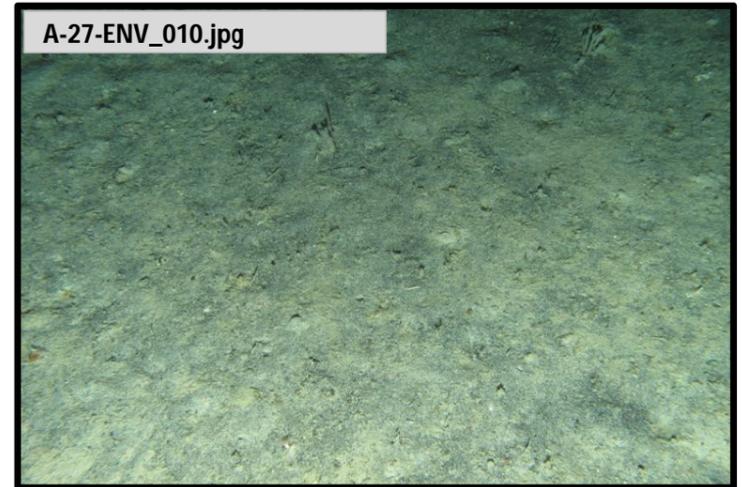


Photo Position: 792427 mE, 4213830 mN

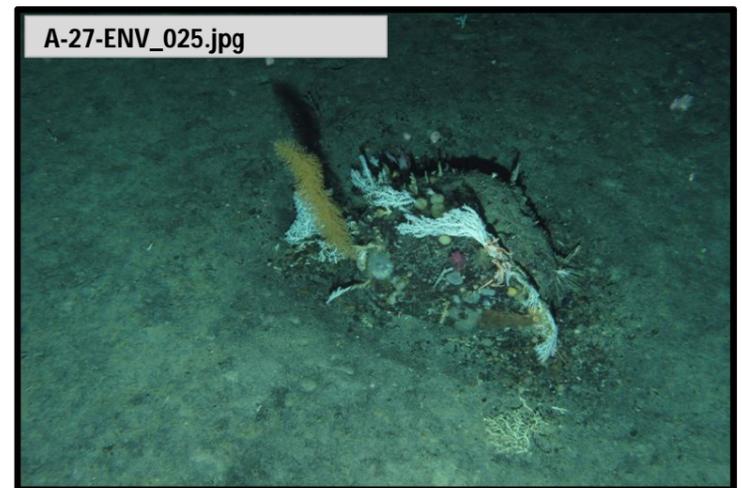
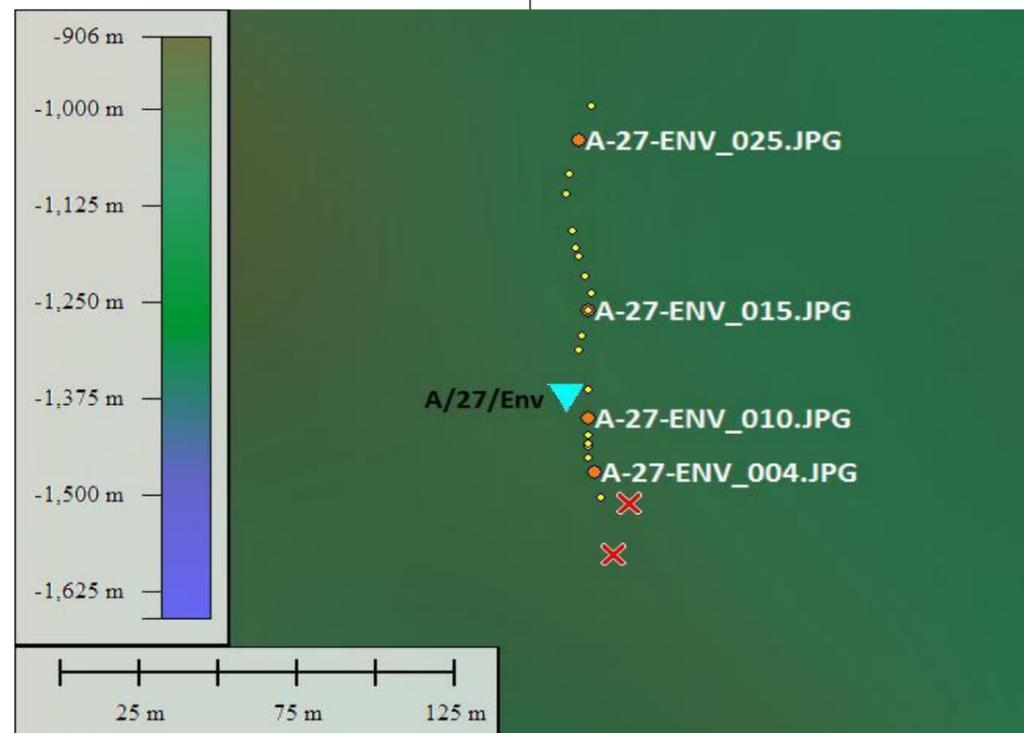


Photo Position: 792424 mE, 4213928 mN



Client
noble energy

Contractor
Benthic Solutions
Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

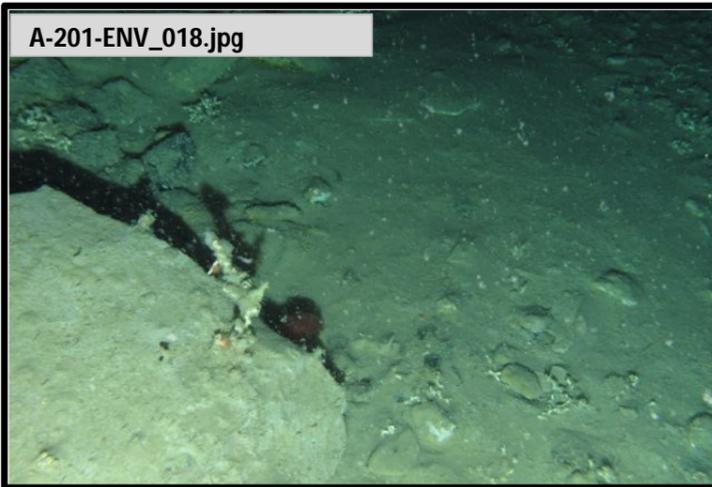


Photo Position: 734346 mE, 4206707 mN

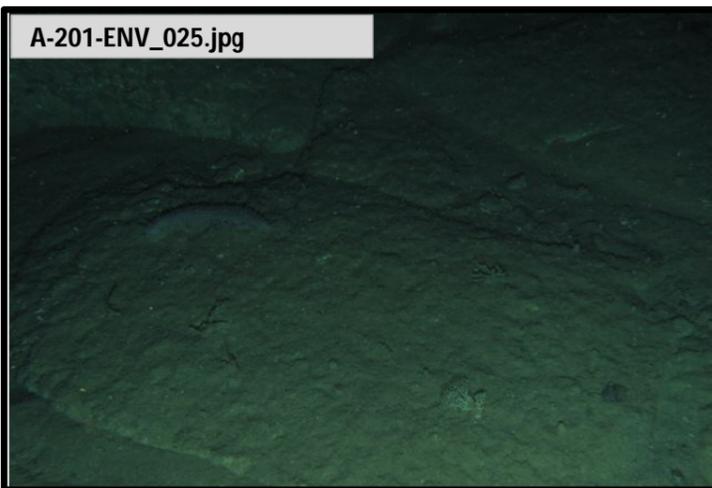


Photo Position: 734400 mE, 4206710 mN

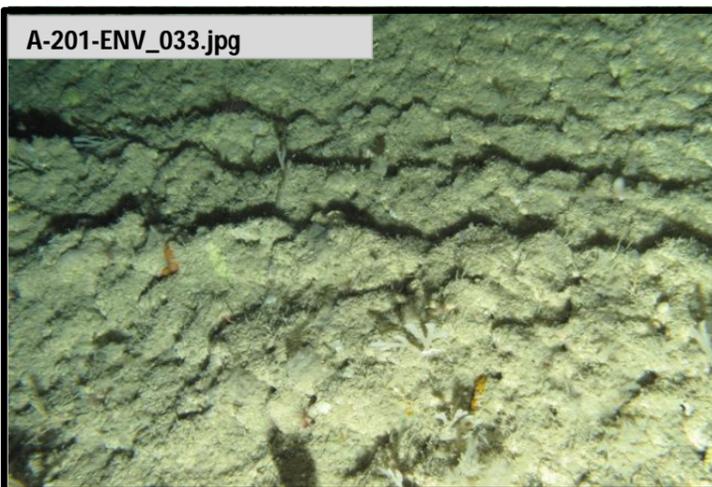


Photo Position: 734514 mE, 4206716 mN

Habitat Summary Information: A/201/ENV

Survey Area: FISA

No. of Stills: 41 Mins of Video: 40 Track Length: 393m

| | |
|---|--|
| <p>Site Selection Criteria Escarpment in 'hole' feature</p> | <p>Analogue Interpretation Escarpment feature</p> |
| <p>Sediment Description Hard compact fine sand with cobbles and boulders</p> | |
| <p>Conspicuous Fauna Lophelia pertusa, Sea Pen: possibly <i>Anthoptilum grandiflorum</i>, Bryozoa sp., Sea Pen: Pennatulacea, Anthozoa: <i>Cerianthus</i> sp., Eelpout: <i>Lycenchelys</i> sp., Tube-building Polychaete, Isopod: possibly <i>Acutiserolis neaera</i>, Brittlestar (Ophiuroid), Gastropoda sp., Relic <i>Lophelia</i> sp., Branched Bryozoa sp., Relic Polychaete Tubes, Moridae sp., Squat Lobster: Galatheididae sp., Sponge: Porifera sp., Funnel Sponge: Demospongiae sp., Branched Stone Coral: <i>Stylaster</i> sp., Hydroid sp., Holothuroidea sp., Cushion Starfish, Gastropoda sp.,</p> | |



Photo Position: 734350 mE, 4206708 mN

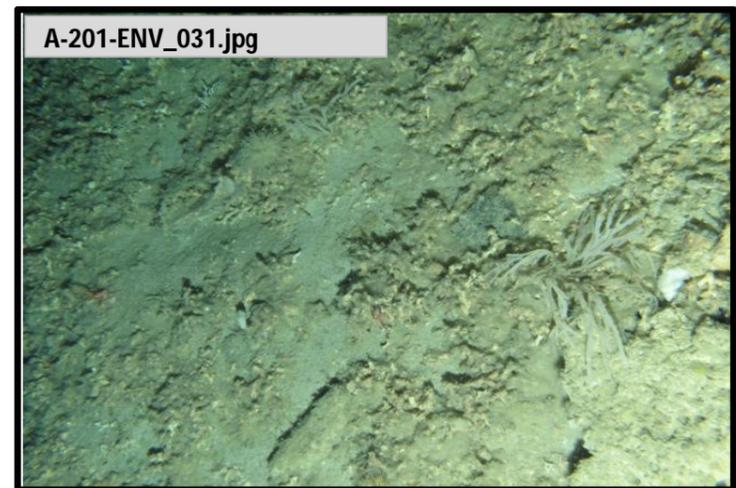


Photo Position: 734452 mE, 4206717 mN

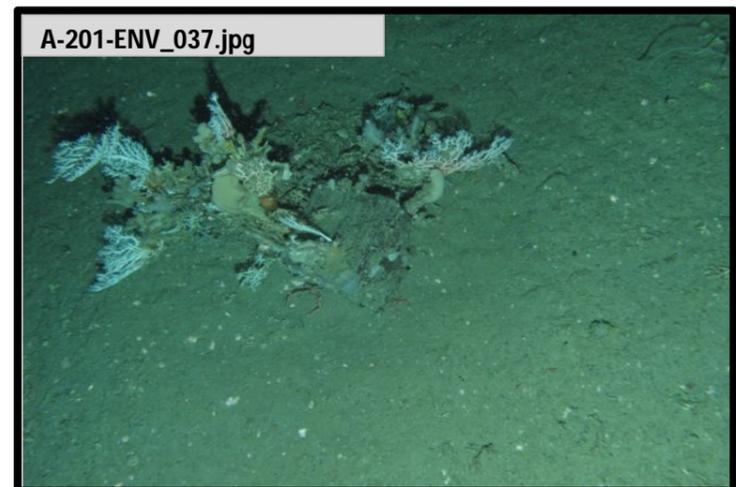
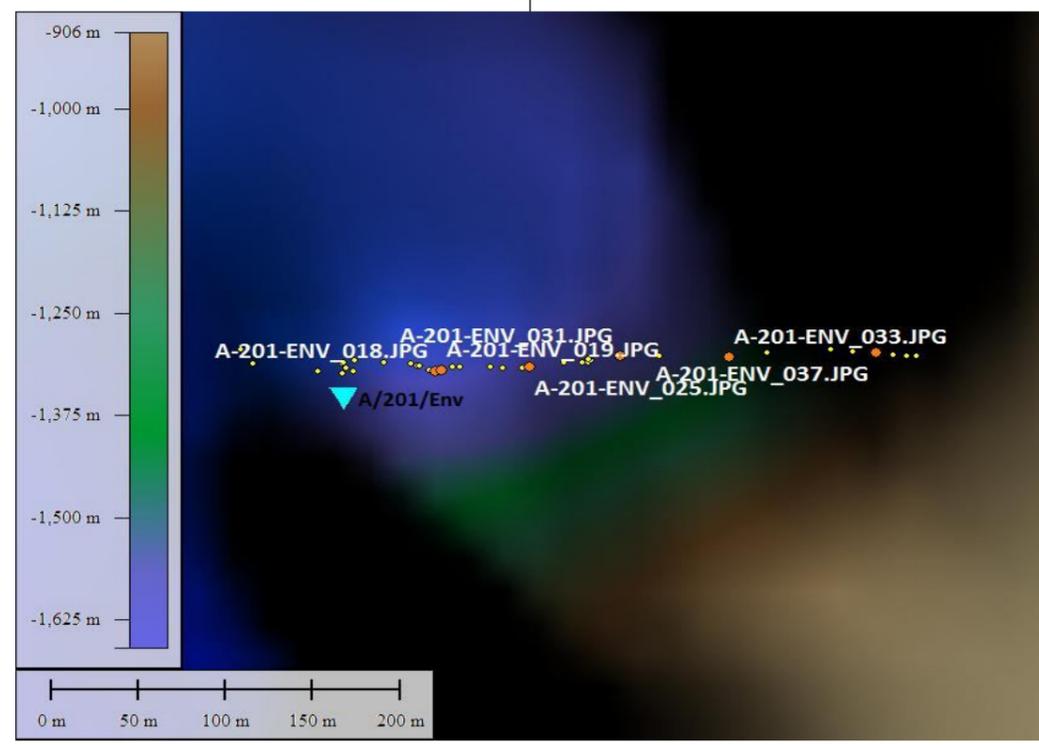


Photo Position: 734598 mE, 4206719 mN



| | |
|--|---|
| <p>Client noble energy</p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
|--|---|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

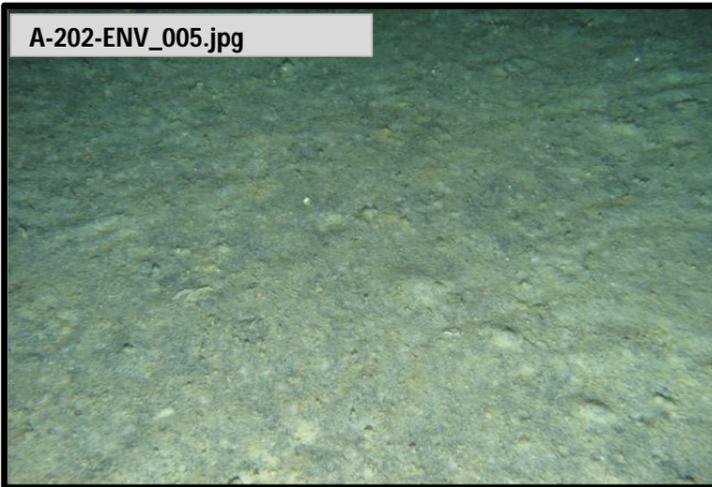


Photo Position: 754157 mE, 4216068 mN

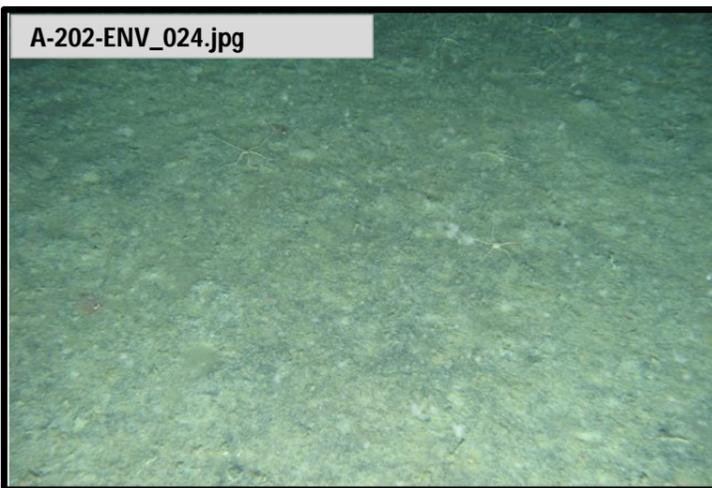


Photo Position: 754186 mE, 4216082 mN



Sediment Example Image

Habitat Summary Information: A/202/ENV

Survey Area: FISA

No. of Stills: 33 Mins of Video: 43 Track Length: 93m

| | |
|--|---|
| <p>Site Selection Criteria Undulating seafloor</p> | <p>Analogue Interpretation Base of slope</p> |
| <p>Sediment Description Hard compact medium sand with clay lumps</p> | |
| <p>Conspicuous Fauna Brittlestar (Ophiuroid), Bryozoa sp., Tube-building Polychaete, Polychaete sp., Isopod: possibly <i>Acutiserolis neaera</i>, Sea Pen: possibly <i>Anthoptilum grandiflorum</i>, Cup Coral: <i>Flabellum</i> sp.,</p> | |



Photo Position: 754186 mE, 4216082 mN

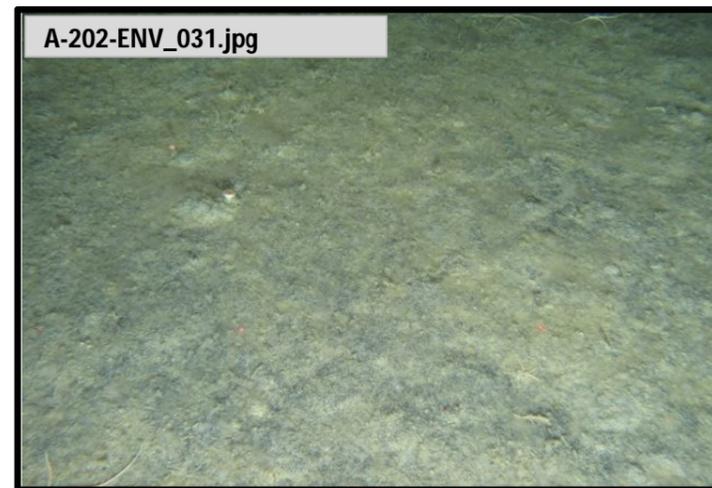
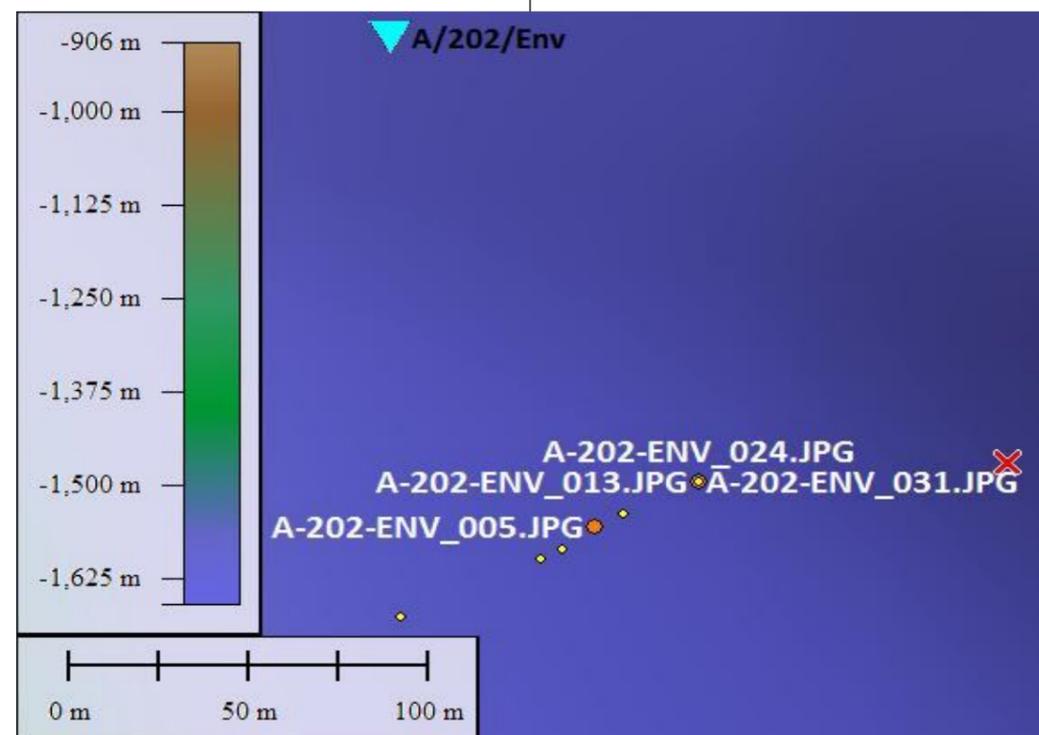
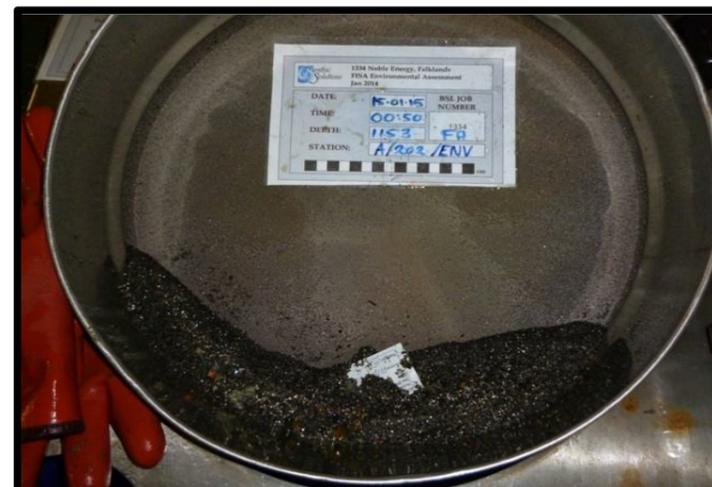


Photo Position: 754186 mE, 4216082 mN



| | |
|---------------|--|
| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
|---------------|--|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

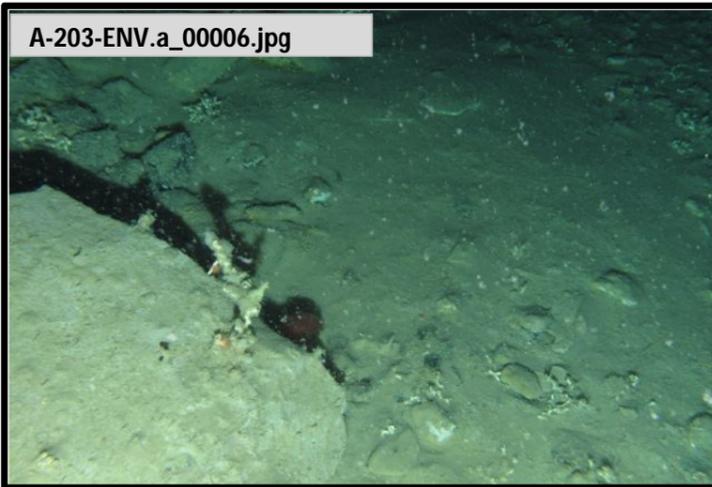


Photo Position: 787586 mE, 4199232 mN

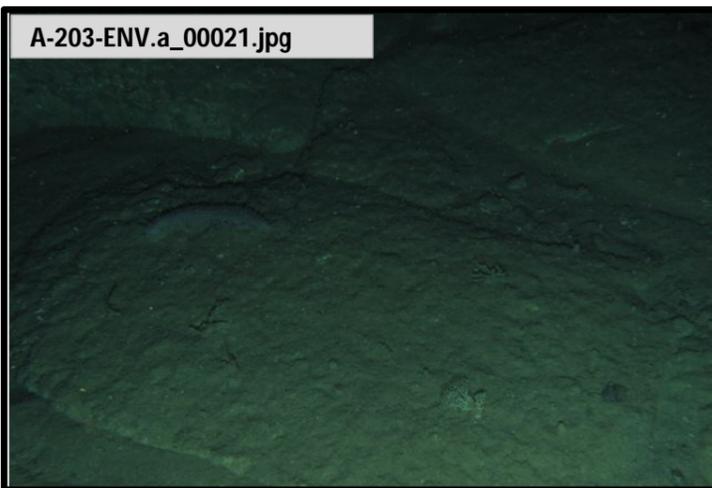


Photo Position: 787566 mE, 4199170 mN

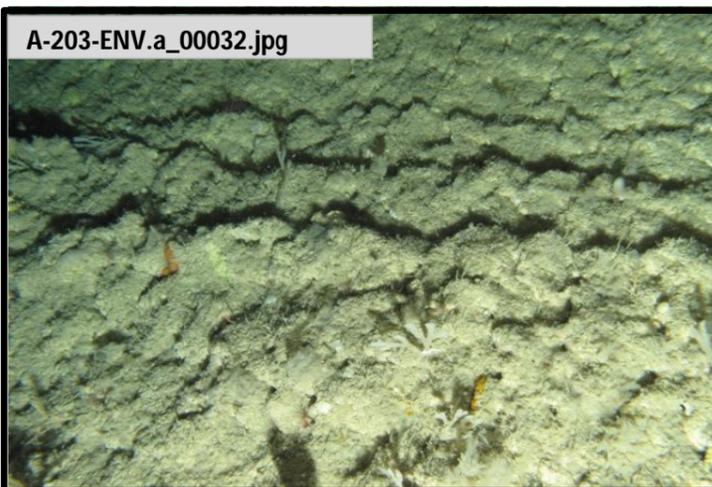


Photo Position: 787555 mE, 4199137 mN

Habitat Summary Information: A/203/ENV

Survey Area: FISA

No. of Stills: 37 Mins of Video: 26 Track Length: 130m

| | |
|---|---|
| <p>Site Selection Criteria Camera transect on isolated feature</p> | <p>Analogue Interpretation Sloped seabed</p> |
| <p>Sediment Description Hard compact fine sand with cobbles and boulders</p> | |
| <p>Conspicuous Fauna Lophelia pertusa, Relic <i>Lophelia</i> sp., Tube-building Polychaete, Brittlestar (Ophiuroid), Gorgonian sp., Squat Lobster: Galatheididae sp., Branched Bryozoa sp., Sea Pen: Pennatulacea, Branched Stone Coral: Stylaster sp., Hydroid sp., Anthozoa: Soft coral, Pencil Urchin: possibly <i>Cidaris</i> sp., Sponge: Porifera sp., Moridae sp., Hake: possibly <i>Merluccius hubbsi</i>, Decapod Shrimp sp.,</p> | |

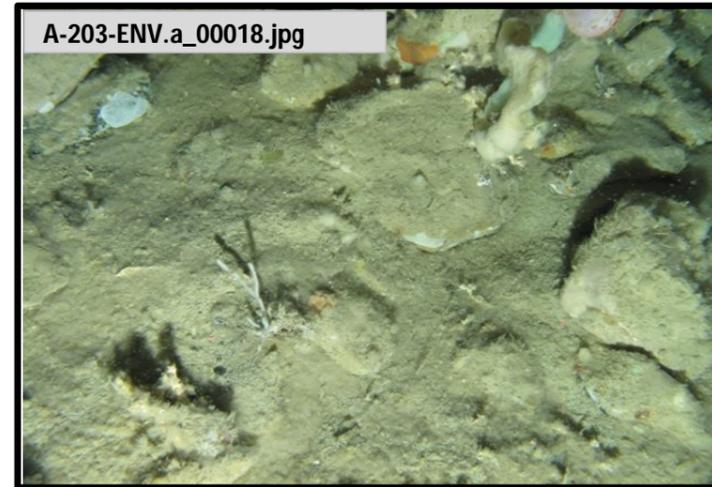


Photo Position: 787570 mE, 4199182 mN

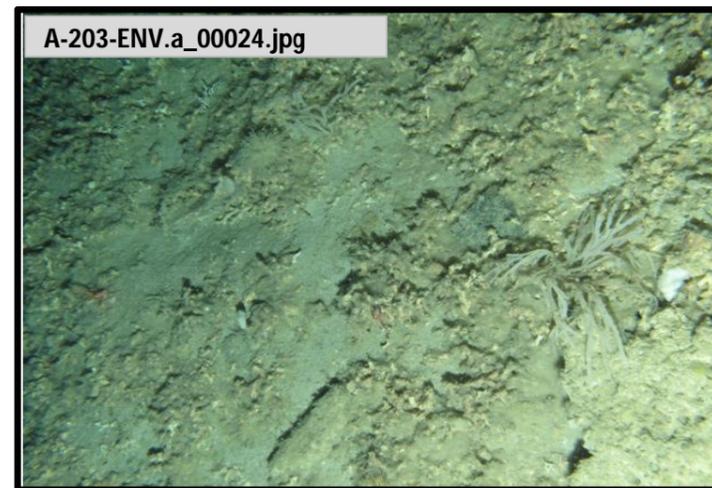


Photo Position: 787563 mE, 4199162 mN

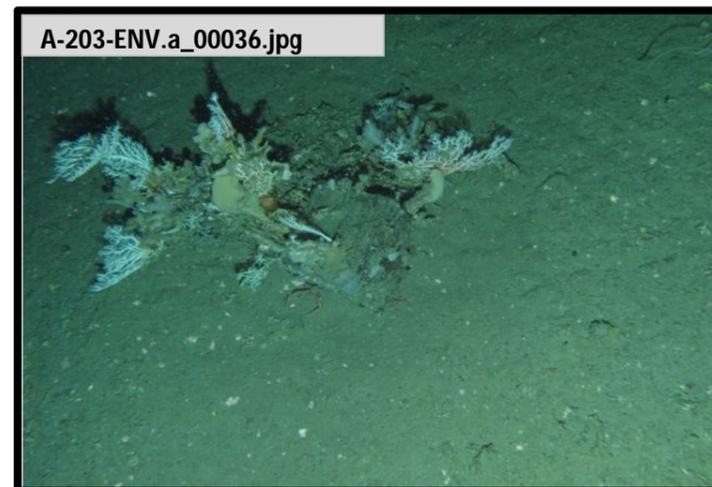
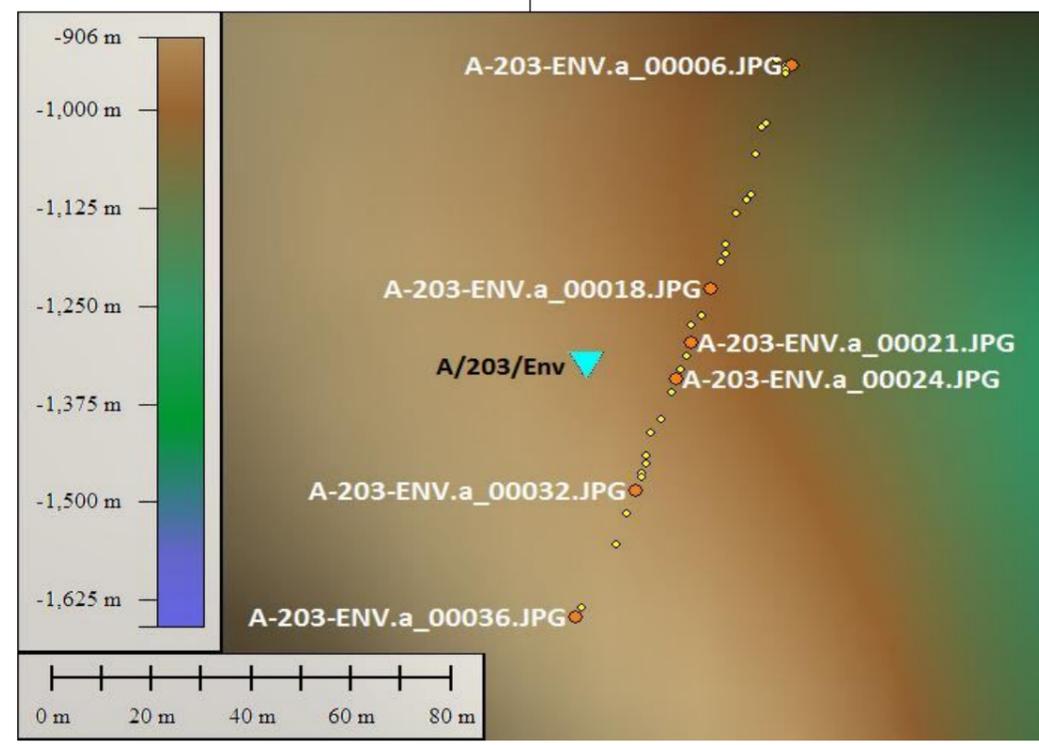


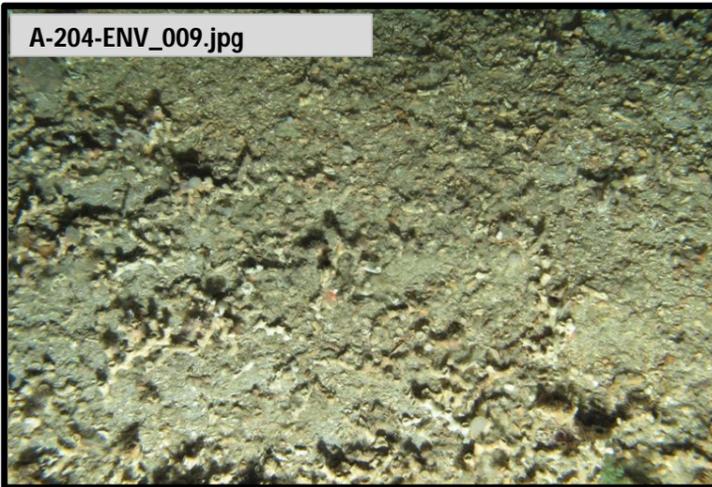
Photo Position: 787543 mE, 4199109 mN



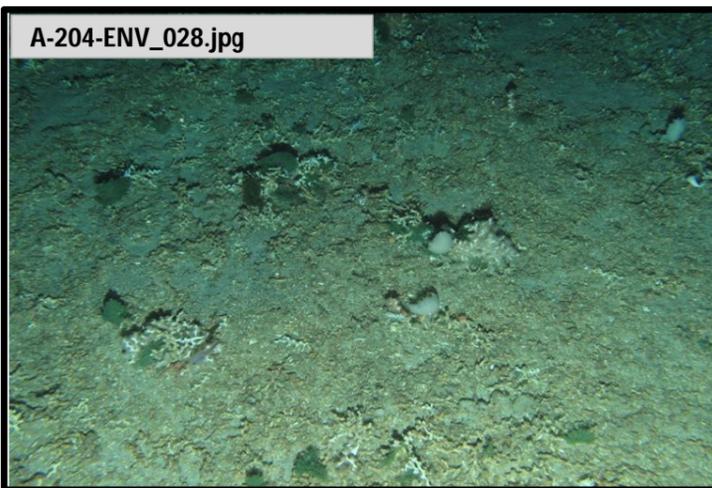
| | |
|--|---|
| <p>Client noble energy</p> | <p>Contractor Benthic Solutions Benthic Solutions Ltd., Marsh Road,</p> |
|--|---|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West



No beacon data acquired



No beacon data acquired



Sediment Example Image

Habitat Summary Information: A/204/ENV

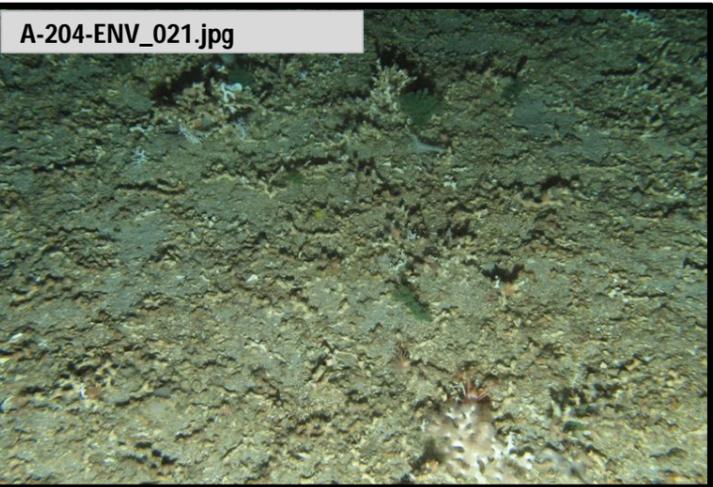
Survey Area: FISA

No. of Stills: 50 Mins of Video: 27 Track Length: No Data

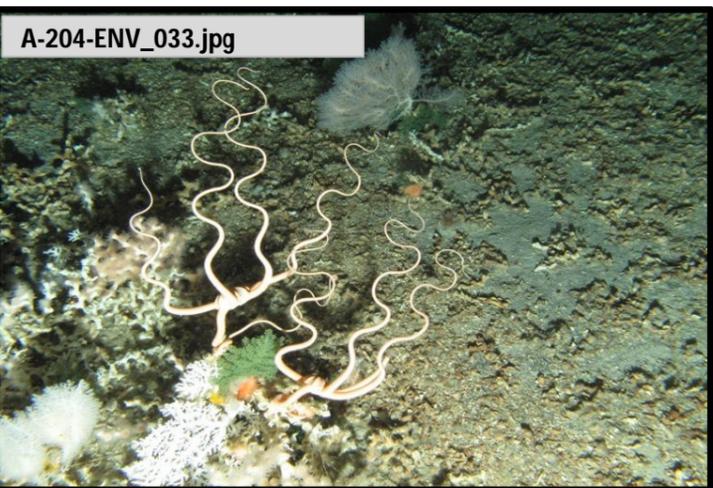
| | |
|---|---|
| <p>Site Selection Criteria Slump feature</p> | <p>Analogue Interpretation Slump feature</p> |
|---|---|

Sediment Description
Coarse beige and dark sediment, larger boulder pile-up/slump feature with multiple collapses

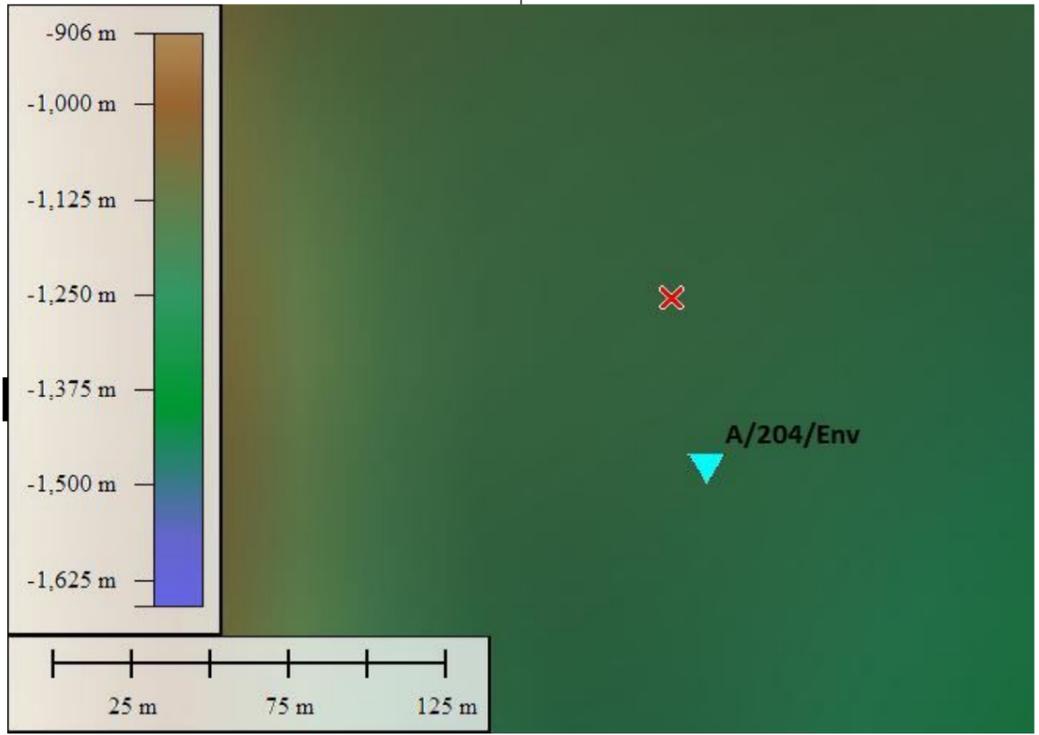
Conspicuous Fauna
Lophelia pertusa, Relic *Lophelia* sp., Anthozoa: Soft coral, Hydroid sp., Sponge: Porifera sp., Branched Stone Coral: *Stylaster* sp., Gorgonian sp., Brittlestar (Ophiuroid), Moridae sp., Squat Lobster: Galatheididae sp., Bryozoa sp., Sea Pen: Pennatulacea, Branched Bryozoa sp., Encrusting Sponge,



No beacon data acquired



No beacon data acquired



| | |
|---------------|--|
| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
|---------------|--|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

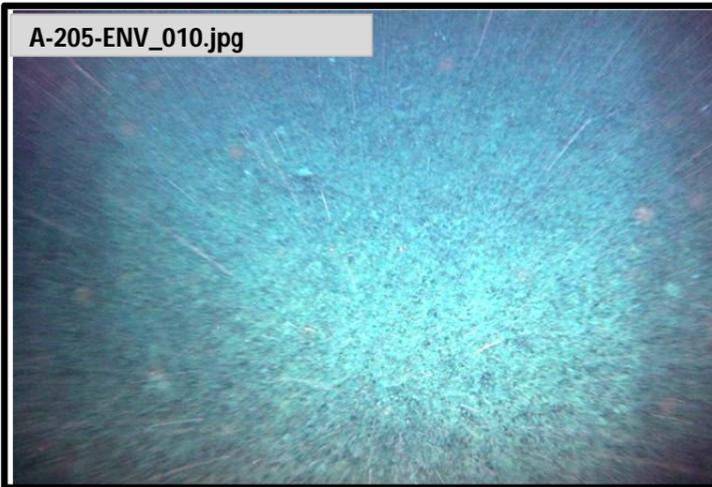


Photo Position: 792218 mE, 4178377 mN

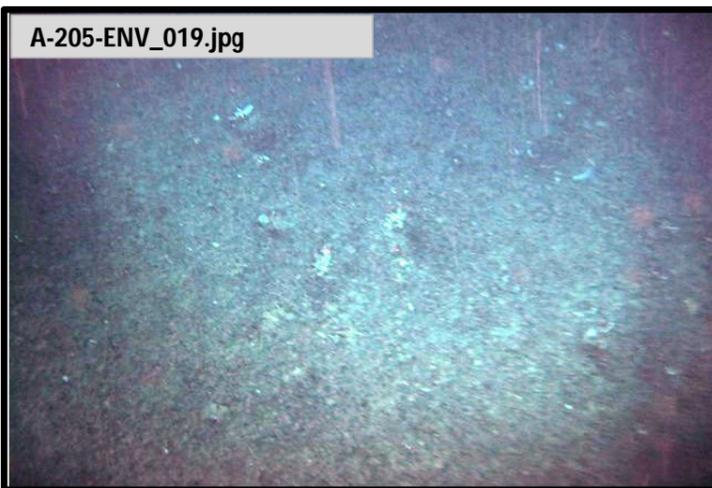


Photo Position: 792218 mE, 4178377 mN

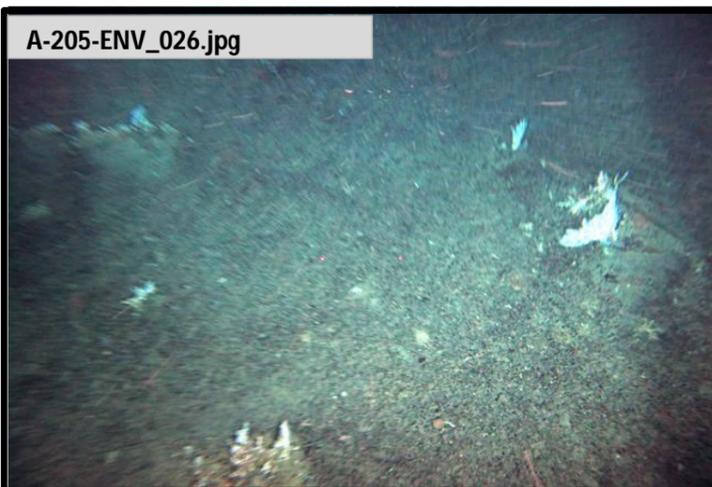


Photo Position: 792218 mE, 4178377 mN

Habitat Summary Information: A/205/ENV

Survey Area: FISA

No. of Stills: 12 Mins of Video: 40 Track Length: 198m

| | |
|--|---|
| <p>Site Selection Criteria Discrete slope failure</p> | <p>Analogue Interpretation Sloped seabed</p> |
| <p>Sediment Description Very large boulder, coarse sediment, rocky outcrops</p> | |
| <p>Conspicuous Fauna <i>Lophelia pertusa</i>, Relic <i>Lophelia</i> sp., Sponge: Porifera sp., Branched Stone Coral: <i>Stylaster</i> sp., Sea Pen: Pennatulacea,</p> | |

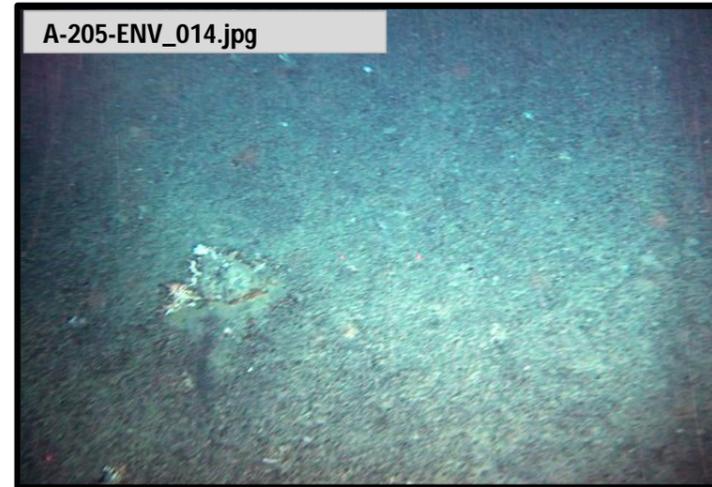


Photo Position: 792218 mE, 4178377 mN

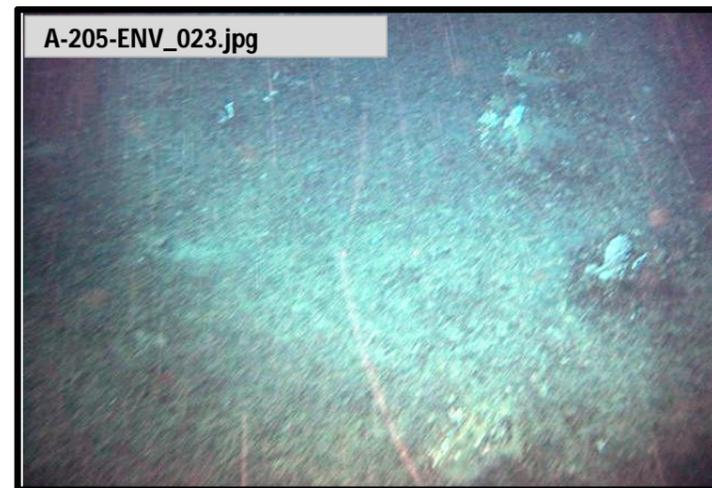


Photo Position: 792218 mE, 4178377 mN

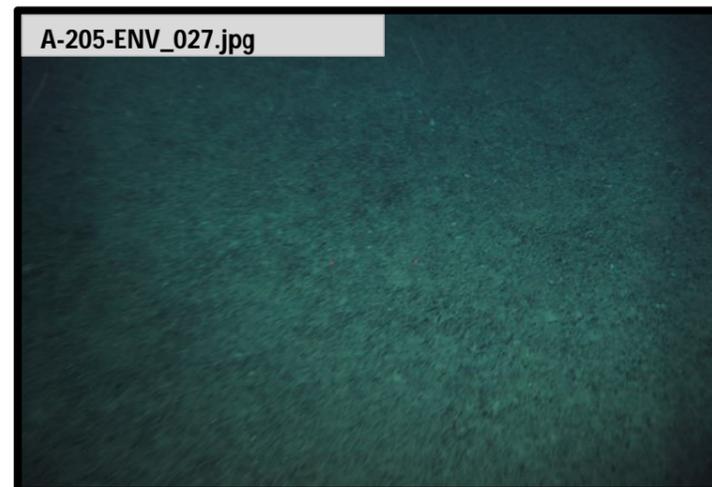
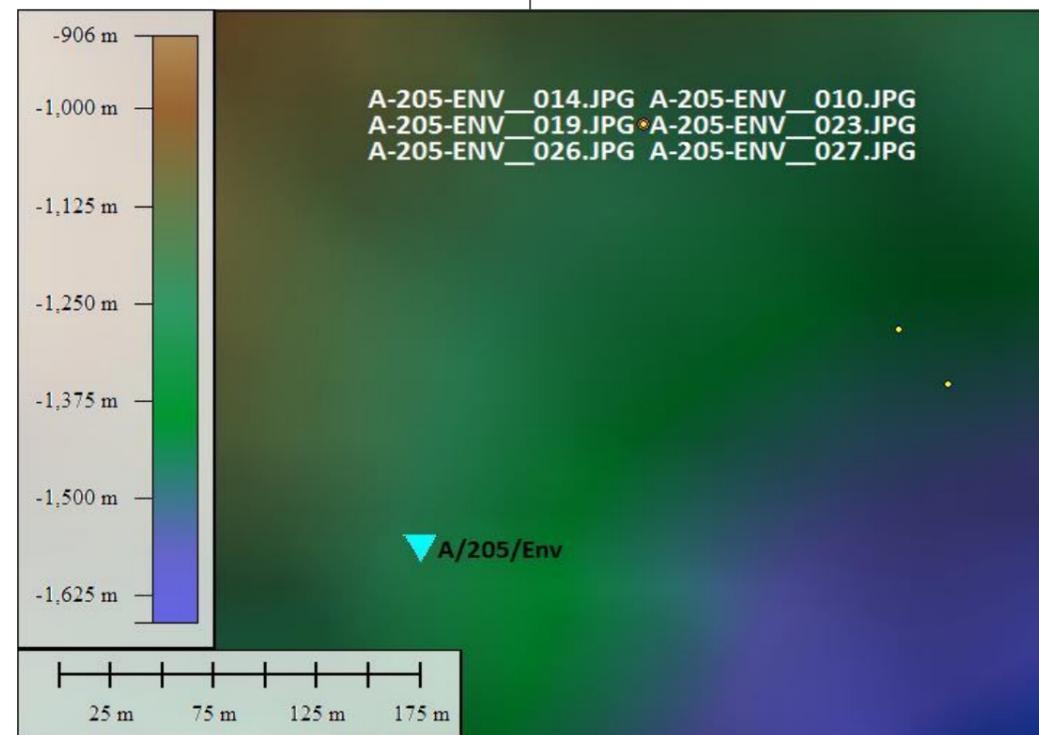


Photo Position: 792218 mE, 4178377 mN



| | |
|---------------------------|---|
| <p>Client </p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
|---------------------------|---|

✕ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

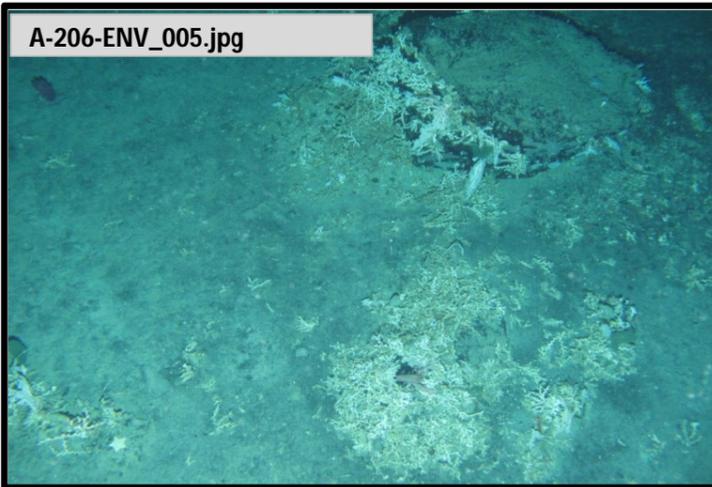


Photo Position: 776850 mE, 4163322 mN

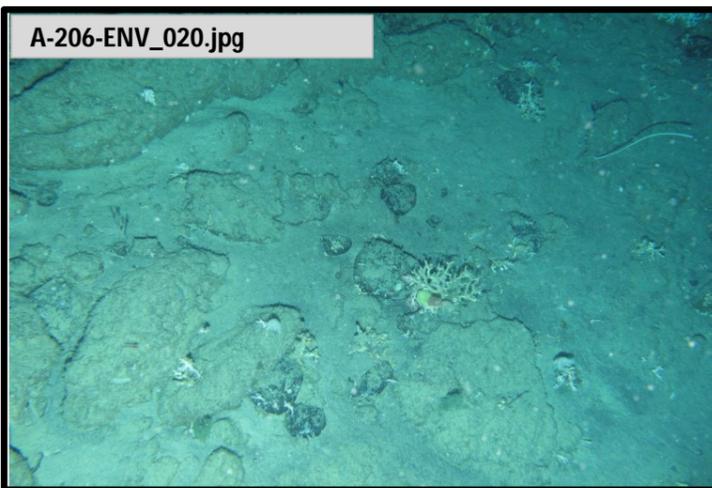


Photo Position: 776867 mE, 4163426 mN

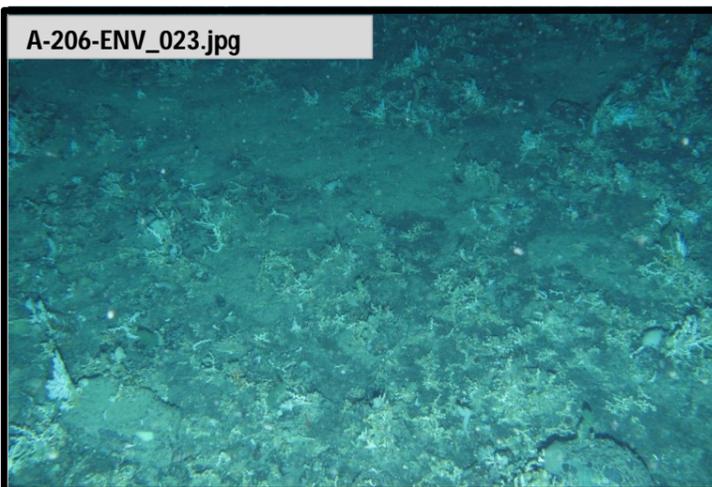


Photo Position: 776857 mE, 4163484 mN

Habitat Summary Information: A/206/ENV

Survey Area: FISA

No. of Stills: 29 Mins of Video: 32 Track Length: 690m

| | |
|---|---|
| <p>Site Selection Criteria Discrete feature</p> <p>Sediment Description Upslope, rocky outcrops</p> <p>Conspicuous Fauna Relic <i>Lophelia</i> sp., Branched Stone Coral: <i>Stylaster</i> sp., Sea Lily: Crinoidea sp., Gorgonian sp., Hydroid sp., Decapod Shrimp sp., Nephropidae sp. <i>Thymops birsteini</i>, <i>Lophelia pertusa</i>, Sea Pen: Pennatulacea, Sponge: Porifera sp.,</p> | <p>Analogue Interpretation Slope feature</p> |
|---|---|

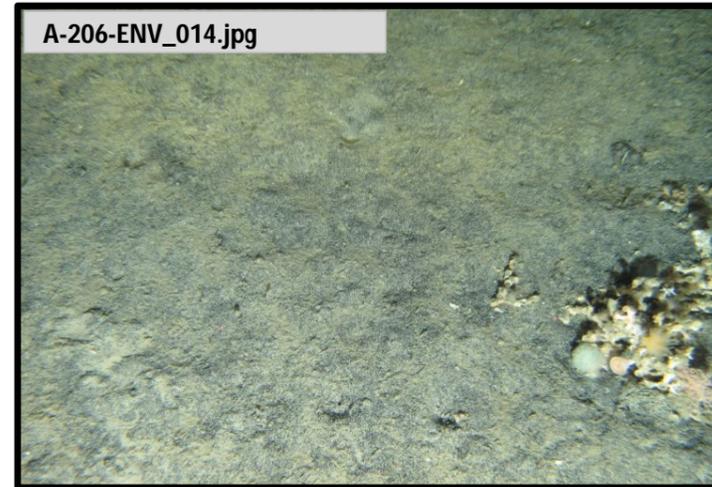


Photo Position: 776850 mE, 4163322 mN

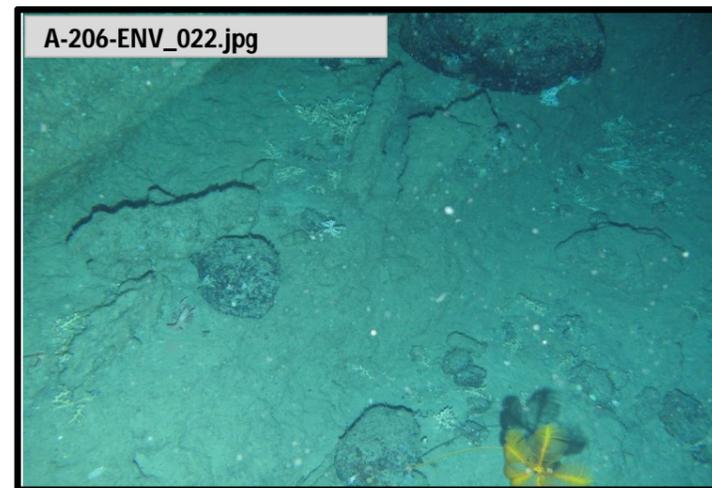


Photo Position: 776689 mE, 4163450 mN

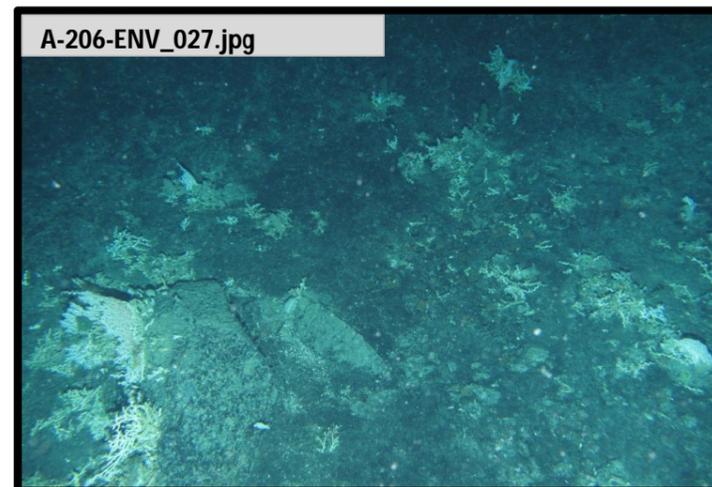
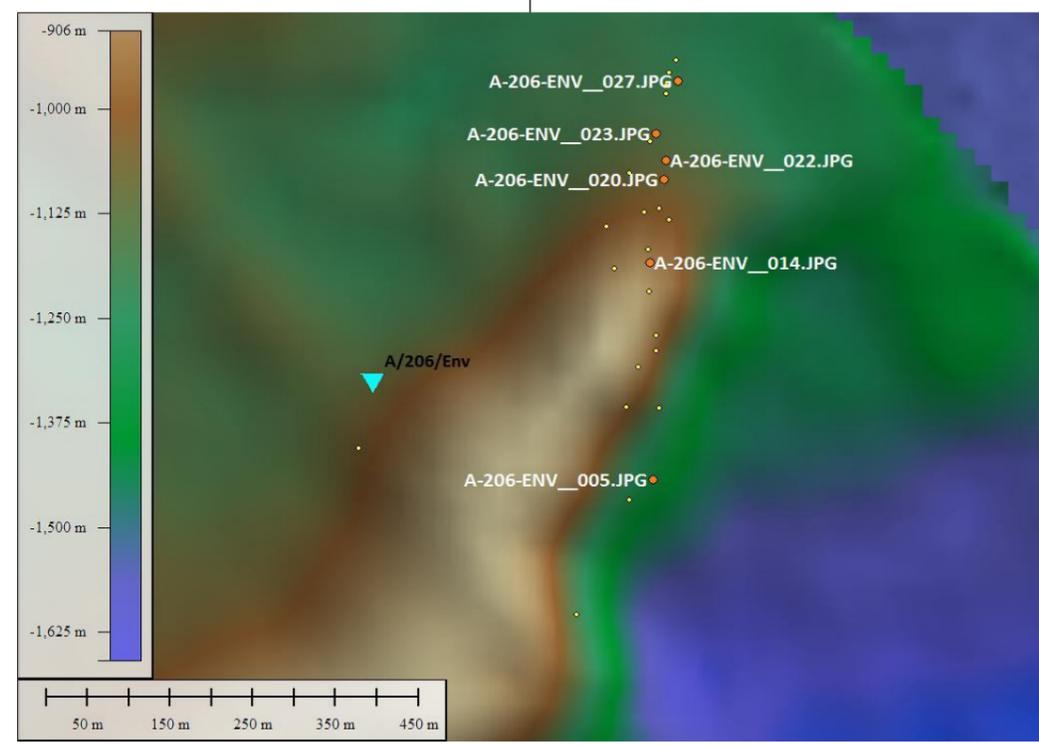


Photo Position: 776884 mE, 4163550 mN



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|----------------------|---|
| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
|----------------------|---|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

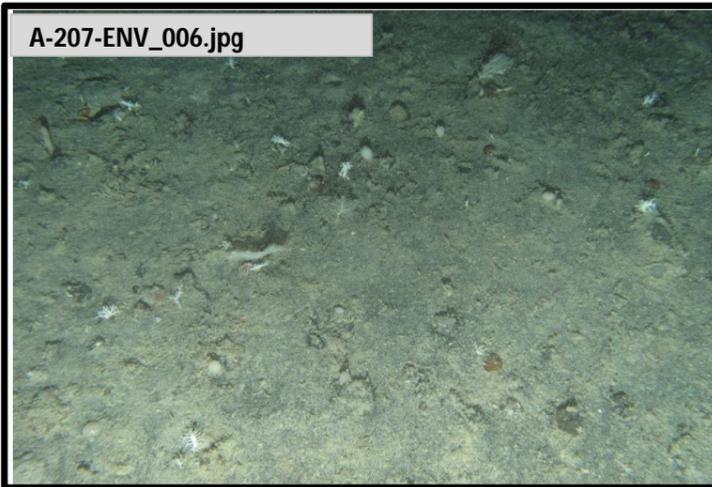


Photo Position: 786682 mE, 4209681 mN

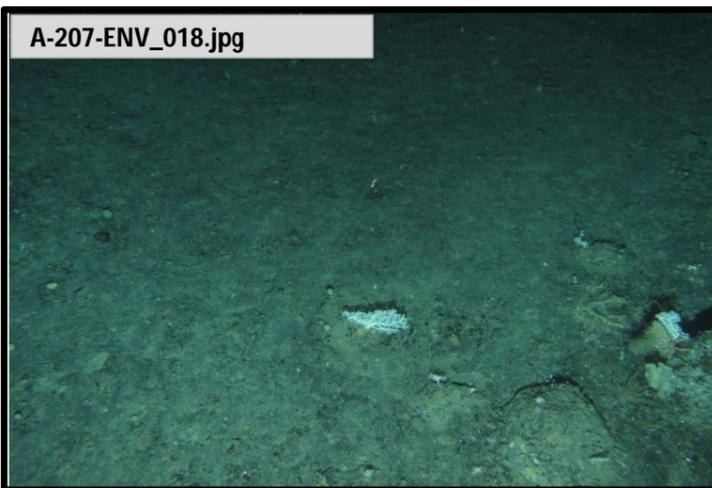


Photo Position: 786670 mE, 4209686 mN

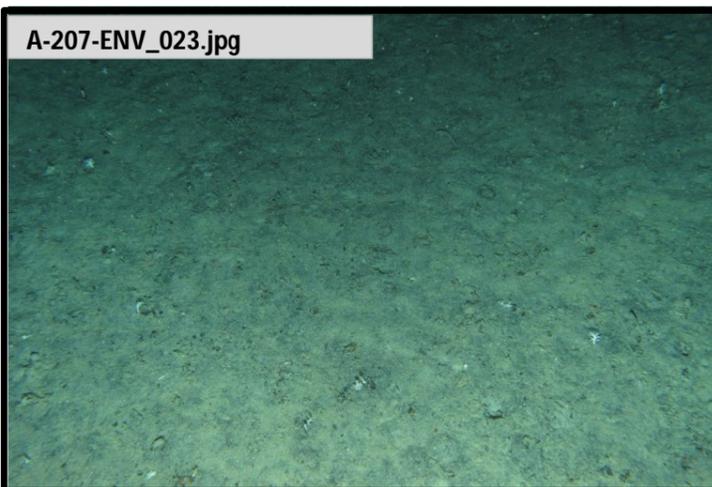


Photo Position: 786606 mE, 42097441 mN

Habitat Summary Information: A/207/ENV

Survey Area: FISA

No. of Stills: 35 Mins of Video: 27 Track Length: 154m

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|---|---|
| <p>Site Selection Criteria Discrete 'reef' area</p> <p>Sediment Description Homogeneous beige sand, gravel and boulders</p> <p>Conspicuous Fauna Branched Stone Coral: <i>Stylaster</i> sp., Squat Lobster: Galatheidae sp., Branched Bryozoa sp., Sponge: possibly <i>Suberites</i> sp., Sea Pen: Pennatulacea, Gorgonian sp., Hydroid sp., Isopod: possibly <i>Acutiserolis neaera</i>, Bryozoa sp.,</p> | <p>Analogue Interpretation Sloped seabed</p> |
|---|---|

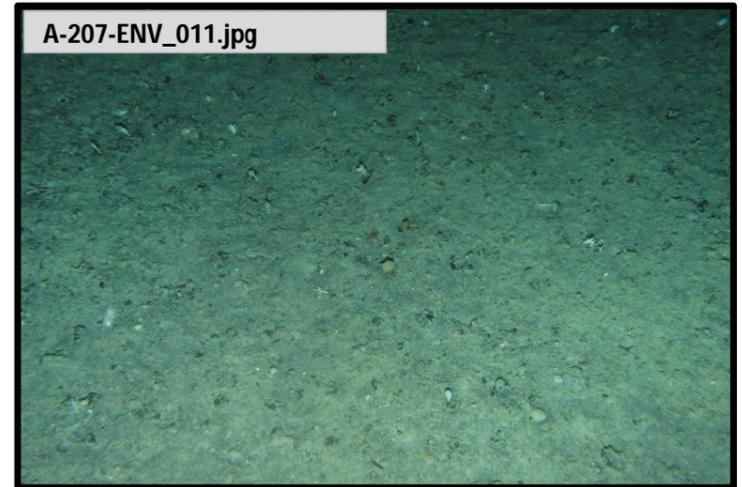


Photo Position: 786682 mE, 4209681 mN

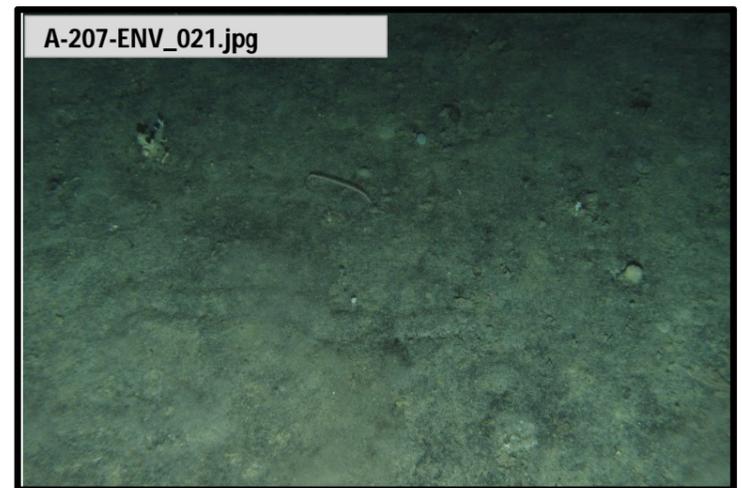


Photo Position: 786606 mE, 4209741 mN

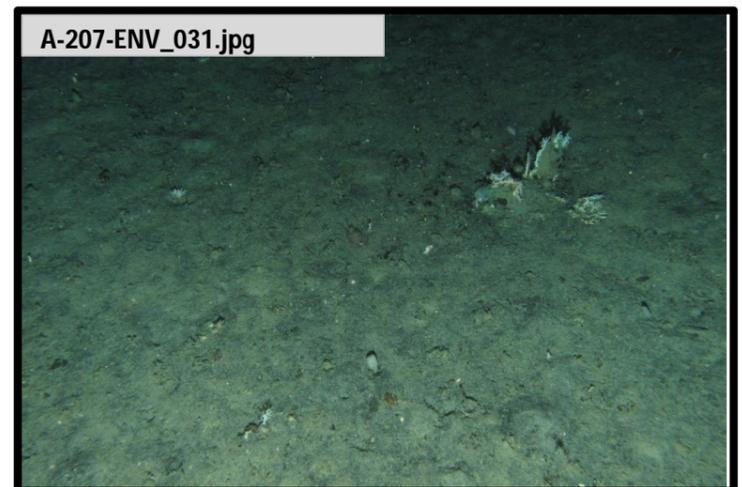
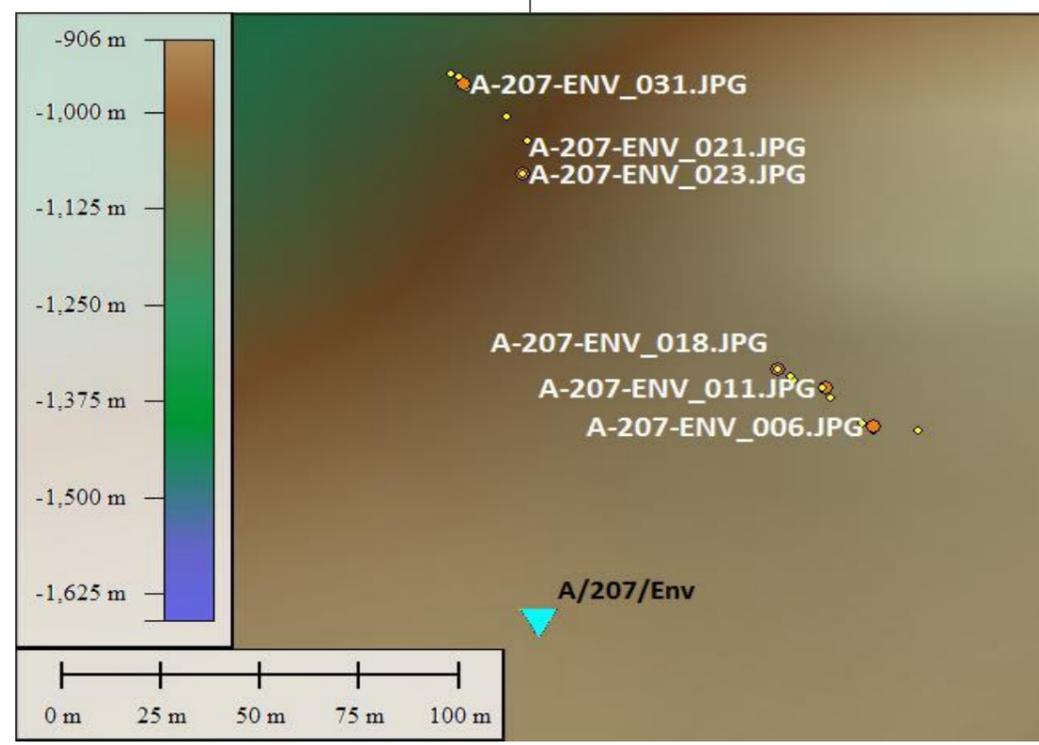


Photo Position: 786591 mE, 4209766 mN



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| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
|----------------------|---|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

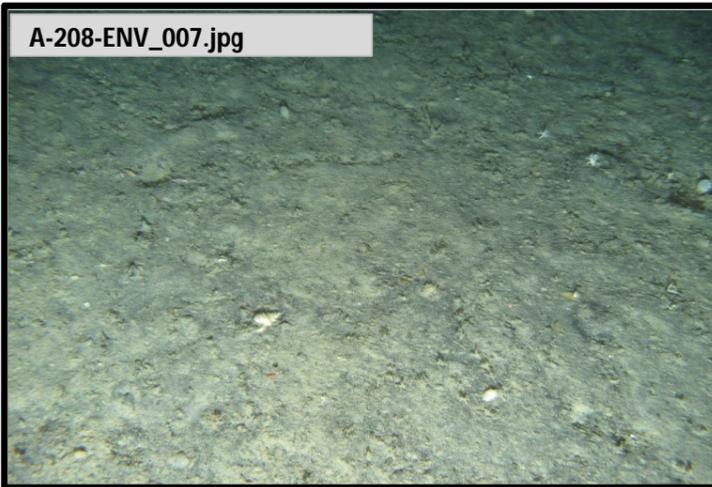


Photo Position: 773671 mE, 4228281 mN

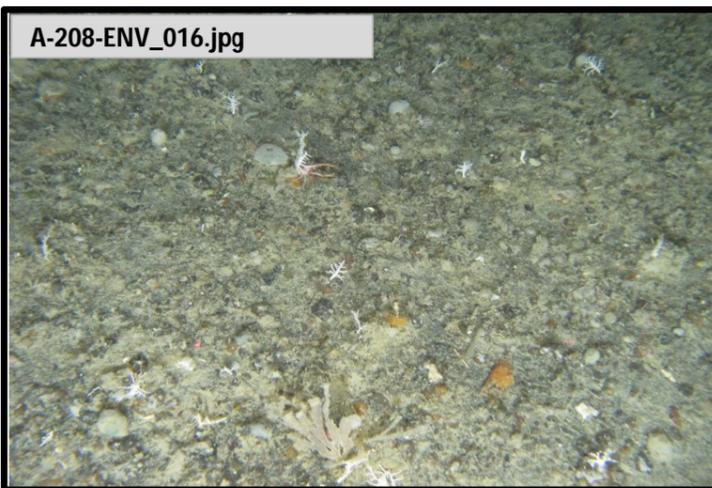


Photo Position: 773698 mE, 4228315 mN

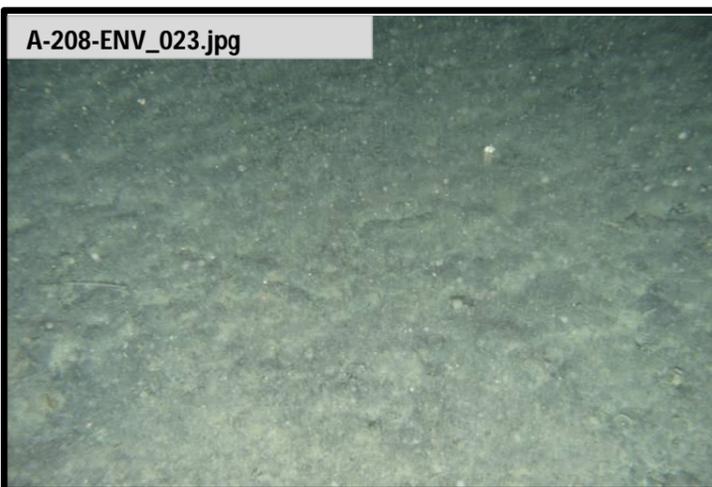


Photo Position: 773729 mE, 4228356 mN

Habitat Summary Information: A/208/ENV

Survey Area: FISA

No. of Stills: 25 Mins of Video: 40 Track Length: 124m

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| <p>Site Selection Criteria Geographical coverage</p> | <p>Analogue Interpretation Slightly sloped seabed</p> |
| <p>Sediment Description Homogeneous beige sand with gravel, occasional cobbles</p> | |
| <p>Conspicuous Fauna Branched Stone Coral: <i>Stylaster</i> sp., Isopod: possibly <i>Acutiserolis neaera</i>, Sponge: Porifera sp., Holothuroidea sp., Bryozoa sp., Squat Lobster: Galatheidae sp., Starfish: Asteroidea sp., Cup Coral: <i>Flabellum</i> sp., Sea Pen: Pennatulacea,</p> | |

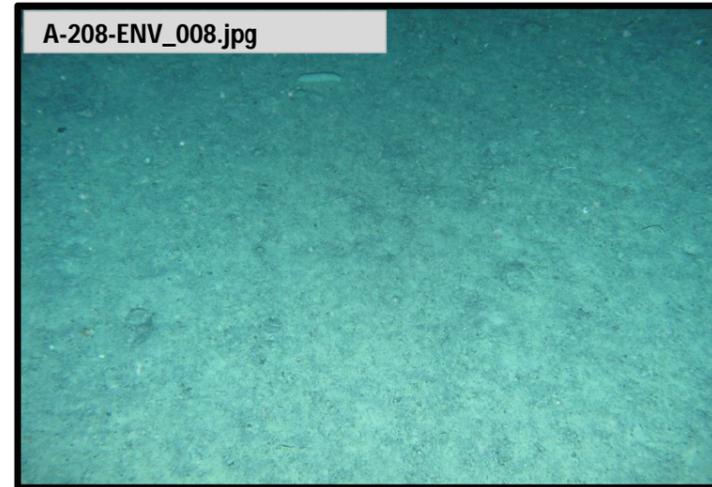


Photo Position: 773671 mE, 4228281 mN

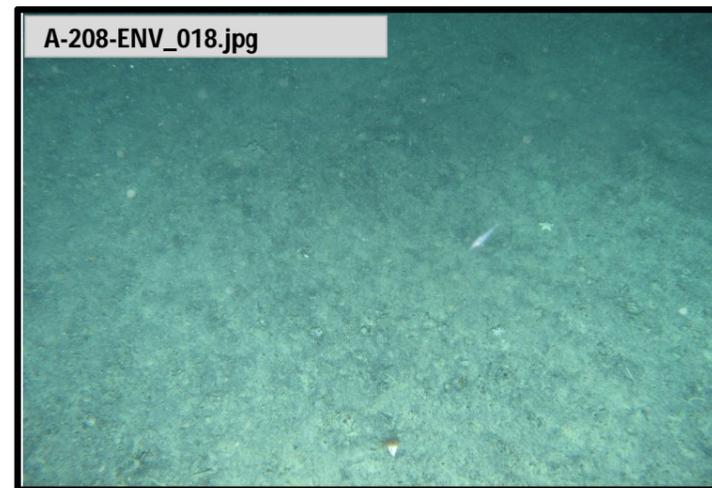


Photo Position: 773705 mE, 4228325 mN

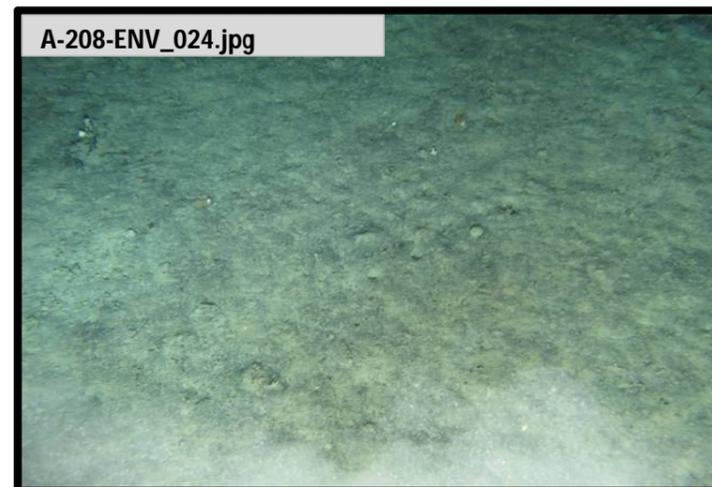
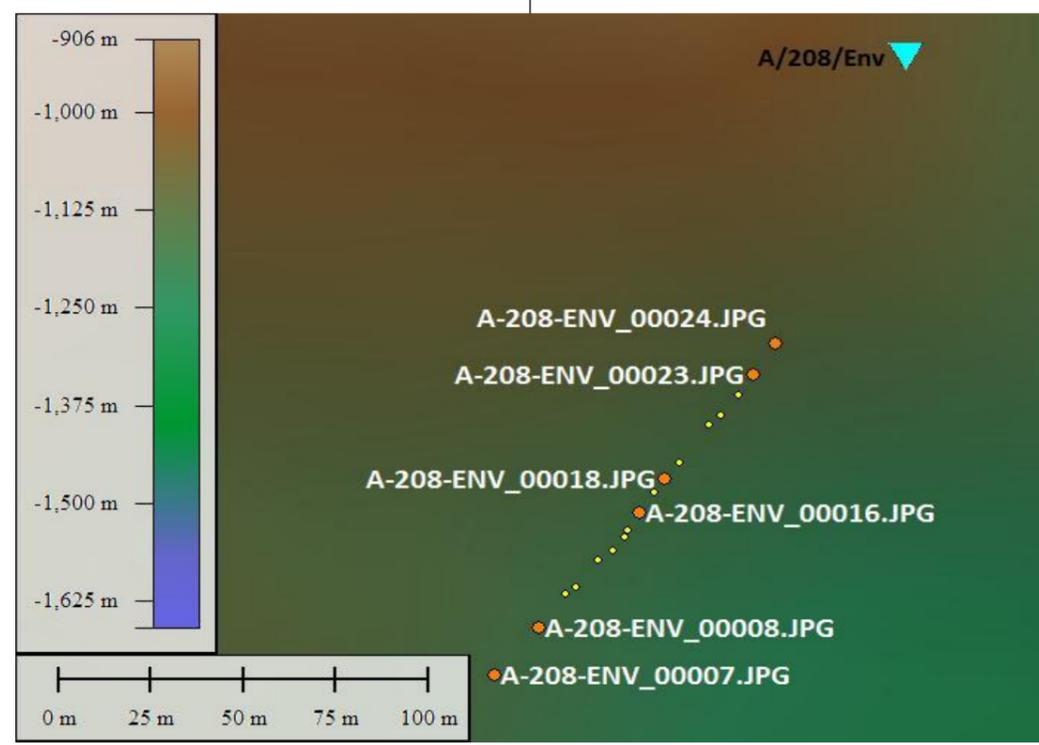


Photo Position: 773735 mE, 4228365 mN



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| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
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✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

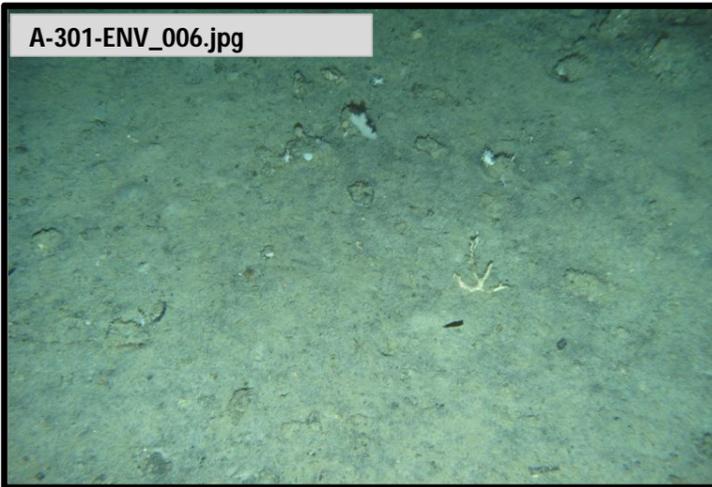


Photo Position: 790831 mE, 4215005 mN

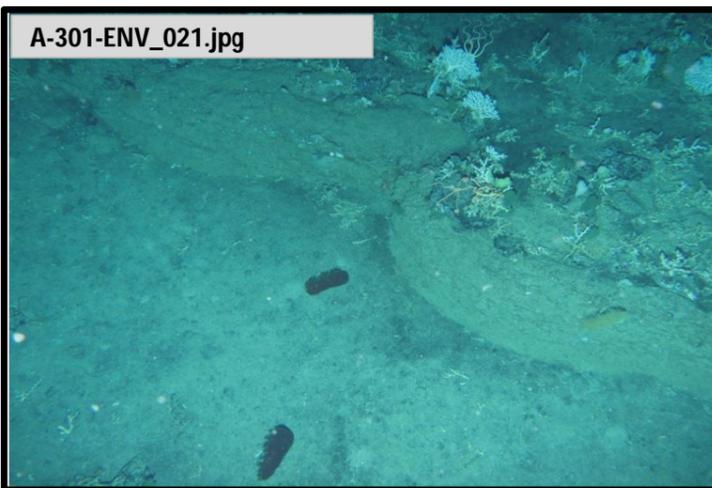


Photo Position: 790834 mE, 4214991 mN



Sediment Example Image

Habitat Summary Information: A/301/ENV

Survey Area: FISA

No. of Stills: 35 Mins of Video: 29 Track Length: 56m

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| <p>Site Selection Criteria Slope feature</p> | <p>Analogue Interpretation Slope feature</p> |
|---|---|

Sediment Description
Paving slabs, homogeneous beige sand and cobbles with evidence of bioturbation

Conspicuous Fauna
Lophelia pertusa, Relic *Lophelia* sp., Sea Pen: Pennatulacea, Cup Coral: *Flabellum* sp., Gorgonian sp., Hydroid sp., Brittlestar (Ophiuroid), Sponge: Porifera sp., Moridae sp., Bryozoa sp., Branched Bryozoa sp., Isopod: possibly *Acutiserolis neaera*, Squat Lobster: Galatheidae sp., Encrusting Sponge, Glass Sponge: Hexactinellida sp., Gastropoda sp., Branched Stone Coral: *Stylaster* sp.,

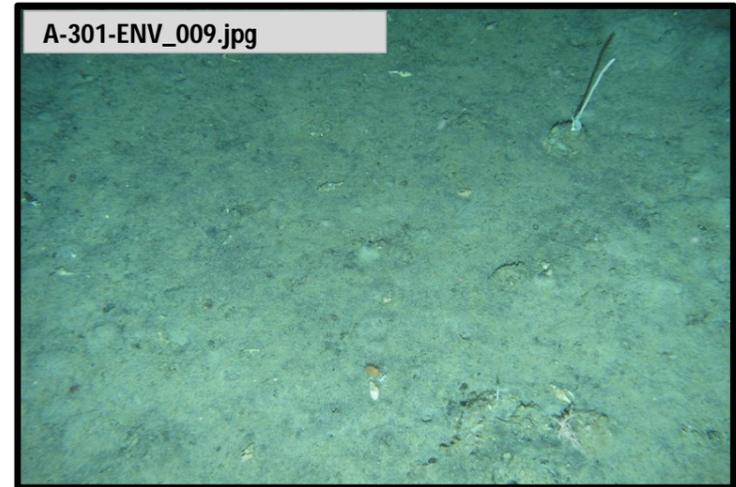


Photo Position: 790831 mE, 4215005 mN

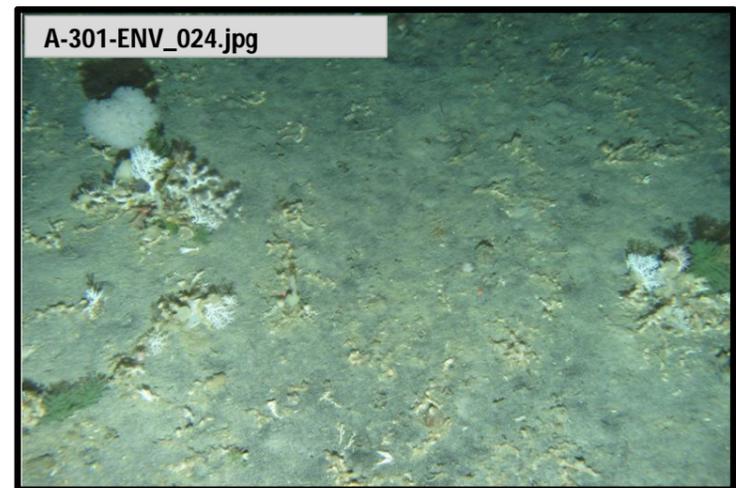
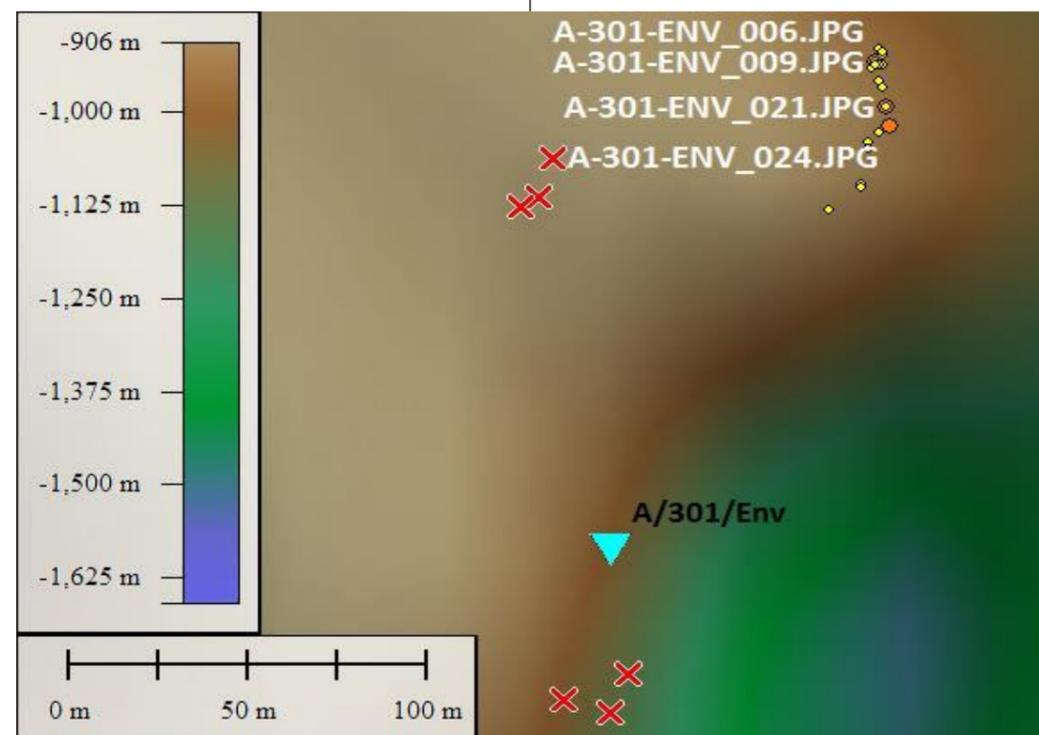


Photo Position: 790835 mE, 4214985 mN



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| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
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✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

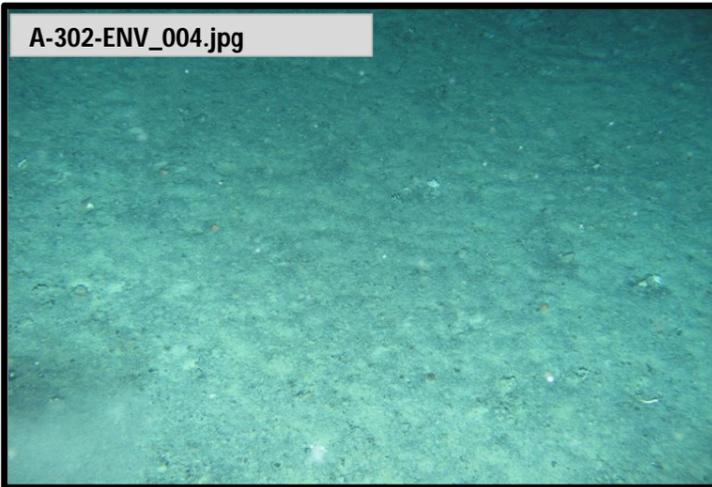


Photo Position: 804643 mE, 4210660 mN



Photo Position: 804631 mE, 4210621 mN



Sediment Example Image

Habitat Summary Information: A/302/ENV

Survey Area: FISA

No. of Stills: 24 Mins of Video: 22 Track Length: 92m

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| <p>Site Selection Criteria Regional infill station</p> | <p>Analogue Interpretation Slightly sloped seabed</p> |
| <p>Sediment Description Beige and black sand with gravels, occasional cobbles and evidence of bioturbation</p> | |
| <p>Conspicuous Fauna Cup Coral: <i>Flabellum</i> sp., Branched Stone Coral: <i>Stylaster</i> sp., Sea Pen: Pennatulacea, Stone Crab: <i>Paralomis</i> sp., Sponge: Porifera sp., Hydroid sp., Nephropidae sp. <i>Thymops birsteini</i>, Glass Sponge: Hexactinellida sp., Bryozoa sp.,</p> | |

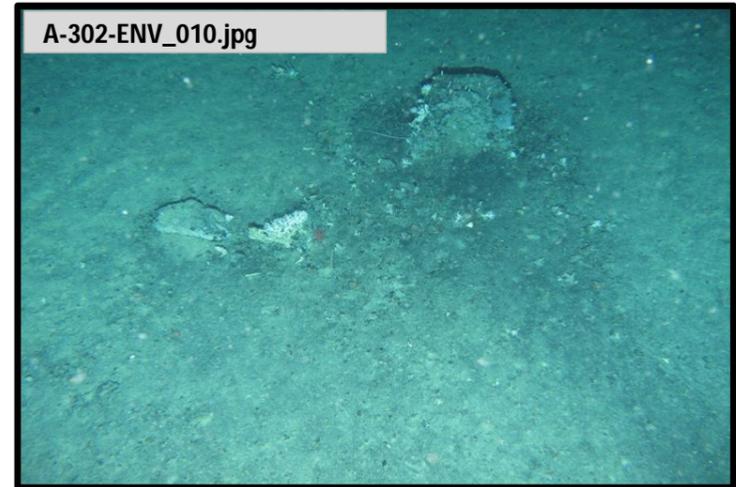


Photo Position: 804647 mE, 4210645 mN

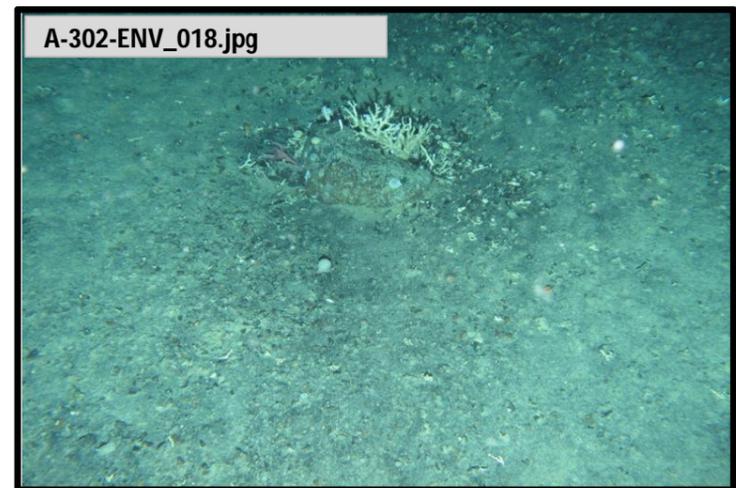
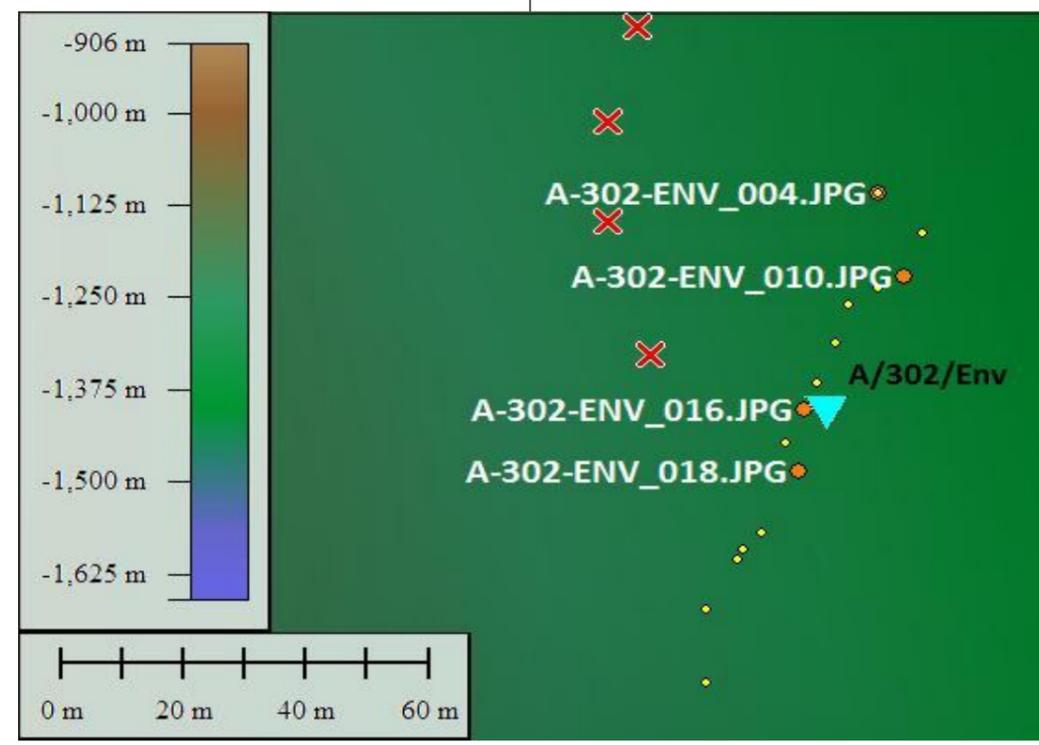


Photo Position: 804630 mE, 4210610 mN



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| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
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✕ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

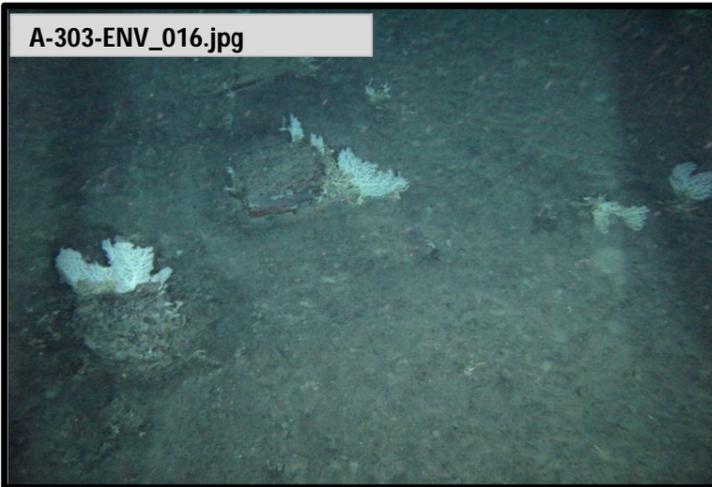


Photo Position: 796078 mE, 4211863 mN



Photo Position: 796089 mE, 4211851 mN



Sediment Example Image

Habitat Summary Information: A/303/ENV

Survey Area: FISA

No. of Stills: 31 Mins of Video: 34 Track Length: 57m

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| <p>Site Selection Criteria Slope feature</p> | <p>Analogue Interpretation Slightly sloped seabed</p> |
| <p>Sediment Description Darker, coarse surface layer over beige fine to medium compact sands</p> | |
| <p>Conspicuous Fauna Cup Coral: <i>Flabellum</i> sp., Branched Stone Coral: <i>Stylaster</i> sp., Sea Pen: Pennatulacea, Relic <i>Lophelia</i> sp., <i>Lophelia pertusa</i>, Glass Sponge: Hexactinellida sp., Brittlestar (Ophiuroid), Sponge: possibly <i>Suberites</i> sp., Bryozoa sp., Tube-building Polychaete, Polychaete sp., Gastropoda sp.,</p> | |

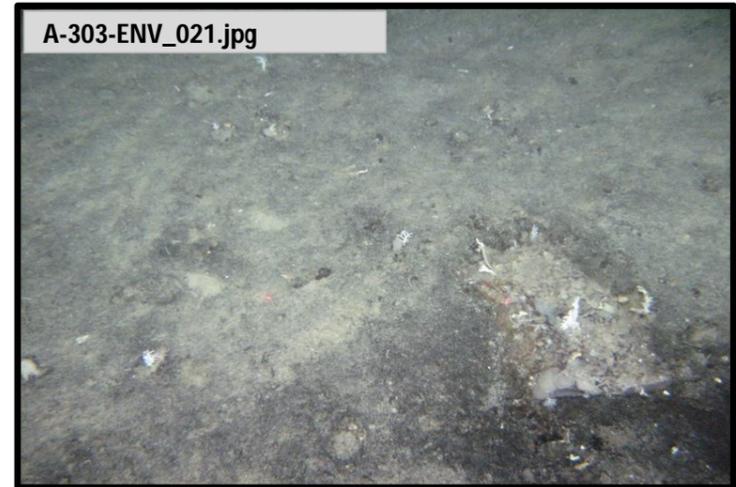
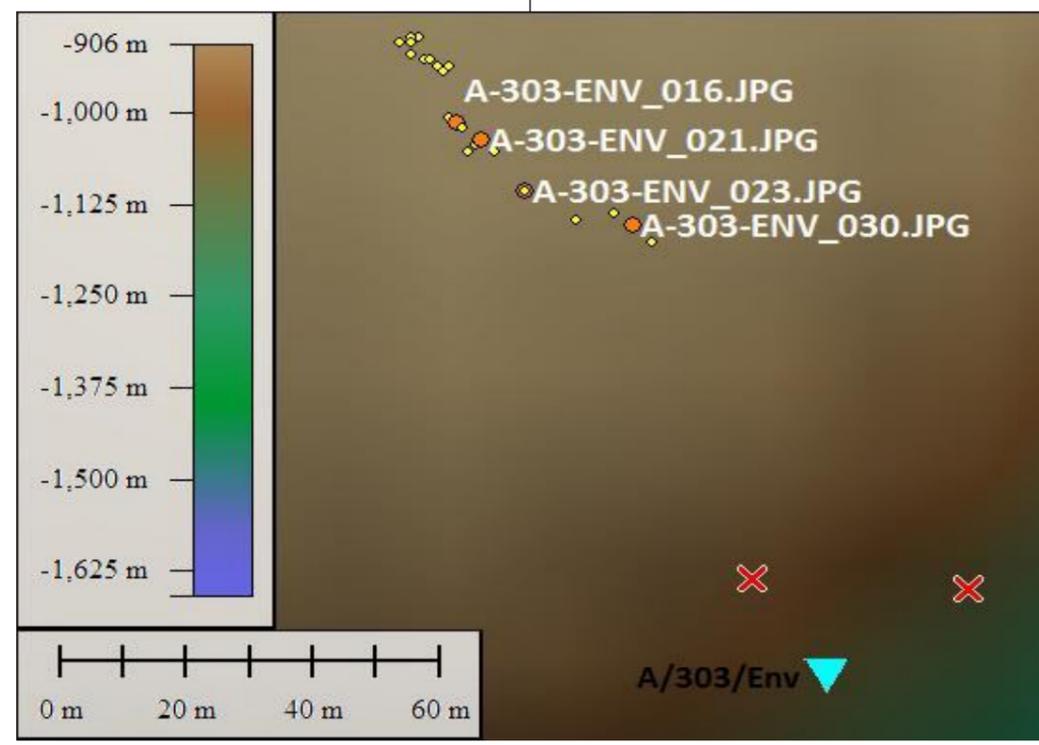


Photo Position: 796082 mE, 4211860 mN



Photo Position: 796106 mE, 4211845 mN



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| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
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✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

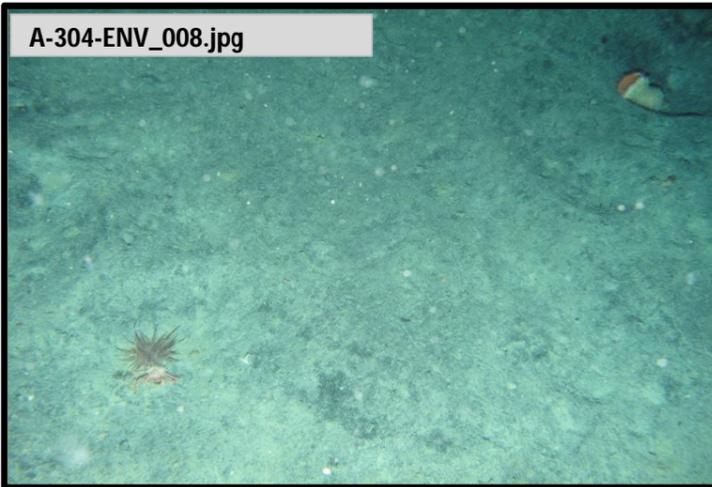


Photo Position: 783332 mE, 4221139 mN

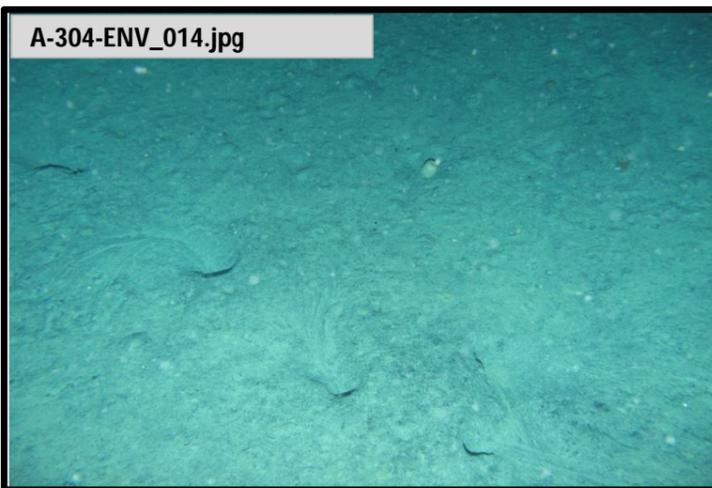


Photo Position: 783301 mE, 4221165 mN



Sediment Example Image

Habitat Summary Information: A/304/ENV

Survey Area: FISA

No. of Stills: 18 Mins of Video: 21 Track Length: 78m

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| <p>Site Selection Criteria Regional infill station</p> | <p>Analogue Interpretation Relatively flat seabed</p> |
| <p>Sediment Description Beige with dark flecks, fine to medium sand with occasional fine gravel</p> | |
| <p>Conspicuous Fauna Stone Crab: <i>Paralomis</i> sp., Branched Stone Coral: <i>Stylaster</i> sp., Sea Pen: Pennatulacea, Relic <i>Lophelia</i> sp., <i>Lophelia pertusa</i>, Glass Sponge: Hexactinellida sp., Venus Fly-trap Anemone: <i>Actinoscyphia</i> sp., Anemone: Actiniaria sp., Nephropidae sp. <i>Thymops birsteini</i>, Cup Coral: <i>Flabellum</i> sp., Decapod Shrimp sp., Tube-building Polychaete, Polychaete sp., Brittlestar (Ophiuroid), Sea Spider: Pycnogonida sp., Tusk Shell: Scaphopoda sp.,</p> | |

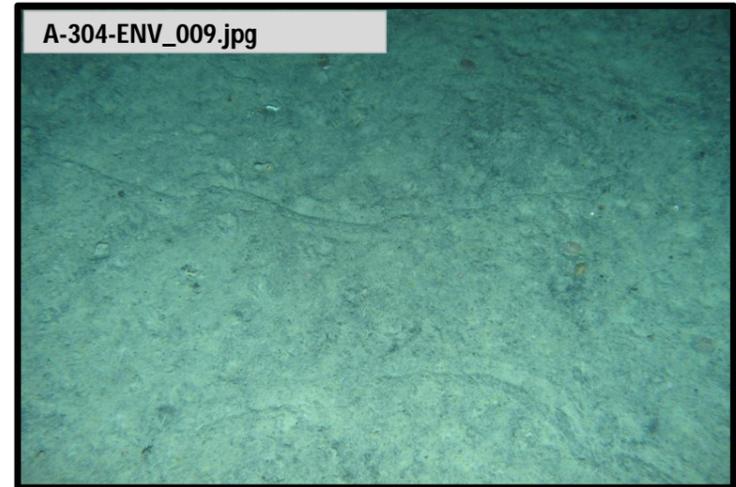


Photo Position: 783330 mE, 4221146 mN

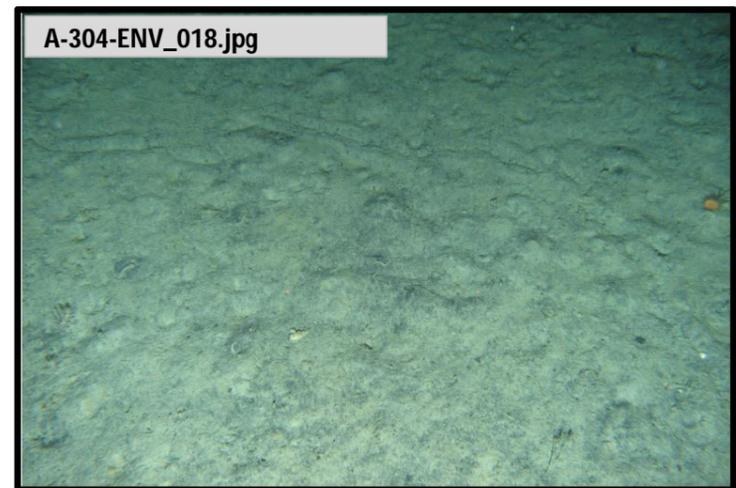
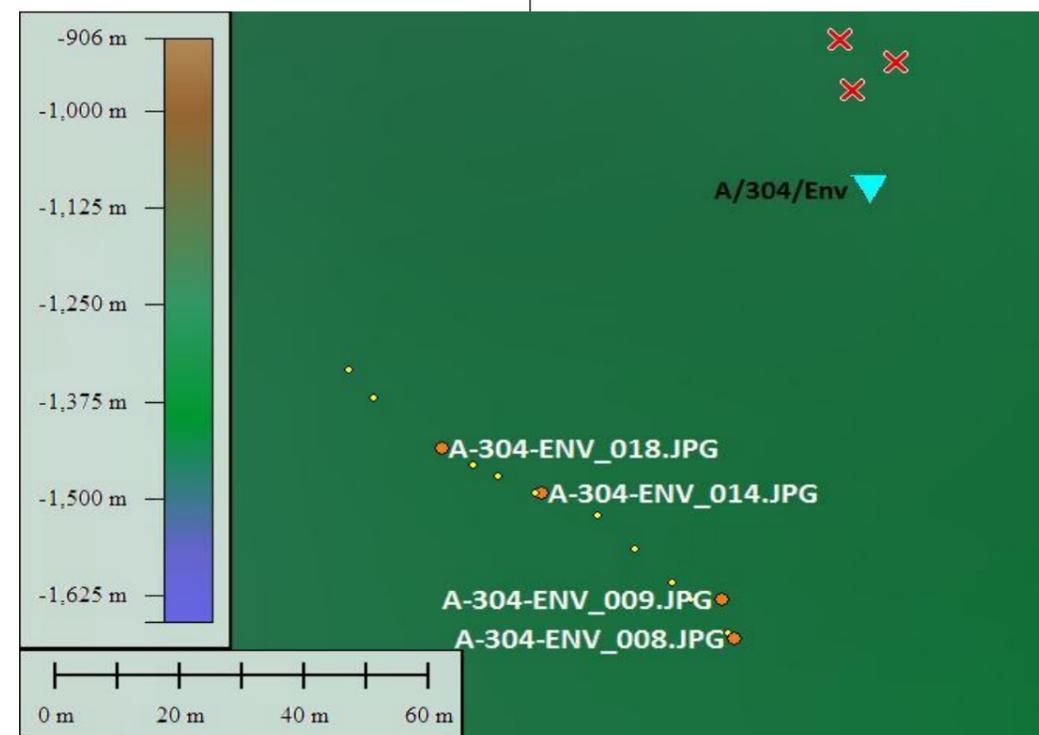
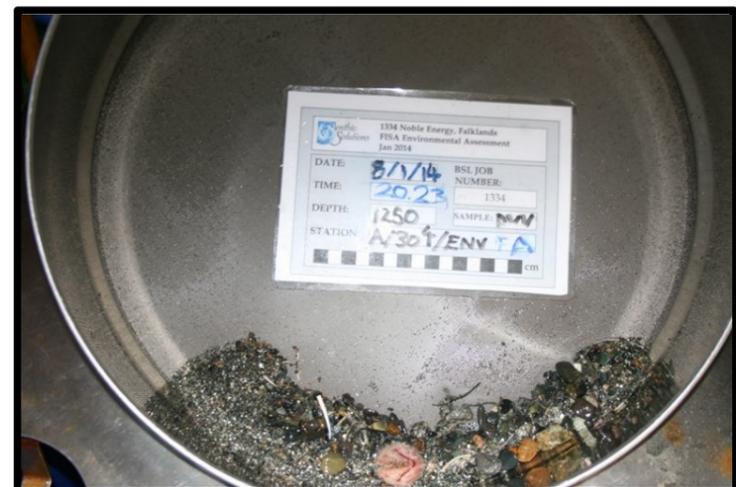


Photo Position: 783285 mE, 4221173 mN



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| <p>Client noble energy</p> | <p>Contractor Benthic Solutions Benthic Solutions Ltd., Marsh Road,</p> |
|--|---|

✕ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

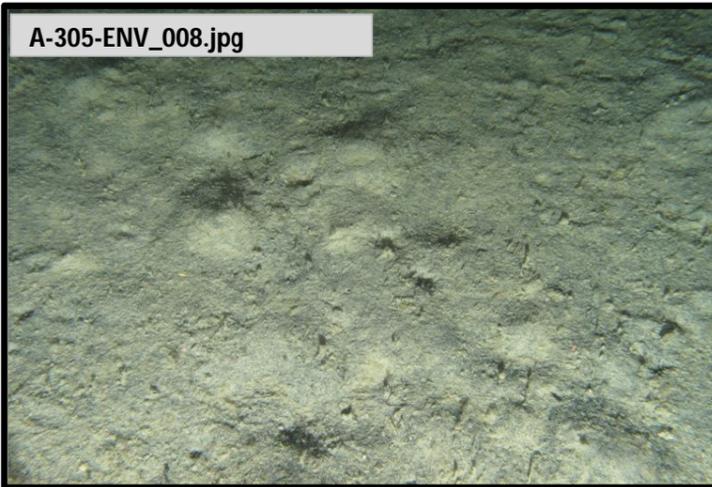


Photo Position: 778086 mE, 4210837 mN

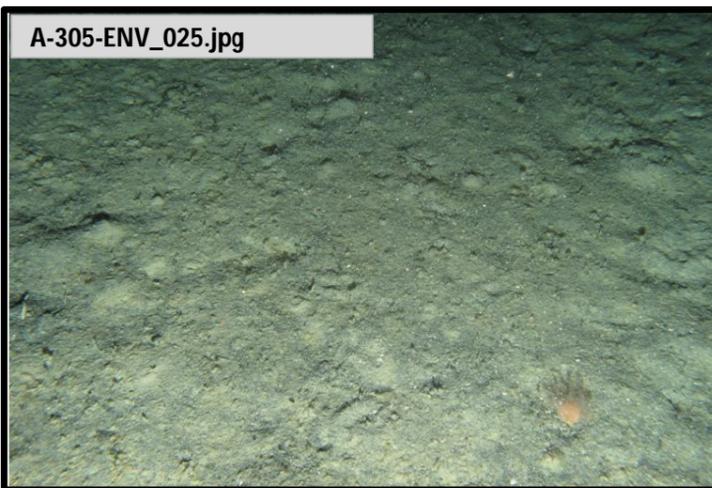


Photo Position: 777955 mE, 4210965 mN



Sediment Example Image

Habitat Summary Information: A/305/ENV

Survey Area: FISA

No. of Stills: 19 Mins of Video: 32 Track Length: 375m

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| <p>Site Selection Criteria Regional infill station</p> | <p>Analogue Interpretation Slightly sloped seabed</p> |
| <p>Sediment Description Darker, coarse surface layer over beige fine to medium compact sands</p> | |
| <p>Conspicuous Fauna Isopod: possibly <i>Acutiserolis neaera</i>, Cup Coral: <i>Flabellum</i> sp., Decapod Shrimp sp., Sea Pen: Pennatulacea, Tube-building Polychaete, Polychaete sp.,</p> | |

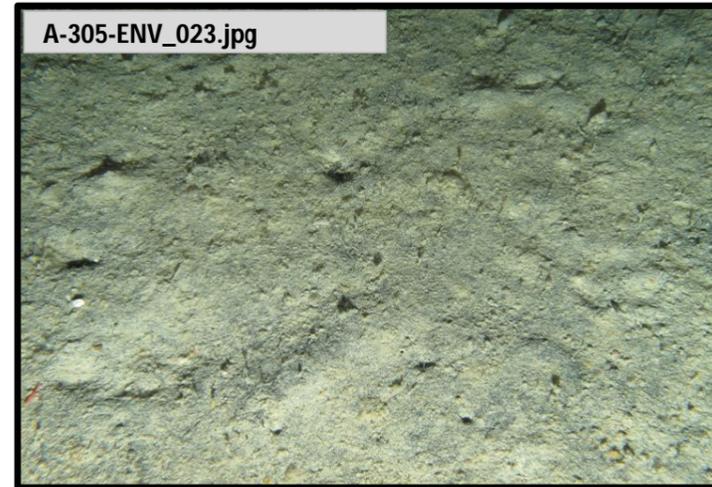


Photo Position: 777972 mE, 4210943 mN

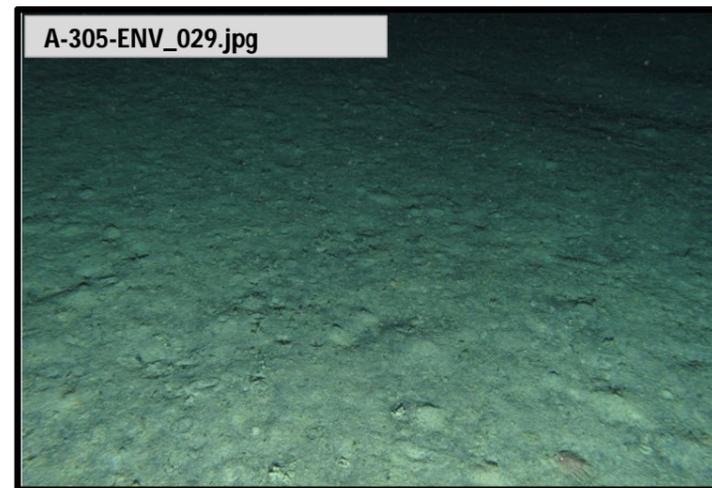
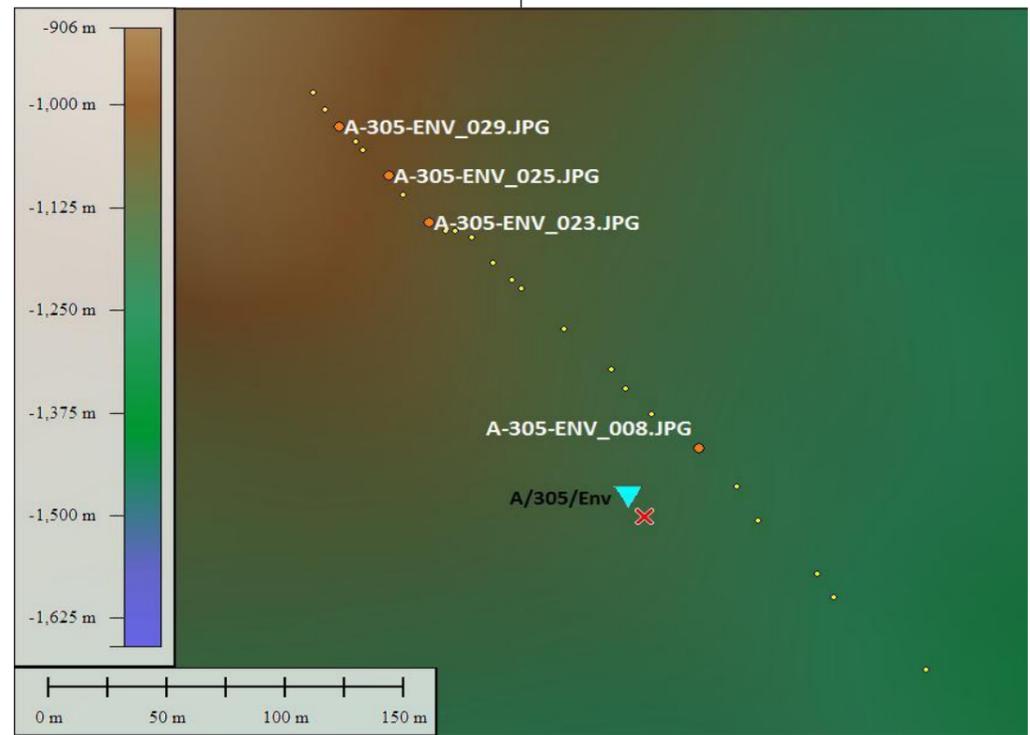


Photo Position: 777934 mE, 4210988 mN



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| <p>Client </p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
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✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

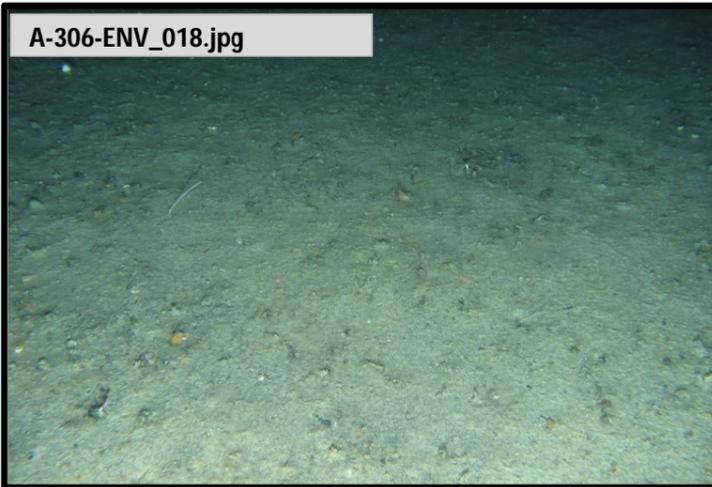


Photo Position: 796857 mE, 4206311 mN



Photo Position: 796803 mE, 4206390 mN



Sediment Example Image

Habitat Summary Information: A/306/ENV

Survey Area: FISA

No. of Stills: 32 Mins of Video: 33 Track Length: 158m

| | |
|---|--|
| <p>Site Selection Criteria Trench feature</p> | <p>Analogue Interpretation Base of trench feature</p> |
| <p>Sediment Description Coarse pebbles and rocks suspended in clay</p> | |
| <p>Conspicuous Fauna Cup Coral: <i>Flabellum</i> sp., Sea Pen: Pennatulacea, Hake: possibly <i>Merluccius hubbsi</i>, Sponge: Porifera sp., Tube-building Polychaete, Polychaete sp., Decapod Shrimp sp., Relic <i>Lophelia</i> sp., Bryozoa sp., Branched Stone Coral: <i>Stylaster</i> sp., Brittlestar (Ophiuroid),</p> | |

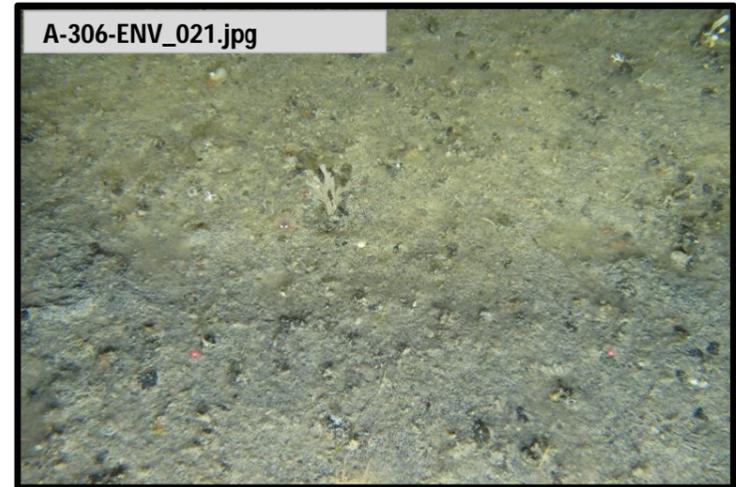


Photo Position: 796857 mE, 4206311 mN

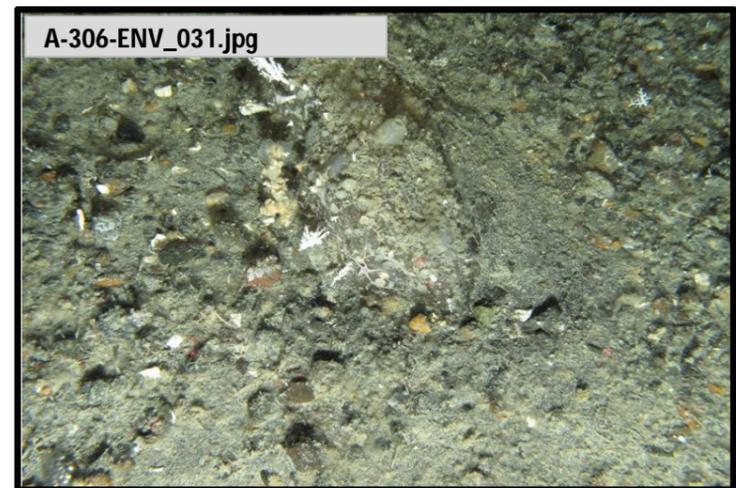
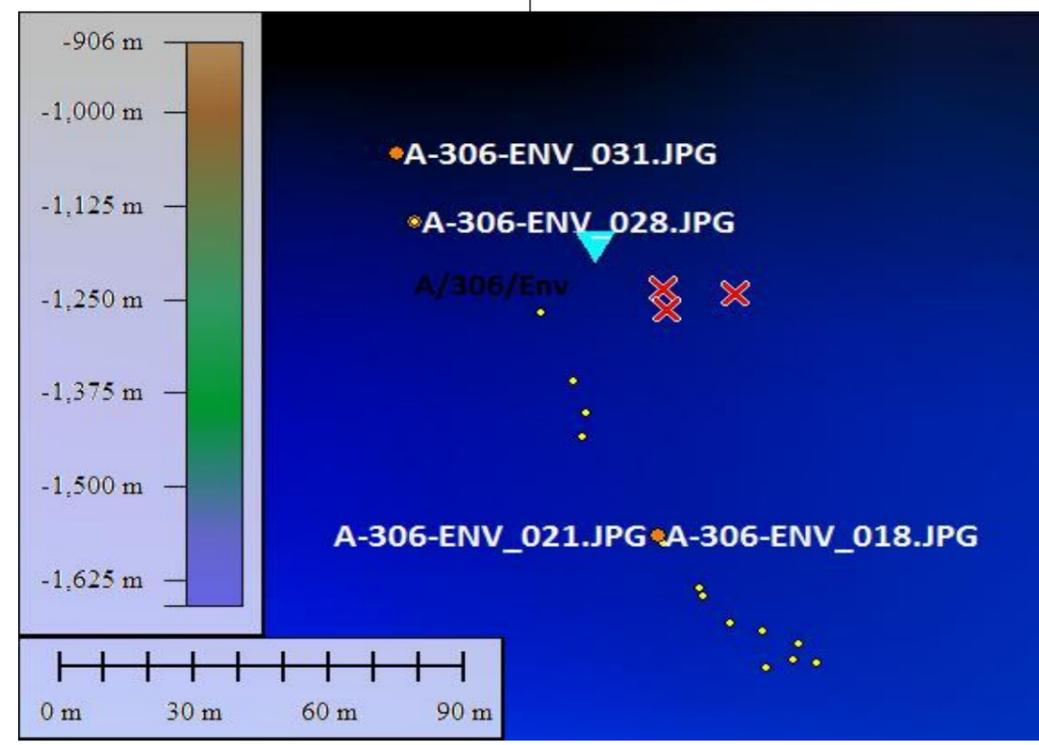


Photo Position: 796799 mE, 4206407 mN



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| <p>Client noble energy</p> | <p>Contractor Benthic Solutions Benthic Solutions Ltd., Marsh Road,</p> |
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✕ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

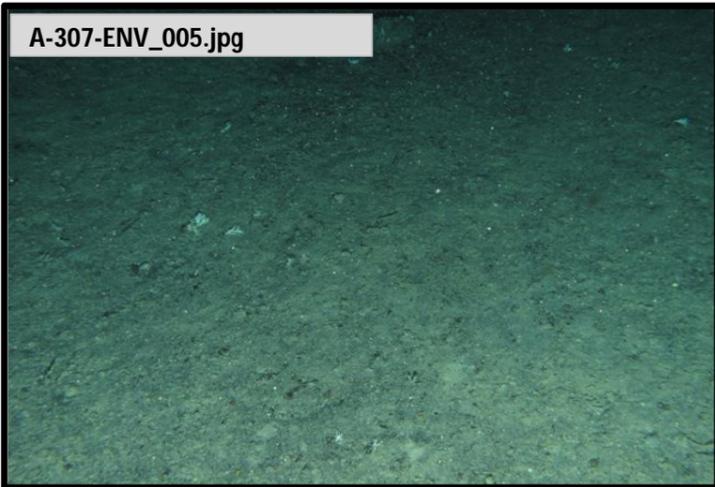


Photo Position: 793337 mE, 4188006 mN

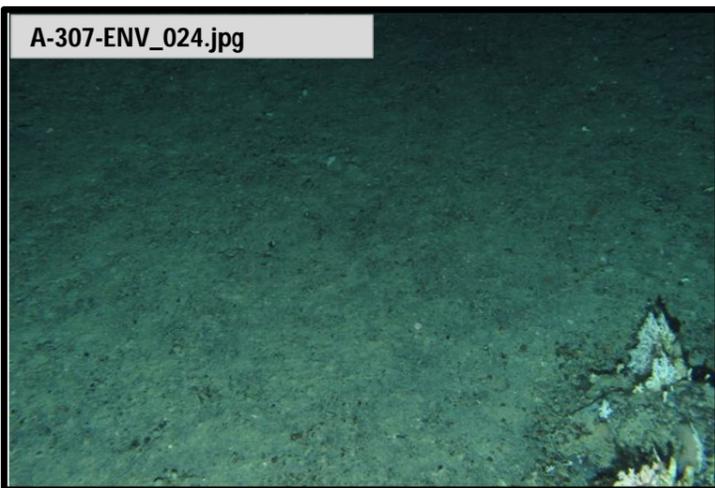


Photo Position: 793246 mE, 4188021 mN



Sediment Example Image

Habitat Summary Information: A/307/ENV

Survey Area: FISA

No. of Stills: 30 Mins of Video: 30 Track Length: 142m

| | |
|--|--|
| <p>Site Selection Criteria Regional infill station in deepest area</p> | <p>Analogue Interpretation Relatively flat seabed</p> |
| <p>Sediment Description Coarse sand, pebbles and rocks suspended in clay with evidence of bioturbation</p> | |
| <p>Conspicuous Fauna Branched Stone Coral: <i>Stylaster</i> sp., Bryozoa sp., Sponge: Porifera sp., Cup Coral: <i>Flabellum</i> sp., Sea Spider: Pycnogonida sp., Starfish: Asteroidea sp.,</p> | |

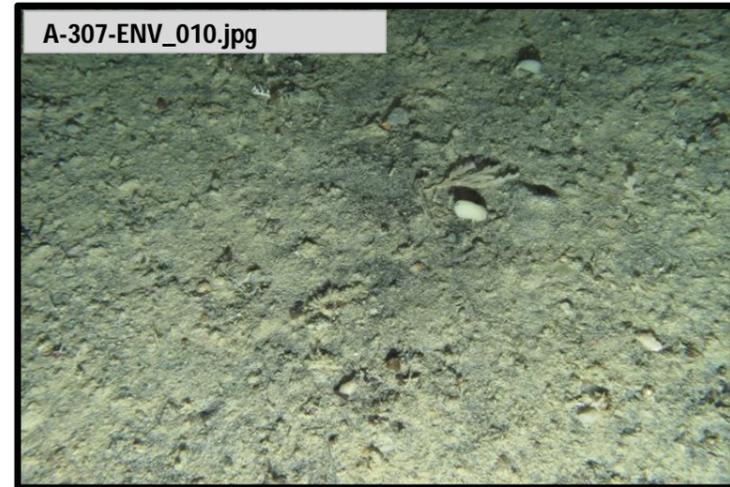


Photo Position: 793327 mE, 4188006 mN

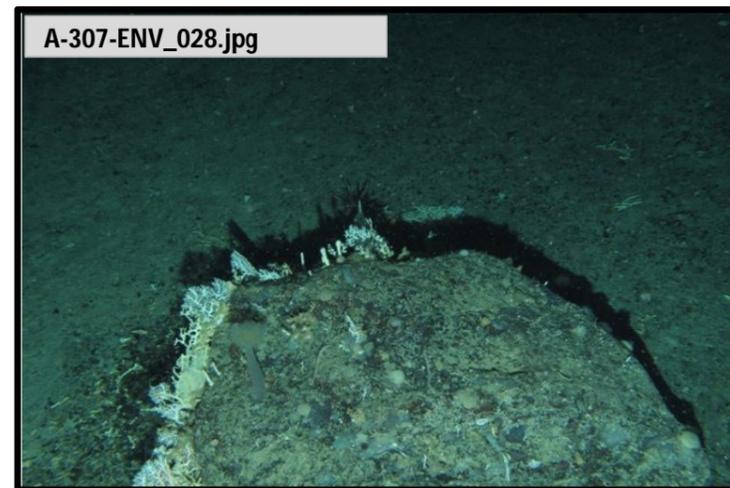
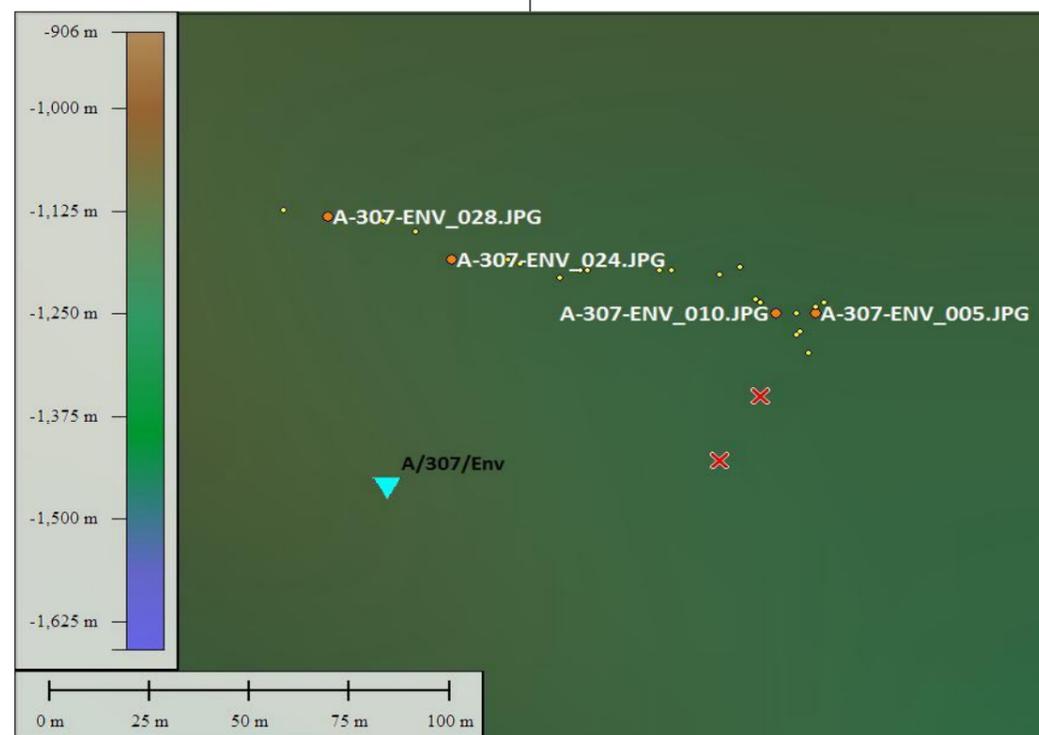


Photo Position: 793215 mE, 4188033 mN



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|---------------------------|---|
| <p>Client </p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
|---------------------------|---|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

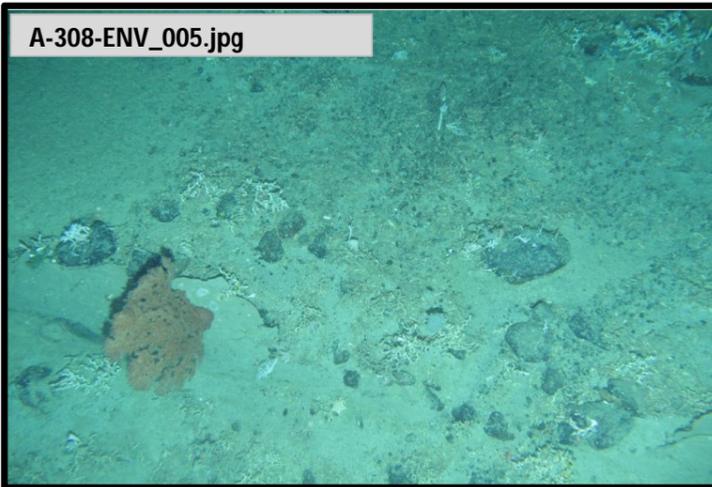


Photo Position: 784240 mE, 4192294 mN

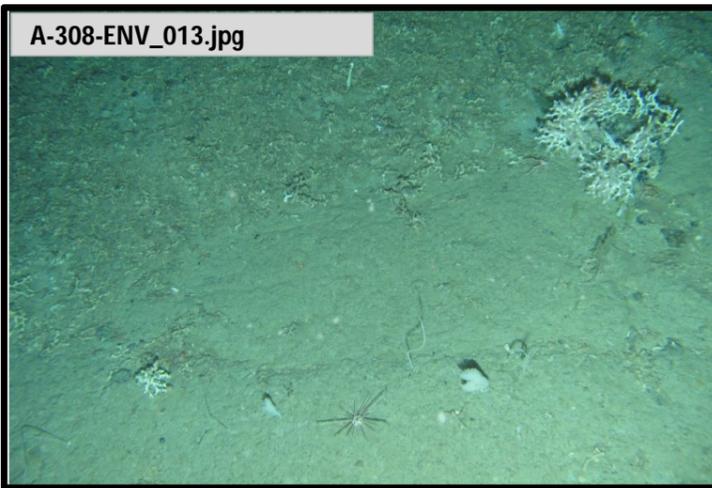


Photo Position: 784252 mE, 4192327 mN

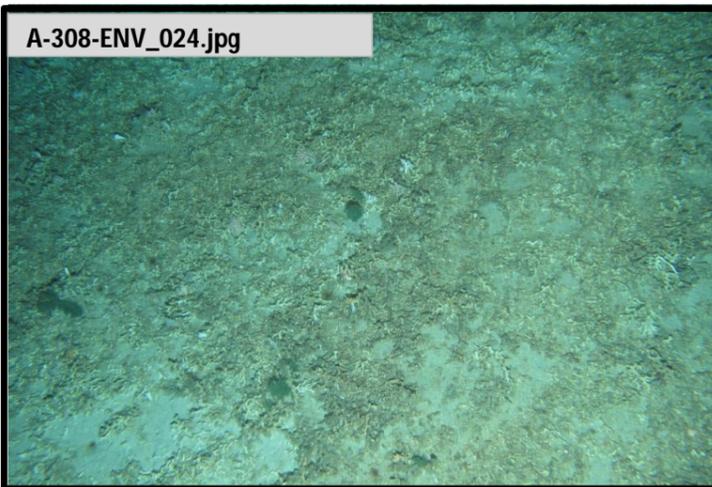


Photo Position: 784287 mE, 4192439 mN

Habitat Summary Information: A/308/ENV

Survey Area: FISA

No. of Stills: 26 Mins of Video: 24 Track Length: 217m

| | |
|---|---|
| <p>Site Selection Criteria Slump at base of escarpment</p> <p>Sediment Description Coarse pebbles and rocks suspended in clay</p> <p>Conspicuous Fauna <i>Lophelia pertusa</i>, Relic <i>Lophelia</i> sp., Sponge: Porifera sp., Branched Stone Coral: <i>Stylaster</i> sp., Sea Pen: Pennatulacea, Gorgonian sp., Starfish: Asteroidea sp., Hydroid sp., Glass Sponge: Hexactinellida sp., Squat Lobster: Galatheididae sp., Bryozoa sp., Decapod Shrimp sp., Sea Pen: Pennatulacea, Moridae sp., Octocoral sp., Branched Bryozoa sp., Brittlestar (Ophiuroid),</p> | <p>Analogue Interpretation Slump feature</p> |
|---|---|

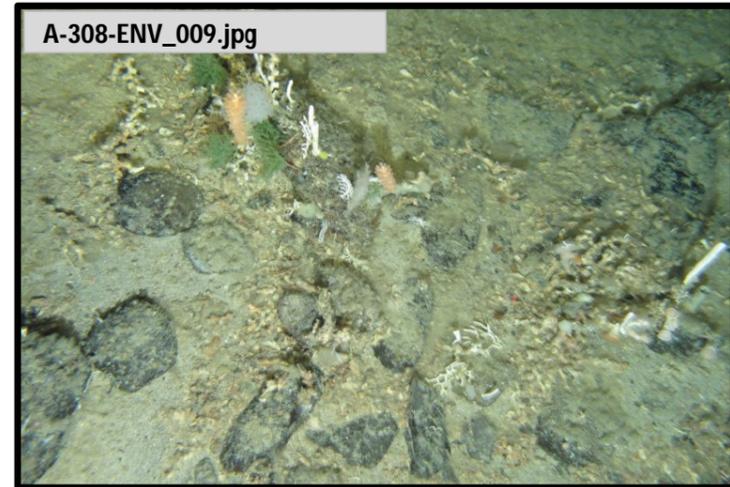


Photo Position: 784240 mE, 4192294 mN

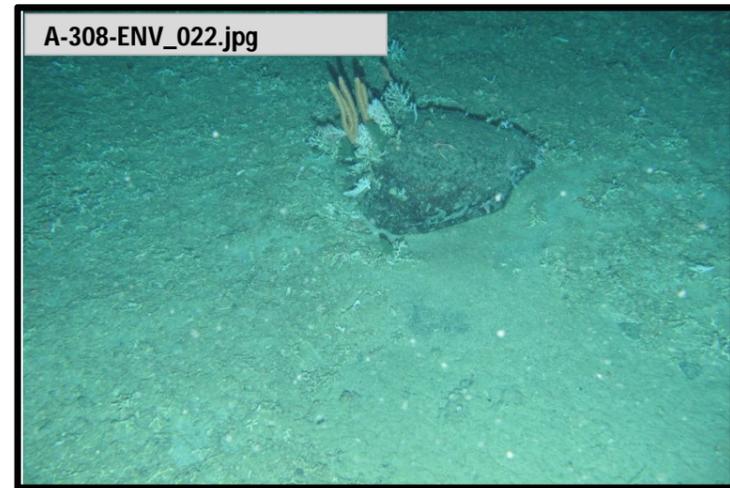


Photo Position: 784275 mE, 4192401 mN

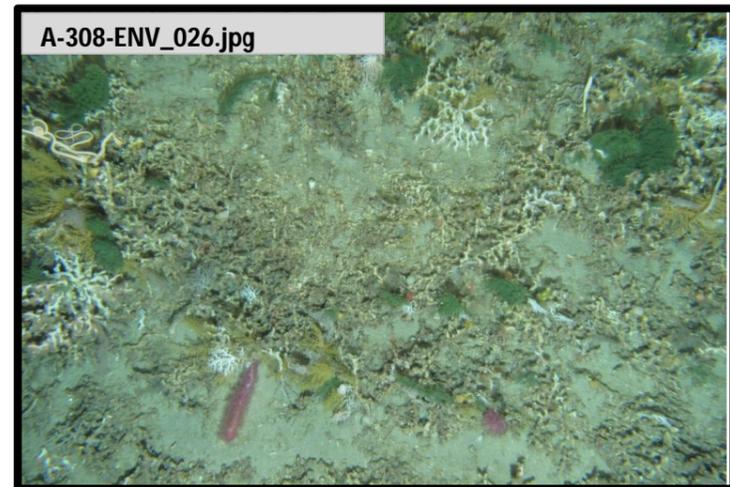
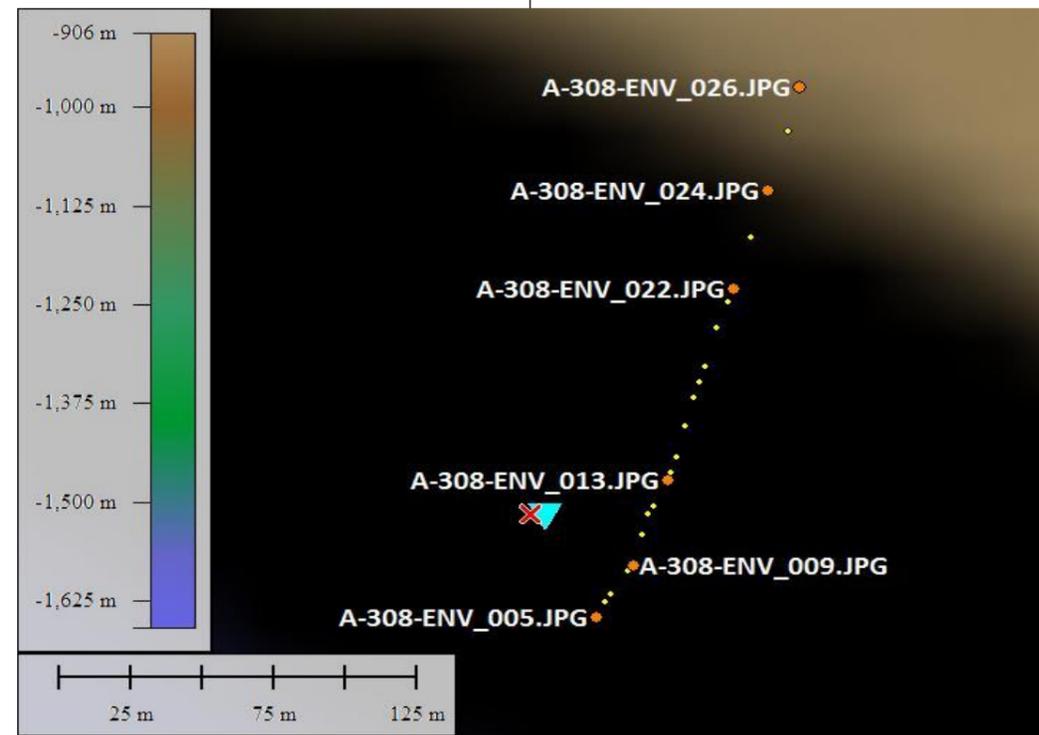


Photo Position: 784298 mE, 4192479 mN



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|---------------------------|---|
| <p>Client </p> | <p>Contractor Benthic Solutions Ltd., Marsh Road,</p> |
|---------------------------|---|

✕ Sample Location
 ● UW Photo
 ● UW Photo (featured)

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West



Photo Position: 791040 mE, 4193466 mN

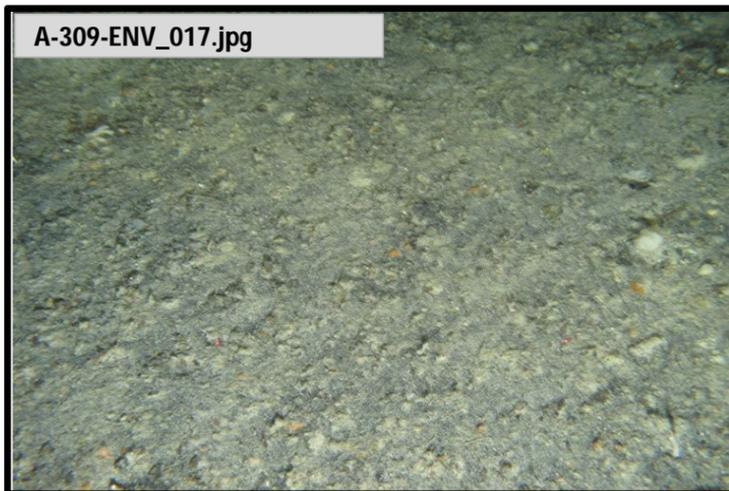


Photo Position: 790980 mE, 4193578 mN



Sediment Example Image

Habitat Summary Information: A/309/ENV

Survey Area: FISA

No. of Stills: 20 Mins of Video: 27 Track Length: 189m

| | |
|---|--|
| <p>Site Selection Criteria Base of trench feature</p> | <p>Analogue Interpretation Base of trench feature</p> |
| <p>Sediment Description Dark compact medium sand</p> | |
| <p>Conspicuous Fauna Cup Coral: <i>Flabellum sp.</i>, Bryozoa sp., Sponge: Porifera sp., Branched Stone Coral: <i>Stylaster sp.</i>, Isopod: possibly <i>Acutiserolis neaera</i>, Sea Pen: Pennatulacea,</p> | |

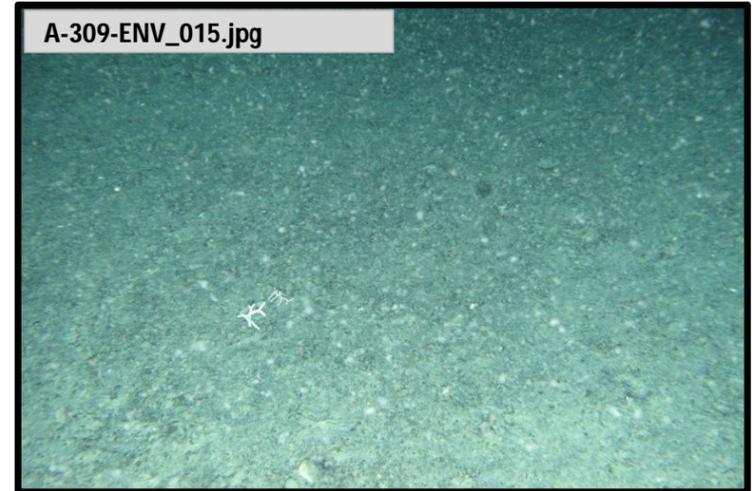


Photo Position: 790993 mE, 4193560 mN

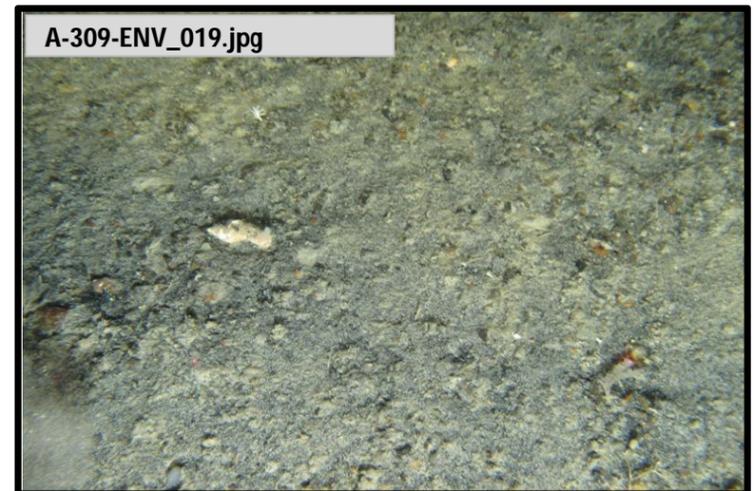
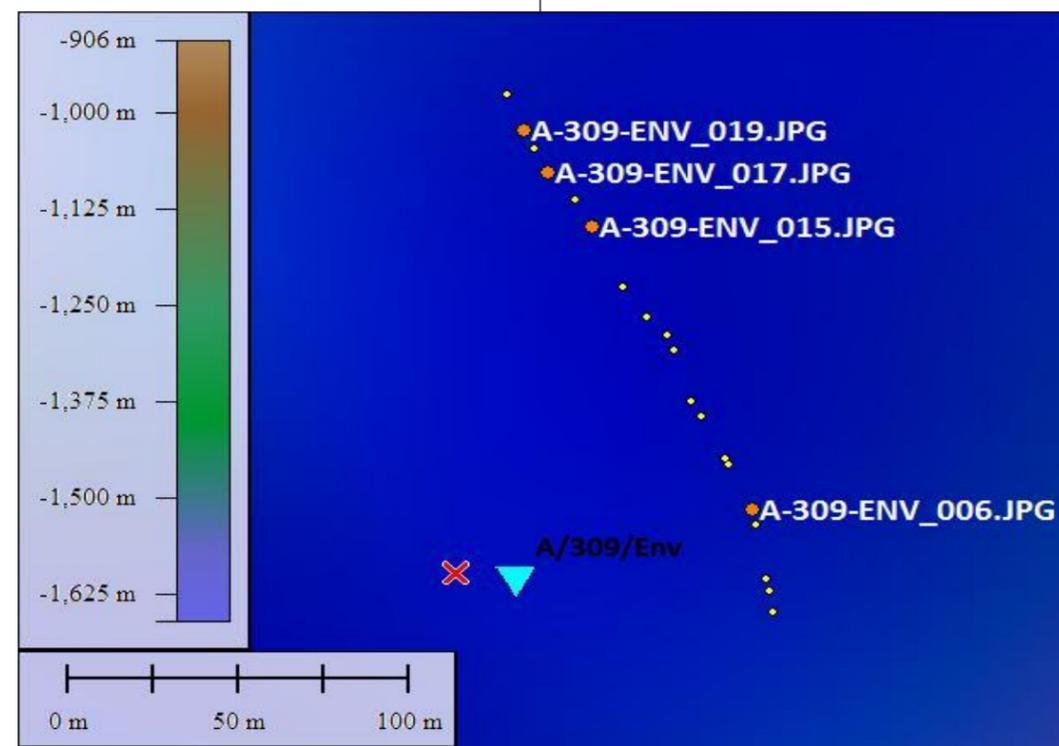


Photo Position: 790973 mE, 4193592 mN



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|---------------|--|
| <p>Client</p> | <p>Contractor</p> <p>Benthic Solutions Ltd., Marsh Road,</p> |
|---------------|--|

✗ Sample Location
 ● UW Photo
 ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

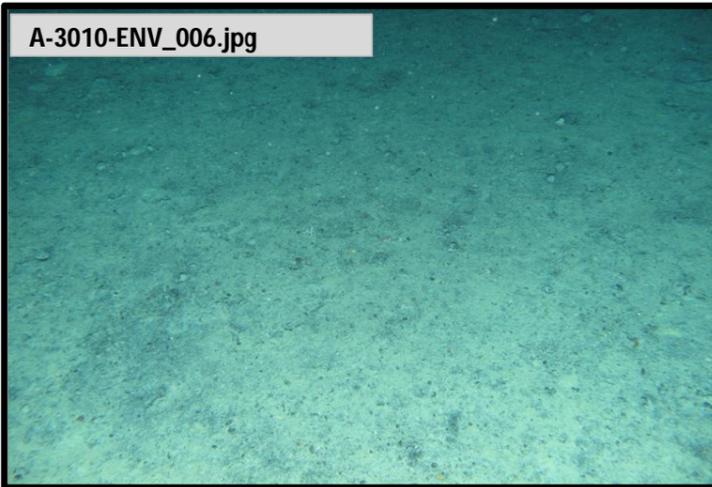


Photo Position: 791040 mE, 4193466 mN

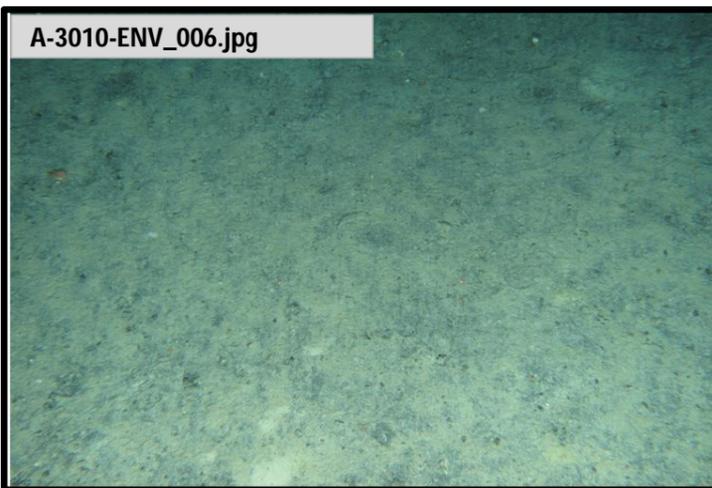


Photo Position: 790980 mE, 4193578 mN



Sediment Example Image

Habitat Summary Information: A/3010/ENV

Survey Area: FISA

No. of Stills: 20 Mins of Video: 27 Track Length: 432m

Site Selection Criteria

Repeat sample from 2011 survey (08666.1 FOGL Vinson West EBS)

Analogue Interpretation

Undulating seabed

Sediment Description

Olive grey fine to medium sand with fine gravel and a dark surface layer

Conspicuous Fauna

Isopod: possibly *Acutiserolis neaera*, Branched Stone Coral: *Stylaster sp.*, Cup Coral: *Flabellum sp.*, Sponge: *Porifera sp.*, Sea Pen: *Pennatulacea*,

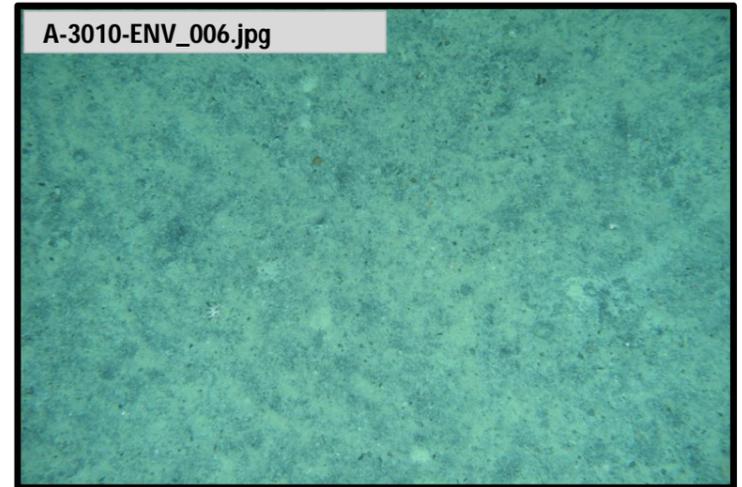


Photo Position: 790993 mE, 4193560 mN

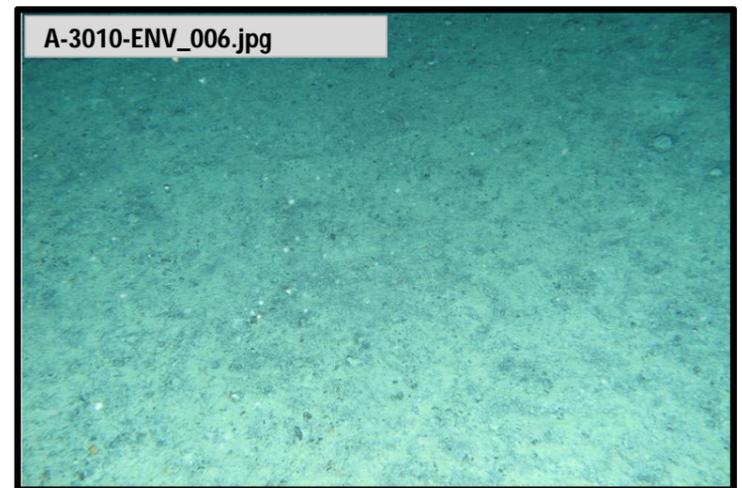
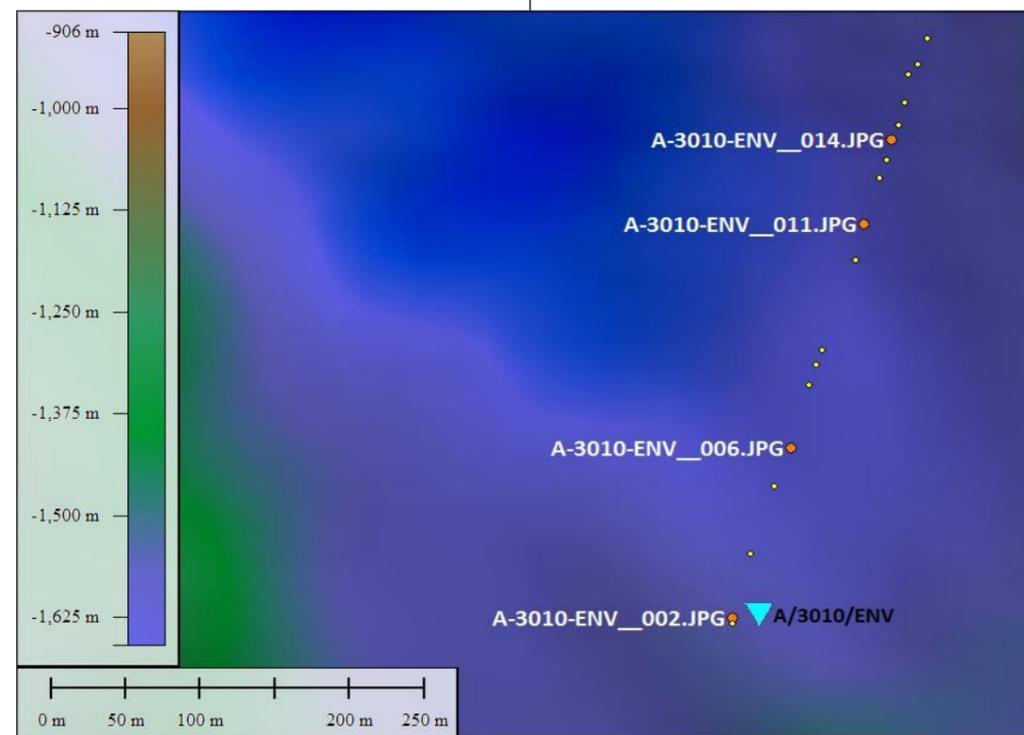


Photo Position: 790973 mE, 4193592 mN



Client
noble energy

Contractor
Benthic Solutions
Benthic Solutions Ltd., Marsh Road,

✗ Sample Location ● UW Photo ● UW Photo (featured)



Sieved Sample Image

Geodetic Information: Datum: WGS84 Projection: Transverse Mercator Central Meridian: 60° West

APPENDIX IX - SEABED PHOTOGRAPHIC POSITIONS

*Due to hardware limitations (beacon type and behaviour), navigation accuracy varied between sites. The accuracy has been given a rating of 1 to 10. 10 is considered good (under 30seconds between photo being taken and nearest beacon communication), and 0 is considered very bad (no communication with the beacon at all). A rating of 5 would mean around 15 minutes delay of beacon position, so accuracy would be ~100m in this case. Although not ideal – given the scale of the area being covered this inaccuracy is manageable.

| Site | Location | Pic Filename | Time | Date | Easting | Northing | Position rating |
|------|------------|--------------------|----------|------------|---------|----------|-----------------|
| FISA | A/03/ENV | A-03-ENV//002.JPG | 12:15:53 | 02/02/2014 | 736983 | 4188568 | 10 |
| FISA | A/03/ENV | A-03-ENV//003.JPG | 12:17:38 | 02/02/2014 | 737011 | 4188539 | 10 |
| FISA | A/03/ENV | A-03-ENV//004.JPG | 12:19:03 | 02/02/2014 | 736997 | 4188592 | 10 |
| FISA | A/03/ENV | A-03-ENV//005.JPG | 12:22:04 | 02/02/2014 | 736999 | 4188638 | 10 |
| FISA | A/03/ENV | A-03-ENV//006.JPG | 12:27:15 | 02/02/2014 | 737023 | 4188626 | 10 |
| FISA | A/03/ENV | A-03-ENV//007.JPG | 12:28:11 | 02/02/2014 | 736960 | 4188761 | 10 |
| FISA | A/03/ENV | A-03-ENV//008.JPG | 12:29:48 | 02/02/2014 | 737024 | 4188675 | 10 |
| FISA | A/03/ENV | A-03-ENV//009.JPG | 12:31:45 | 02/02/2014 | 737007 | 4188758 | 10 |
| FISA | A/03/ENV | A-03-ENV//010.JPG | 12:32:26 | 02/02/2014 | 737005 | 4188771 | 10 |
| FISA | A/03/ENV | A-03-ENV//011.JPG | 12:36:07 | 02/02/2014 | 736967 | 4188873 | 10 |
| FISA | A/03/ENV | A-03-ENV//012.JPG | 12:37:28 | 02/02/2014 | 736998 | 4188865 | 10 |
| FISA | A/03/ENV | A-03-ENV//013.JPG | 12:38:13 | 02/02/2014 | 736677 | 4188807 | 10 |
| FISA | A/03/ENV | A-03-ENV//014.JPG | 12:39:13 | 02/02/2014 | 737018 | 4188879 | 10 |
| FISA | A/03/ENV | A-03-ENV//015.JPG | 12:40:50 | 02/02/2014 | 737030 | 4188872 | 10 |
| FISA | A/03/ENV | A-03-ENV//016.JPG | 12:41:46 | 02/02/2014 | 737012 | 4188931 | 10 |
| FISA | A/03/ENV | A-03-ENV//017.JPG | 12:42:23 | 02/02/2014 | 737018 | 4188932 | 10 |
| FISA | A/03/ENV | A-03-ENV//018.JPG | 12:43:43 | 02/02/2014 | 737020 | 4188964 | 10 |
| FISA | A/03/ENV | A-03-ENV//019.JPG | 12:45:08 | 02/02/2014 | 737020 | 4188995 | 10 |
| FISA | A/03/ENV | A-03-ENV//020.JPG | 12:46:49 | 02/02/2014 | 737003 | 4189047 | 10 |
| FISA | A/03/ENV | A-03-ENV//021.JPG | 12:48:22 | 02/02/2014 | 737031 | 4189034 | 10 |
| FISA | A/03/ENV | A-03-ENV//022.JPG | 12:49:34 | 02/02/2014 | 737019 | 4189084 | 10 |
| FISA | A/03/ENV | A-03-ENV//023.JPG | 12:50:51 | 02/02/2014 | 737009 | 4189125 | 10 |
| FISA | A/03/ENV | A-03-ENV//024.JPG | 12:52:20 | 02/02/2014 | 737022 | 4189140 | 10 |
| FISA | A/03/ENV | A-03-ENV//025.JPG | 12:53:24 | 02/02/2014 | 737026 | 4189151 | 10 |
| FISA | A/03/ENV | A-03-ENV//026.JPG | 12:54:45 | 02/02/2014 | 737018 | 4189208 | 10 |
| FISA | A/09/ENV | A-09-ENV/00002.JPG | 00:54:20 | 01/01/2014 | 774026 | 4201278 | 10 |
| FISA | A/09/ENV | A-09-ENV/00003.JPG | 00:55:29 | 01/01/2014 | 774025 | 4201276 | 10 |
| FISA | A/09/ENV | A-09-ENV/00004.JPG | 00:57:18 | 01/01/2014 | 774027 | 4201279 | 10 |
| FISA | A/09/ENV | A-09-ENV/00005.JPG | 00:57:51 | 01/01/2014 | 774028 | 4201280 | 10 |
| FISA | A/09/ENV | A-09-ENV/00006.JPG | 00:58:40 | 01/01/2014 | 774028 | 4201281 | 10 |
| FISA | A/09/ENV | A-09-ENV/00007.JPG | 00:59:04 | 01/01/2014 | 774028 | 4201281 | 10 |
| FISA | A/09/ENV | A-09-ENV/00008.JPG | 00:59:59 | 01/01/2014 | 774027 | 4201284 | 10 |
| FISA | A/09/ENV | A-09-ENV/00009.JPG | 01:03:27 | 01/01/2014 | 774025 | 4201300 | 10 |
| FISA | A/09/ENV | A-09-ENV/00010.JPG | 01:05:07 | 01/01/2014 | 774025 | 4201302 | 10 |
| FISA | A/09/ENV | A-09-ENV/00011.JPG | 01:05:10 | 01/01/2014 | 774025 | 4201302 | 10 |
| FISA | A/09/ENV | A-09-ENV/00012.JPG | 01:05:43 | 01/01/2014 | 774025 | 4201304 | 10 |
| FISA | A/09/ENV | A-09-ENV/00013.JPG | 01:07:02 | 01/01/2014 | 774027 | 4201313 | 10 |
| FISA | A/09/ENV | A-09-ENV/00014.JPG | 01:07:38 | 01/01/2014 | 774028 | 4201317 | 10 |
| FISA | A/09/ENV | A-09-ENV/00015.JPG | 01:07:51 | 01/01/2014 | 774028 | 4201318 | 10 |
| FISA | A/09/ENV | A-09-ENV/00016.JPG | 01:08:21 | 01/01/2014 | 774028 | 4201322 | 10 |
| FISA | A/09/ENV | A-09-ENV/00017.JPG | 01:09:03 | 01/01/2014 | 774030 | 4201325 | 10 |
| FISA | A/09/ENV | A-09-ENV/00018.JPG | 01:09:49 | 01/01/2014 | 774032 | 4201330 | 10 |
| FISA | A/09/ENV | A-09-ENV/00019.JPG | 01:10:49 | 01/01/2014 | 774035 | 4201337 | 10 |
| FISA | A/09/ENV | A-09-ENV/00020.JPG | 01:11:37 | 01/01/2014 | 774035 | 4201343 | 10 |
| FISA | A/09/ENV | A-09-ENV/00021.JPG | 01:13:14 | 01/01/2014 | 774038 | 4201361 | 10 |
| FISA | A/09/ENV | A-09-ENV/00022.JPG | 01:14:54 | 01/01/2014 | 774039 | 4201367 | 10 |
| FISA | A/09/ENV | A-09-ENV/00023.JPG | 01:15:00 | 01/01/2014 | 774039 | 4201368 | 10 |
| FISA | A/09/ENV | A-09-ENV/00024.JPG | 01:15:43 | 01/01/2014 | 774040 | 4201374 | 10 |
| FISA | A/09/ENV | A-09-ENV/00025.JPG | 01:16:01 | 01/01/2014 | 774042 | 4201377 | 10 |
| FISA | A/09/ENV | A-09-ENV/00026.JPG | 01:18:11 | 01/01/2014 | 774044 | 4201394 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/002.JPG | 03:42:37 | 13/01/2014 | 784320 | 4202061 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/003.JPG | 03:43:41 | 13/01/2014 | 784321 | 4202062 | 10 |

| Site | Location | Pic Filename | Time | Date | Easting | Northing | Position rating |
|------|------------|--------------------|----------|------------|---------|----------|-----------------|
| FISA | A/1008/ENV | A-1008-ENV/004.JPG | 03:44:46 | 13/01/2014 | 784322 | 4202064 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/005.JPG | 03:46:07 | 13/01/2014 | 784324 | 4202064 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/006.JPG | 03:46:39 | 13/01/2014 | 784325 | 4202063 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/007.JPG | 03:47:03 | 13/01/2014 | 784325 | 4202063 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/008.JPG | 03:47:47 | 13/01/2014 | 784325 | 4202062 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/009.JPG | 03:48:20 | 13/01/2014 | 784325 | 4202061 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/010.JPG | 03:50:49 | 13/01/2014 | 784327 | 4202048 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/011.JPG | 03:51:13 | 13/01/2014 | 784327 | 4202049 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/012.JPG | 03:51:45 | 13/01/2014 | 784327 | 4202049 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/013.JPG | 03:52:26 | 13/01/2014 | 784327 | 4202046 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/014.JPG | 03:52:54 | 13/01/2014 | 784327 | 4202043 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/015.JPG | 03:54:31 | 13/01/2014 | 784326 | 4202037 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/016.JPG | 03:55:15 | 13/01/2014 | 784325 | 4202032 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/017.JPG | 03:56:08 | 13/01/2014 | 784326 | 4202029 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/018.JPG | 03:56:36 | 13/01/2014 | 784327 | 4202027 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/019.JPG | 03:58:29 | 13/01/2014 | 784327 | 4202013 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/020.JPG | 03:59:21 | 13/01/2014 | 784328 | 4202009 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/021.JPG | 03:59:46 | 13/01/2014 | 784327 | 4202007 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/022.JPG | 04:01:31 | 13/01/2014 | 784326 | 4201989 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/023.JPG | 04:02:11 | 13/01/2014 | 784325 | 4201981 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/024.JPG | 04:02:39 | 13/01/2014 | 784325 | 4201976 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/025.JPG | 04:03:11 | 13/01/2014 | 784324 | 4201970 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/026.JPG | 04:03:44 | 13/01/2014 | 784324 | 4201966 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/027.JPG | 04:04:52 | 13/01/2014 | 784324 | 4201962 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/028.JPG | 04:05:57 | 13/01/2014 | 784324 | 4201953 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/029.JPG | 04:06:49 | 13/01/2014 | 784323 | 4201941 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/030.JPG | 04:07:42 | 13/01/2014 | 784324 | 4201931 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/031.JPG | 04:09:31 | 13/01/2014 | 784322 | 4201913 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/032.JPG | 04:10:55 | 13/01/2014 | 784322 | 4201900 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/033.JPG | 04:12:24 | 13/01/2014 | 784323 | 4201891 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/034.JPG | 04:13:33 | 13/01/2014 | 784323 | 4201880 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/035.JPG | 04:14:09 | 13/01/2014 | 784321 | 4201870 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/036.JPG | 04:14:49 | 13/01/2014 | 784320 | 4201862 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/037.JPG | 04:16:06 | 13/01/2014 | 784320 | 4201850 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/038.JPG | 04:17:07 | 13/01/2014 | 784320 | 4201840 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/039.JPG | 04:17:47 | 13/01/2014 | 784318 | 4201833 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/040.JPG | 04:18:19 | 13/01/2014 | 784318 | 4201828 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/041.JPG | 04:20:36 | 13/01/2014 | 784316 | 4201800 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/042.JPG | 04:21:21 | 13/01/2014 | 784317 | 4201792 | 10 |
| FISA | A/1008/ENV | A-1008-ENV/043.JPG | 04:21:53 | 13/01/2014 | 784317 | 4201786 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/002.JPG | 12:19:41 | 05/01/2014 | 784839 | 4213104 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/003.JPG | 12:20:53 | 05/01/2014 | 784828 | 4213108 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/004.JPG | 12:23:27 | 05/01/2014 | 784828 | 4213105 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/005.JPG | 12:25:12 | 05/01/2014 | 784824 | 4213107 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/006.JPG | 12:25:56 | 05/01/2014 | 784826 | 4213107 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/007.JPG | 12:27:09 | 05/01/2014 | 784823 | 4213108 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/008.JPG | 12:27:53 | 05/01/2014 | 784820 | 4213112 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/009.JPG | 12:29:30 | 05/01/2014 | 784818 | 4213110 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/010.JPG | 12:31:11 | 05/01/2014 | 784805 | 4213111 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/011.JPG | 12:31:51 | 05/01/2014 | 784800 | 4213111 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/012.JPG | 12:34:49 | 05/01/2014 | 784782 | 4213109 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/013.JPG | 12:35:41 | 05/01/2014 | 784774 | 4213113 | 10 |
| FISA | A/1011/ENV | A-1011-ENV/014.JPG | 12:36:29 | 05/01/2014 | 784777 | 4213115 | 10 |
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| FISA | A/1011/ENV | A-1011-ENV/016.JPG | 12:41:56 | 05/01/2014 | 784734 | 4213121 | 10 |
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| FISA | A/1013/ENV | A-1013-ENV/004.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 2 |

| Site | Location | Pic Filename | Time | Date | Easting | Northing | Position rating |
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| FISA | A/1013/ENV | A-1013-ENV/008.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 4 |
| FISA | A/1013/ENV | A-1013-ENV/009.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 4 |
| FISA | A/1013/ENV | A-1013-ENV/010.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 5 |
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| FISA | A/1013/ENV | A-1013-ENV/013.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 6 |
| FISA | A/1013/ENV | A-1013-ENV/014.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 6 |
| FISA | A/1013/ENV | A-1013-ENV/015.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 6 |
| FISA | A/1013/ENV | A-1013-ENV/016.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 7 |
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| FISA | A/1013/ENV | A-1013-ENV/018.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 7 |
| FISA | A/1013/ENV | A-1013-ENV/019.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 8 |
| FISA | A/1013/ENV | A-1013-ENV/020.JPG | 21:03:33 | 12/01/2014 | 790314 | 4204430 | 8 |
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| FISA | A/1015/ENV | A-1015-ENV/016.JPG | 09:09:16 | 13/01/2014 | 786502 | 4198430 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/017.JPG | 09:10:04 | 13/01/2014 | 786493 | 4198431 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/018.JPG | 09:10:36 | 13/01/2014 | 786487 | 4198430 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/019.JPG | 09:11:53 | 13/01/2014 | 786477 | 4198430 | 10 |
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| FISA | A/1015/ENV | A-1015-ENV/021.JPG | 09:12:45 | 13/01/2014 | 786469 | 4198431 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/022.JPG | 09:13:18 | 13/01/2014 | 786464 | 4198434 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/023.JPG | 09:13:58 | 13/01/2014 | 786459 | 4198437 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/024.JPG | 09:15:11 | 13/01/2014 | 786447 | 4198436 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/025.JPG | 09:16:11 | 13/01/2014 | 786435 | 4198434 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/026.JPG | 09:16:52 | 13/01/2014 | 786428 | 4198434 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/027.JPG | 09:17:36 | 13/01/2014 | 786420 | 4198435 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/028.JPG | 09:18:00 | 13/01/2014 | 786415 | 4198436 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/029.JPG | 09:18:45 | 13/01/2014 | 786407 | 4198436 | 10 |
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| FISA | A/1015/ENV | A-1015-ENV/031.JPG | 09:20:17 | 13/01/2014 | 786386 | 4198439 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/032.JPG | 09:20:54 | 13/01/2014 | 786378 | 4198440 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/033.JPG | 09:21:34 | 13/01/2014 | 786370 | 4198440 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/034.JPG | 09:22:18 | 13/01/2014 | 786361 | 4198439 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/035.JPG | 09:22:51 | 13/01/2014 | 786355 | 4198439 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/036.JPG | 09:23:11 | 13/01/2014 | 786351 | 4198438 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/037.JPG | 09:24:19 | 13/01/2014 | 786334 | 4198438 | 10 |
| FISA | A/1015/ENV | A-1015-ENV/038.JPG | 09:25:16 | 13/01/2014 | 786321 | 4198438 | 10 |

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| FISA | A/106/ENV | A-106-ENV/003.JPG | 21:14:50 | 16/01/2014 | 743498 | 4180226 | 10 |
| FISA | A/106/ENV | A-106-ENV/004.JPG | 21:18:14 | 16/01/2014 | 743498 | 4180209 | 10 |
| FISA | A/106/ENV | A-106-ENV/005.JPG | 21:27:45 | 16/01/2014 | 743339 | 4180057 | 10 |
| FISA | A/106/ENV | A-106-ENV/006.JPG | 21:29:01 | 16/01/2014 | 743323 | 4180032 | 10 |
| FISA | A/106/ENV | A-106-ENV/007.JPG | 21:29:51 | 16/01/2014 | 743304 | 4180018 | 10 |
| FISA | A/106/ENV | A-106-ENV/008.JPG | 21:31:09 | 16/01/2014 | 743285 | 4179996 | 10 |
| FISA | A/106/ENV | A-106-ENV/009.JPG | 21:32:14 | 16/01/2014 | 743262 | 4179981 | 10 |
| FISA | A/106/ENV | A-106-ENV/010.JPG | 21:34:14 | 16/01/2014 | 743229 | 4179939 | 10 |
| FISA | A/106/ENV | A-106-ENV/011.JPG | 21:34:41 | 16/01/2014 | 743221 | 4179933 | 10 |
| FISA | A/106/ENV | A-106-ENV/012.JPG | 21:36:19 | 16/01/2014 | 743193 | 4179903 | 10 |
| FISA | A/106/ENV | A-106-ENV/013.JPG | 21:37:20 | 16/01/2014 | 743180 | 4179886 | 10 |
| FISA | A/106/ENV | A-106-ENV/014.JPG | 21:38:17 | 16/01/2014 | 743162 | 4179871 | 10 |
| FISA | A/106/ENV | A-106-ENV/015.JPG | 21:38:56 | 16/01/2014 | 743149 | 4179859 | 10 |
| FISA | A/106/ENV | A-106-ENV/016.JPG | 21:39:54 | 16/01/2014 | 743133 | 4179841 | 10 |
| FISA | A/106/ENV | A-106-ENV/017.JPG | 21:40:40 | 16/01/2014 | 743119 | 4179827 | 10 |
| FISA | A/106/ENV | A-106-ENV/018.JPG | 21:41:12 | 16/01/2014 | 743112 | 4179819 | 10 |
| FISA | A/106/ENV | A-106-ENV/019.JPG | 21:41:55 | 16/01/2014 | 743101 | 4179805 | 10 |
| FISA | A/106/ENV | A-106-ENV/020.JPG | 21:42:35 | 16/01/2014 | 743088 | 4179792 | 10 |
| FISA | A/106/ENV | A-106-ENV/021.JPG | 21:44:08 | 16/01/2014 | 743075 | 4179781 | 10 |
| FISA | A/106/ENV | A-106-ENV/022.JPG | 21:46:15 | 16/01/2014 | 743055 | 4179786 | 10 |
| FISA | A/106/ENV | A-106-ENV/023.JPG | 21:48:10 | 16/01/2014 | 743056 | 4179779 | 10 |
| FISA | A/106/ENV | A-106-ENV/024.JPG | 21:48:40 | 16/01/2014 | 743053 | 4179780 | 10 |
| FISA | A/10/ENV | A-10-ENV/001.JPG | 14:31:28 | 04/01/2014 | 774452 | 4205142 | 10 |
| FISA | A/10/ENV | A-10-ENV/002.JPG | 14:32:35 | 04/01/2014 | 774452 | 4205141 | 10 |
| FISA | A/10/ENV | A-10-ENV/003.JPG | 14:34:03 | 04/01/2014 | 774452 | 4205141 | 10 |
| FISA | A/10/ENV | A-10-ENV/004.JPG | 14:35:27 | 04/01/2014 | 774452 | 4205141 | 10 |
| FISA | A/10/ENV | A-10-ENV/005.JPG | 14:36:22 | 04/01/2014 | 774453 | 4205140 | 10 |
| FISA | A/10/ENV | A-10-ENV/006.JPG | 14:38:17 | 04/01/2014 | 774449 | 4205139 | 10 |
| FISA | A/10/ENV | A-10-ENV/007.JPG | 14:39:17 | 04/01/2014 | 774447 | 4205138 | 10 |
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| FISA | A/10/ENV | A-10-ENV/011.JPG | 14:43:32 | 04/01/2014 | 774435 | 4205137 | 10 |
| FISA | A/10/ENV | A-10-ENV/012.JPG | 14:45:27 | 04/01/2014 | 774426 | 4205136 | 10 |
| FISA | A/10/ENV | A-10-ENV/013.JPG | 14:47:03 | 04/01/2014 | 774419 | 4205134 | 10 |
| FISA | A/10/ENV | A-10-ENV/014.JPG | 14:47:46 | 04/01/2014 | 774415 | 4205133 | 10 |
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| FISA | A/12/ENV | A-12-ENV/007.JPG | 20:26:59 | 04/01/2014 | 783879 | 4203855 | 10 |
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| FISA | A/12/ENV | A-12-ENV/010.JPG | 20:30:24 | 04/01/2014 | 783878 | 4203855 | 10 |
| FISA | A/12/ENV | A-12-ENV/011.JPG | 20:30:24 | 04/01/2014 | 783878 | 4203855 | 10 |
| FISA | A/12/ENV | A-12-ENV/012.JPG | 20:32:01 | 04/01/2014 | 783871 | 4203856 | 10 |
| FISA | A/12/ENV | A-12-ENV/013.JPG | 20:32:45 | 04/01/2014 | 783868 | 4203856 | 10 |
| FISA | A/12/ENV | A-12-ENV/014.JPG | 20:32:57 | 04/01/2014 | 783867 | 4203855 | 10 |
| FISA | A/12/ENV | A-12-ENV/015.JPG | 20:36:36 | 04/01/2014 | 783851 | 4203854 | 9 |
| FISA | A/12/ENV | A-12-ENV/016.JPG | 20:38:25 | 04/01/2014 | 783838 | 4203858 | 9 |
| FISA | A/12/ENV | A-12-ENV/017.JPG | 20:42:57 | 04/01/2014 | 783806 | 4203857 | 9 |
| FISA | A/12/ENV | A-12-ENV/018.JPG | 20:42:57 | 04/01/2014 | 783806 | 4203857 | 10 |
| FISA | A/12/ENV | A-12-ENV/019.JPG | 20:42:57 | 04/01/2014 | 783806 | 4203857 | 10 |
| FISA | A/12/ENV | A-12-ENV/020.JPG | 20:42:57 | 04/01/2014 | 783806 | 4203857 | 10 |
| FISA | A/12/ENV | A-12-ENV/021.JPG | 20:43:42 | 04/01/2014 | 783799 | 4203860 | 10 |
| FISA | A/12/ENV | A-12-ENV/022.JPG | 20:43:42 | 04/01/2014 | 783799 | 4203860 | 10 |

| Site | Location | Pic Filename | Time | Date | Easting | Northing | Position rating |
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| FISA | A/14/ENV | A-14-ENV/005.JPG | 14:51:25 | 13/01/2014 | 782524 | 4194716 | 10 |
| FISA | A/14/ENV | A-14-ENV/006.JPG | 14:52:45 | 13/01/2014 | 782528 | 4194723 | 10 |
| FISA | A/14/ENV | A-14-ENV/007.JPG | 14:54:30 | 13/01/2014 | 782541 | 4194740 | 10 |
| FISA | A/14/ENV | A-14-ENV/008.JPG | 14:56:31 | 13/01/2014 | 782549 | 4194761 | 10 |
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| FISA | A/14/ENV | A-14-ENV/012.JPG | 15:01:58 | 13/01/2014 | 782572 | 4194810 | 10 |
| FISA | A/14/ENV | A-14-ENV/013.JPG | 15:03:39 | 13/01/2014 | 782585 | 4194830 | 10 |
| FISA | A/14/ENV | A-14-ENV/014.JPG | 15:04:47 | 13/01/2014 | 782585 | 4194838 | 10 |
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| FISA | A/18/ENV | A-18-ENV//003.JPG | 13:22:14 | 01/02/2014 | 768169 | 4173962 | 10 |
| FISA | A/18/ENV | A-18-ENV//004.JPG | 13:24:13 | 01/02/2014 | 768149 | 4173917 | 10 |
| FISA | A/18/ENV | A-18-ENV//005.JPG | 13:28:04 | 01/02/2014 | 768101 | 4173839 | 10 |
| FISA | A/18/ENV | A-18-ENV//006.JPG | 13:29:09 | 01/02/2014 | 768088 | 4173816 | 10 |
| FISA | A/18/ENV | A-18-ENV//007.JPG | 13:31:49 | 01/02/2014 | 768053 | 4173757 | 10 |
| FISA | A/18/ENV | A-18-ENV//008.JPG | 13:32:12 | 01/02/2014 | 768048 | 4173748 | 10 |
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| FISA | A/18/ENV | A-18-ENV//010.JPG | 13:35:29 | 01/02/2014 | 768005 | 4173670 | 10 |
| FISA | A/18/ENV | A-18-ENV//011.JPG | 13:35:52 | 01/02/2014 | 768000 | 4173664 | 10 |
| FISA | A/18/ENV | A-18-ENV//012.JPG | 13:38:46 | 01/02/2014 | 767962 | 4173601 | 10 |
| FISA | A/18/ENV | A-18-ENV//013.JPG | 13:39:23 | 01/02/2014 | 767954 | 4173587 | 10 |
| FISA | A/18/ENV | A-18-ENV//014.JPG | 13:40:30 | 01/02/2014 | 767941 | 4173567 | 10 |
| FISA | A/18/ENV | A-18-ENV//015.JPG | 13:40:58 | 01/02/2014 | 767936 | 4173558 | 10 |
| FISA | A/18/ENV | A-18-ENV//016.JPG | 13:41:25 | 01/02/2014 | 767930 | 4173550 | 10 |
| FISA | A/18/ENV | A-18-ENV//017.JPG | 13:41:47 | 01/02/2014 | 767925 | 4173540 | 10 |
| FISA | A/18/ENV | A-18-ENV//018.JPG | 13:42:38 | 01/02/2014 | 767915 | 4173524 | 10 |
| FISA | A/18/ENV | A-18-ENV//019.JPG | 13:44:23 | 01/02/2014 | 767894 | 4173482 | 10 |
| FISA | A/18/ENV | A-18-ENV//020.JPG | 13:44:47 | 01/02/2014 | 767888 | 4173473 | 10 |
| FISA | A/18/ENV | A-18-ENV//021.JPG | 13:45:21 | 01/02/2014 | 767881 | 4173461 | 10 |
| FISA | A/18/ENV | A-18-ENV//022.JPG | 13:45:41 | 01/02/2014 | 767877 | 4173453 | 10 |
| FISA | A/18/ENV | A-18-ENV//023.JPG | 13:46:15 | 01/02/2014 | 767871 | 4173444 | 10 |
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| FISA | A/201/ENV | A-201-ENV/004.JPG | 04:46:55 | 15/01/2014 | 734242 | 4206712 | 10 |
| FISA | A/201/ENV | A-201-ENV/005.JPG | 04:53:39 | 15/01/2014 | 734279 | 4206707 | 10 |
| FISA | A/201/ENV | A-201-ENV/006.JPG | 04:53:39 | 15/01/2014 | 734279 | 4206707 | 10 |
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| FISA | A/201/ENV | A-201-ENV/012.JPG | 04:57:49 | 15/01/2014 | 734300 | 4206714 | 10 |
| FISA | A/201/ENV | A-201-ENV/013.JPG | 04:58:37 | 15/01/2014 | 734317 | 4206713 | 10 |
| FISA | A/201/ENV | A-201-ENV/014.JPG | 04:59:34 | 15/01/2014 | 734332 | 4206712 | 10 |
| FISA | A/201/ENV | A-201-ENV/015.JPG | 05:00:18 | 15/01/2014 | 734336 | 4206711 | 10 |
| FISA | A/201/ENV | A-201-ENV/016.JPG | 05:00:42 | 15/01/2014 | 734337 | 4206711 | 10 |
| FISA | A/201/ENV | A-201-ENV/017.JPG | 05:02:11 | 15/01/2014 | 734343 | 4206708 | 10 |
| FISA | A/201/ENV | A-201-ENV/018.JPG | 05:02:47 | 15/01/2014 | 734346 | 4206707 | 10 |
| FISA | A/201/ENV | A-201-ENV/019.JPG | 05:03:20 | 15/01/2014 | 734350 | 4206708 | 10 |
| FISA | A/201/ENV | A-201-ENV/020.JPG | 05:03:56 | 15/01/2014 | 734356 | 4206710 | 10 |
| FISA | A/201/ENV | A-201-ENV/021.JPG | 05:04:20 | 15/01/2014 | 734360 | 4206710 | 10 |
| FISA | A/201/ENV | A-201-ENV/022.JPG | 05:05:45 | 15/01/2014 | 734378 | 4206710 | 10 |
| FISA | A/201/ENV | A-201-ENV/023.JPG | 05:06:17 | 15/01/2014 | 734385 | 4206709 | 10 |
| FISA | A/201/ENV | A-201-ENV/024.JPG | 05:07:10 | 15/01/2014 | 734396 | 4206709 | 10 |

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| FISA | A/201/ENV | A-201-ENV/027.JPG | 05:09:39 | 15/01/2014 | 734430 | 4206713 | 10 |
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| FISA | A/202/ENV | A-202-ENV/004.JPG | 23:14:08 | 14/01/2014 | 754148 | 4216061 | 10 |
| FISA | A/202/ENV | A-202-ENV/005.JPG | 23:15:12 | 14/01/2014 | 754157 | 4216068 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00003.JPG | 19:40:44 | 03/01/2014 | 787585 | 4199231 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00004.JPG | 19:41:09 | 03/01/2014 | 787585 | 4199231 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00008.JPG | 19:42:45 | 03/01/2014 | 787585 | 4199230 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00009.JPG | 19:46:19 | 03/01/2014 | 787581 | 4199219 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00010.JPG | 19:46:39 | 03/01/2014 | 787580 | 4199218 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00011.JPG | 19:47:44 | 03/01/2014 | 787579 | 4199212 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00012.JPG | 19:49:33 | 03/01/2014 | 787578 | 4199203 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00019.JPG | 19:53:31 | 03/01/2014 | 787568 | 4199176 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00020.JPG | 19:54:03 | 03/01/2014 | 787566 | 4199174 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00021.JPG | 19:54:39 | 03/01/2014 | 787566 | 4199170 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00022.JPG | 19:55:04 | 03/01/2014 | 787565 | 4199167 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00023.JPG | 19:55:24 | 03/01/2014 | 787564 | 4199164 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00024.JPG | 19:55:36 | 03/01/2014 | 787563 | 4199162 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00026.JPG | 19:56:40 | 03/01/2014 | 787560 | 4199153 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00027.JPG | 19:57:13 | 03/01/2014 | 787558 | 4199150 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00029.JPG | 19:57:57 | 03/01/2014 | 787557 | 4199143 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00030.JPG | 19:58:13 | 03/01/2014 | 787556 | 4199141 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00032.JPG | 19:58:49 | 03/01/2014 | 787555 | 4199137 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00033.JPG | 19:59:30 | 03/01/2014 | 787553 | 4199132 | 10 |
| FISA | A/203/ENV | A-203-ENV.a/00034.JPG | 20:00:26 | 03/01/2014 | 787551 | 4199125 | 10 |
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| FISA | A/203/ENV | A-203-ENV.a/00036.JPG | 20:02:19 | 03/01/2014 | 787543 | 4199109 | 10 |
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| FISA | A/206/ENV | A-206-ENV//005.JPG | 23:17:42 | 01/02/2014 | 776854 | 4163049 | 10 |
| FISA | A/206/ENV | A-206-ENV//006.JPG | 23:25:09 | 01/02/2014 | 776836 | 4163191 | 10 |
| FISA | A/206/ENV | A-206-ENV//007.JPG | 23:27:02 | 01/02/2014 | 776825 | 4163024 | 10 |
| FISA | A/206/ENV | A-206-ENV//008.JPG | 23:27:27 | 01/02/2014 | 776861 | 4163139 | 10 |
| FISA | A/206/ENV | A-206-ENV//009.JPG | 23:29:03 | 01/02/2014 | 776858 | 4163211 | 10 |
| FISA | A/206/ENV | A-206-ENV//010.JPG | 23:29:48 | 01/02/2014 | 776857 | 4163230 | 10 |
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| FISA | A/206/ENV | A-206-ENV//014.JPG | 23:33:22 | 01/02/2014 | 776850 | 4163322 | 10 |
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| FISA | A/206/ENV | A-206-ENV//018.JPG | 23:37:24 | 01/02/2014 | 776861 | 4163390 | 10 |
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| FISA | A/206/ENV | A-206-ENV//022.JPG | 23:40:33 | 01/02/2014 | 776869 | 4163450 | 10 |
| FISA | A/206/ENV | A-206-ENV//023.JPG | 23:41:10 | 01/02/2014 | 776857 | 4163484 | 10 |
| FISA | A/206/ENV | A-206-ENV//024.JPG | 23:44:15 | 01/02/2014 | 776870 | 4163534 | 10 |
| FISA | A/206/ENV | A-206-ENV//025.JPG | 23:44:43 | 01/02/2014 | 776870 | 4163545 | 10 |
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| FISA | A/206/ENV | A-206-ENV//027.JPG | 23:46:20 | 01/02/2014 | 776884 | 4163550 | 10 |
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| FISA | A/207/ENV | A-207-ENV//005.JPG | 09:39:01 | 05/01/2014 | 786694 | 4209669 | 10 |
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| FISA | A/207/ENV | A-207-ENV//009.JPG | 09:45:01 | 05/01/2014 | 786683 | 4209678 | 9 |
| FISA | A/207/ENV | A-207-ENV//010.JPG | 09:45:01 | 05/01/2014 | 786683 | 4209678 | 10 |
| FISA | A/207/ENV | A-207-ENV//011.JPG | 09:45:22 | 05/01/2014 | 786682 | 4209681 | 10 |
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| FISA | A/207/ENV | A-207-ENV//013.JPG | 09:47:35 | 05/01/2014 | 786674 | 4209683 | 10 |
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| FISA | A/207/ENV | A-207-ENV//034.JPG | 10:00:30 | 05/01/2014 | 786588 | 4209769 | 10 |
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| FISA | A/208/ENV | A-208-ENV//00012.JPG | 15:08:57 | 31/12/2013 | 773687 | 4228301 | 10 |
| FISA | A/208/ENV | A-208-ENV//00013.JPG | 15:10:20 | 31/12/2013 | 773691 | 4228304 | 10 |
| FISA | A/208/ENV | A-208-ENV//00014.JPG | 15:11:22 | 31/12/2013 | 773694 | 4228308 | 10 |
| FISA | A/208/ENV | A-208-ENV//00015.JPG | 15:11:57 | 31/12/2013 | 773695 | 4228310 | 10 |
| FISA | A/208/ENV | A-208-ENV//00016.JPG | 15:12:55 | 31/12/2013 | 773698 | 4228315 | 10 |
| FISA | A/208/ENV | A-208-ENV//00017.JPG | 15:14:16 | 31/12/2013 | 773702 | 4228321 | 10 |
| FISA | A/208/ENV | A-208-ENV//00018.JPG | 15:15:06 | 31/12/2013 | 773705 | 4228325 | 10 |
| FISA | A/208/ENV | A-208-ENV//00019.JPG | 15:16:01 | 31/12/2013 | 773709 | 4228330 | 10 |
| FISA | A/208/ENV | A-208-ENV//00020.JPG | 15:18:14 | 31/12/2013 | 773717 | 4228341 | 10 |
| FISA | A/208/ENV | A-208-ENV//00021.JPG | 15:18:50 | 31/12/2013 | 773720 | 4228344 | 10 |
| FISA | A/208/ENV | A-208-ENV//00022.JPG | 15:20:01 | 31/12/2013 | 773725 | 4228350 | 10 |
| FISA | A/208/ENV | A-208-ENV//00023.JPG | 15:20:57 | 31/12/2013 | 773729 | 4228356 | 10 |
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| FISA | A/20/ENV | A-20-ENV/00006.JPG | 08:36:04 | 03/01/2014 | 791007 | 4195278 | 10 |
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| FISA | A/301/ENV | A-301-ENV/034.JPG | 00:46:00 | 09/01/2014 | 790818 | 4214959 | 10 |
| FISA | A/302/ENV | A-302-ENV/002.JPG | 20:22:16 | 09/01/2014 | 804643 | 4210660 | 8 |
| FISA | A/302/ENV | A-302-ENV/003.JPG | 20:22:16 | 09/01/2014 | 804643 | 4210660 | 9 |
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| FISA | A/302/ENV | A-302-ENV/005.JPG | 20:22:16 | 09/01/2014 | 804643 | 4210660 | 9 |
| FISA | A/302/ENV | A-302-ENV/006.JPG | 20:22:16 | 09/01/2014 | 804643 | 4210660 | 10 |
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| FISA | A/302/ENV | A-302-ENV/008.JPG | 20:27:23 | 09/01/2014 | 804647 | 4210645 | 10 |
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| FISA | A/302/ENV | A-302-ENV/011.JPG | 20:28:52 | 09/01/2014 | 804643 | 4210643 | 10 |
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| FISA | A/302/ENV | A-302-ENV/013.JPG | 20:30:25 | 09/01/2014 | 804638 | 4210640 | 10 |
| FISA | A/302/ENV | A-302-ENV/014.JPG | 20:31:57 | 09/01/2014 | 804636 | 4210633 | 10 |

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| FISA | A/302/ENV | A-302-ENV/018.JPG | 20:36:28 | 09/01/2014 | 804630 | 4210610 | 10 |
| FISA | A/302/ENV | A-302-ENV/019.JPG | 20:38:12 | 09/01/2014 | 804624 | 4210599 | 10 |
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| FISA | A/302/ENV | A-302-ENV/021.JPG | 20:39:49 | 09/01/2014 | 804620 | 4210594 | 10 |
| FISA | A/302/ENV | A-302-ENV/022.JPG | 20:41:34 | 09/01/2014 | 804615 | 4210585 | 10 |
| FISA | A/302/ENV | A-302-ENV/023.JPG | 20:42:31 | 09/01/2014 | 804615 | 4210572 | 10 |
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| FISA | A/303/ENV | A-303-ENV/007.JPG | 13:22:19 | 09/01/2014 | 796071 | 4211877 | 10 |
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| FISA | A/304/ENV | A-304-ENV/011.JPG | 16:12:08 | 08/01/2014 | 783322 | 4221149 | 10 |
| FISA | A/304/ENV | A-304-ENV/012.JPG | 16:13:20 | 08/01/2014 | 783316 | 4221155 | 10 |
| FISA | A/304/ENV | A-304-ENV/013.JPG | 16:14:21 | 08/01/2014 | 783310 | 4221161 | 10 |
| FISA | A/304/ENV | A-304-ENV/014.JPG | 16:15:54 | 08/01/2014 | 783301 | 4221165 | 10 |
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| FISA | A/304/ENV | A-304-ENV/016.JPG | 16:16:50 | 08/01/2014 | 783294 | 4221168 | 10 |
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| FISA | A/304/ENV | A-304-ENV/019.JPG | 16:19:23 | 08/01/2014 | 783274 | 4221182 | 10 |
| FISA | A/304/ENV | A-304-ENV/020.JPG | 16:20:04 | 08/01/2014 | 783270 | 4221187 | 10 |
| FISA | A/305/ENV | A-305-ENV/003.JPG | 06:59:48 | 12/01/2014 | 778182 | 4210733 | 10 |
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| FISA | A/305/ENV | A-305-ENV/017.JPG | 07:17:04 | 12/01/2014 | 778007 | 4210916 | 10 |
| FISA | A/305/ENV | A-305-ENV/018.JPG | 07:17:48 | 12/01/2014 | 777999 | 4210924 | 10 |
| FISA | A/305/ENV | A-305-ENV/020.JPG | 07:18:45 | 12/01/2014 | 777990 | 4210936 | 10 |
| FISA | A/305/ENV | A-305-ENV/021.JPG | 07:19:17 | 12/01/2014 | 777983 | 4210939 | 10 |
| FISA | A/305/ENV | A-305-ENV/022.JPG | 07:19:53 | 12/01/2014 | 777979 | 4210939 | 10 |
| FISA | A/305/ENV | A-305-ENV/023.JPG | 07:20:19 | 12/01/2014 | 777972 | 4210943 | 10 |
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| FISA | A/305/ENV | A-305-ENV/028.JPG | 07:23:09 | 12/01/2014 | 777941 | 4210981 | 10 |
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| FISA | A/305/ENV | A-305-ENV/030.JPG | 07:24:29 | 12/01/2014 | 777928 | 4210996 | 10 |
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| FISA | A/306/ENV | A-306-ENV/021.JPG | 13:59:34 | 12/01/2014 | 796857 | 4206311 | 9 |
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| FISA | A/306/ENV | A-306-ENV/029.JPG | 14:11:12 | 12/01/2014 | 796803 | 4206390 | 10 |
| FISA | A/306/ENV | A-306-ENV/030.JPG | 14:11:12 | 12/01/2014 | 796803 | 4206390 | 10 |
| FISA | A/306/ENV | A-306-ENV/031.JPG | 14:12:00 | 12/01/2014 | 796799 | 4206407 | 10 |
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| FISA | A/307/ENV | A-307-ENV/006.JPG | 07:10:28 | 14/01/2014 | 793339 | 4188009 | 10 |
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| FISA | A/307/ENV | A-307-ENV/013.JPG | 07:17:15 | 14/01/2014 | 793318 | 4188019 | 10 |
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| FISA | A/307/ENV | A-307-ENV/021.JPG | 07:24:03 | 14/01/2014 | 793273 | 4188016 | 10 |
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| FISA | A/307/ENV | A-307-ENV/023.JPG | 07:26:24 | 14/01/2014 | 793260 | 4188021 | 10 |
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| FISA | A/308/ENV | A-308-ENV/007.JPG | 18:24:04 | 13/01/2014 | 784232 | 4192283 | 10 |
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| FISA | A/308/ENV | A-308-ENV/019.JPG | 18:33:33 | 13/01/2014 | 784265 | 4192371 | 10 |
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| FISA | A/308/ENV | A-308-ENV/026.JPG | 18:40:35 | 13/01/2014 | 784298 | 4192479 | 10 |
| FISA | A/309/ENV | A-309-ENV/002.JPG | 22:11:39 | 13/01/2014 | 791046 | 4193432 | 10 |
| FISA | A/309/ENV | A-309-ENV/003.JPG | 22:13:15 | 13/01/2014 | 791045 | 4193439 | 10 |
| FISA | A/309/ENV | A-309-ENV/004.JPG | 22:14:08 | 13/01/2014 | 791044 | 4193443 | 10 |
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| FISA | A/309/ENV | A-309-ENV/006.JPG | 22:18:14 | 13/01/2014 | 791040 | 4193466 | 10 |
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| FISA | A/309/ENV | A-309-ENV/009.JPG | 22:22:48 | 13/01/2014 | 791025 | 4193497 | 10 |
| FISA | A/309/ENV | A-309-ENV/010.JPG | 22:23:08 | 13/01/2014 | 791022 | 4193502 | 10 |
| FISA | A/309/ENV | A-309-ENV/011.JPG | 22:24:37 | 13/01/2014 | 791017 | 4193519 | 10 |
| FISA | A/309/ENV | A-309-ENV/012.JPG | 22:25:14 | 13/01/2014 | 791015 | 4193524 | 10 |
| FISA | A/309/ENV | A-309-ENV/013.JPG | 22:26:02 | 13/01/2014 | 791009 | 4193530 | 10 |
| FISA | A/309/ENV | A-309-ENV/014.JPG | 22:27:02 | 13/01/2014 | 791002 | 4193540 | 10 |
| FISA | A/309/ENV | A-309-ENV/015.JPG | 22:28:51 | 13/01/2014 | 790993 | 4193560 | 10 |

| Site | Location | Pic Filename | Time | Date | Easting | Northing | Position rating |
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| FISA | A/309/ENV | A-309-ENV/018.JPG | 22:31:13 | 13/01/2014 | 790976 | 4193586 | 10 |
| FISA | A/309/ENV | A-309-ENV/019.JPG | 22:31:49 | 13/01/2014 | 790973 | 4193592 | 10 |
| FISA | A/309/ENV | A-309-ENV/020.JPG | 22:32:45 | 13/01/2014 | 790968 | 4193604 | 10 |

APPENDIX X - AQC CERTIFICATION OF LABORATORIES

Schedule of Accreditation

issued by

United Kingdom Accreditation Service

21 - 47 High Street, Feltham, Middlesex, TW13 4UN, UK

| | | |
|---|---|---|
|  <p>Accredited to ISO/IEC 17025:2005</p> | ENVIRONMENTAL SCIENTIFICS GROUP LIMITED Trading as ESG Environmental Chemistry Issue No: 059 Issue date: 30 July 2012 | |
| | Environmental Chemistry PO Box 100 Bretby Business Park Burton-on-Trent Staffordshire DE15 0XD | Contact: Mr Andy Peirce Tel: +44 (0)1283 554542 Fax: +44 (0)1283 554422 E-Mail: andy.peirce@esgl.co.uk Website: |
| Testing performed at the above address only | | |

DETAIL OF ACCREDITATION

| Materials/Products tested | Type of test/Properties measured/Range of measurement | Standard specifications/ Equipment/Techniques used |
|------------------------------|--|--|
| SILICACIOUS MATERIALS | <u>Chemical Tests</u> Oxides of: Aluminium Calcium Iron Magnesium Manganese Phosphorus Potassium Silicon Sodium Sulphur Titanium | Documented In-House Method based on Analyst: June 1985: Vol 110 by ICP-OES, No ICPASH |
| SOILS, SEDIMENTS and SLUDGES | <u>Chemical Tests</u> Aluminium Arsenic Cadmium Chromium Cobalt Copper Iron Lead Manganese Molybdenum Nickel Vanadium Zinc | Documented In-House Method based on Blue Book Methods for the Examination of Waters and Associated Materials. Determination of Metals in Soils, Sediments and Sewage Sludge and Plants using ICP-OES, No ICPSOIL |

| | |
|--|---|
|  <p>1252 Accredited to ISO/IEC 17025:2005</p> | Schedule of Accreditation issued by United Kingdom Accreditation Service 21 - 47 High Street, Feltham, Middlesex, TW13 4UN, UK |
| | ENVIRONMENTAL SCIENTIFICS GROUP LIMITED Trading as ESG Environmental Chemistry Issue No: 059 Issue date: 30 July 2012 |
| Testing performed at main address only | |

| Materials/Products tested | Type of test/Properties measured/Range of measurement | Standard specifications/ Equipment/Techniques used |
|---------------------------------------|---|---|
| SOILS, SEDIMENTS and SLUDGES (cont'd) | <u>Chemical Tests</u> (cont'd) | |
| SOILS ONLY (includes made ground) | Calcium Magnesium Sodium Potassium Strontium Phosphorus | |
| SOILS ONLY | Antimony Arsenic Cadmium Chromium Cobalt Copper Lead Manganese Mercury Molybdenum Nickel Selenium Thallium Uranium Vanadium Zinc | Documented In-House Method using ICPMS, No ICPMSS |
| MARINE SEDIMENTS | Arsenic Tin Lead Copper Lithium Aluminium Barium Iron Manganese Strontium Chromium Nickel Vanadium Zinc | Documented in house method: ICPSEEXT - Hydrofluoric acid digestion followed by ICPMSSED analysis by ICPMS Documented in house method: ICPSEEXT - Hydrofluoric acid digestion followed by ICPSED analysis by ICPOES |

United Kingdom Accreditation Service

ACCREDITATION CERTIFICATE



**TESTING LABORATORY
No. 1205**

**Environmental Scientifics Group Limited
Trading as TES Bretby**

is accredited in accordance with the recognised International Standard ISO/IEC 17025:2005
General Requirements for the competence of testing and calibration laboratories.

This accreditation demonstrates technical competence for a defined scope as detailed in and at the locations specified in the schedule to this certificate, and the operation of a laboratory quality management system (refer joint ISO-ILAC-IAF Communiqué dated 18 June 2005).

The schedule to this certificate is an essential accreditation document and from time to time may be revised and reissued by the United Kingdom Accreditation Service. The most recent issue of the schedule of accreditation, which bears the same accreditation number as this certificate, is available from the UKAS website www.ukas.org.

This accreditation is subject to continuing conformity with United Kingdom Accreditation Service requirements. The absence of a schedule on the UKAS website indicates that the accreditation is no longer in force.



Accreditation Manager, United Kingdom Accreditation Service

**Initial Accreditation date
12 October 1992**

**This certificate issued on
01 November 2010**

The Department for Innovation, Universities and Skills (DIUS) has entered into a memorandum of understanding with the United Kingdom Accreditation Service (UKAS) through which UKAS is recognised as the national body responsible for assessing and accrediting the competence of organisations in the fields of calibration, testing, inspection and certification of systems, products and persons



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31/08/14

Dear Ian,

Please find below the results of the Falklands macrobenthic sample AQC process.

R.E. Falklands AQC Results

| Benthic Solutions' sample code | Taxa found by Aquatic Environments | Aquatic Environments comments |
|---------------------------------------|---|---|
| A18 | 1x Copepoda | Very clean sample, well sorted. |
| A202 | Nothing | Very clean sample, well sorted. |
| A302 | 1x Melinninae 1x Syllidae | Very small specimens. Very clean sample well sorted. |
| T10 | 1x Copepod 1x Ostracoda 1x Gastropoda | Very clean sample well sorted. Gastropod similar size to a UK Hydrobia. |
| T301 | 1x Copepoda | Very clean sample, well sorted. |
| T305 | Nothing | Very clean sample, well sorted. |

Yours sincerely

Tom Mercer
 Principal Consultant

Aquatic Environments is a specialised aquatic environmental consultancy providing services to the public and private sectors.
 VAT No. 708 9820 07.

APPENDIX XI – PEARSONS CORRELATION

APPENDIX XII - SERVICE WARRANTY

This report, with its associated works and services, has been designed solely to meet the requirements of the contract agreed with you, our client. If used in other circumstances, some or all of the results may not be valid and we can accept no liability for such use. Such circumstances include different or changed objectives, use by third parties, or changes to, for example, site conditions or legislation occurring after completion of the work. In case of doubt, please consult Benthic Solutions Limited.

Appendix L: FISA12 and FIST13 MetOcean Study Report

RPS | Falkland Islands (Blocks FISA12 and FIST13). Preliminary MetOcean Study

ASA Team: Stefanie Zamorski, Eric Comerma (PM)

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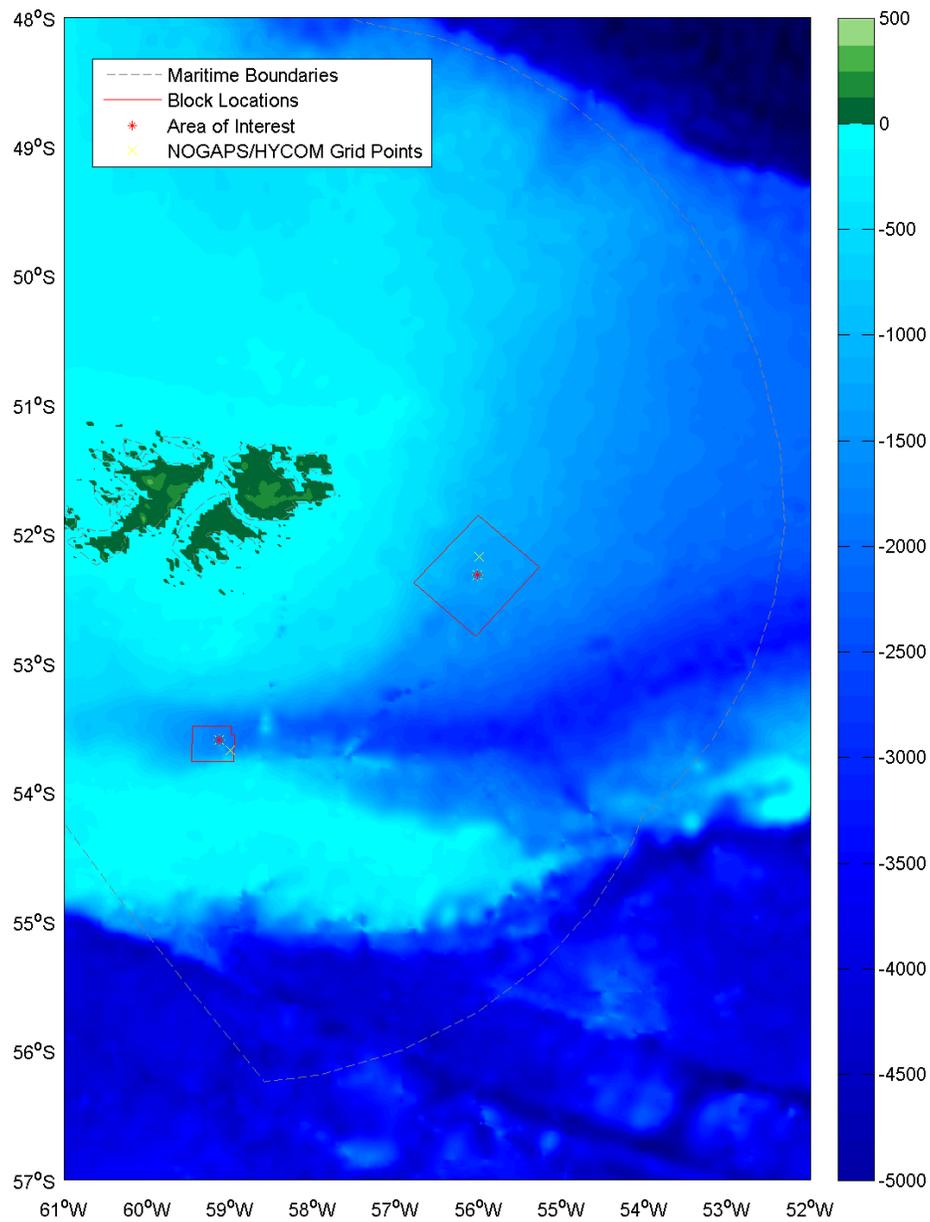


Figure 1. Location of the sites of interest with bathymetry (m).

Table 1. Coordinates of the sites of interest.

| Site Name | Type / Block | Latitude | Longitude | Approximate Distance from land (km) | Approximate Water Depth (m) |
|-----------|--------------|------------|------------|-------------------------------------|-----------------------------|
| FISA12 | Block center | 52.3228° S | 56.0229° W | 137 (Cape Pembroke) | 1,177 |
| FIST13 | Block center | 53.6032° S | 59.135° W | 76 (Beauchene Island) | 1,527 |

2. Environmental Data Analysis

2.1. Bathymetry

The Falkland Islands are situated on a projection of the Patagonian continental shelf that is bounded to the north by a steep slope, called the Falklands Escarpment. The continental shelf extends around 200 km beyond the Falklands coast to the north, about 50 km to the south-west, and approximately 50-100 km offshore on the eastern side (Otley, 2008). The North Falklands Basin is a gradually north-eastward sloping area between the Falkland Islands and the Falklands Escarpment, with depths between 150 and 1,500 m (Figure 2). The South Falkland Basin is located about 200 km south of the Falkland Islands and 600 km east of the southeastern coast of South America. It lies at the southern edge of the Falkland Plateau, which is a shallow continental promontory (Bry, 2004). The Falkland Plateau extends about 1,800 km east of the Falkland Islands, bordered by the Falklands Escarpment to the north and the North Scotia Ridge to the south. The Falkland Trough lies between the plateau and the ridge, which begins as a slight depression in the continental shelf and deepens to 3,700 m (Ludwig, 1983).

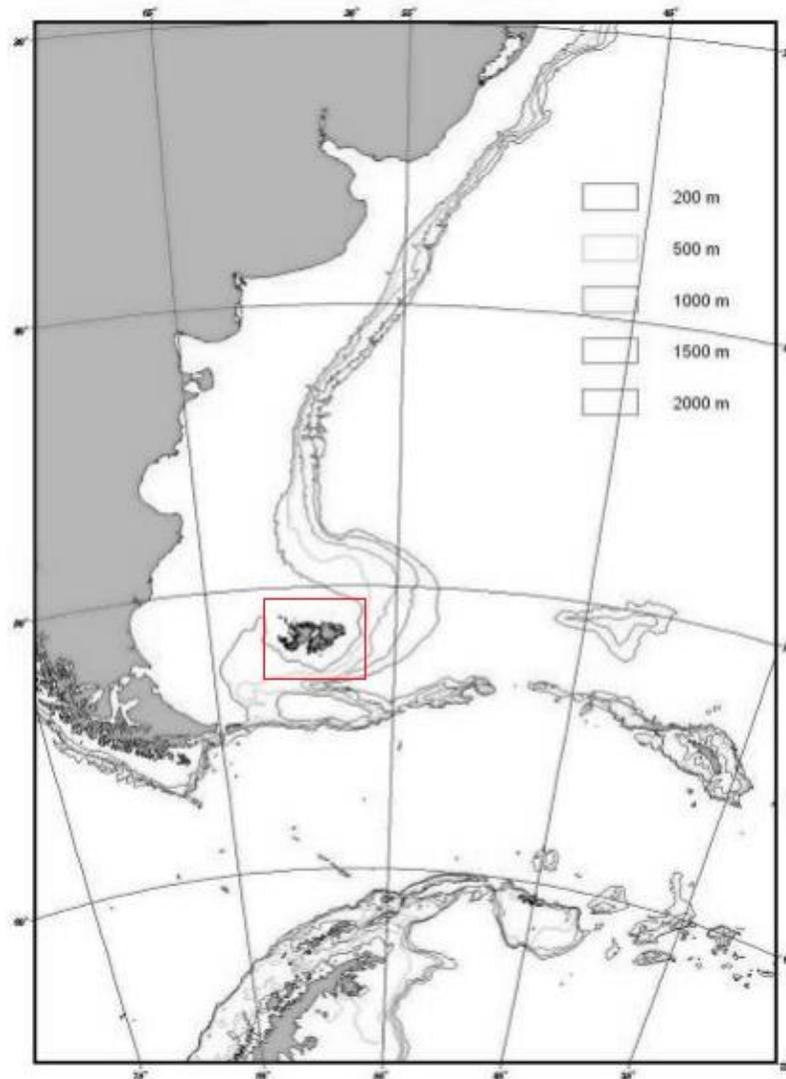


Figure 2. The location of Falkland Islands relative to South America, South Georgia and Antarctica (Otley, 2008).

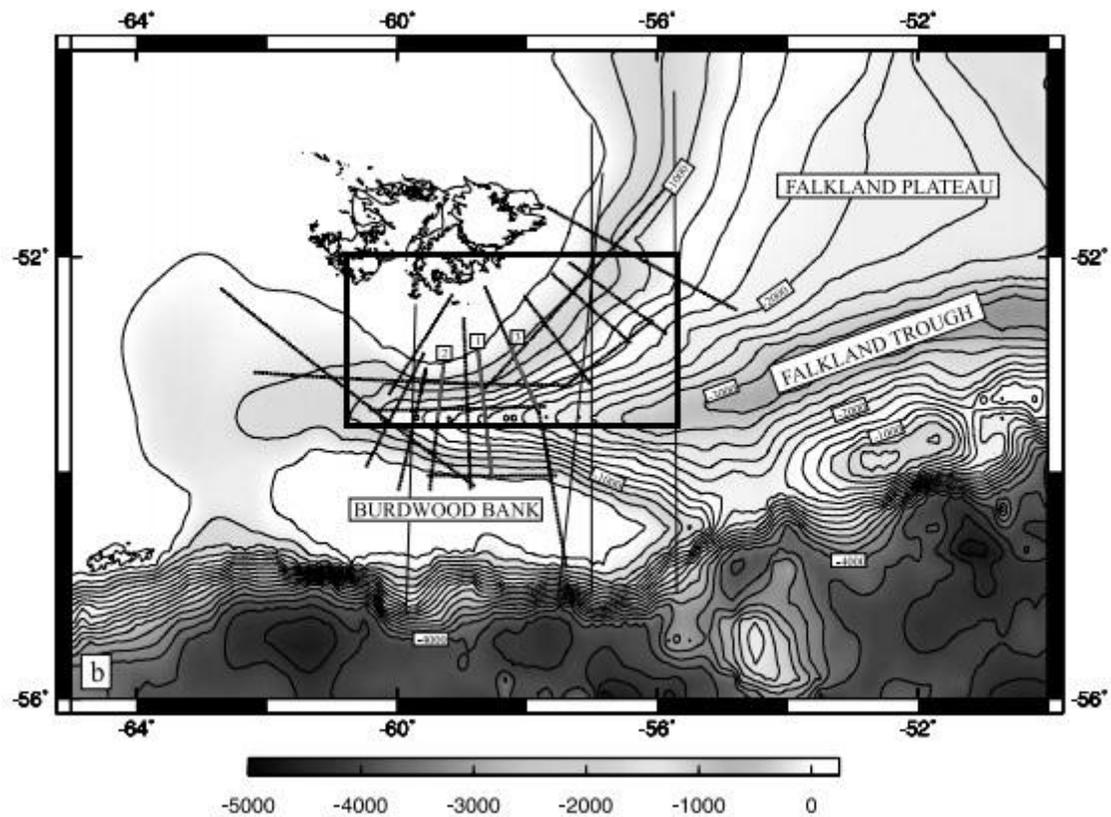


Figure 3. Bathymetry surrounding the Falkland Islands and notable bathymetry features. The South Falkland Basin is indicated by the black box (Source: Bry, 2004).

2.2. Seasonality

The weather regime in the Falkland Islands is influenced by the close proximity of the Andes to the west, the surrounding waters, particularly the cold East Falkland current moving northwards, and the presence of the Antarctic Peninsula over 1,000 km to the south. The annual mean maximum temperature is approximately 10°C and annual mean minimum temperature is approximately 3°C. The predominant wind direction is westerly, however, depressions and associated fronts move through the area frequently with wind speeds greater than 30 knots, thus causing variability in the regime. The prevailing wind direction falls in a broad arc from south-southwesterly to north-northwesterly for about 70% of the time (Upton and Shaw, 2002). When low pressure troughs accelerate towards the Falkland Islands, this leads to strong northerly winds and enhanced precipitation. This occurs more commonly during austral winter months. In general, the precipitation rate in the Falkland Islands is low, with around 625 mm annually.

2.3. Wind Dataset – NOGAPS

For this study, in the absence of an extended spatial coverage of long term observed winds, wind data was obtained from the output of the U.S. Navy Operational Global Atmospheric Prediction System (NOGAPS). The version of the NOGAPS dataset used for this modeling study is originally derived from

the publically available version hosted by the U.S. Global Ocean Data Assimilation Experiment (GODAE) and subsequently has a QuikSCAT correction applied by the HYCOM Consortium. This dataset of winds at 10 m above the surface is provided at 0.5 degree horizontal resolution with a 3-hour time step provided from 2009 through 2012. NOGAPS winds were also included as one of the main driving forces used in HYCOM, the global hydrodynamic currents dataset also used in this study (Section 2.5, Current Dataset – *HYCOM Hindcast*)

Figure 4 illustrates the spatial variability of the yearly average NOGAPS wind field represented by rose diagrams. Overall, wind direction is primarily from the west and fluctuates between southwest to northwesterly winds. Wind speeds tend to be greater than 15 knots, decreasing slightly closer to the Falkland Islands.

Figure 5 provides monthly wind speed statistics (average and 95th percentile) between 2009 and 2012 for the NOGAPS wind grid point at an offshore location in the center of the FISA12 block. This site shows average monthly wind speeds between 16-21 knots, reaching peak speeds in June. The highest wind speeds occur during late austral fall-winter, with lesser values throughout austral summer. Annual and monthly wind roses for years 2009 through 2012 for the selected location in Block FISA12 are shown in Figure 6 and Figure 7. Analysis of the wind roses at this site indicates little seasonality the wind regime. The wind direction throughout the year is primarily westerly, expanding in a northwest and southwesterly arc, with an average speed around 18.5 knots. The strongest monthly winds occur in late austral fall through winter (April-August) with speeds averaging 18-20 knots and average maximum monthly values reaching 48 knots during this time. Winds weaken during the spring and summer.

The same monthly wind speed statistics (average and 95th percentile) were performed for a central grid point in the southern block, FIST13. The same trend as the northern site is seen in Figure 8. Yearly (Figure 9) and monthly (Figure 10) wind roses at the southern site show wind direction primarily from the west-northwest with a yearly average of 18.7 knots. Similar seasonality is seen compared with the northern site. Winds are stronger during the winter and predominantly from the west, with northwesterly and southwesterly components as well. During the summer (December-February), monthly averaged speeds decrease and shift direction to predominantly westerly and southwesterly. The highest monthly averages are during April through August (18.5-20 knots).

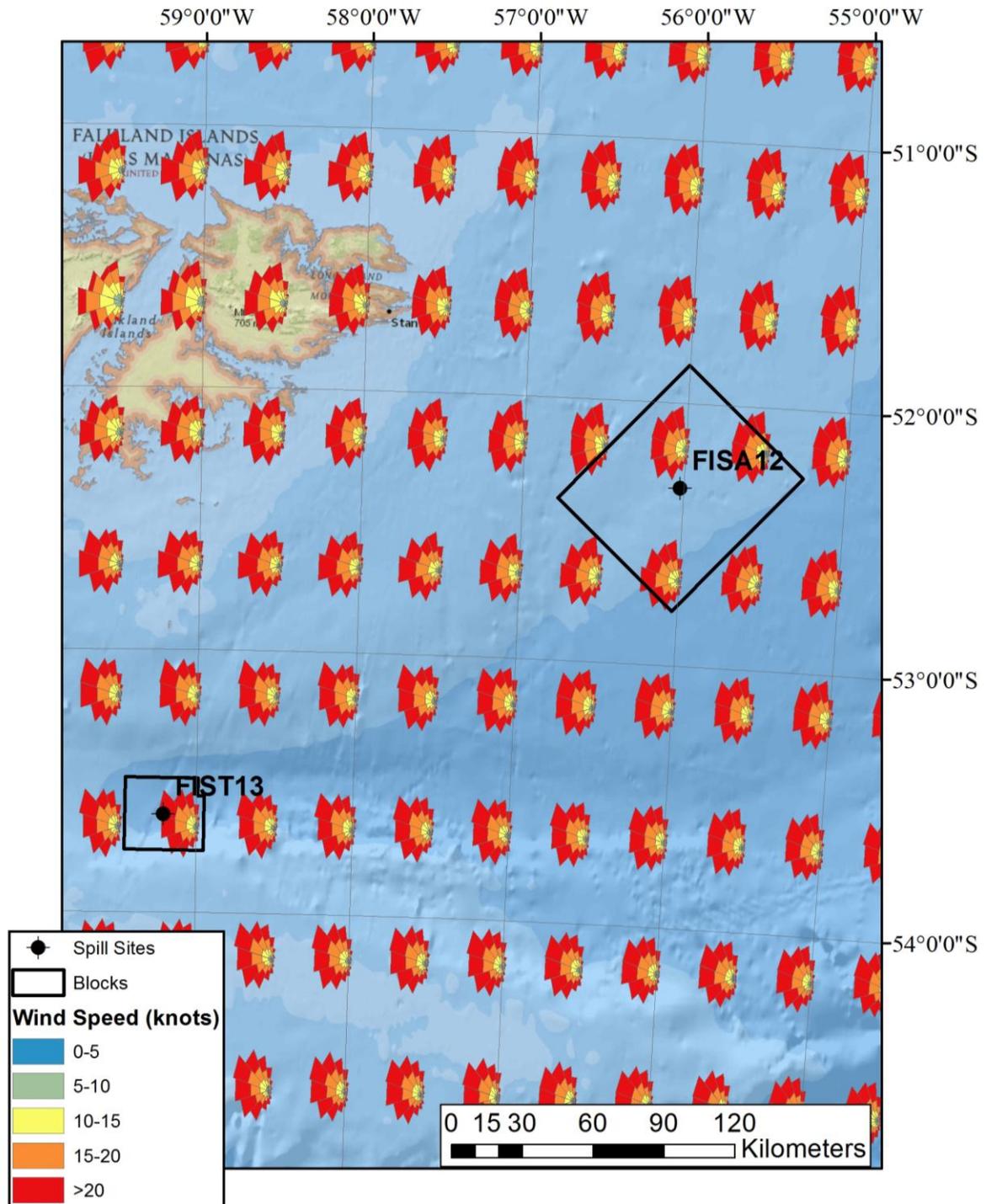


Figure 4. Spatial variability of the average NOGAPS wind field represented by rose diagrams; wind speeds in knots, using meteorological convention (i.e. direction wind is coming from).

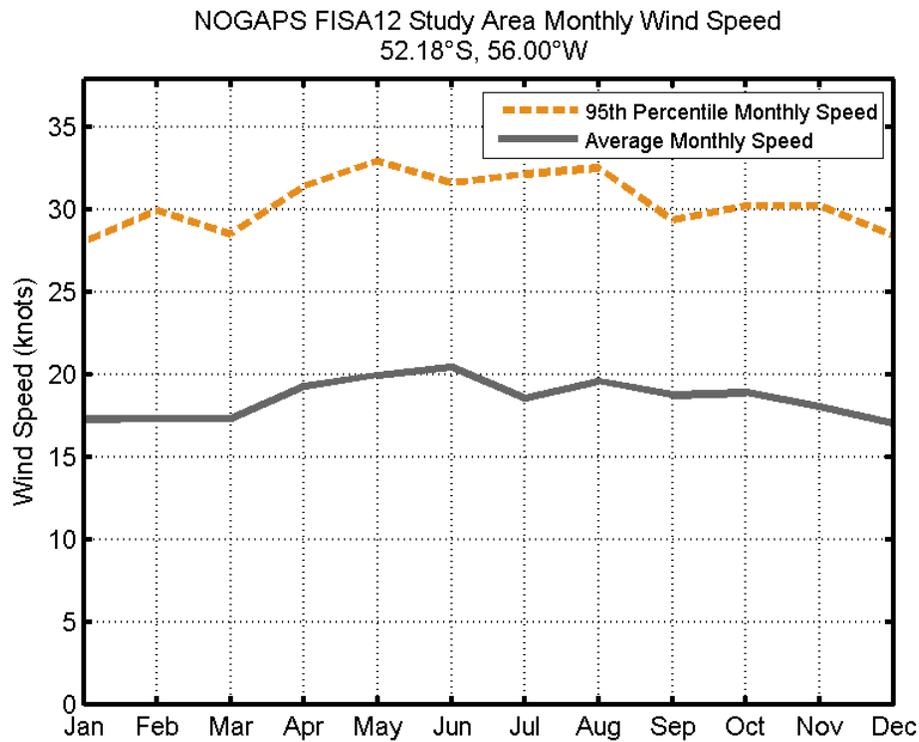


Figure 5. NOGAPS wind speed statistics: monthly average (grey solid) and 95th Percentile (orange dashed) wind speed in the FISA12 block offshore Falklands for the period of 2009-2012.

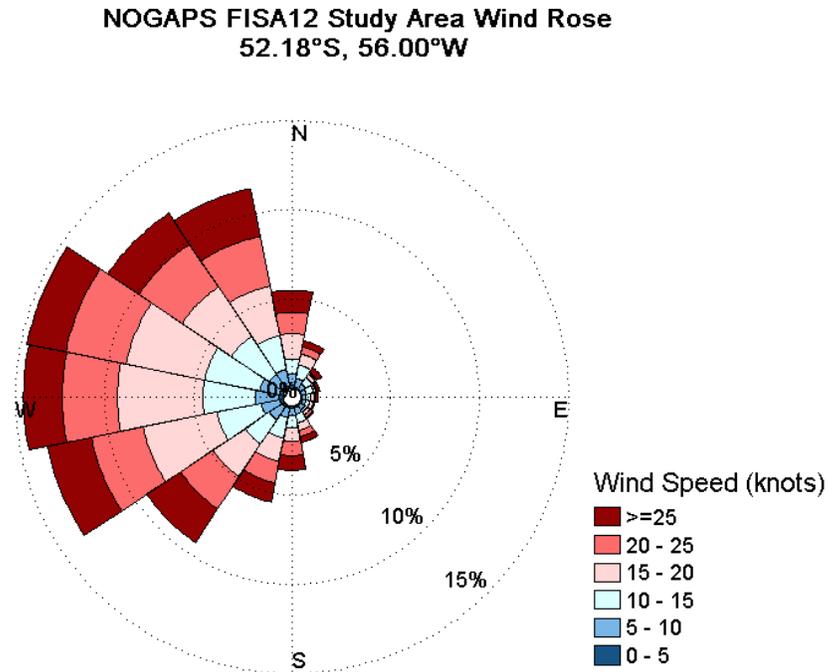


Figure 6. Yearly NOGAPS wind roses in FISA12 block offshore Falklands. Wind speeds are in knots, using meteorological convention (i.e., direction wind is coming from).

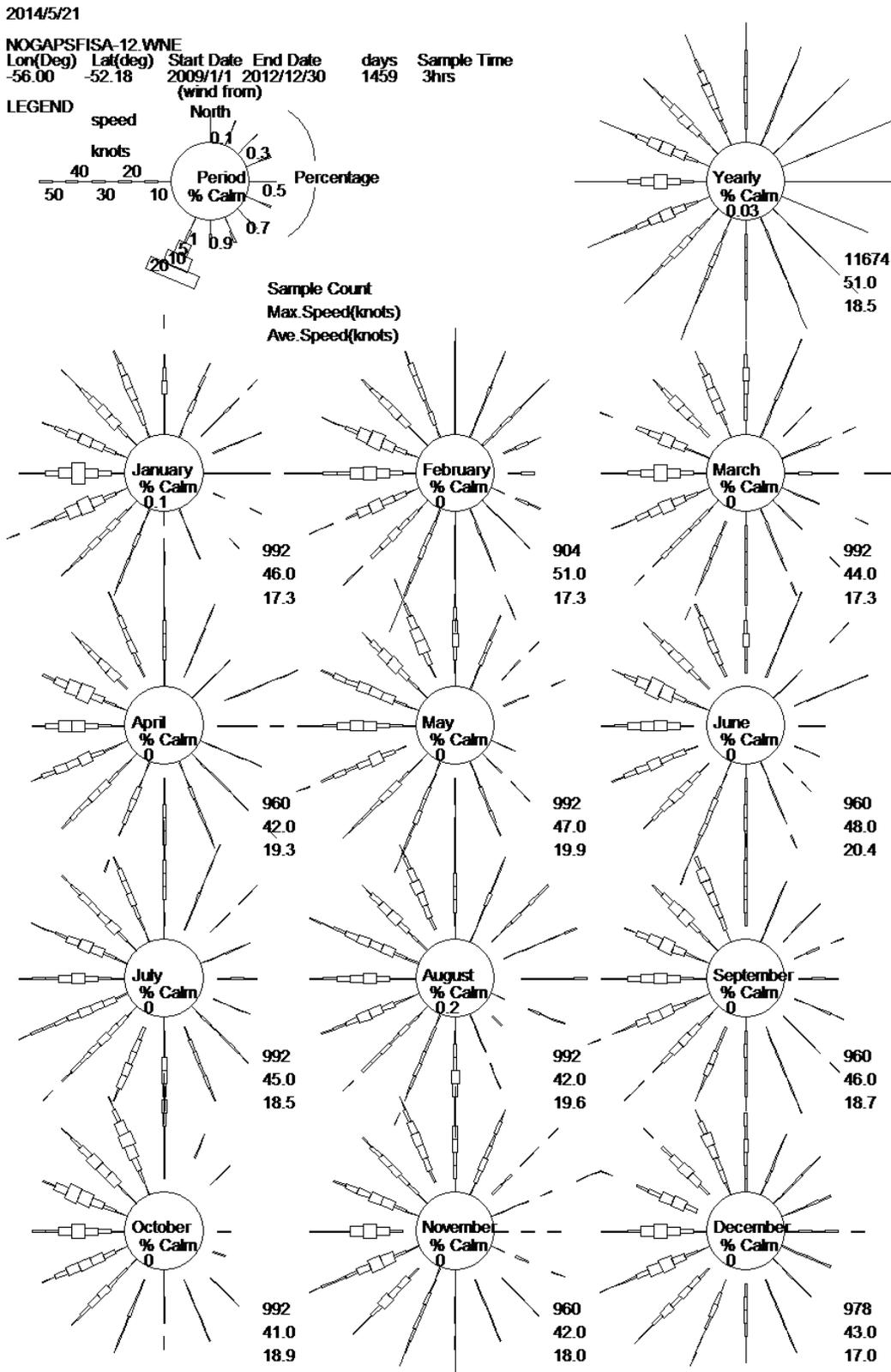


Figure 7. NOGAPS monthly and yearly wind roses for the northern grid point in FISA12 block offshore Falklands. Wind speeds are in knots, using meteorological convention (i.e., direction wind is coming from).

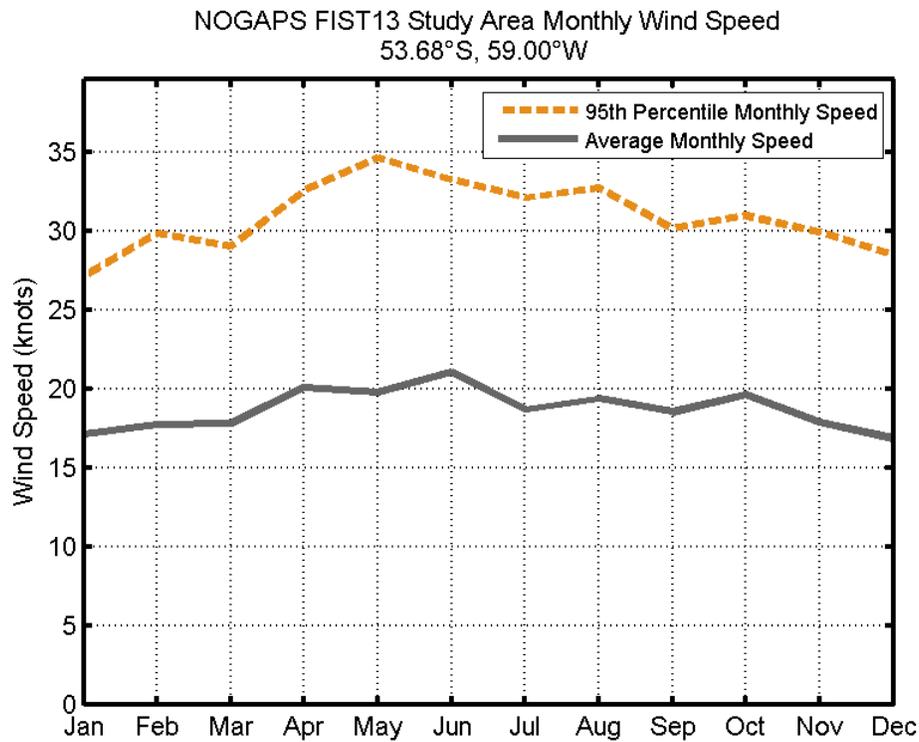


Figure 8. NOGAPS wind speed statistics: monthly average (grey solid) and 95th Percentile (orange dashed) wind speed in the FIST13 block offshore Falklands for the period of 2009-2012.

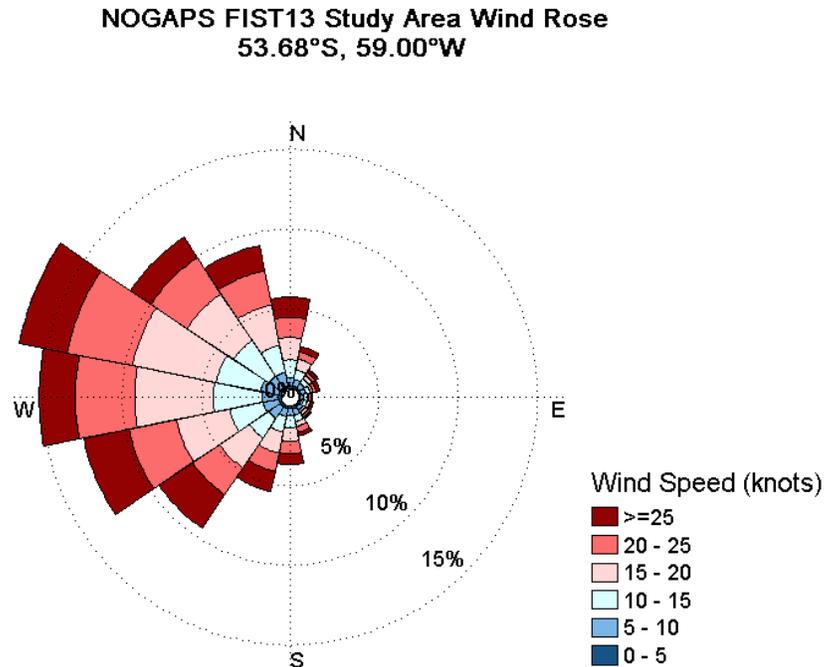


Figure 9. Yearly NOGAPS wind roses in FIST13 block offshore Falklands. Wind speeds are in knots, using meteorological convention (i.e., direction wind is coming from).

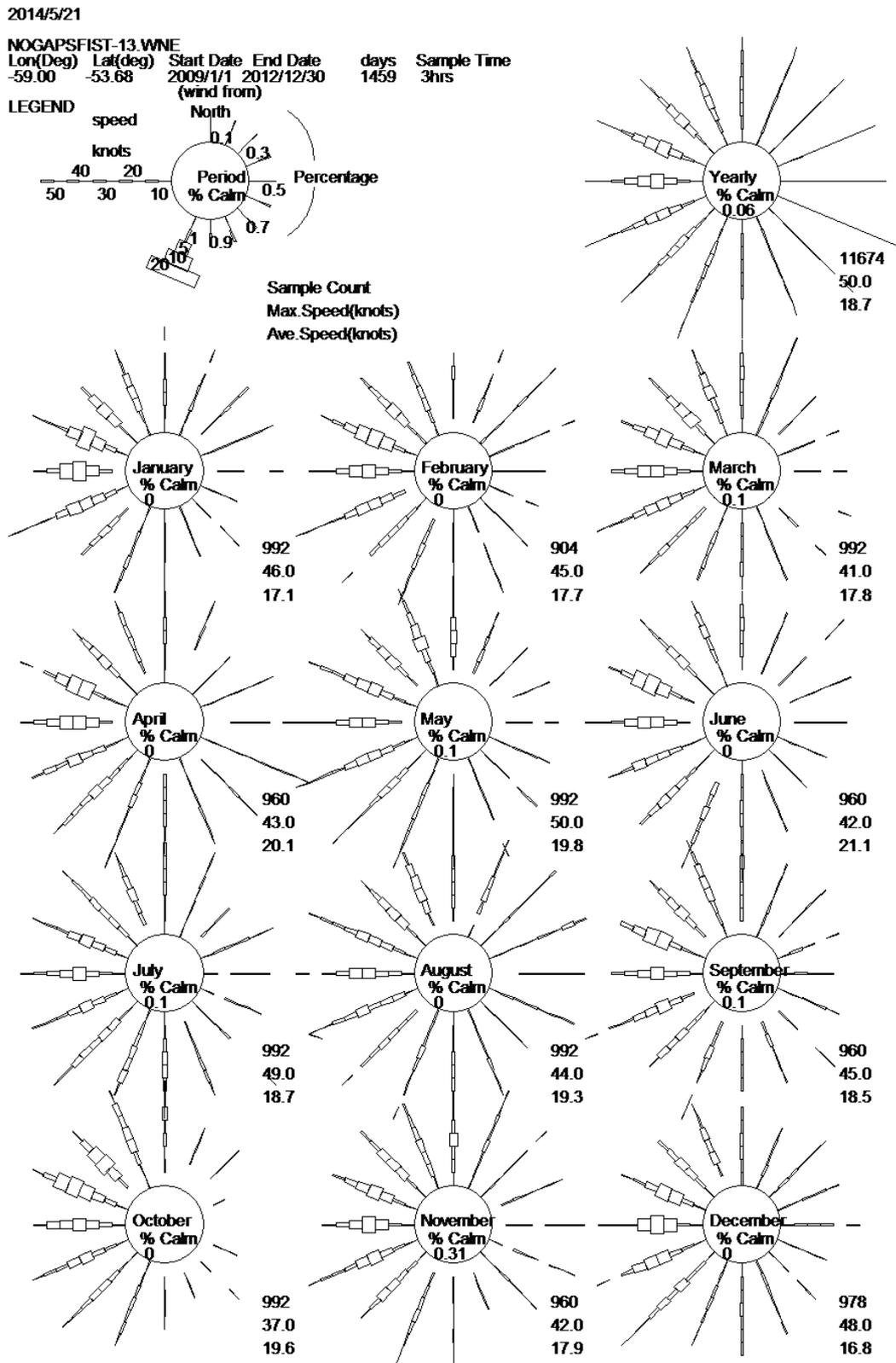


Figure 10. NOGAPS monthly and yearly wind roses for the northern grid point in FIST13 block offshore Falklands. Wind speeds are in knots, using meteorological convention (i.e., direction wind is coming from).

2.4. Ocean Circulation and Physical Attributes

The Falkland Islands consist of two large Islands (East and West Falkland) and at least 780 smaller islands and islets (Otley, 2008). Circulation off the Falkland Islands is primarily influenced by the Falkland, or Malvinas, Current (Figure 11). The Falkland Current is strong, relatively fresh (typically between 33.5-34.5 psu), and cold with sea-surface temperatures ranging from around 2-6°C in austral winter, to 10-14°C during austral summer (Brandini et al., 2000). The Falkland Current is a branch of the Antarctic Circumpolar Current (ACC) that flows northwards along the continental shelf of Argentina until it reaches the Brazil Current. The Brazil Current is a poleward (southward) flowing current that branches off from the South Equatorial Current around 8-10°S.

The Antarctic Polar Front (APF) forms the boundary between the cold Antarctic surface water and the warmer-sub-Antarctic water (Otley, 2008). This convergence zone marks the location where Antarctic surface waters moving northward sink below sub-Antarctic waters (Moore, 1999). Although the boundaries vary, typically this occurs between 50-60°S and is defined by water at 200 m depth with temperatures around 2°C (Park et al., 1993). The water between the APF and the sub-Antarctic Front is defined as the Antarctic Polar Frontal Zone (APFZ), which lies to the south and southeast of the Falkland Islands. The ACC diverts sharply northwards after Cape Horn and at the Burdwood Bank and the Falkland Islands splits into two branches. The larger current branch passes to the east of the islands and propagates northward along the continental shelf. North of the Falkland Islands, an anti-cyclonic (counter-clockwise) ring typically forms. The East and West Falkland Currents meet again around 49°S, where the current continues northward roughly parallel to the 200 m isobath around 0.5 knots and 100 km width.

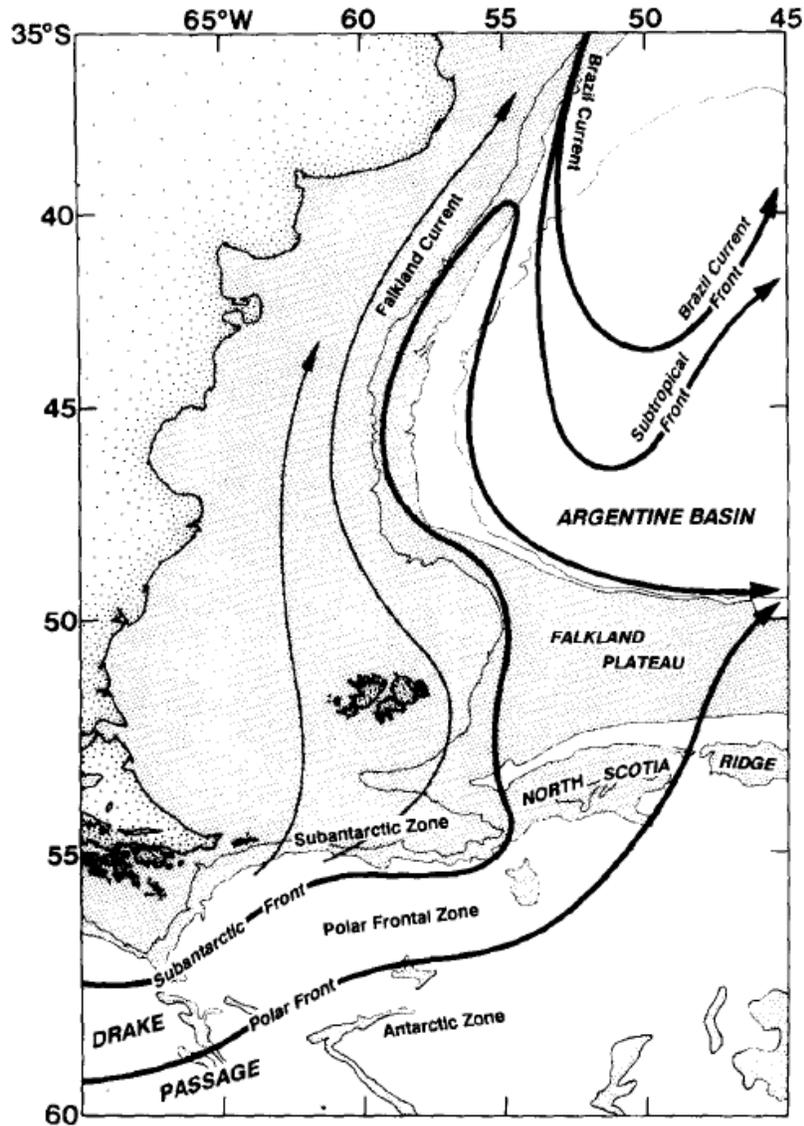


Figure 11. A schematic of the large-scale geostrophic currents and fronts in the southwestern South Atlantic Ocean. Depths less than 3,000 m are shaded (Source: Peterson and Stramma, 1990).

When the warm Brazil Current reaches around 33-38°S, it collides with the cold, northward flowing Falkland Current, thus causing a sharp temperature and salinity gradient and diverging both currents offshore (Otley, 2008). The Brazil Current is then deflected to the east into the region known as the Brazil-Malvinas Confluence Zone (Brazil-Falkland Confluence Zone), which is one of the most energetic regions in all the oceans (Figure 12) (Sarceno et al., 2004). The confluence varies meridionally, and is farthest north during austral winter and spring. This seasonality is presumed to be related to the general seasonal shift of wind systems and seasonal meridional shift of the subtropical gyre (Peterson and Stramma, 1990). After the Brazil Current reaches the confluence region, it bifurcates into two branches; one turning northward forming a recirculation cell, while the other continues southward and veers northeast around 45°S to become the South Atlantic Current (Boebel et al., 1999). At this confluence zone, the northward flowing Falkland Current retroflects cyclonically back toward the south, while the lesser portions continue north along the inner portions of the shelf. On the eastern side of the cyclonic

trough is the combined southward flow of the Falkland and Brazil Current that extend to about 45°S before turning east and north to form the poleward limits of the subtropical gyre. The Falkland waters continue south to the southern margin of the Argentine Basin (49°S) before turning east with the circumpolar regime (Campos, 1995). The warm-core eddies formed from the poleward extension of the Brazil current can be up to 300-400 km in diameter, and they transport heat and salt to the northern realm of the ACC.

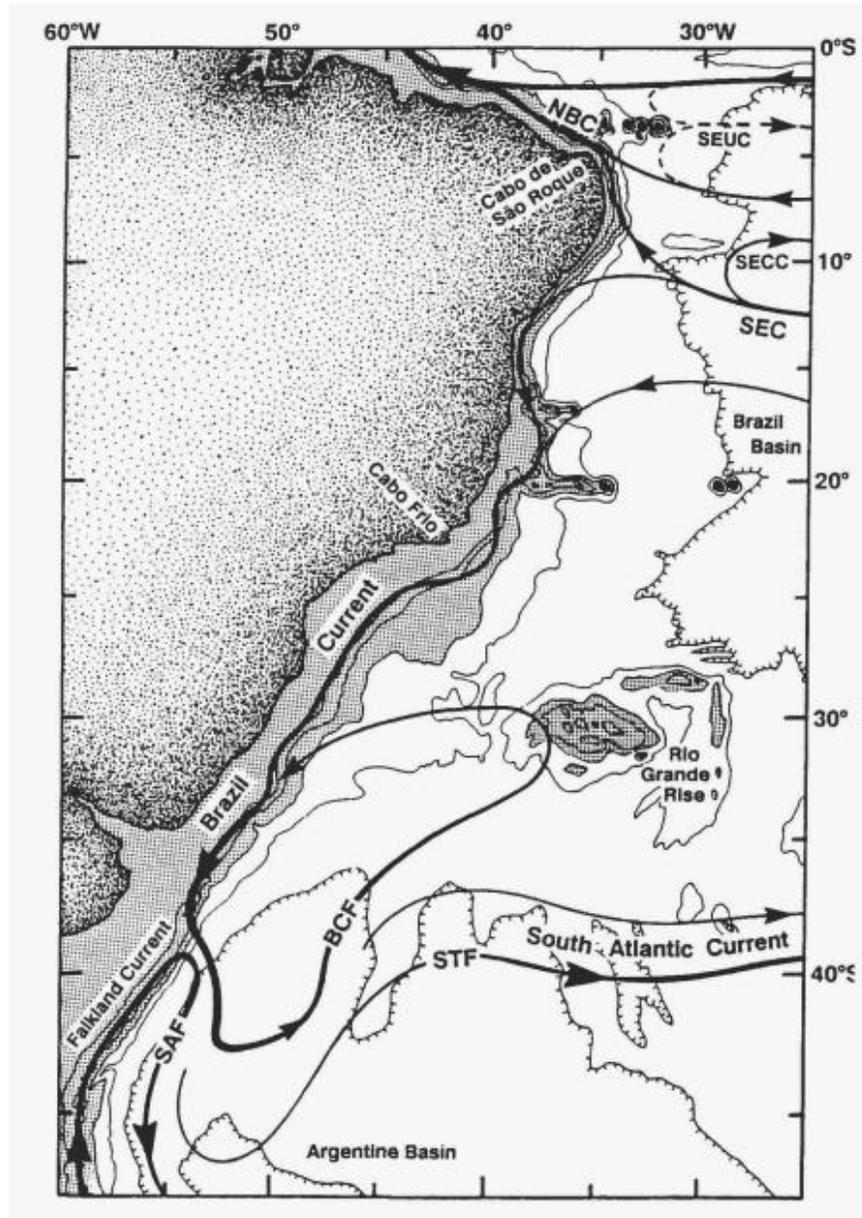


Figure 12. Schematic of flow in the South West Atlantic. At mid-latitudes are the Brazil and Falkland (or Malvinas) Currents along the western boundary and the South Atlantic Current in the interior. Equatorial current systems are also indicated. The bathymetry contour interval is 1,000 m (Source: Campos, 1995).

There are few direct measurements of the velocity of the Falkland Current. Peterson (1992) deployed surface drifters in the Falkland Current and found speeds around 40 cm/s. The along-shelf flow of the Falkland Current is variable year to year with no distinct seasonal cycle. However, the cross-shelf flow exhibits an annual cycle that is associated with the position of the sub-Antarctic front (Gyory, 2013, Vivier and Provost, 1999).

A 2009 FUGRO report and accompanying data has been provided to ASA by the client. The data consist of real-time current measurements from two Acoustic Doppler Current Profilers (ADCP) that were collected offshore the Falkland islands between December 6th, 2008 and October 21st, 2009. The two data collection locations and time periods are listed in Table 2 (Figure 13). The site locations were chosen in relation to the Falklands (Malvinas) Current. Loligo was positioned in deeper water that was in the predominantly northward flow of the Falklands Current. This site experienced higher current speeds than Toroa (about 25% higher) and several eddy events that lasted from days to weeks. The more southern site, Toroa, was positioned to the southwest of Loligo in shallower water along the shelf slope. Here, there were weaker currents, smaller scale eddy events and some influence from tidal current streams. At Loligo, the current direction was predominantly north to northeast, while Toroa was primarily northeastward. At both sites, the water column was well-mixed and had little annual variation in temperature (FUGRO, 2009).

Table 2. ADCP locations, depth and time period.

| Site Name | Latitude | Longitude | Approximate Water Depth (m) | Time Period |
|-----------|------------|------------|-----------------------------|--------------------------|
| Loligo | 51.2372° S | 54.6978° W | 1,421 | 2008-12-09 to 2009-10-21 |
| Toroa | 53.0327° S | 57.9527° W | 691 | 2009-05-02 to 2009-10-20 |

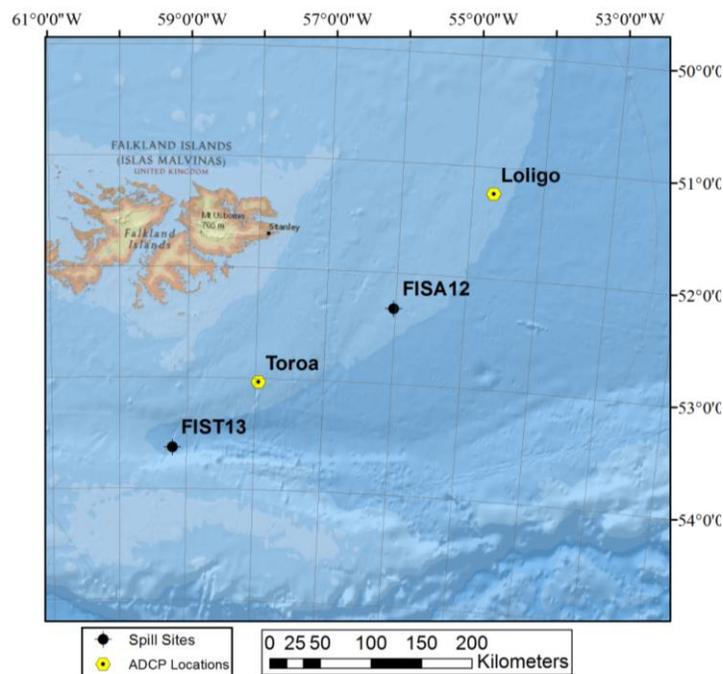


Figure 13. ADCP locations and FISA12 and FIST13 center points.

2.5. Current Dataset – HYCOM Hindcast

For this study, regional currents for the area were obtained from a hindcast analysis using inputs from the HYCOM (HYbrid Coordinate Ocean Model) 1/12 degree global simulation assimilated with NCODA (Navy Coupled Ocean Data Assimilation) from the U.S. Naval Research Laboratory (<http://www.hycom.org>). The model domain has a spatial resolution defined by a 1/12 degree grid in the horizontal direction and a daily temporal resolution, which for this study was obtained for the period of 2009-2012.

Comparisons between HYCOM data and supplied ADCP data (mentioned previously) were performed and demonstrated good agreement between the two data sources (Figure 14). Both the HYCOM and the ADCP data indicated the current speed increases to the north, as seen by the lower current speeds at the FIST13 site (Figure 15). The dominant direction is generally northeastward around 15-30 cm/s.

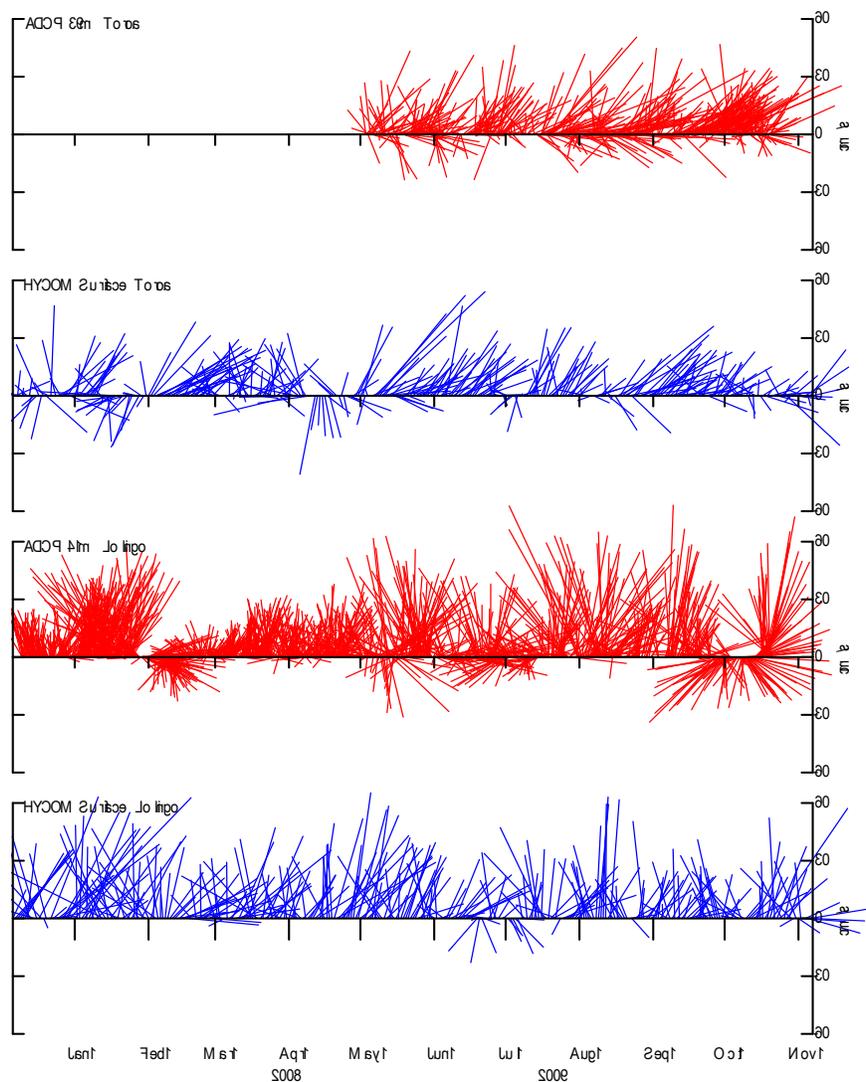


Figure 14. Comparisons of HYCOM and ADCP data at Toroa and Loliga locations.

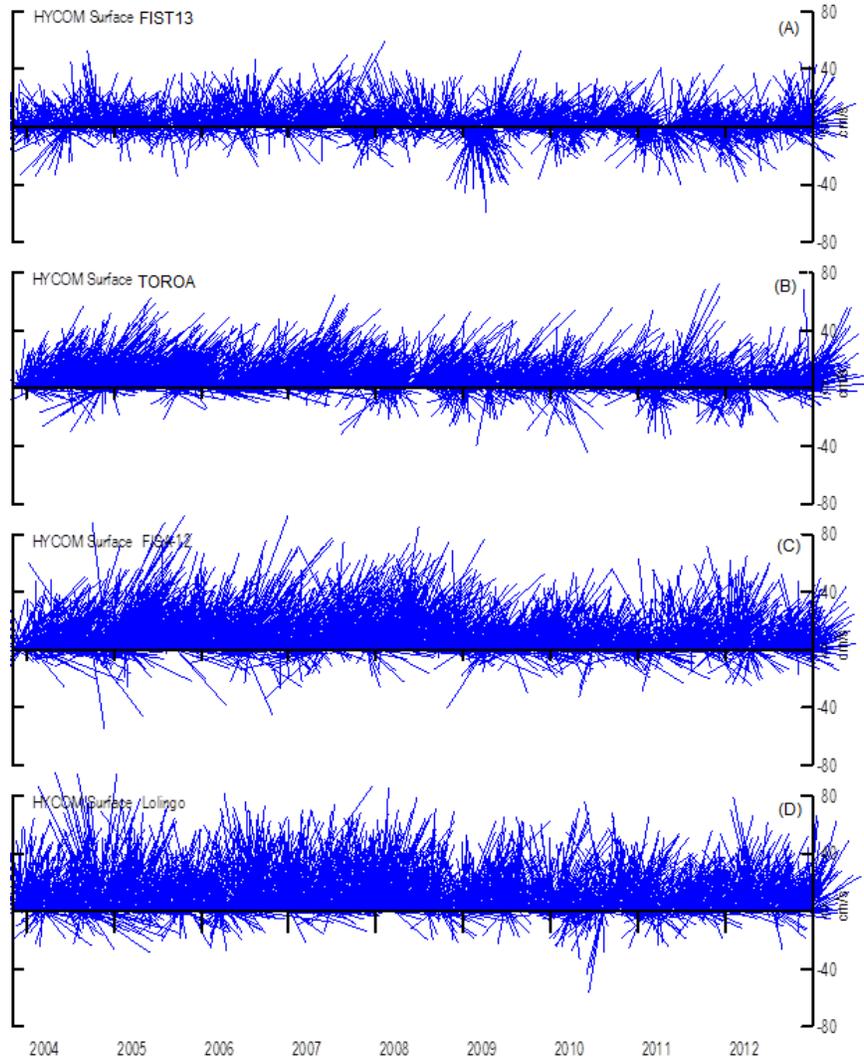


Figure 15. HYCOM surface velocities for FIST13 (A), TOROA (B), FISA12 (C) AND LOLIGO (D). Velocities are in cm/s.

2.6. Monthly Statistics

Figure 16 presents monthly statistics (average and 95th percentile) of the HYCOM current speeds at the surface and 1,500 m depth for a central site in FISA12. Throughout the period of interest, average monthly surface currents vary between 20-30 cm/s, while currents expectantly decrease with depth. Monthly averaged surface currents reach a maximum in August with average speeds around 30 cm/s, and a slightly lesser peak in June around 28 cm/s. The current decreases rapidly after its maximum to a minimum in October around 20 cm/s. Subsequently, current speeds increase to near 30 cm/s in January, with some variability in the following months. At depth, around 1,500 m, current speeds are fairly consistent, varying between 0.7-1.5 cm/s (Figure 16).

Figure 17 presents HYCOM monthly statistics in a central location of the southern block, FIST13. There are differences in the seasonality at this site based on the monthly statistics. In general, current speeds are lower and less variable at this site. The minimum occurs in March, around 14 cm/s, while the first maximum occurs in May around 20 cm/s, with a slightly smaller peak in October, near 19 cm/s. At depth, around 1,500 m, the current speeds are consistently around 2 cm/s throughout the year.

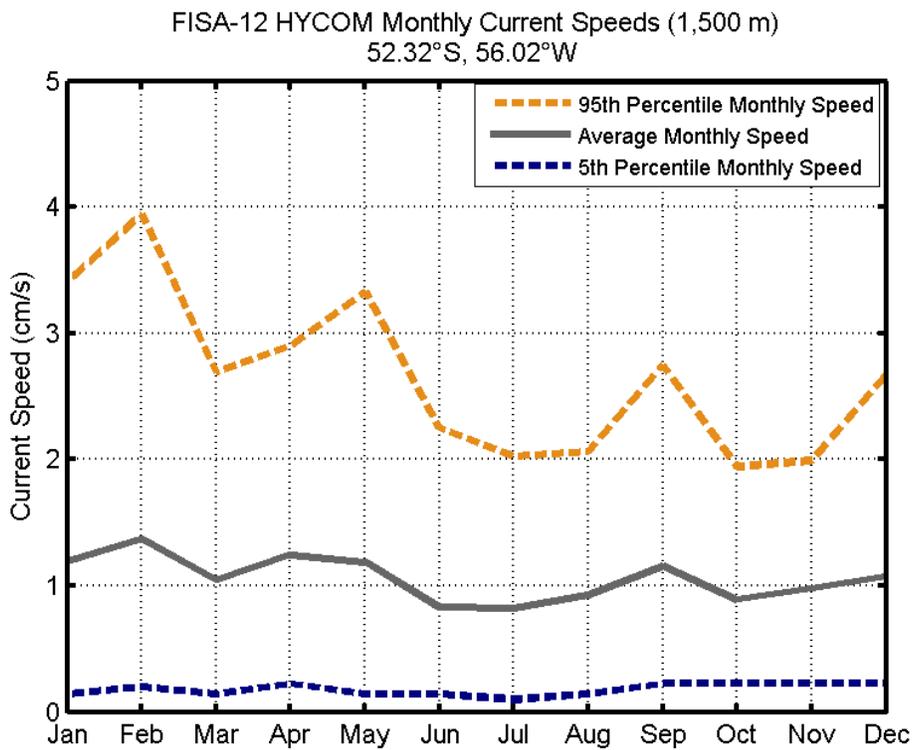
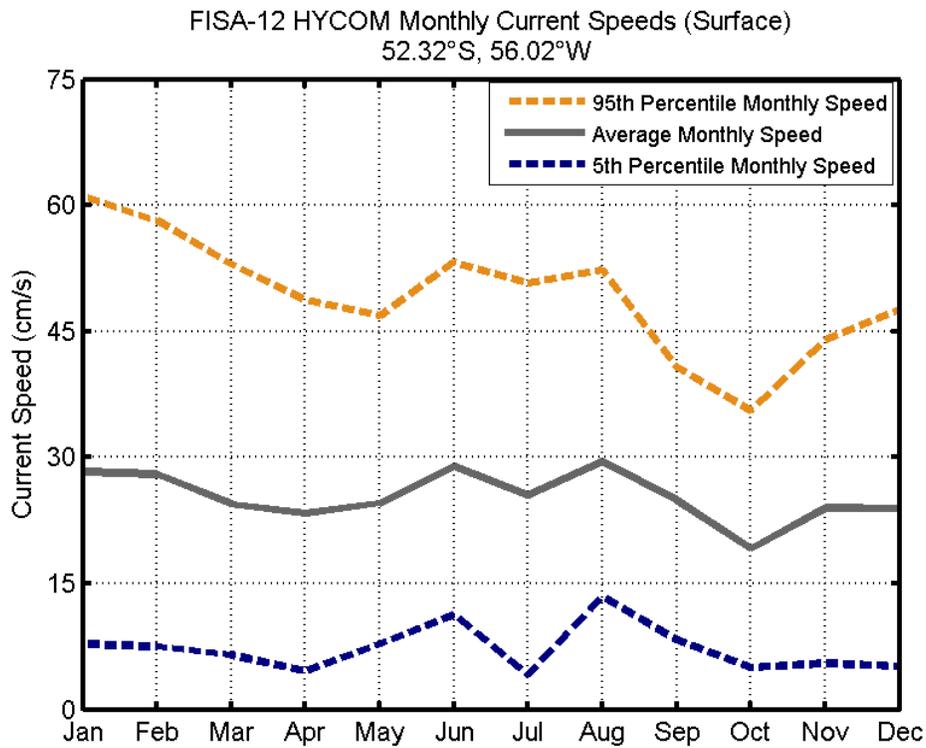


Figure 16. HYCOM current statistics: monthly average (grey solid) and 95th percentile (orange) current speed at the surface (top) and 1,500 m (bottom) in the center of the FISA12 block for the period of 2009-2012.

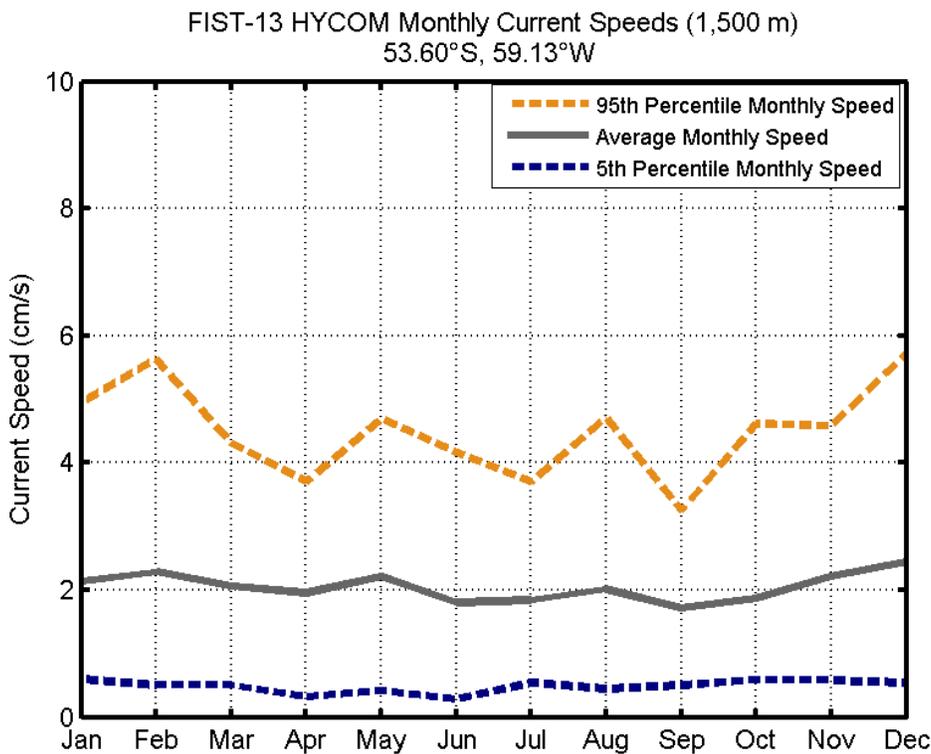
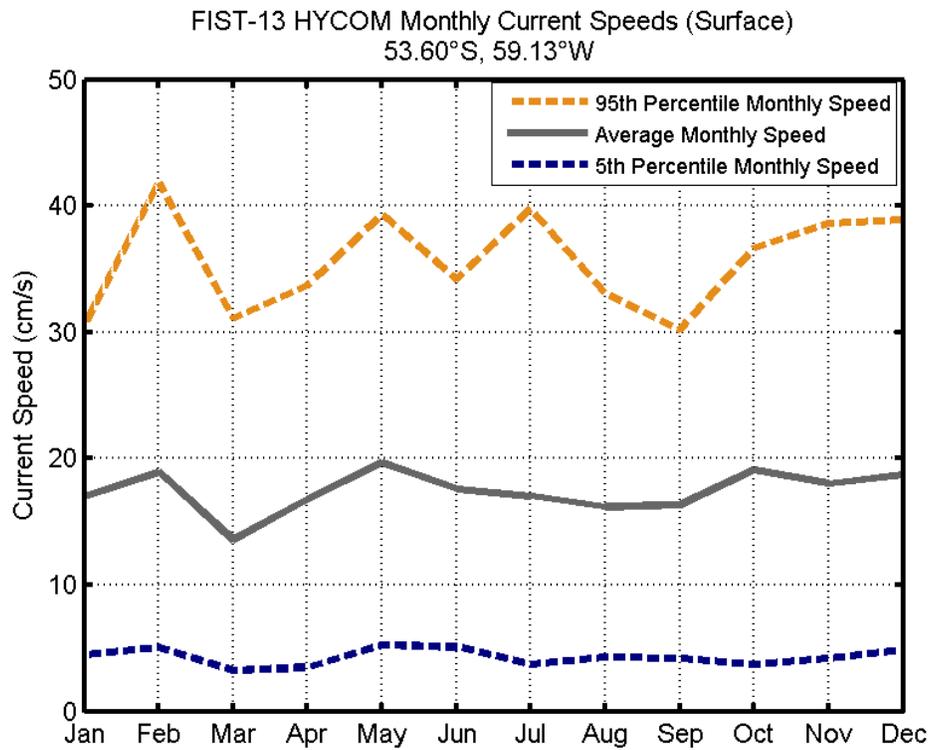


Figure 17. HYCOM current statistics: monthly average (grey solid) and 95th percentile (orange) current speed at the surface (top) and 1,500 m (bottom) in the center of the FIST13 block for the period of 2009-2012.

2.7. Current Roses

Figure 18 illustrates the spatial variability of the yearly average HYCOM current field represented by rose diagrams. Overall, current direction is primarily east-northeastwards. Current speeds are variable and differ depending on the location of interest. Offshore, there is a strong north-northeastward current with speeds frequently greater than 60 cm/s. Moving west towards Falklands, speeds are more commonly between 15-30 cm/s, and increase slightly as you near the eastern coast.

Figure 19 presents HYCOM current roses in the center of FISA12 averaged over the period of interest, for both the surface and 1,500 m water depths. The yearly surface current rose illustrates that the flow is predominantly northeastward between 15-30 cm/s. Currents at 1,500 m are typically less than 2 cm/s with a predominantly east-northeastward flow. Monthly current roses for the surface illustrate some slight seasonality in the area (Figure 20). Currents have a more northward component during summer through early fall (December-May). During winter and spring (June through October), direction is primarily to the northeast.

At the FIST13 site, direction is primarily northeastward with slightly more variability in direction, with a southeastward component observed as well (Figure 21). At depth, flow is dominated by both westward and eastward movement, with average speeds between 2-3 cm/s that can reach over 5 cm/s. The seasonality seen in the monthly surface current roses in Figure 22 illustrates changes in direction rather than speed. During mid-summer through early fall (January-April), the dominant flow is to the south. From May through winter and spring (June-October), the flow regime shifts to northeastward with speeds between 15-30 cm/s.

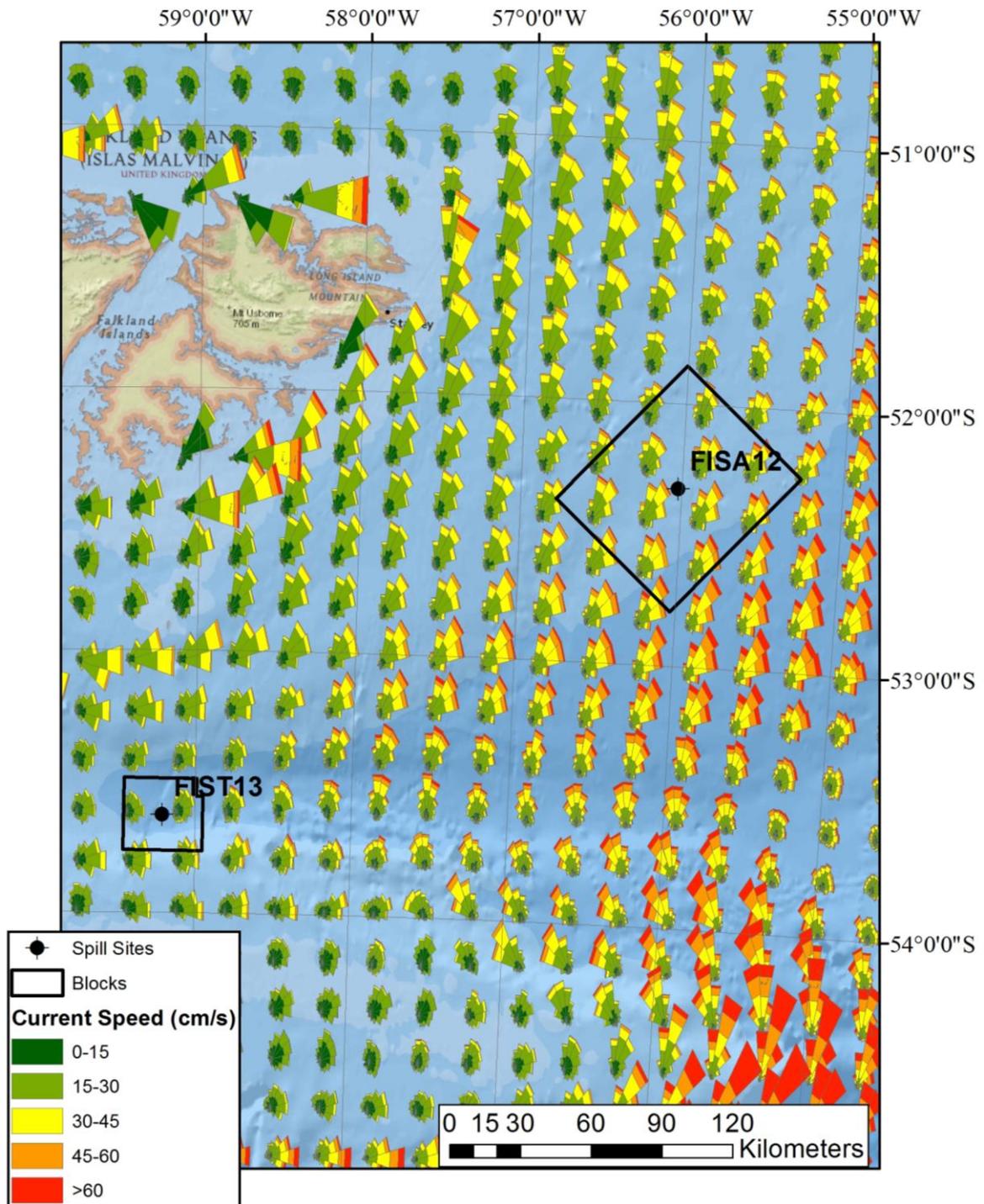
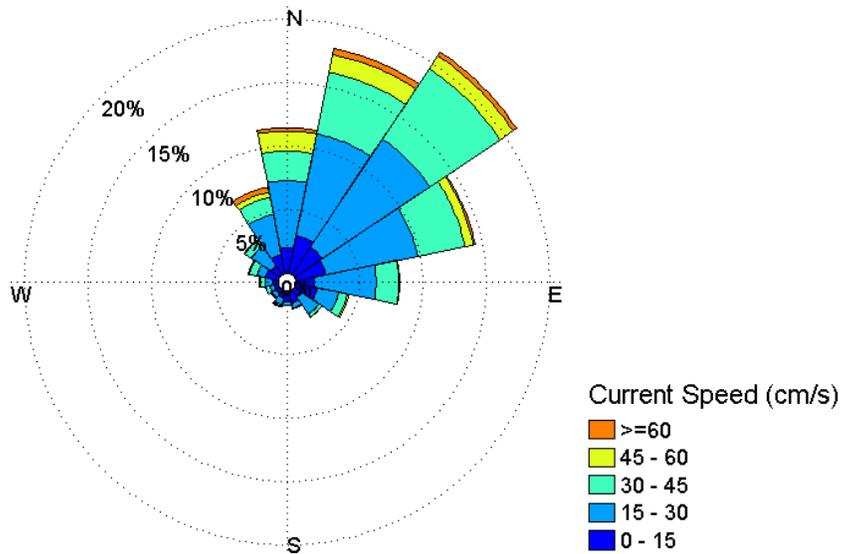


Figure 18. Spatial variability of the average HYCOM current field represented by rose diagrams; current speed is in cm/s, using standard convention (i.e. direction currents are going to).

HYCOM Current Rose near FISA12 (Surface)
52.32°S, 56.02°W



HYCOM Current Rose near FISA12 (1,500 m)
52.32°S, 56.02°W

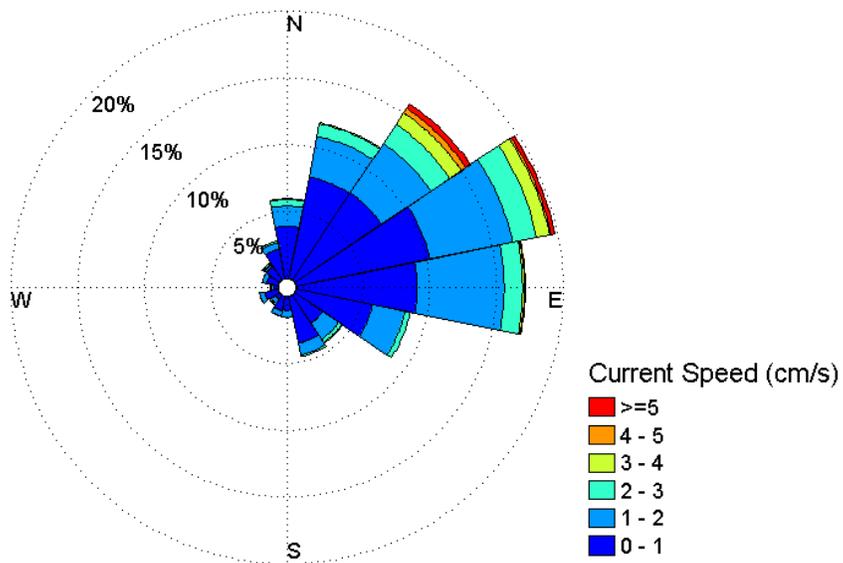


Figure 19. HYCOM current roses in the center of the FISA12 block averaged over the period of 2009-2012. Current roses presented for the surface (top) and 1,500 m (bottom) water depths. Direction convention is standard (i.e., direction currents are moving to).

Monthly current roses at FISA12 (Surface)
52.32°S, 56.02°W

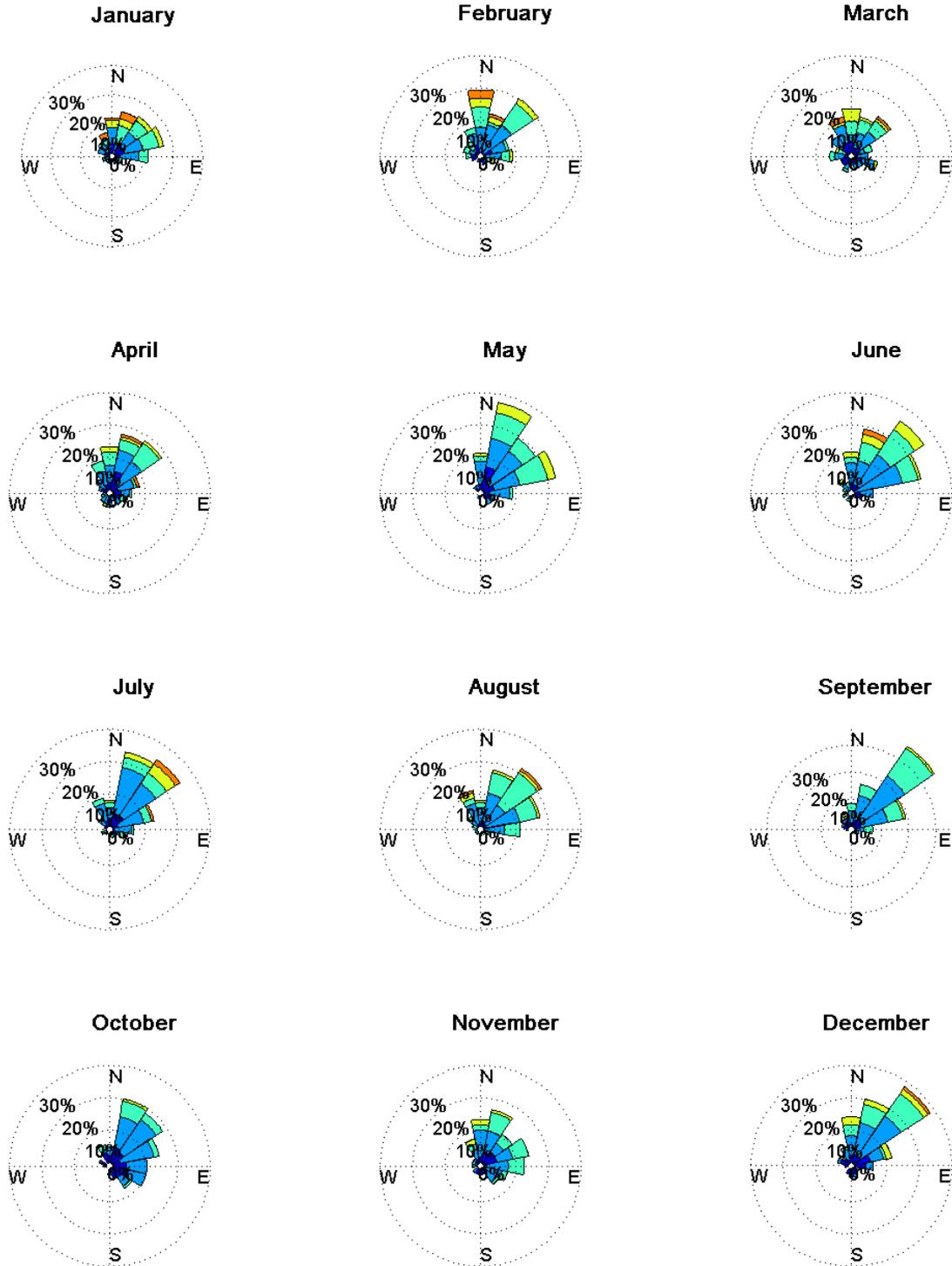
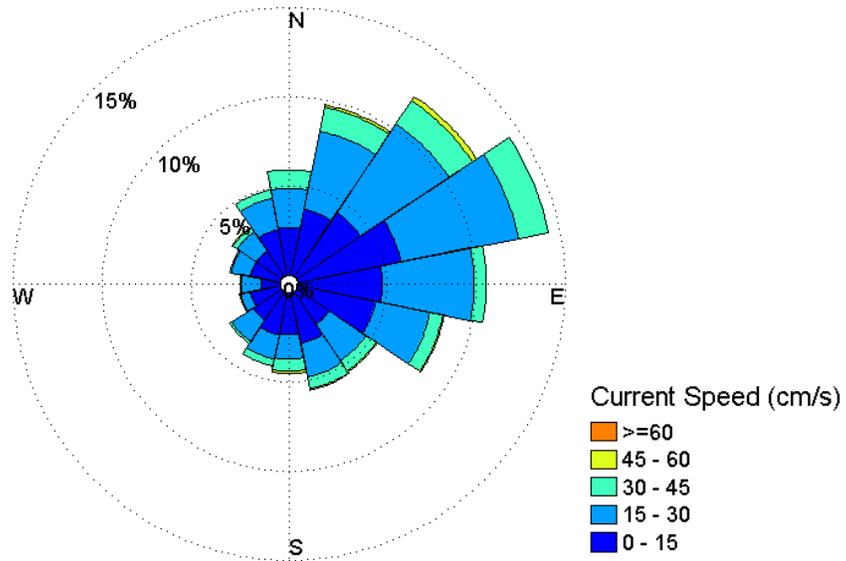


Figure 20. HYCOM monthly averaged surface current roses in the center of the FISA12 block averaged over the period of 2009-2012. Direction convention is standard (i.e., direction currents are moving to).

HYCOM Current Rose near FIST13 (Surface)
53.60°S, 59.13°W



HYCOM Current Rose near FIST13 (1,500 m)
53.60°S, 59.13°W

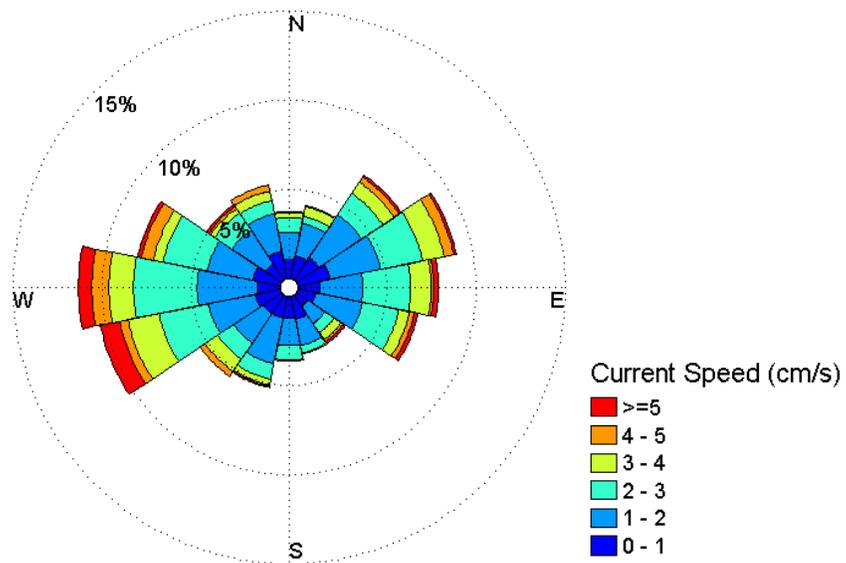


Figure 21. HYCOM current roses in the center of the FIST13 block averaged over the period of 2009-2012. Current roses presented for the surface (top) and 1,500 m (bottom) water depths. Direction convention is standard (i.e., direction currents are moving to).

Monthly current roses at FIST13 (Surface)
53.60°S, 59.13°W

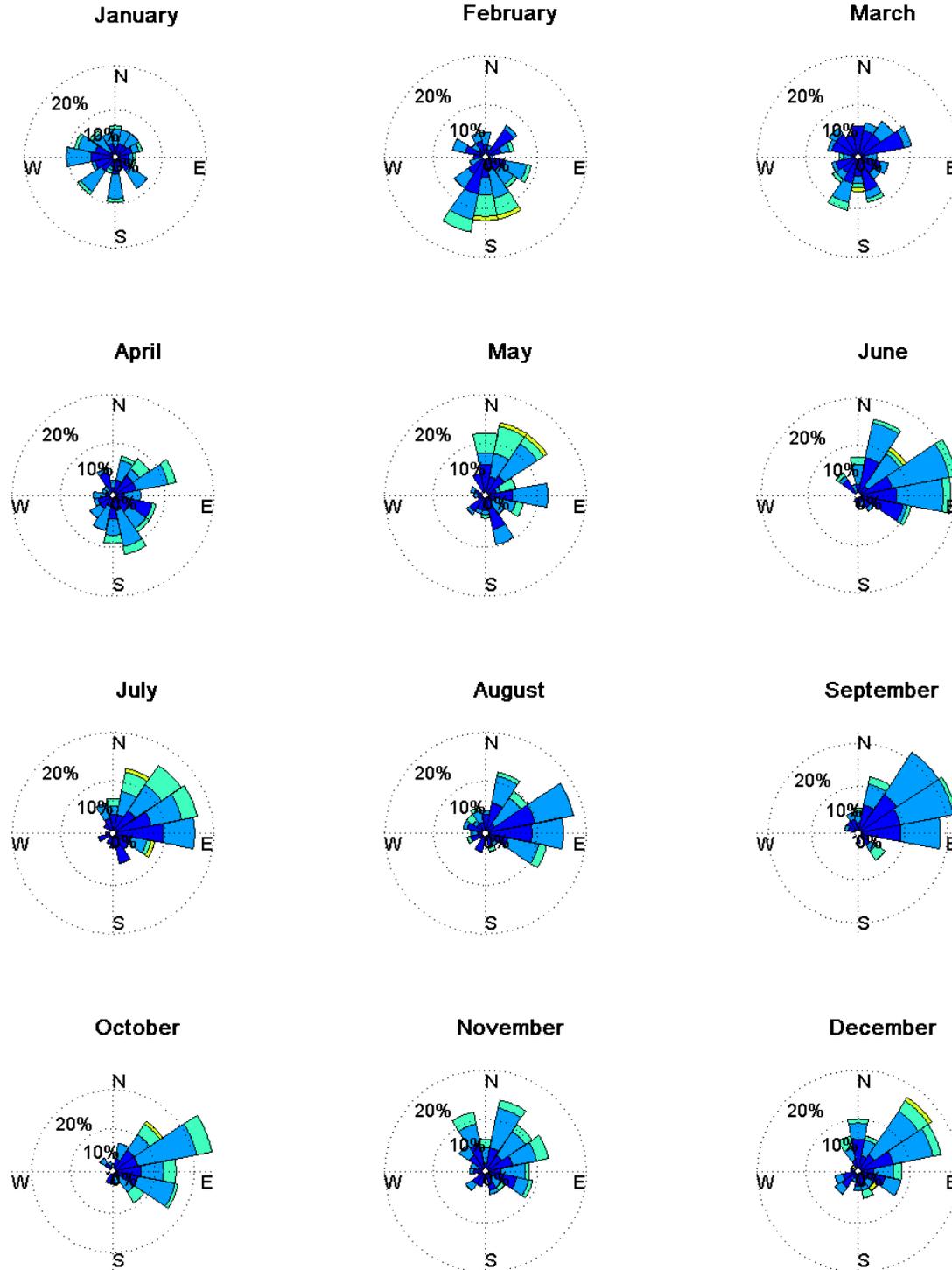


Figure 22. HYCOM monthly averaged surface current roses in the center of the FIST13 block averaged over the period of 2009-2012. Direction convention is standard (i.e., direction currents are moving to).

2.8. Mapped Current Fields

Figure 23 shows a spatial view of HYCOM mean surface currents for years 2009-2012. These figures clearly indicate the edge of the strong, eastward flowing branch of the ACC south of the areas of interest. As discussed above, the current that separates from the polar current to flow northward is known as the Falklands Current, and propagates northeastward through the FISA12 block. The FIST13 block does not appear to be directly impacted by the sub-polar northward current.

Around the coastal areas of the Falkland Islands, current speeds vary largely between 5-35 cm/s. The current direction around the southern part of the Islands is eastward, shifting northeastward as currents round the eastern side of the Islands. Current speeds near the coast are typically less than offshore. Monthly spatial maps for 2009-2012 are seen in Figure 24. Overall, the monthly averages look similar, though the northwards Falklands Current changes zonal position based on the season. It shifts westward, towards the Falkland Islands, during austral summer, which can cause an increase in current velocity through FISA12. Yearly averaged surface currents are seen in Figure 25 through Figure 28. Flow through the FIST13 block is primarily east-northeastward each year with consistent speeds. Within FISA12, current direction is primarily northeastward each year, with some slight changes in speed depending on the northward current location and if it passes directly through the block. After the current passes through the areas of interest, propagation is northward until around 49°S, where part of the current flows westward, while also branching eastward.

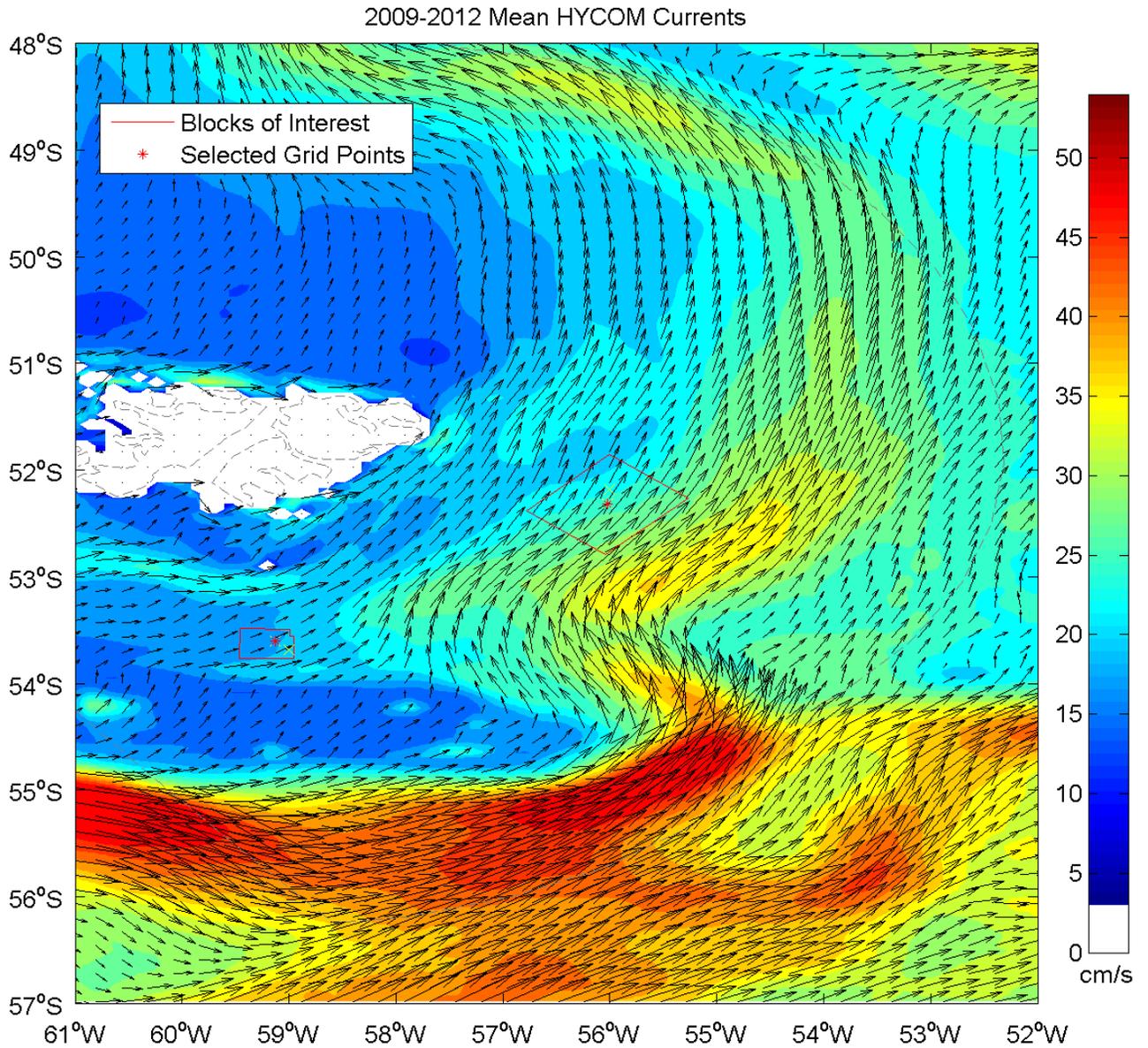


Figure 23. Mean HYCOM surface currents from 2009-2012. The blocks of interest are outlined in red and red stars indicate points selected for analysis.

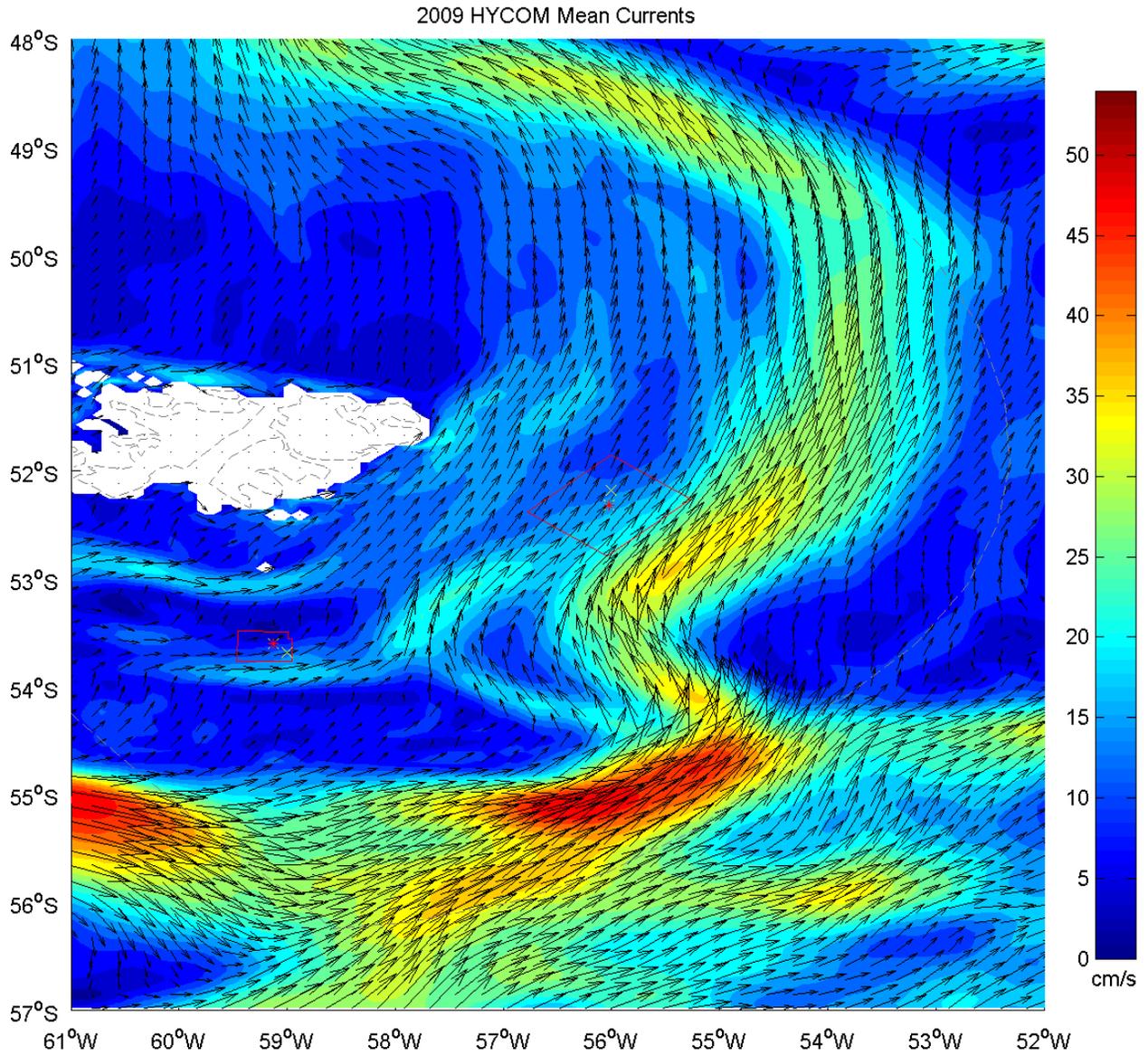


Figure 25. 2009 mean HYCOM surface currents. Blocks of interest are outlined in red. Red stars indicate points selected for analysis.

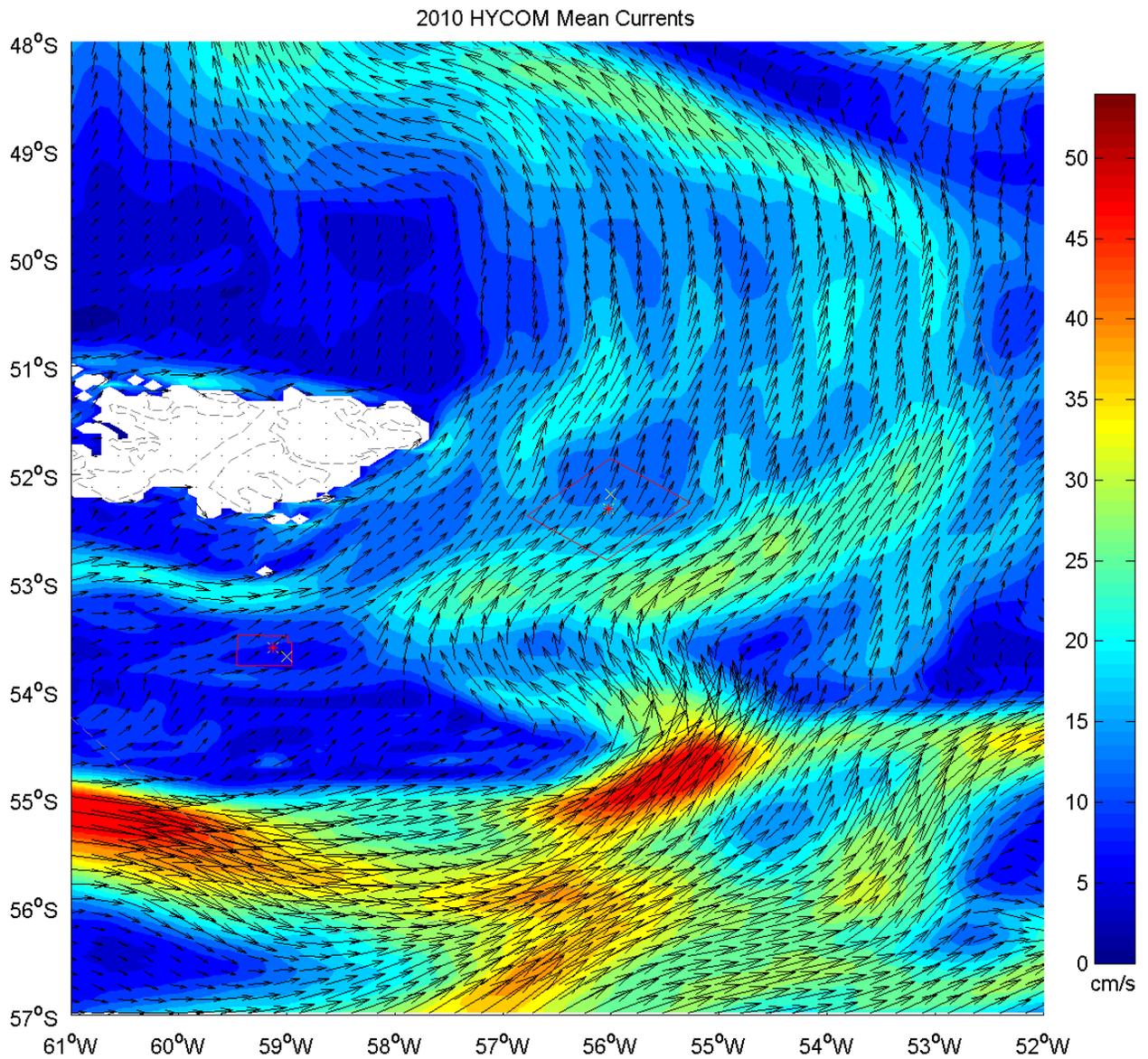


Figure 26. 2010 mean HYCOM surface currents. Blocks of interest are outlined in red. Red stars indicate points selected for analysis.

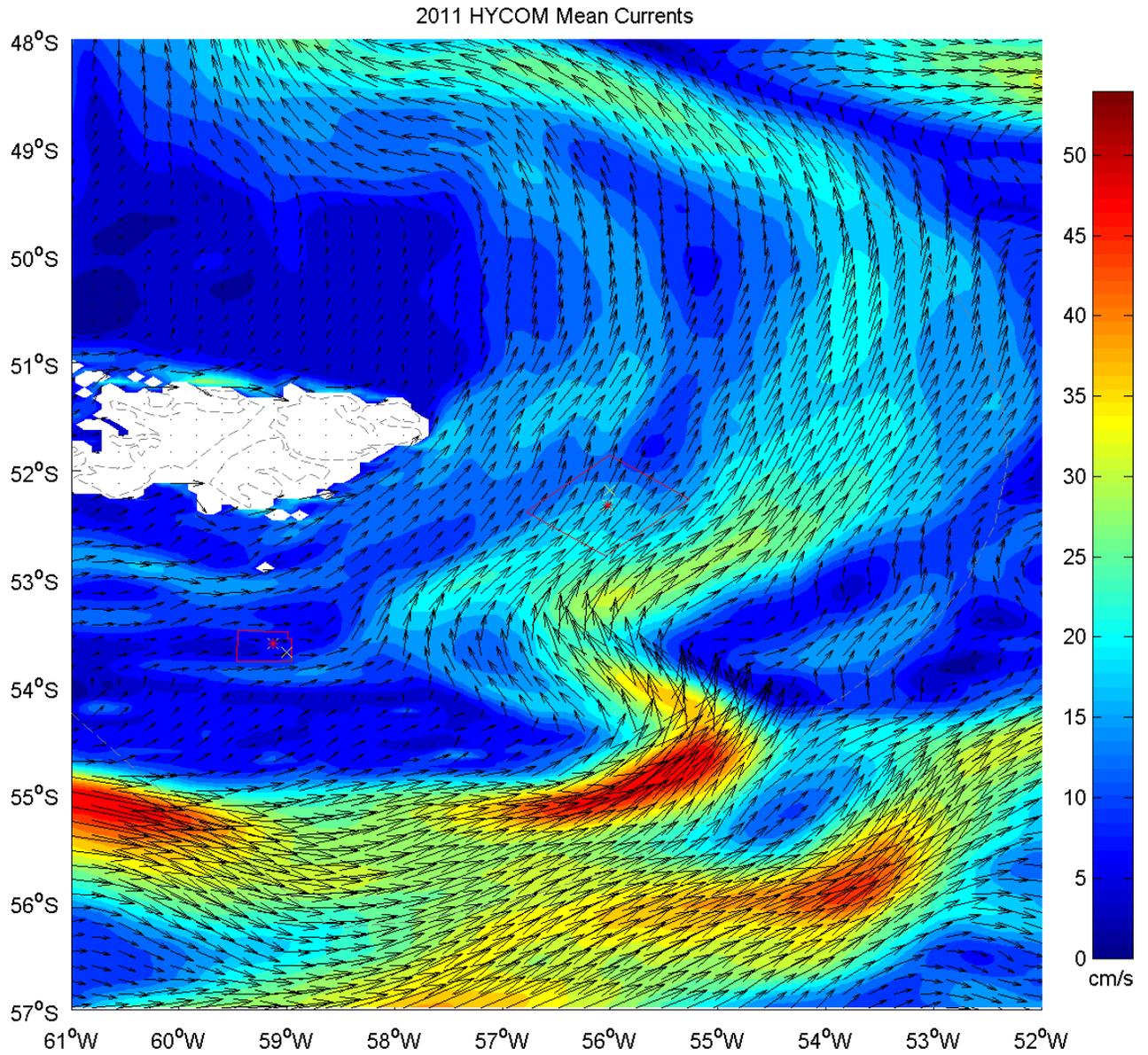


Figure 27. 2011 mean HYCOM surface currents. Blocks of interest are outlined in red. Red stars indicate points selected for analysis.

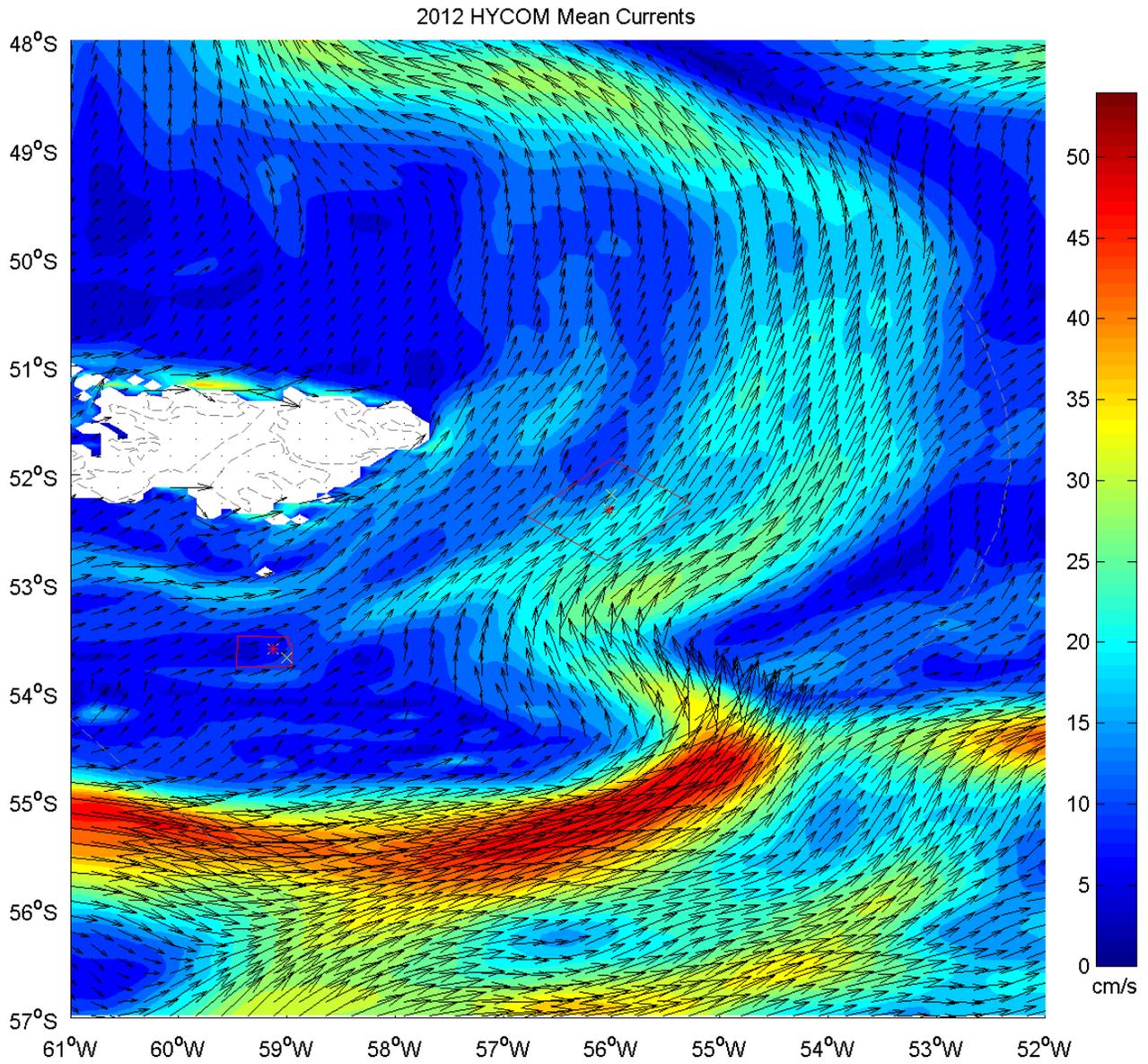


Figure 28. 2012 mean HYCOM surface currents. Blocks of interest are outlined in red. Red stars indicate points selected for analysis.

2.9. Water Column Vertical Structure

Figure 29 shows the vertical profile of currents obtained from the HYCOM model outputs for the area of interest averaged from 2009 through 2012 (FISA12 site). Near the surface, the current velocity can exceed 50 cm/s. On average, however, surface speeds are around 25 cm/s. Velocity decreases with depth to a minimum of about 1 cm/s near the seabed. In the FIST13 block, currents at the surface exceed 35 cm/s, but typically average 17 cm/s (Figure 30). Near the bottom, the current flows around 2-3 cm/s.

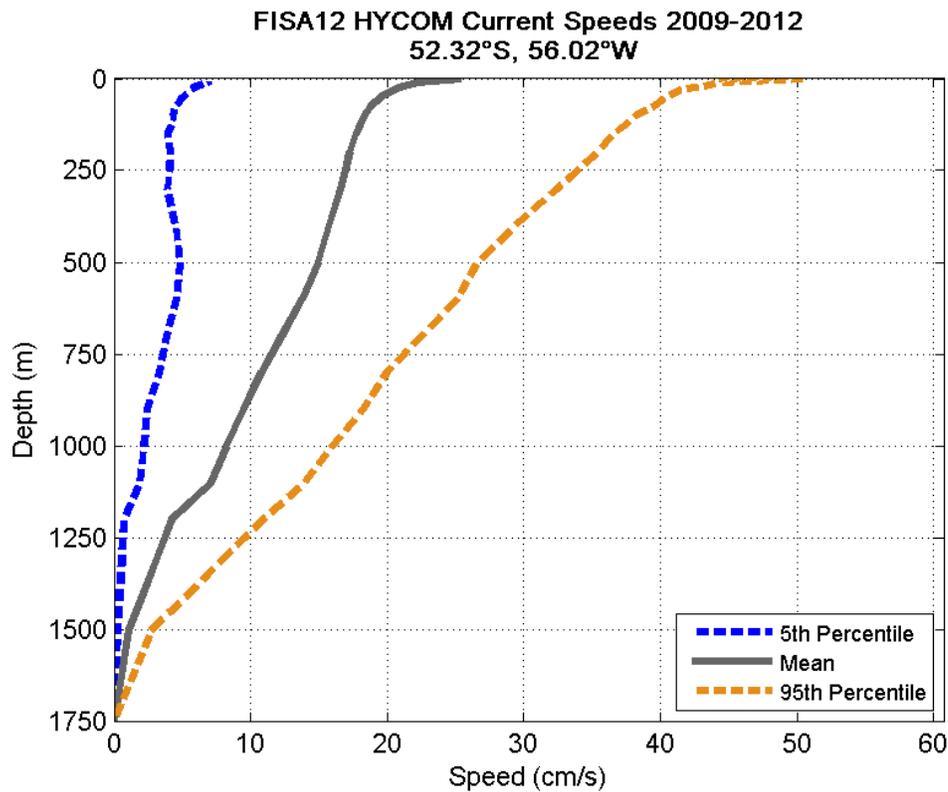


Figure 29. HYCOM 5th percentile (blue), average (solid grey), and 95th percentile (dashed orange) current speed with depth in FISA12 block for the period of 2009-2012.

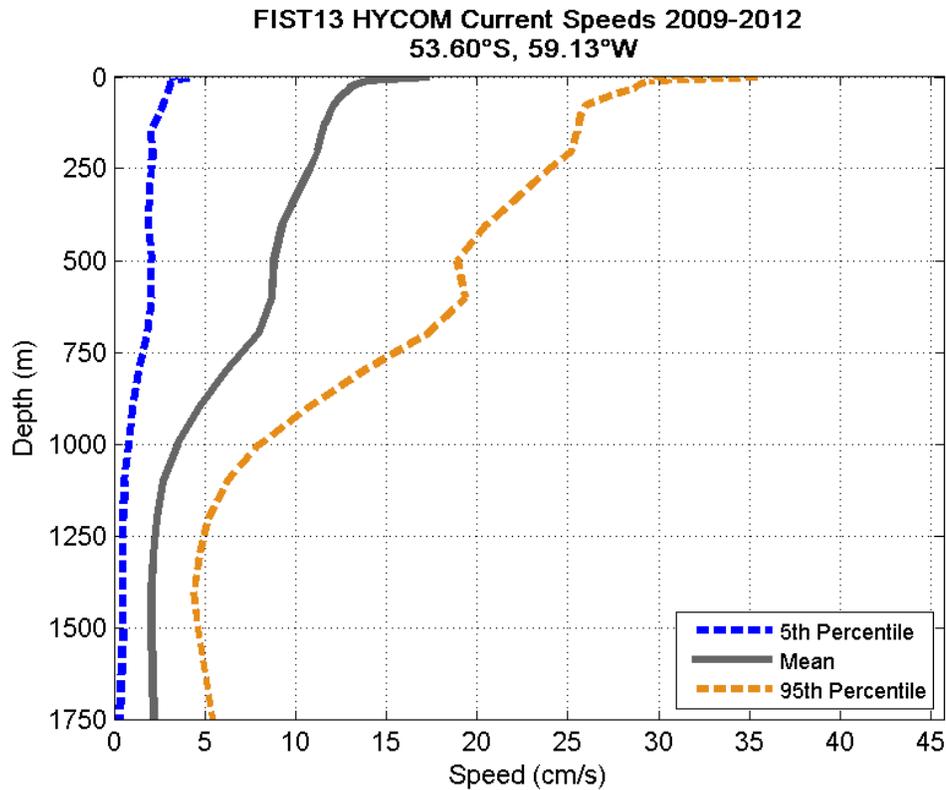


Figure 30. HYCOM 5th percentile (blue), average (solid grey), and 95th percentile (dashed orange) current speed with depth in FIST13 block for the period of 2009-2012.

Figure 31 and Figure 32 illustrate the yearly-averaged vertical profile of temperature, salinity, and density near the locations of interest. Temperature and salinity data were obtained from the World Ocean Atlas NODC NOAA data product (Levitus, 1982). Surface temperatures at the FISA12 site are approximately 6.5°C and decline gradually before reaching a minimum of approximately 3°C at depth. Salinity values are steady, with an average of about 34 psu at the water surface and increase slightly to 34.25 psu at depth. Density is around 26.7 kg/m³ at the surface and increases to 28 kg/m³ at depth. At the FIST13 site the water column properties are similar, with surface temperatures are near 6.5°C at the surface and less than 2.5°C at depth. Salinity values are around 34 psu at the surface and increase to over 34.5 psu at depth. Density is around 26.7 kg/m³ at the surface and increases to 28.5 kg/m³ at depth.

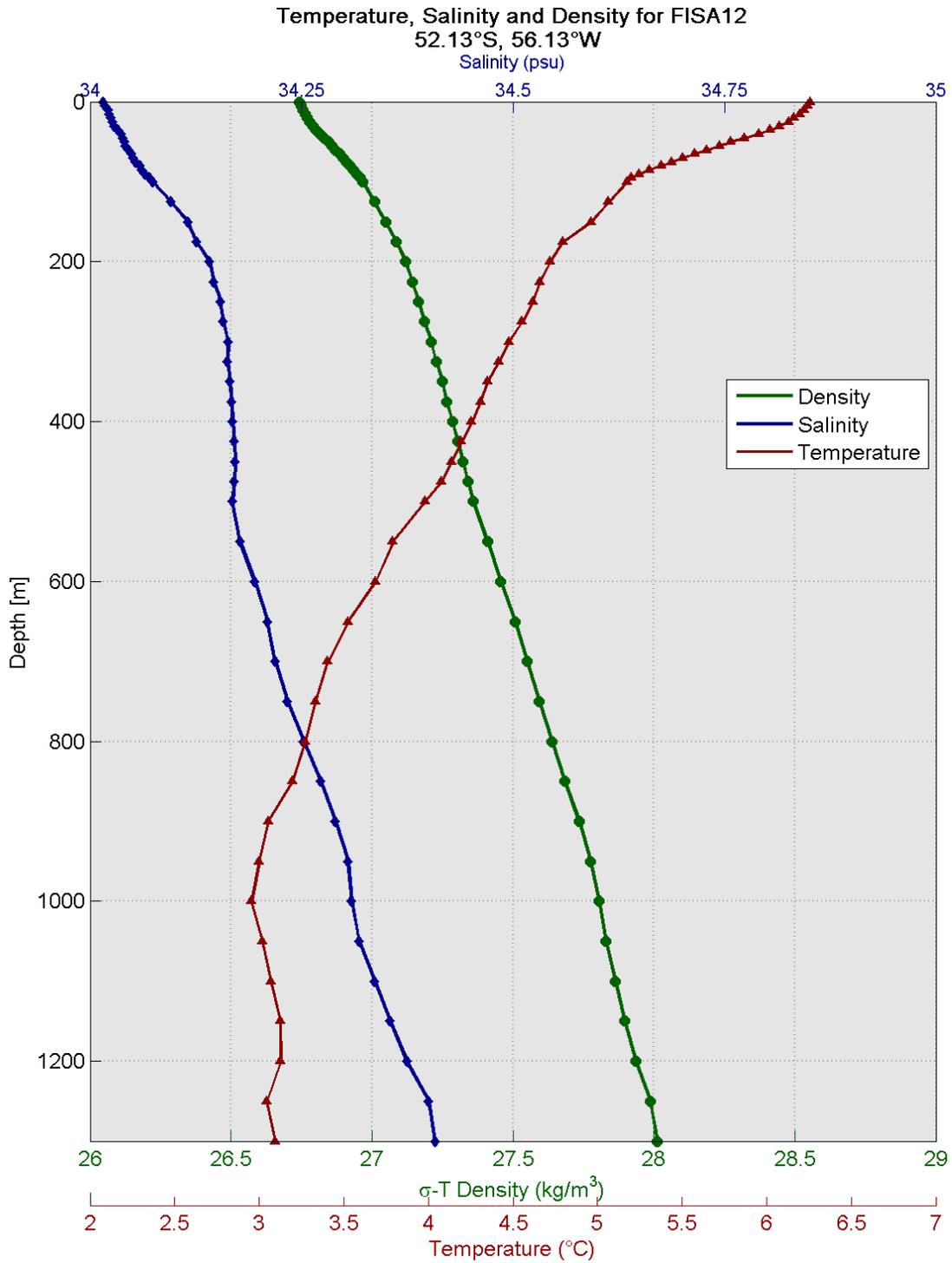


Figure 31. Yearly averaged temperature and salinity vertical profiles in the center of the FISA12 block.

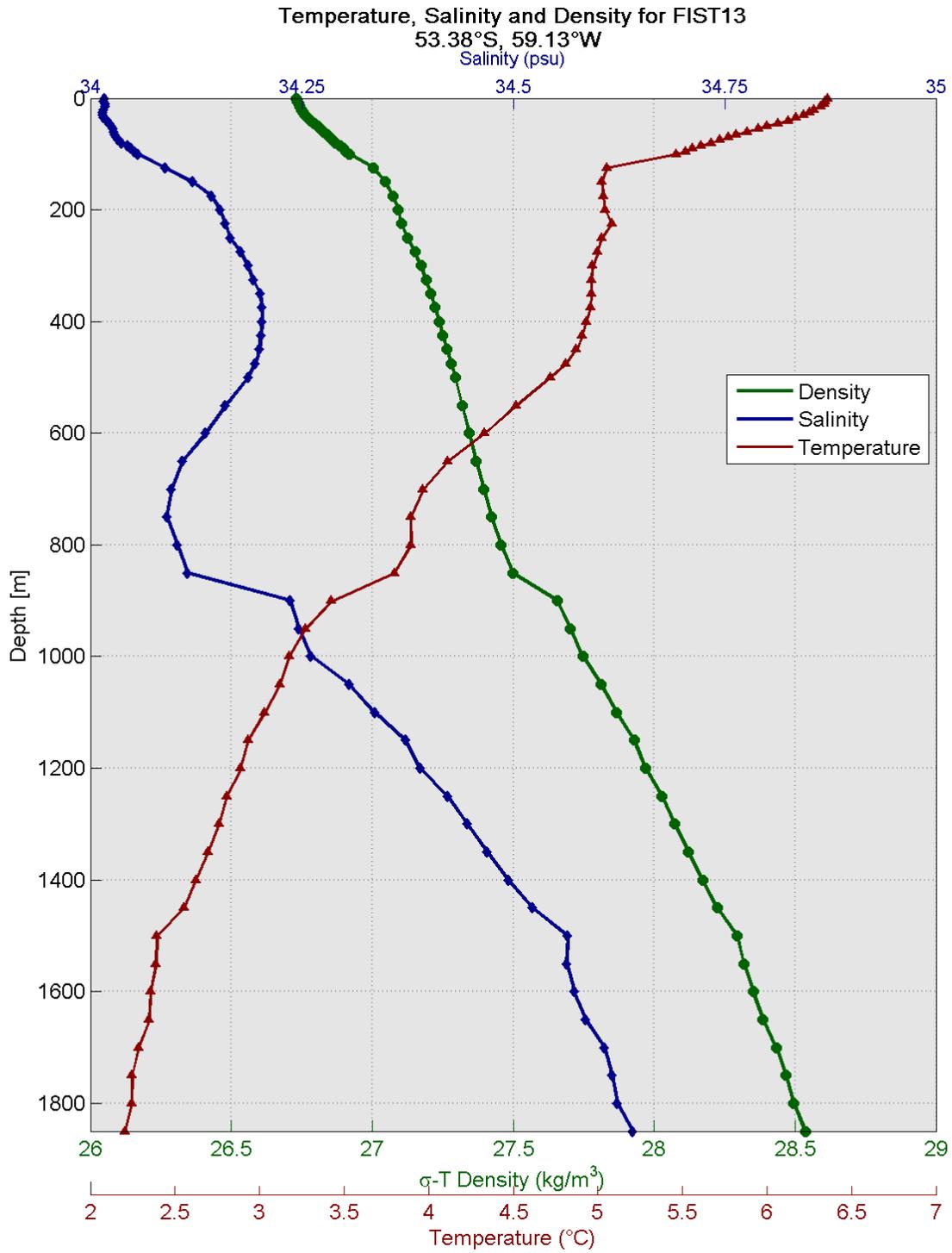


Figure 32. Yearly averaged temperature and salinity vertical profiles in the center of the FIST13 block.

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Appendix M: MMO Monitoring Reports

Protected Species Monitoring Report, FISA12 and FIST13, 29th November 2012 – 4th June 2013 (excluding Appendices).M-3 to M-39.

Protectde Species Observer Report, FINA13, 5th November 2013 – 18th February 2014 (excluding Appendix C)..... M41 to M92



PROTECTED SPECIES MONITORING REPORT

Noble Energy Falklands Ltd.

PGS Ramform Sterling

PON3

**Falkland Islands Southern Phase A 2012 &
Falkland Islands Southern Tilt 2013 (FIST13)**

East Falkland Island Basin & Southern Tilt Falkland Island Basin

29 November 2012 – 4 June 2013

| | | |
|-----------------------|---|--|
| Project No. | UOS01285M | RPS |
| Report No. | 1 | 411 N Sam Houston Parkway E |
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1. EXECUTIVE SUMMARY

The following report details the protected species monitoring and seismic survey operations undertaken during the Falkland Islands Southern Phase A 2012 and Falkland Islands Southern Tilt 3D surface seismic surveys aboard the Ramform Sterling from 29 November 2012 through 4 June 2013. This survey was conducted by PGS Geophysical for Noble Energy Falkland Islands Ltd., operated within a permitted area as mentioned in Falklands Islands permit Petroleum Operations Notice 3 (PON3).

Two trained visual protected species observers (PSOs) and a dedicated passive acoustic monitoring (PAM) operator, contracted through RPS, were on board to fulfill the regulatory requirements and reporting mandated by the Joint Nature Conservation Committee. Mitigation measures were to be implemented to minimize potential impacts to marine mammals throughout the duration of the survey. Mitigation measures included the use of PSOs for both visual and acoustic monitoring, establishment of safety radii, and implementation of ramp-up delays if necessary.

The PAM system was utilized during night time hours as well as for exclusion zone clearance during periods of extreme fog as outlined within the PGS Geophysical/Noble Energy Falklands Limited Project Plan section 9.3 the Environmental Management Plan (EMP). Mitigation guidelines for this project are set forth in the Joint Nature Conservation Committee's current *JNCC guidelines for minimising the risk of injury and disturbance to marine mammals from seismic surveys*, August 2010.

On 8 December 2012, the vessel began airgun testing prior to commencing on the Southern Phase A survey on 10 December 2012. The Southern phase A survey was completed on 10 May 2013 at which time the vessel recovered all seismic gear and transited to port for crew change. Upon completion of crew change the vessel transited to the second survey area, the Southern Tilt 2013 survey. This survey began with gun testing just prior to production on 19 May 2013. The Southern Tilt survey was completed on 4 June 2013 and seismic gear began being recovered on this day. All seismic gear was recovered and stored on deck by 5 June 2013.

PSOs undertook a combination of visual and acoustic watches, conducting a total of 1723 hours 32 minutes of visual observations and 732 hours 21 minutes of acoustic monitoring over the course of the survey.

This visual monitoring effort produced a project total of 231 protected species detection records; 228 for cetaceans and three for pinnipeds. Of the 228 cetacean records collected, 101 records were collected for mysticetes, 98 records of odontocetes, 28 records of unidentified large whales, and one record of an unidentified cetacean. All three pinniped detections were of unidentified pinnipeds. There were no detections of sea turtles during the survey. There was one acoustic detection using the passive acoustic monitoring (PAM) system of an unidentified Delphinid. There were no correlating visual and acoustic detections.

On 19 January 2013, there was intent for airgun testing immediately after completion of the survey line during an unmonitored period at night. However, there was delay due to technicalities of changing airgun configurations for testing which resulted in the airguns going silent for duration of four minutes between survey line and airgun testing.

Detections of protected species resulted in two mitigation actions being implemented. On 25 March 2013 a soft start was delayed eight minutes due to long-finned pilot whales (detection 198) being observed within the exclusion zone during the pre-soft start survey. The soft start was being performed prior to testing the airguns so the mitigation action did not result in any

production loss. The second mitigation action occurred on 19 May 2013, a soft start prior to gun testing was delayed 15 minutes due to Sei whales (detection 210) heading toward the exclusion zone. The soft start was being performed prior to testing the airguns so the mitigation action did not result in any production loss.

1.1. PROJECT LOCATION AND OPERATION PARAMETERS

There were two prospect areas surveyed during this period. The first prospect area was the Falkland Islands Southern Phase A 2012 Survey located in the East Falkland Islands basin, where water depths ranged from approximately 900 to 1,700 meters (Figure 1). The survey consisted of 109 survey lines with a grid orientation of northeast. The survey was of the Southern Quadrants 52 (block numbers 3-5, 7-10, 12-15, 18-20, and 24-25) and 53 (block numbers 1-3, 6-9, 11-14, 16-17 and 21) covering 2,808 square kilometers. The survey lines were 79 kilometers long with line spacing of 720 meters. Line changes were approximately three to four hours in duration. The *Ramform Sterling* worked with two support vessels, acting as both chase and occasional supply boats, the *Windward* and the *Christina Debora*. On 12 February the *Christina Debora* was replaced by the *Thor Pioneer*.

The second prospect area was the Falkland Islands Southern Tilt 2013 Survey located in the Southern Falkland Islands basin, where water depths ranged from approximately 660 to 2,100 meters (Figure 1). The survey was positioned in the Southern Quadrants 60 (block numbers 13a-15a, 18a-19a, 18b-19b, 20, 23, 24a-25a and 24b-25b) and 61 (block numbers 11, 16 and 21) covering 1,160 square kilometres. The survey consisted of 49 survey lines shot in the East to West direction, averaging 32 kilometres in length with line spacing of 720 meters. Line changes were approximately three to five hours in duration.

The *Ramform Sterling* towed two airgun arrays, separated by 60 meters, each comprised of three sub-arrays, separated by 8 meters. Each sub-array consisted of ten (2 strings) to twelve airguns ranging in volume from 40 in³ to 250 in³. Each string was equipped with one spare airgun. The airguns were towed at an average depth of 7 meters. The full source included a total of 62 airguns, alternating arrays for a maximum source volume of 4130 in³ and a pressure of 2000 psi. Intensity of the source was 69 bar meters at a towed depth of 7 meters. The frequency range is a broadband frequency from 0-240 Hz with an intent to survey at 10-70 Hz. The center of the seismic source array was 750 meters from the vessel reference point (VRP) which was located above the PSO viewing station. The shot point interval was 25 meters or approximately every 10.4 seconds. The vessel towed 12 streamers of 6,600 meters in length, each separated by 120 meters and towed at a depth of approximately 10 meters.

Soft starts were conducted by firing the smallest airgun, and then gradually adding airguns until the airguns reached full power. Current JNCC regulations required all soft starts to be completed within 20 to 40 minutes. The mitigation source is the continuous firing of the smallest airgun at the same shot point interval as production shots. During this survey, the mitigation source was not utilized. Except during the occasional airgun testing after the end of line, airguns were immediately silenced after completion of survey lines. As per the JNCC, during periods of airgun silence lasting less than 10 minutes no soft start is required to return to full power, providing that PSOs are monitoring continuously (visual or PAM) throughout the silence and no protected species are observed within the exclusion zone.

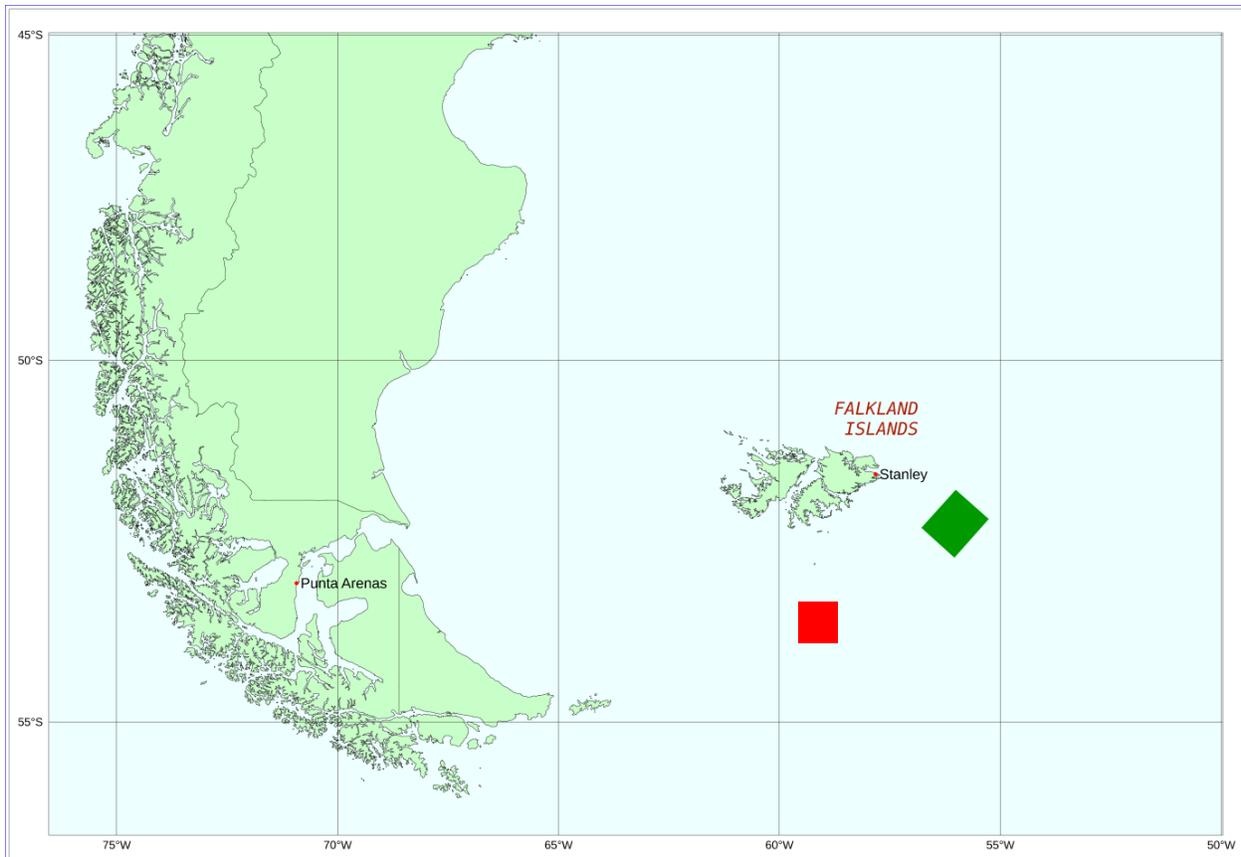


Figure 1: General location of the surveys in relation to the Falkland Islands. The green square represents the Southern Phase A 2012 survey and the red square represents the Southern Tilt 2013 survey.

1.2. PASSIVE ACOUSTIC MONITORING (PAM)

A *Seiche Measurements* PAM system was installed by the first PAM operator to join the vessel shortly after the start up meeting in Punta Arenas, Chile on 25 November 2012. The PAM system was onboard during all survey operations and was uninstalled by the PAM operator upon completion of the survey on 4 June 2013.

The PAM system consisted of two 250m conventional array cable, three 20m airgun-towed (AGT) array cables (one in use with two spares), two 100m deck cables (one for each type of array), a data processing unit with a computer, dual monitors, an acoustic analysis software package, headphones for aural monitoring, various leads and adapters, and a complete spare set of electronics and other hardware (other than cables).

The distance between the area of deployment and the monitoring station are often greater than the length of hydrophone tow cable that remains after deployment and the deck cable serves as an extension that directly interfaces with the data processing unit and the hydrophone tow cable.

The data processing unit processes the raw data from the hydrophones through two external sound cards, a *National Instruments DAQ* card and an *ASIO Fireface* card (contained within a *RME Fireface 800* unit). *National Instruments DAQ* sound cards sample raw audio at 500kHz and are used to detect beaked whale, *Kogia* species, porpoise, and delphinid

(echolocation) clicks up to up to 250kHz. *ASIO Fireface* sound cards sample audio at 96kHz and are used to detect mysticete, delphinid, and non-delphinid odontocetes (including sperm whale) vocalizations up to 48kHz. The data processing unit also contains a *Measurements and Computing* data logger for the depth gauge, digital signal amplifiers, an *UltraLink Pro* audio mixer, and an *UltraCurve Pro* graphic equalizer.

The two sound cards supply low and high frequency digital audio feeds to a computer that contains a suite of software for monitoring cetacean acoustics. *Pamguard* (Beta version 1.11.02) is the primary software utilized on this survey. The International Federation on Animal Welfare (IFAW) software including *Logger 2000*, *Rainbow Click*, *Whistle*, and *Rainbow Click Porpoise* is the secondary software and will only be used if the *Pamguard* software fails.

The computer receives both the raw audio from the *National Instruments DAQ* sound card as well as the raw audio from the *ASIO Fireface* sound card. *Pamguard* modules including a high and low frequency click detector, whistle and moan detector, spectrogram, map with a direct GPRMC GPS feed from the *Ramform Sterling's* navigation system, and high and low frequency sound recorder.

Raw audio from the *ASIO Fireface* sound card is monitored aurally with *Sennheiser* headphones.

The 250m conventional array cable was installed by the PAM operator and seismic personnel, and was used as a backup means of PAM in the event that the AGT cable became damaged or the gun string it was attached to was brought onboard for maintenance, preventing acoustic monitoring. The conventional array was first deployed and monitored on 06 January 2013 after the second AGT cable was damaged. On 18 January 2013 the conventional array was audibly damaged and subsequently retrieved. It became apparent that the free end of the array had wrapped around the lead in and caused damage to the wiring. The conventional became entangled with the lead in on 17 March 2013 and was snapped during retrieval. The second conventional cable was installed with more p-links to prevent future entanglement. On 25 March 2013 the second conventional array cable was entangled around 2 separate lead ins, with the possibility of an additional entanglement around a third lead in under the water's surface. The cable snapped above the p-links during retrieval. During this time, the first conventional cable was repaired for future deployments.

The 20m AGT array cable was installed by the PAM operator and airgun mechanics on gun string 3, aft of the guns and float, and was first used on 09 December 2012 at the start of seismic survey operations. The cable remained deployed until it became kinked on 01 January 2012. The deployment method was revised and a second cable was installed in place of the first. The second cable was damaged on 05 January and was replaced with the third cable. The third cable became damaged on 10 January in much the same manner as the second cable. The deployment method was reverted to the original plan, but with additional spiral wrap and tape for support, and the first array (since repaired) was deployed. The repaired cable remained deployed and was undamaged for the rest of the survey.

1.3. VISUAL SURVEY METHODOLOGY

There were two trained and experienced PSOs on board to visually monitor for marine mammals, record and report on observations, and request mitigation actions in accordance with the *JNCC guidelines for minimising the risk of injury and disturbance to marine mammals from seismic surveys*, August 2010. The PSOs on board held certifications from a recognized JNCC course and approved Bureau of Ocean Management (BOEM) course. From the beginning of

the project Lynn Henneberger was on board as the Lead PSO and Monica Arancibia-Colgain as the support PSO. On 23 January 2013 there was a crew change and Heidi Ingram (Lead) and Tatiana Moreno (support) joined the vessel. On 3 April 2013 Megan McManus joined the vessel as Lead PSO and Alexandria Denby as support PSO. On 16 May 2013 Lacey Price joined the vessel as Lead PSO and Monica Arancibia-Colgain returned as support PSO until the completion of the project on 4 June 2013. Visual monitoring was carried out from the “sky lounge” located above the bridge, approximately 22.4 meters above the water surface which provided the PSOs with a 360° viewpoint around the acoustic source.

Binoculars (7x50 and 10x30 magnifications) were used for visual monitoring in addition to the naked eye. Inside the “sky lounge” the PSOs used a laptop for data collection as well as a telephone for communication. There was also a monitor that displayed current information about the vessel’s position, speed, and heading, along with water depth, and source activity. Wind speed and direction were obtained from the ship’s instrumentation on the bridge. PSOs were on a rotating schedule. Watches varied in duration, ranging from two to four hours.

When a protected species was observed range estimations were made using reticle binoculars, the naked eye, and by relating the animal to an object at a known distance, such as the acoustic array located 750 meters from the PSO observation deck. Specific species identifications were made whenever distance, length of sighting, and visual observation conditions allowed. PSOs observed physical features of animals sighted and noted behavior of the animal or group. From late January to early April photographs were taken during most sightings. Sometimes photographs were not taken due to the brevity of a sighting. The camera used was a Canon EOS 60D with a 300 millimeter telephoto lens. Marine mammal identification manuals were consulted and photos were examined during visual watch breaks to confirm identifications.

During each sighting event PSOs recorded the position, time at first and last sighting, number of animals present (adults and juveniles), the initial and any subsequent behaviors observed, the initial range, bearing and movement of the animal(s), the source activity at the initial and final detections and any mitigation measures that were applied. Specific information regarding the animal(s) closest approach to the vessel, acoustic source and the acoustic source output at the closest approach were recorded. Additionally, the vessel position, water depth, vessel speed, the wind speed and direction, Beaufort scale, swell level, visibility, glare, and precipitation were recorded every hour at minimum or every time environmental conditions or seismic activity changed. Each sighting event was linked to an entry on a datasheet such that environmental conditions were available for each sighting event.

The PSOs communicated with the seismic crew in the instrument room via internal ship’s phone. Communication through the internal phone system directly to the instrument room has assisted in prompt, clear communication of any changes in airgun activity or in the event a mitigation action needed to be implemented. The support vessels could also be reached by both UHF and VHF two-way radio communication.

When the acoustic source was activated from silence, PSOs maintained watch for 60 minutes prior to the activation of the source. Visual watches commenced each day before sunrise, beginning as soon as the exclusion zone was visible, and continued past sunset until the exclusion zone became obscured. Start of observation times ranged from 7:17 to 10:40 UTC, while end of observation times ranged from 20:00 to 00:45 UTC.

1.4. PASSIVE ACOUSTIC MONITORING (PAM) SURVEY METHODOLOGY

This project utilized PAM operators to acoustically monitor during the night and during times of reduced daytime visibility; in order to clear the exclusion zone prior to ramp-up when applicable.

The PAM system was monitored on a nightly basis with a start time ranging from 20:30-00:15 UTC and an end time ranging from 07:45-11:20 UTC. PAM monitoring began on the night of 09 December 2012 and continued for the duration of operations ending on 04 June 2013.

Acoustic surveys were conducted by the PAM operator in two to four hour shifts separated by one to two hour breaks. Acoustic monitoring overlapped visual monitoring during the periods just before sunset and just after sunrise. The overlap allowed for continuous monitoring of the exclusion zone for protected species and would permit operations to resume with minimal lost production time in the event of a late operational delay. In addition, the overlap of acoustic and visual monitoring allowed for potential correlations between acoustic and visual detections.

Acoustic monitoring for marine mammals was completed aurally with *Sennheiser* headphones and visually with *Pamguard Beta 1.11.02*. Delphinid whistles, clicks, and burst pulses as well as sperm whale clicks may be viewed on a spectrogram display within *Pamguard*. Sperm whale, beaked whale, *Kogia* species, and delphinid echolocation clicks may be viewed on low and high frequency click detector displays.

Distances for acoustic detections are primarily based upon a noise or detection score system developed by Gannier *et al* (2002). Gannier *et al* monitored sperm whales (*Physeter macrocephalus*) in the Mediterranean both visually and acoustically. A subjective scale was developed based upon the strength or intensity of the sperm whale clicks at various distances that were measured visually when the sperm whales surfaced. Although the scale is subjective and sounds produced in marine environments will vary according to local conditions, the scale provides a measure for approximating distances when using a single, linear hydrophone array.

Another method to determine range of an animal was to use the *Pamguard* Map module along with the Click Detector in the LF *Pamguard* configuration file. In order to successfully utilize this function of *Pamguard* the detection needs to be several minutes long, and the successful determination and selection of “click trains” in the Click Detector module. Due to the ambient noise levels and minimal hydrophone separation, the PAM operator was not able to utilize this function of *Pamguard*.

During a detection of a vocalizing animal, information regarding position, distance, heading of vessel, water depth, and range of animal, if applicable, was recorded, along with any recordings of vocalizations. A detection report was completed, utilizing information gathered from not only the navigation department, but also from the *Pamguard* modules. All information gathered was input into the Detection Report forms and the Daily Passive Acoustic Forms. This information is available in Appendix B, and Appendix C respectively. All detections were reported on a daily basis to onboard client representatives and the vessel Party Chief. Details of these detections were submitted in Weekly Reports along with the combined visual MMO data, and can be found in Appendix B. Recordings made of detections have a corresponding word document that describes the conditions, and details of the recording.

1.5. SURVEY DATA SUMMARY

The *Ramform Sterling* departed Punta Arenas, Chile on 29 November, 2012 for the Southern Phase A 2012 survey area near the Falkland Islands. The seismic gear was deployed and use of the acoustic source commenced at 23:48 UTC on 8 December 2012 and continued until the prospect was completed at 00:17 UTC on 10 May 2013. Gun operations commenced on the second survey, the Southern Tilt 2013 survey, at 11:44 UTC on 19 May 2013 and continued until 16:57 UTC on 4 June 2013 when the project was completed. All seismic gear began being

brought on board on 4 June 2013 and was stored and secured on deck by 5 June 2013.

The acoustic source was active throughout the two surveys, with multiple periods of source silence, for a total of 1515 hours 12 minutes of source activity. This includes ramp-up of the airguns, full power and partial power firing both online and during airgun testing (Figure 2). Full power source operations, while online, accounted for 88 percent (1345 hours 37 minutes) of airgun activity during the project. There was 54 hours 45 minutes of non-production full-volume firing, and 27 hours 32 minutes of airgun testing. Additionally, the airguns were active for a total of 87 hours 18 minutes during ramp ups. There was a total of 2948 hours and 48 minutes of airgun silence throughout the survey.

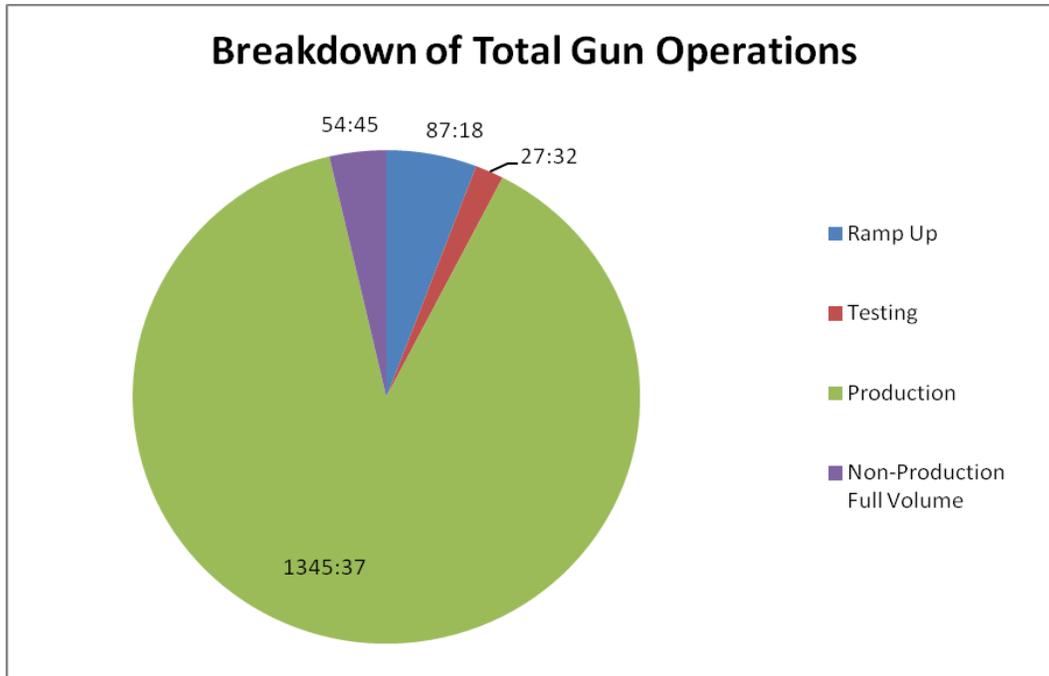


Figure 2: Breakdown of airgun operations.

There were 263 soft starts performed over the course of the survey in order to commence full power survey operations. The soft start ranged from 21 to 37 minutes in duration; with four soft starts being abandoned midway due to computer or compressor problems or bad weather. Nineteen soft starts were performed prior to airgun testing with the remaining soft starts performed to resume production after periods of airgun silence. The soft starts were conducted using an automated gun controller program, which adds guns sequentially to achieve full source over the required period of time. Prior to soft starts and airgun testing the area was monitored for 60 minutes either visually, during daytime, or acoustically, during nighttime or periods of limited visibility. The exclusion zone was cleared prior to airgun activity 139 times through visual monitoring and 124 times through PAM (Table 1).

Table 1. Total acoustic source operations during the Falkland Islands Southern Phase A & Southern Tilt 2013.

| Acoustic Source Operations | Number | Duration (hh:mm) |
|---|------------|------------------|
| Airgun Tests | | 27:32 |
| Soft start | 263 | 87:18 |
| Day time ramp-ups cleared by visual observation | 139 | |
| Day time soft starts cleared by PAM | 14 | |
| Night time soft starts cleared by PAM | 110 | |
| Full power survey acquisition | | 1345:37 |
| Full power non-production | | 54:45 |
| Total time acoustic source was active | | 1515:12 |

1.5.1. Visual Monitoring Survey Summary

Visual monitoring was conducted by two PSOs each day between just before dawn until just after dusk, when it was too dark for the entire safety radius to be visible. The duration of daily observations ranged from approximately 8 hours 40 minutes to 17 hours 19 minutes. Observation times were adjusted as required for seasonal changes in available daylight, as well as daily changes due to cloud cover and fog. Over the course of the project a total of 1723 hours 32 minutes of visual observation were conducted. The acoustic source was active during 902 hours 3 minutes (52%) during visual monitoring and silent during 821 hours 29 minutes (48%) of visual monitoring, as shown in Figure 3. However, when visibility was reduced and the exclusion zone was not visible during daylight hours due to thick fog, passive acoustic monitoring was used to grant clearance prior to soft starts. Daytime PAM was used to clear the exclusion zone 14 times over the course of the project.

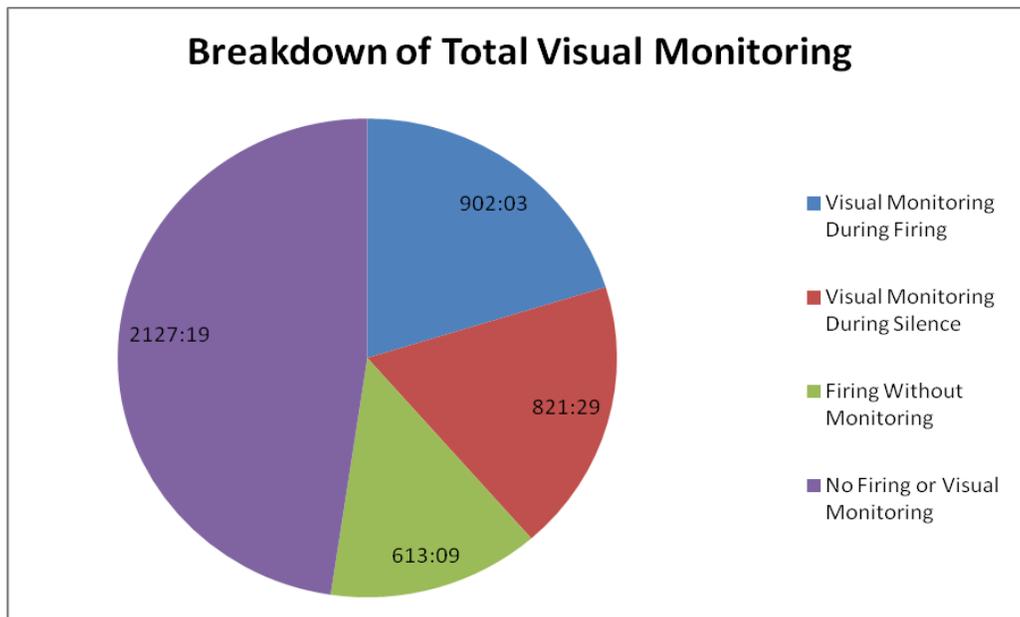


Figure 3: Breakdown of visual monitoring.

1.5.2. Acoustic Monitoring Survey Summary

Acoustic monitoring began on 09 December at 00:15 for pre-watch prior to and during a series of airgun tests and continued throughout the project with the PAM operator monitoring the

hydrophones aurally and monitoring the *Pamguard* detection software. Acoustic monitoring for the project ended at 11:30 UTC on 4 June 2013 when acquisition of the final survey line was completed and the hydrophone cable was retrieved. Over the course of the project, the PAM operator and PSOs conducted 732 hours 21 minutes of acoustic monitoring. The acoustic source was active during 424 hours 42 minutes (58%) during acoustic monitoring and silent during 307 hours 39 minutes (42%) of acoustic monitoring (Figure 4).

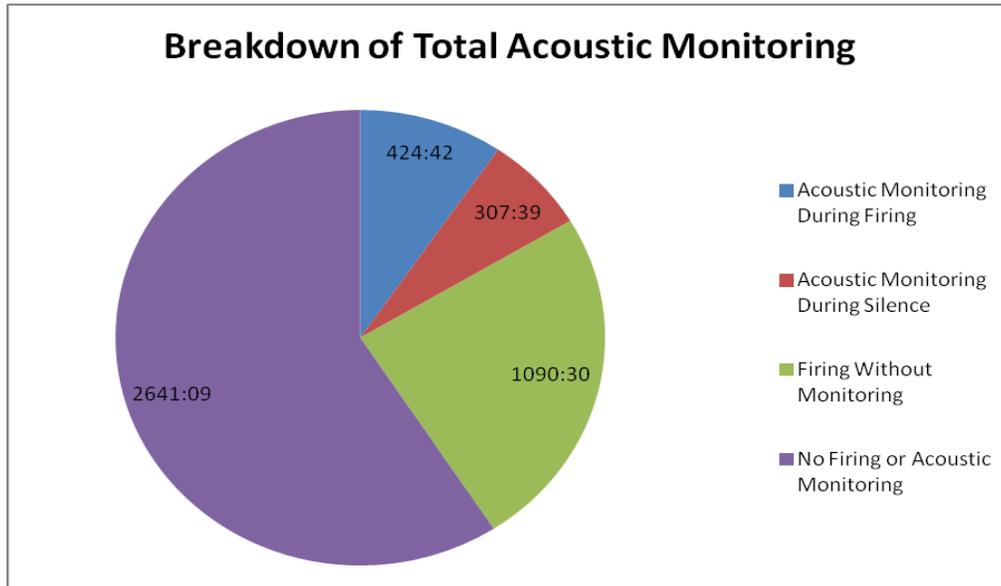


Figure 4: Breakdown of acoustic monitoring.

1.6. MITIGATION ACTIONS

There were two mitigation actions implemented during the Falkland Islands Southern Phase A & Southern Tilt seismic surveys. On 25 March 2013 (detection 198) a pod of long-finned pilot whales were observed approximately 250 meters from the acoustic source during the one hour survey period prior to soft start. The soft start was delayed by eight minutes to ensure the animals had been outside of the exclusion zone for 20 minutes before the airguns were activated. Hourglass dolphins were also observed with the pilot whales but were not observed inside the exclusion zone. The soft start was conducted prior to an airgun test so there was no loss of production due to the mitigation action.

The second mitigation action occurred on 19 May 2013, a soft start prior to gun testing was delayed 15 minutes due to Sei whales (detection 210) heading toward the exclusion zone. The soft start was being performed prior to testing the airguns so the mitigation action did not result in any production loss.

Passive acoustic monitoring was implemented during this project to clear soft starts during times of limited visibility including episodes of heavy fog, in addition to, the night-time operations.

2. WILDLIFE SUMMARY

2.1. VISUAL DETECTIONS

Visual monitoring conducted during the Falkland Islands Southern Phase A 2012 and Southern Tilt 2013 surveys resulted in the collection of 231 records of detection for protected species. Forms describing each detection can be found in Appendix B. Twelve species of marine mammal were positively identified, along with unidentified baleen whales, unidentified whales, unidentified cetaceans, unidentified dolphins, and unidentified pinnipeds. The total number of detection events and total number of animals recorded by species is described in Table 2 and shown in figures 5-7.

The wildlife detections from this project can be separated into two categories: birds and fish. A single sighting of an unidentified shark occurred during a feeding frenzy on 17 February in close proximity to a pod of long-finned pilot whales and hourglass dolphins. Birds from a total of 12 families, including 32 positively identified species, were observed over the course of this project. The most abundant species observed throughout the project was the cape petrel (*Daption capense*), although this is due to a particularly dense number observed in the month of December with numbers tapering dramatically proceeding that month. Other species with similar observation patterns are the giant petrel (*Macronectes giganteus*), southern fulmar (*Fulmarus glacialisoides*), and the black-browed albatross (*Thalassarche melanophrys*). The latter of which has its highest number of observed individuals in December and January. The species most regularly observed throughout the project is the black-browed albatross, followed by the wandering albatross (*Diomedea exulans*). Outliers include sooty albatross (*Phoebastria fusca*), gray-headed albatross (*Thalassarche chrysostoma*), Chilean skua (*Stercorarius chilensis*), white-bellied storm-petrel (*Fregetta grallaria*), black-chinned siskin (*Carduelis barbata*), an unidentified penguin species, and an unidentified tern species, each of which represents one to two individuals observed on one to two days.

A complete list of bird species and other marine life observed and identified in addition to the approximate number of individuals observed and the number of days on which they were observed can be found in Tables 4-6.

Table 2. Number of visual detection records recorded for each protected species.

| | Total Number of Detection Records | Total Number of Animals Recorded |
|---------------------------|-----------------------------------|----------------------------------|
| Cetaceans | | |
| Unidentified whale | 28 | 33 |
| Unidentified cetacean | 1 | 1 |
| Mysticetes | | |
| Fin whale | 25 | 54 |
| Sei whale | 21 | 47 |
| Minke whale | 8 | 10 |
| Southern right whale | 5 | 5 |
| Humpback whale | 1 | 1 |
| Unidentified baleen whale | 42 | 66 |
| Odontocetes | | |
| Sperm whale | 1 | 2 |
| Orca | 2 | 2 |
| Long-finned pilot whale | 14 | 431 |

| | | |
|-----------------------|------------|-------------|
| Hourglass dolphin | 58 | 797 |
| Peale's dolphin | 4 | 54 |
| Dusky dolphin | 1 | 1 |
| Commerson's dolphin | 1 | 7 |
| Unidentified dolphin | 16 | 106 |
| Pinnipeds | | |
| Unidentified pinniped | 3 | 3 |
| TOTAL | 231 | 1621 |

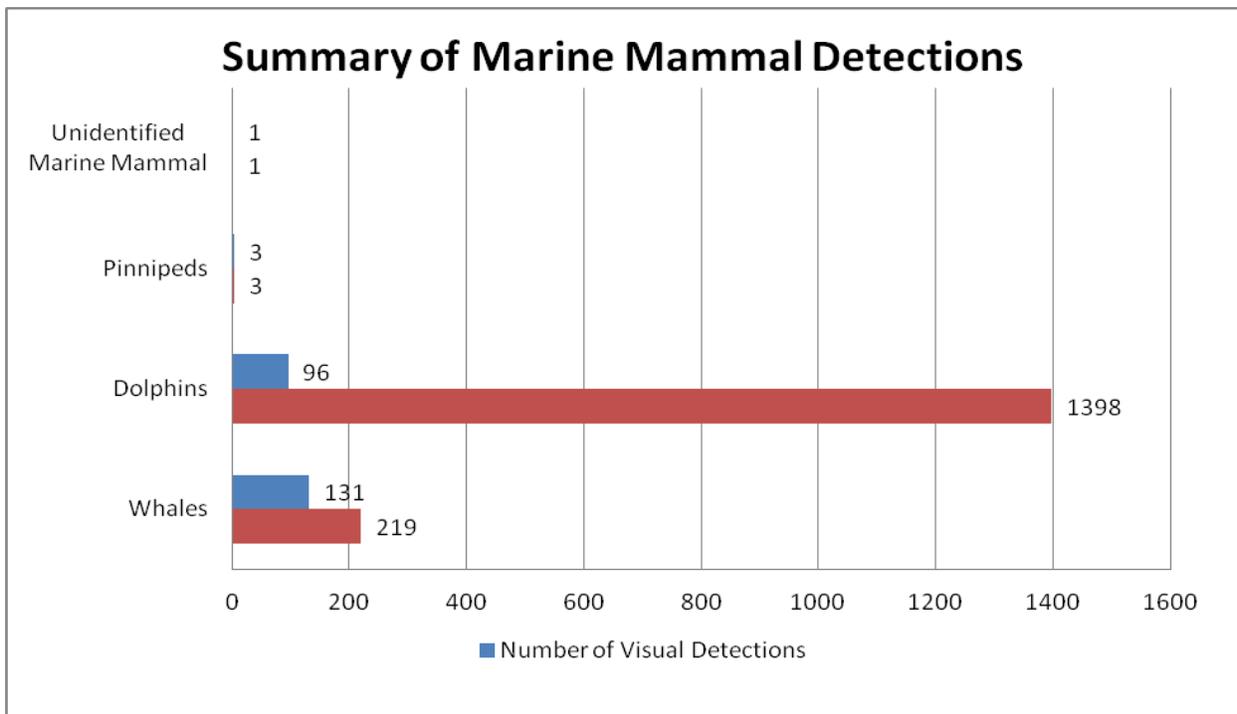


Figure 5: Breakdown of protected species detected.

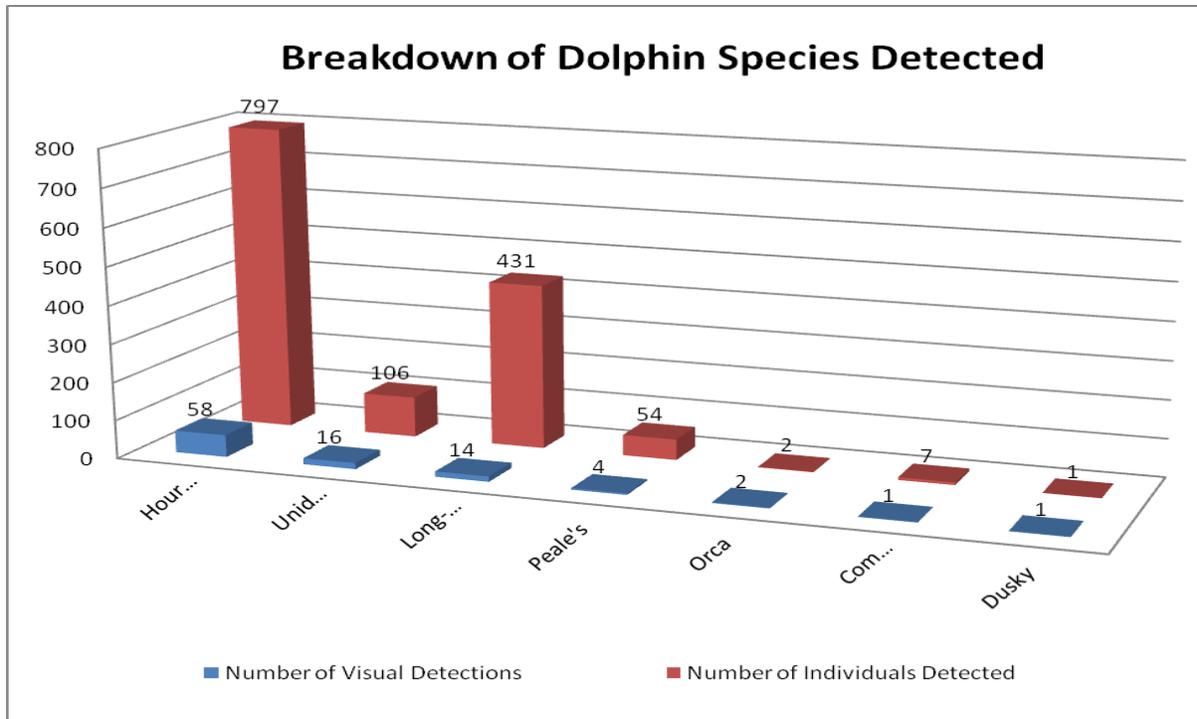


Figure 6: Breakdown of dolphin species detected.

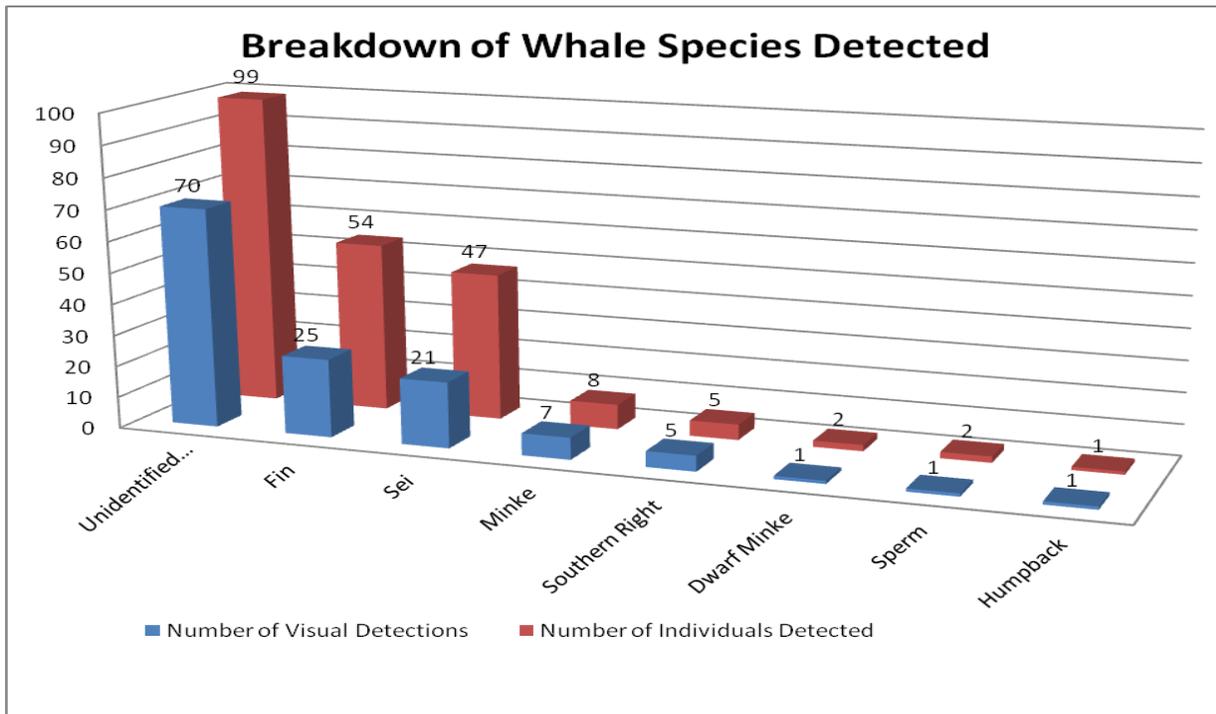


Figure 7. Breakdown of whale species detected during the survey.

Of the 231 visual protected species detection events during the survey, 130 detections occurred while the acoustic source was active and 101 detections occurred while the acoustic source was silent. Table 3 demonstrates the average closest approach of each species to the source at various source volumes. Figure 8 shows the average closest distance of each category of

marine mammals to the source while firing and during silence.

Table 3. Average closest approach of protected species to the acoustic source at various volumes.

| Species Detected | Full Power (4130 in ³) | | Soft start | | Reduced power | | Not Firing | |
|---------------------------|------------------------------------|---|----------------------|---|----------------------|---|----------------------|---|
| | Number of detections | Average closest approach to source (meters) | Number of detections | Average closest approach to source (meters) | Number of detections | Average closest approach to source (meters) | Number of detections | Average closest approach to source (meters) |
| Fin whale | 17 | 1208 | - | - | - | - | 8 | 1356 |
| Sei whale | 7 | 1160 | - | - | - | - | 14 | 2300 |
| Minke whale | 5 | 913 | - | - | - | - | 2 | 1100 |
| Dwarf minke whale | - | - | - | - | - | - | 1 | 20 |
| Southern right whale | 4 | 968 | - | - | - | - | 1 | 800 |
| Humpback whale | - | - | - | - | - | - | 1 | 100 |
| Sperm whale | 1 | 5000 | - | - | - | - | - | - |
| Orca | 1 | 800 | - | - | - | - | 1 | 250 |
| Long-finned pilot whale | 4 | 513 | - | - | - | - | 10 | 293 |
| Hourglass dolphin | 31 | 1145 | 1 | 1500 | - | - | 26 | 925 |
| Peale's dolphin | - | - | - | - | - | - | 4 | 481 |
| Dusky dolphin | - | - | - | - | - | - | 1 | 50 |
| Commerson's dolphin | 1 | 850 | - | - | - | - | - | - |
| Unidentified baleen whale | 26 | 2240 | 1 | 1200 | 1 | 1500 | 14 | 2083 |
| Unidentified whale | 17 | 2303 | 3 | 1083 | - | - | 8 | 2413 |
| Unidentified dolphin | 8 | 1634 | - | - | - | - | 8 | 1586 |
| Unidentified cetacean | 1 | 895 | - | - | - | - | - | - |
| Unidentified pinniped | 3 | 838 | - | - | - | - | - | - |

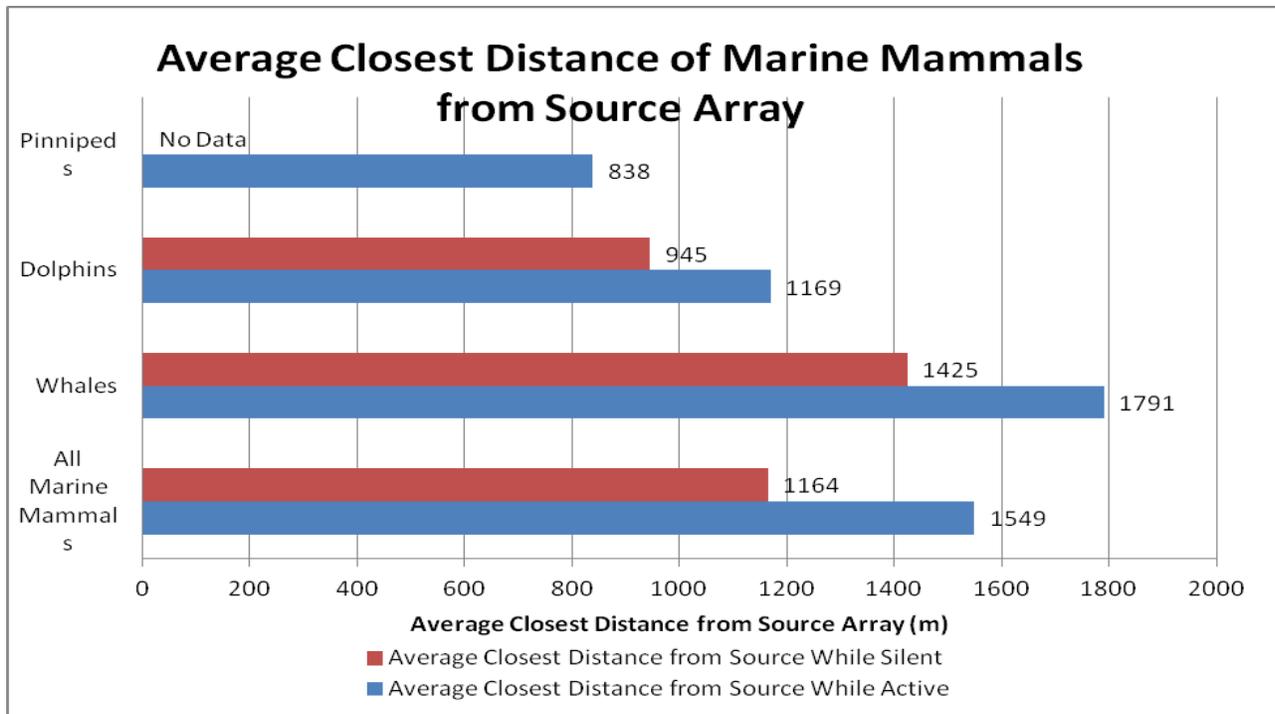


Figure 8. Average closest distance of protected species to source array.

2.1.1. Cetacean Detections

2.1.1.1. Sei whale

There were 21 visual detections of sei whales (*Balaenoptera borealis*) totalling 47 animals, five of the animals were identified as juveniles. The sightings occurred in water depths ranging from 160 to 1,941 meters and were observed in group sizes ranging from one to five animals. The majority of detections (11 detections) occurred during the end of November and December. During one sighting two sei whales were observed with a pod of hourglass dolphins.

2.1.1.2. Fin whale

There were 25 visual detections of fin whales (*Balaenoptera physalus*) totalling 54 animals; observed in group sizes ranging from one to eight animals. Three of the animals were identified as juveniles. The majority of fin whale detections (19 detections) occurred during the end of November through January. The sightings occurred in water depths ranging from 128 to 1,994 meters.

2.1.1.3. Dwarf minke whale

There was one visual detection of two dwarf minke whales (*Balaenoptera acutorostrata*). This sighting occurred at 560 meters of water depth. The whales were observed with a pod of long-finned pilot whales.

2.1.1.4. Minke whale

There were seven visual detections of Antarctic minke whales (*Balaenoptera bonaerensis*) totalling nine animals, observed in group sizes of one to two animals. The sightings occurred in water depths ranging from 482 to 1,641 meters.

2.1.1.5. Humpback whale

There was one visual detection of a humpback whale (*Balaenoptera novaeangliae*). This sighting occurred at 500 meters of water depth.

2.1.1.6. Sperm whale

There was one visual detection of two sperm whales (*Physeter macrocephalus*). This sighting occurred at 1,413 meters of water depth.

2.1.1.7. Southern right whale

There were five visual detections of Southern right whales (*Eubalaena australis*), all sightings of individual animals. All sightings of Southern right whales occurred in February and March. The sightings occurred in water depths ranging from 1,318 to 1,606 meters. During one sighting a pod of five hourglass dolphins were observed swimming alongside the whale.

2.1.1.8. Orca

There were two visual detections of orcas (*Orcinus orca*), both sightings of individual animals. Both sightings occurred in the month of December. The sightings occurred in water depths of 727 and 1,321 meters.

2.1.1.9. Long-finned pilot whale

There were 14 visual detections of long-finned pilot whales (*Globicephala melas*) totalling 431

animals, observed in group sizes of eight to 50 animals. Twenty-three of the animals were identified as juveniles. The sightings occurred in water depths ranging from 305 to 1,754 meters. During five of the detections the pilot whales were observed with other species such as a dwarf minke whale, an unidentified baleen whale, and hourglass dolphins.

2.1.1.10. Peale's dolphin

There were four visual detections of Peale's dolphins (*Lagenorhynchus australis*) totalling 54 animals. The sightings occurred in water depths ranging from 129 to 918 meters.

2.1.1.11. Hourglass dolphin

Hourglass dolphins (*Lagenorhynchus cruciger*) were the most abundant species observed with 58 detections totalling 797 animals. The majority of hourglass dolphin sightings (48 detections) occurred in February and March. The dolphins were observed in group sizes of two to 80 animals, with one sighting of a large pod of approximately 250 animals. Twenty three of the animals were identified as juveniles. The sightings occurred in water depths ranging from 291 to 1,980 meters. Hourglass dolphins were frequently observed bowriding; they were also observed on one occasion with a Southern right whale and on three occasions with long-finned pilot whales.

2.1.1.12. Dusky dolphin

There was one visual detection of a single dusky dolphin (*Lagenorhynchus obscurus*). This sighting occurred at a water depth of 1,786 meters.

2.1.1.13. Commerson's dolphin

There was one visual detection of Commerson's dolphins (*Cephalorhynchus commersonii*) totalling seven animals. This sighting occurred at a water depth of 1,547 meters.

2.1.1.14. Unidentified baleen whale

There were 42 visual detections of unidentifiable baleen whales, totalling 66 animals. Two of the animals were identified as juveniles. The animals in six of these sightings were identified as being either fin or sei whales.

2.1.1.15. Unidentified whale

There were 28 visual detections of unidentifiable whales, totalling 33 animals. One of the animals was identified as a juvenile. During the majority of these sightings only the blow was observed.

2.1.1.16. Unidentified dolphin

There were 16 visual detections of unidentifiable dolphins, totalling 106 animals. One of the animals was identified as a juvenile. Additionally, there was one sighting of a single unidentifiable cetacean.

2.1.2. Pinniped Detections

2.1.2.1. Unidentified pinniped

There were three visual detections of unidentifiable pinnipeds. The sightings occurred at water depths of 981 to 1,247 meters.

2.2. ACOUSTIC DETECTIONS

There was one acoustic detection during this survey. On 27 December 2012 at 01:40 an unidentified delphinid species was detected acoustically. Low frequency whistles were noted aurally as well as via Pamguard's spectrogram software. The dominant frequencies of the whistles were observed at 3-4 kHz, with harmonics peaking above 60 kHz. Lack of overlapping vocalizations suggest that only one animal was both present and vocal. The detection lasted for two minutes and ended at 01:42.

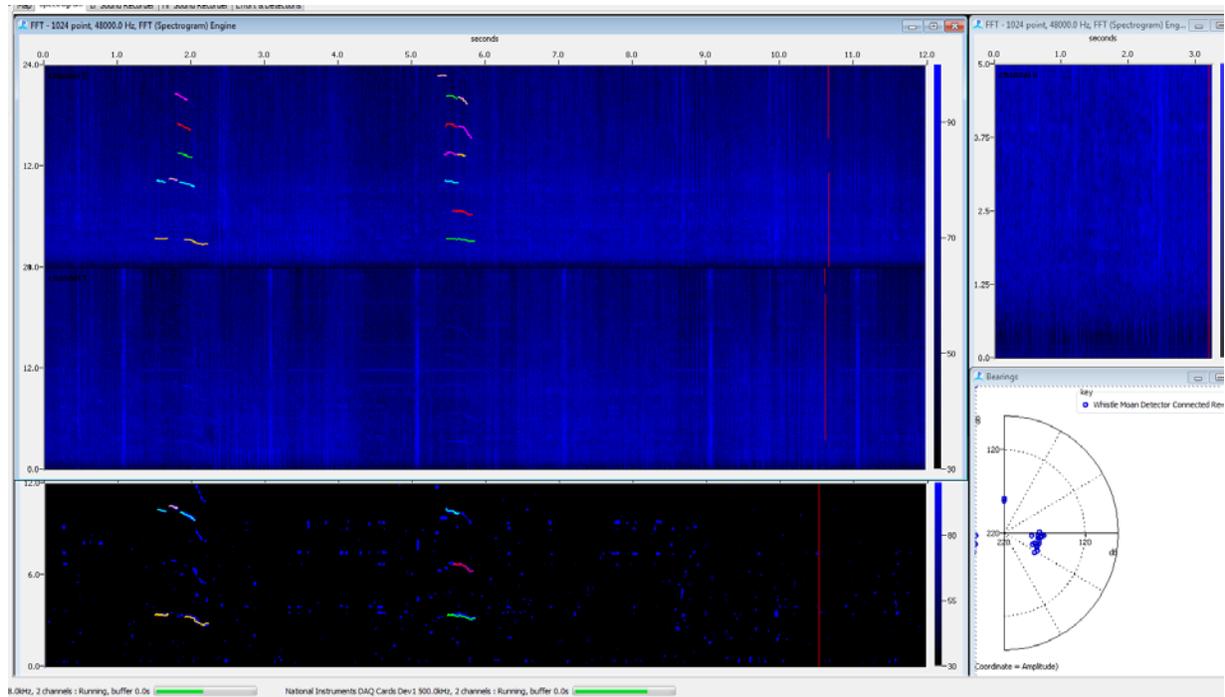


Figure 9. Screen capture of unidentified delphined detection on Pamguard.

2.3. ENVIRONMENTAL CONDITIONS

Visual observations were often hindered by poor environmental conditions. During visual observation the Beaufort scale ranged from zero to nine, with the vast majority of observations occurred with a Beaufort scale of four to six. Wind speed was quite variable from around 1 knot to 45 knots. Sea conditions were greatly affected by the wind changes; the sea state during the majority of watch was slight to choppy. The majority of visual observations were conducted with swells lower than two meters and a significant amount of observations were conducted with moderate swells, ranging from two to four meters. Visibility was good for the majority of visual observations, although there were significant periods of time of dense fog and precipitation, sometimes lasting for several days. Due to frequent cloud cover there was no glare present during the majority of watch with only a few days on glare inhibited sighting conditions.

2.4. WILDLIFE SUMMARY TABLES

2.4.1. Marine Mammals

Table 4: Protected species observed during visual and/or acoustic monitoring.

| Date | Common Name | Scientific Name (Genus species) | Family | Detection Method | Sighting Description | Mitigation Actions |
|------------|---------------------------|-----------------------------------|-----------------|------------------|--|--------------------|
| 29/11/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Four surfaced together | None |
| 30/11/2012 | Peale's Dolphin | <i>Lagenorhynchus australis</i> | Delphinidae | Visual | two barely surfaced and were circling the paravane | None |
| 30/11/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 3 blew in sequence, toward vessel then away | None |
| 01/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Fast swim, approach vssl, swam bw p-vane and vssl | None |
| 01/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | Head out surfacing, ind spread out til last sight | None |
| 01/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Mill, then away | None |
| 01/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Mill, then 2 approached w/in 200m | None |
| 01/12/2012 | Unidentified whale | n/a | n/a | Visual | Traveling slow, surfacing ~8-11 minutes | None |
| 01/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Appeared to be milling, then dove | None |
| 01/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Blew once then 40 seconds later again, then dove | None |
| 02/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Surfaced in breaking wave near bow | None |
| 03/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blow off stbd side, then blow at 09:20 stbd ahead | None |
| 05/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | tight grp trvl fast frm bow to pside, frq srfcng | None |
| 05/12/2012 | Humpback whale | <i>Megaptera novaeangliae</i> | Balaenopteridae | Visual | qk head lob headed W, blow 2-3x, head NE | None |
| 06/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Three blows observed spread out over 8 minutes | None |
| 07/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows were in a tight group from different animals | None |
| 07/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows only | None |
| 09/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows only | None |
| 09/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows only | None |
| 09/12/2012 | Orca | <i>Orcinus orca</i> | Delphinidae | Visual | Crashing through wave | None |
| 09/12/2012 | Dwarf Minke whale | <i>Balaenoptera acutorostrata</i> | Balaenopteridae | Visual | body surfaced @ P Bow, then blew @ S by lines | None |

| | | | | | | |
|------------|---------------------------|---------------------------------|-----------------|--------|--|------|
| 09/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | Lrg. Grp. Spread out by vess. | None |
| 10/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | jpgg clear of lrg waves on star side of vessel | None |
| 10/12/2012 | Unidentified whale | n/a | n/a | Visual | Tight group of blows hdg parallel away | None |
| 10/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | 1 blow smaller nxt to larger; trvl rapid opp dir | None |
| 10/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Blow 2-3x every 7-10 min milling | None |
| 10/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | 2 separate blows at once, blow 2-3x every 5 min. | None |
| 11/12/2012 | Unidentified Delphinidae | n/a | Delphinidae | Visual | brief head when surfacing, 1 lingered w/dorsal vis | None |
| 11/12/2012 | Unidentified whale | n/a | n/a | Visual | One blow sighted | None |
| 11/12/2012 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | 1 grp appchd bow then NE; othr stayd ptsd parallel | None |
| 11/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 2 milling, blow 2-3x/6-8 min. | None |
| 11/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Shallow dive, then surfacd & blows hdg away | None |
| 11/12/2012 | Unidentified Pinniped | n/a | n/a | Visual | leaping near bow 3x | None |
| 11/12/2012 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | passed perpend. To boat NE to SW, leaping | None |
| 11/12/2012 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | passed perpend. To boat NE to SW, leaping | None |
| 11/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | milling ~200-500 m from vess. | None |
| 11/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 3-5 animals, prob feed (lg bubbles) | None |
| 11/12/2012 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Approached w/in 400m then turned perpendicular | None |
| 11/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 2 anim 1 much lrger than othr, sim behav Blow/mill | None |
| 12/12/2012 | Unidentified whale | n/a | n/a | Visual | 1 blow near stbd stern, then 1 blow off sdbd bow | None |
| 12/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | consistent blows every 2-3 min heading ne | None |
| 12/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | 2-3 blows/ 5-7 min, heading SW rapidly | None |
| 12/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | 2-3 blows/5-7 min, heading NE rapidly | None |
| 12/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Mult blows spread over km around vssl -min 8 ind | None |
| 13/12/2012 | Minke Whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | Swimming away, then parallel same | None |
| 13/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | 3 heading 155; 2 off ptsd; 1 ahead pt | None |
| 13/12/2012 | Minke Whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | headed paralell w/ vess then away SW | None |
| 13/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Mill as vssl passed, surfacing w/blow & lg splash | None |
| 14/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | same dir as vess then perpend to NE | None |

| | | | | | | |
|------------|---------------------------|---------------------------------|-----------------|----------|--|------|
| 14/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows (2 to 3 ind together, 1 farther) | None |
| 14/12/2012 | Unidentified Delphinidae | n/a | Delphinidae | Visual | barely broke surf w/leaps, move to vess then away | None |
| 14/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Traveld behnd floats & off pt monowng (not EZ) | None |
| 14/12/2012 | Unidentified Whale | n/a | n/a | Visual | ind. blows ~4 min apart, trvl quickly awy frm vess | None |
| 14/12/2012 | Minke Whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | head awy frm vess NE, blow ~4-5 min 2x | None |
| 14/12/2012 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 2 animals near vssl swam bw streamers; blows ~5min | None |
| 15/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Crossing ahead | None |
| 15/12/2012 | Unidentified whale | n/a | n/a | Visual | 1:blws only;2:clsr paralel vssl,turnd away thn bk | None |
| 16/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | porpoising off stbd bow against vess | None |
| 17/12/2012 | Unidentified whale | n/a | n/a | Visual | blew 1-2x every 4-5 min. mill | None |
| 17/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | One blow sighted | None |
| 17/12/2012 | Unidentified cetacean | n/a | n/a | Visual | Surfaced 2X b4 porpoising twds vssl | None |
| 18/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | var heading near vess. Spread out | None |
| 18/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | blow, v. lrg body | None |
| 18/12/2012 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | tight group heading away frm boat | None |
| 18/12/2012 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Grp split,appchd w/in 50m;pod alngsde 500m | None |
| 18/12/2012 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | surfacing around whales | None |
| 18/12/2012 | Sei Whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Whitewater when surfacing, blows | None |
| 18/12/2012 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 8-10 min,then blow,2 min blow,repeat 8-10 min | None |
| 18/12/2012 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Slowly crossing ahead | None |
| 18/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Both spyhopped together, blows together | None |
| 22/12/2012 | Dusky Dolphin | <i>Lagenorhynchus obscurus</i> | Delphinidae | Visual | Slowly submerged off the bow heading towards stern | None |
| 24/12/2012 | Unidentified Delphinidae | n/a | Delphinidae | Visual | Porpoising parallel creating large splashes | None |
| 27/12/2012 | Unidentified Delphinidae | n/a | Delphinidae | Acoustic | | None |
| 27/12/2012 | Unidentified whale | n/a | n/a | Visual | Blows sighted, travelling fast | None |

| | | | | | | |
|------------|---------------------------------|------------------------------------|-----------------|--------|---|------|
| 27/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Body sighted surfacing off stbd, then 2 blows after it had passed streamers | None |
| 28/12/2012 | Orca | <i>Orcinus orca</i> | Delphinidae | Visual | porpoised once off stbd side, no blows | None |
| 28/12/2012 | Minke whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | Rapid surfacing all around vessel | None |
| 28/12/2012 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | slow-moving, 2 off stbd, 1 new or old sighted ptsd | None |
| 28/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows sighted off stbd deflector (poss 2 animals) | None |
| 29/12/2012 | Unidentified Delphinidae | n/a | Delphinidae | Visual | porpoising ~2-3 min | None |
| 29/12/2012 | Unidentified whale | n/a | n/a | Visual | water disturbance, fast srfcng, no blows | None |
| 29/12/2012 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | 2 blows, simt. Water dist | None |
| 29/12/2012 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Multiple blows, erratic water disturbance w/2 ind | None |
| 31/12/2012 | Peale's dolphin | <i>Lagenorhynchus australis</i> | Delphinidae | Visual | porpoising rapidly | None |
| 31/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Lg whiteH2O created, head briefly observed w/1 blow | None |
| 31/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows, 3 ind side-by-side, 1 blow tiny in relation | None |
| 31/12/2012 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows, animals near each other | None |
| 01/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows observed, no body sighted | None |
| 01/01/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | approachd bw streamers 8&9, then bw vessl & source | None |
| 01/01/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Blows~3 min; parallel, away, cut across chase vssl | None |
| 01/01/2013 | Unidentified large baleen whale | n/a | Balaenopteridae | Visual | Large whiteH2O created, multiple blows | None |
| 01/01/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Light splashing, sighted off stbd milling | None |
| 01/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | One blow sighted | None |
| 01/01/2013 | Unidentified Delphinidae | n/a | Delphinidae | Visual | Light splashing, then jumping ~700m stbd | None |
| 04/01/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | milling @ port, move to stbd ~500m, port stern | None |
| 04/01/2013 | Commerson's dolphin | <i>Cephalorhynchus commersonii</i> | Delphinidae | Visual | Splashes, birds over, vssl approachd, riding waves | None |
| 05/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blows | None |
| 05/01/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | Logging spread out as vessel approached | None |
| 05/01/2013 | Unidentified Delphinidae | n/a | Delphinidae | Visual | Porpoising dolphins | None |

| | | | | | | |
|------------|---------------------------------|---------------------------------|-----------------|--------|---|------|
| 05/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | 1-2 tall blows every 4-6 min | None |
| 06/01/2013 | Unidentified Delphinidae | n/a | Delphinidae | Visual | Splashes on wave crests, 1 surfacing sighted | None |
| 07/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Lrg blows sighted ptsd; blended w/fog & lost sight | None |
| 07/01/2013 | Unidentified whale | n/a | n/a | Visual | Blows sighted off bow as vessel passed ind | None |
| 07/01/2013 | Unidentified Delphinidae | n/a | Delphinidae | Visual | Breach mult xs, then porpoising twds vssl | None |
| 08/01/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Partial breach, head out of water splashing | None |
| 15/01/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | whiteH2O ahead of blow; blow then lg dorsl exposed | None |
| 15/01/2013 | Unidentified whale | n/a | n/a | Visual | blows sighted; hdg slightly parallel to angled away | None |
| 15/01/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | surfaced exposing dorsal | None |
| 16/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | sequential blows far apart, tall columnar, no body | None |
| 17/01/2013 | Unidentified whale | n/a | n/a | Visual | Milling, blows obs every 1-2min, then every 4 | None |
| 19/01/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | srfgcd w/o blow, head awy frm ship blw ~3-5min | None |
| 19/01/2013 | Minke whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | Multiple full body breaches | None |
| 19/01/2013 | Unidentified whale | n/a | n/a | Visual | Blows obs side-by-side, & then ~300m apart | None |
| 20/01/2013 | Unidentified Delphinidae | n/a | Delphinidae | Visual | Splashes, breaches and extremely high jumps | None |
| 20/01/2013 | Unidentified Pinniped | n/a | n/a | Visual | Surfaced, swam undr, surfaced-repeatd creatd v in H2O | None |
| 21/01/2013 | Unidentified whale | n/a | n/a | Visual | one lrg blow then disappeared | None |
| 21/01/2013 | Fin Whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | hdg parallel opp, shallow, right under surface | None |
| 24/01/2013 | Sperm Whale | <i>Physeter macrocephalus</i> | Physeteridae | Visual | 2 blows side by side traveling away from vessel | None |
| 27/01/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | 2 blows every 2/3 min, crossing behind vssl | None |
| 31/01/2013 | Unidentified whale | n/a | n/a | Visual | 3 blows observed, one every 6 minutes | None |
| 09/02/2013 | Small unidentified whale | n/a | n/a | Visual | 2 blows ~12 seconds apart | None |
| 10/02/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | one blow observed | None |
| 10/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | swimming quickly, changing course frequently, porpoising occasionally | None |
| 14/02/2013 | Unidentified large baleen whale | n/a | Balaenopteridae | Visual | 1-2 blows every several minutes, traveling parallel in the opposite direction before turning and traveling away from the vessel | None |

| | | | | | | |
|------------|---------------------------|--------------------------------|-----------------|--------|---|------|
| 14/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | swam slowly crossing bow before turning and swimming parallel to the vessel, turned and swam towards stern before traveling quickly away from vessel creating a lot of spray | None |
| 15/02/2013 | Unidentified whale | n/a | n/a | Visual | three blows observed ~10 seconds apart | None |
| 15/02/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 16:09 two blows 1 km off bow, 16:13 one blow dive towards vessel 200m off stbd, 16:15 one blow 300m behind stbd stern, 16:18 one blow whale changed direction traveling same as ship near streamers, 16:21 one blow 150m outside of streamers | None |
| 15/02/2013 | Unidentified whale | n/a | n/a | Visual | 18:58 one blow observed 900m off port bow, another within a minute. Two more blows seen side-by-side at 19:08 2km off port stern. One blow significantly larger than the other suggesting a juvenile | None |
| 15/02/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | First blow observed at 19:26 2.5 km from port bow. Blows seen every 2-3min. Traveled parallel to the vessel, in the opposite direction. Last seen at 19:33 | None |
| 15/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | swam quickly towards vessel, disappeared below bow for 7 minutes, observed swimming away from bow | None |
| 16/02/2013 | Unidentified small whale | n/a | n/a | Visual | first blow low and bushy, subsequent blows every 20-30 seconds very diffuse, barely visible | None |
| 16/02/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | First blow observed 2.2 km off bow, 15 minutes later another blow was observed 1.8 km directly off port side | None |
| 16/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | briefly observed off stbd bow porpoising towards vessel | None |
| 16/02/2013 | Unidentified whale | n/a | n/a | Visual | 1-2 blows observed every 3-4 minutes | None |
| 17/02/2013 | Unidentified whale | n/a | n/a | Visual | One blow observed 3 km off bow. One more blow observed at 9:56. Not seen again | None |
| 17/02/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | 16+ swimming slowly towards vssl. Appear to be herding bait ball. Swam btwn streamer buoys. 650m from guns | None |
| 17/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Swimming quickly towards vessel. | None |

| | | | | | | |
|------------|---------------------------|---------------------------------|-----------------|--------|---|------|
| 17/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | sighted swimming quickly towards bow (20:03), bow riding for a few minutes, swam away from vessel in same direction approached. Last seen at 20:18 | None |
| 17/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Observed off stbd bow swimming towards vessel, rarely breaking water surface, crossed bow of vessel not seen again. | None |
| 20/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins briefly observed porpoising towards vessel, creating lots of spray | None |
| 21/02/2013 | Unidentified whale | n/a | n/a | Visual | observed 3.5km off port stern. 2-3 blows every 7-10 minutes. Last seen 18:49 | None |
| 21/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | observed off stbd, swimming quickly at surface in various directions, traveled up towards bow, then away from vessel. | None |
| 21/02/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | multiple blows observed from 21:47 to 21:50 (every 15-20 seconds), resurfaced 3.5 km away observed several more blows | None |
| 22/02/2013 | Minke whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | multiple blows observed from 16:16-16:18, observed surfacing at 16:24, last seen 16:25 | None |
| 22/02/2013 | Southern right whale | <i>Eubalaena australis</i> | Balaenidae | Visual | originally observed splashes, 17:01 head of whale visible observed two blows, remained at surface of water before diving at 17:04, did not show flukes | None |
| 23/02/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | animals briefly observed porpoising while traveling quickly, creating a lot of spray | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | animals swam quickly towards vessel, creating a lot of spray, ~300m from vessel animals turned and began traveling parallel to vessel | None |
| 23/02/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | First observed by FRC crew, following them. Swam towards stern of vssl. 550m closest to guns. Drifted back and off port. | None |
| 23/02/2013 | Unidentified whale | n/a | Balaenopteridae | Visual | Large, dark whale observed at 14:55, logging for several minutes. No blow observed, no dorsal visible. Last seen at 15:00 | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Dolphins observed swimming slowly in the same direction as the vessel but angled away | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Traveling parallel to vessel towards port bow. Swimming quickly & porpoising. Last seen at 18:38 UTC | None |
| 23/02/2013 | Southern right whale | <i>Eubalaena australis</i> | Balaenidae | Visual | Two breaths every time animal surfaced. First obs 3km off bow, travelled parallel to vssl in opp direction. Dove at 18:49, reappeared then dove and showed flukes at 18:57 2km off port stern. Observed with hrglass dolphins | None |

| | | | | | | |
|------------|----------------------|--------------------------------|-----------------|--------|--|------|
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | 5 appeared alongside right whale bowing and flipping, then traveled towards the vessel. Lost in severe glare at 18:51. | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Pod trvlng towards bow from 2.5km off port at 18:55. Very acrobatic. Converged with another pod while bowriding then trvld perp, away from vssl. Last seen 19:26 | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Obsvd 18:55 trvlng twds bow from 11 o'clock. Very acrobatic. Converged with other pod while bow riding. Split into two groups. 8 cont bowrdng, the rest swam 800 m from stbd bow, circling as if feeding. Regrouped & traveled away from bow lost at 19:36 | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Obs at 20:51 traveling parallel to vessel, then perpendicular, away from vessel. Lots of splashing and appear to be circling. Last observed at 20:56. | None |
| 23/02/2013 | Unidentified whale | n/a | Balaenopteridae | Visual | Multiple blows observed from 21:20 to 21:23 (every 10-18 seconds), shallow dive, surfaced 21:31. Last seen 21:33 | None |
| 23/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Observed off bow swimming quickly towards vessel, before reaching vessel turned and swam away from vessel off port bow | None |
| 24/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Observed briefly at bow before swimming quickly away from vessel off stbd bow | None |
| 25/02/2013 | Unidentified whale | n/a | n/a | Visual | only one blow observed off bow | None |
| 25/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed swimming slowly with only dorsal fins above the water | None |
| 25/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins appeared to be feeding, leaping and spinning, briefly began porpoising towards vessel | None |
| 25/02/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | observed porpoising parallel in the opposite direction of the vessel creating a lot of spray | None |
| 25/02/2013 | Southern right whale | <i>Eubalaena australis</i> | Balaenidae | Visual | surfaced ~200m from port bow swimming away from vessel, dove just below surface - still visible from water disturbance, surfaced again just before streamers, last seen surfacing between two streamers | None |
| 25/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | 5 indiv obs at 19:58 ~700m from vessel, swimming parallel in the same direction. Changed direction as if crossing in front of vssl. Lost at 20:07 off pt bow. Seen 2km off port bow at 20:14 jumping and breaching. Last obs at 20:30, 500m off port bow | None |

| | | | | | | |
|------------|---------------------------|---------------------------------|-----------------|--------|--|------|
| 26/02/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | Multiple groups of ~8-15 animals spread out over ~3 km. One group sighted just off bow traveling in the opposite direction down the port side of the vessel. Other groups remained ~2 km away from the vessel. | None |
| 26/02/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Sighted with pilot whales. Swam parallel in the same direction as the vessel just off port side briefly before turning and continuing in same direction as pilot whales. | None |
| 27/02/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | observed blowing regularly making remaining near surface of water, traveling in opposite direction of vessel. Adult and juvenile ahead whit second adult following ~600m behind | None |
| 27/02/2013 | Peale's dolphin | <i>Lagenorhynchus australis</i> | Delphinidae | Visual | dolphins observed porpoising towards vessel to bowride, lost sight of them at bow | None |
| 28/02/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | whales observed in inlet near Stanley, observed blowing 1-3X every 3-6 minutes. | None |
| 02/03/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | 11:25 one tall blow observed, 11:28 two blows observed, lost sight of whale in severe glare. Whale re-sighted 11:52 on other side of glare, blowing 3-5x every 5-9 minutes | None |
| 02/03/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | dolphins briefly observed porpoising | None |
| 02/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins swimming quickly changing course frequently | None |
| 02/03/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | dolphins swimming quickly creating spray | None |
| 02/03/2013 | Minke whale | <i>Balaenoptera bonaerensis</i> | Balaenopteridae | Visual | observed blowing 2-3x every 2-7 minutes | None |
| 02/03/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | porpoising creating spray | None |
| 02/03/2013 | Unidentified whale | n/a | n/a | Visual | three low blows observed | None |
| 02/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | traveling fast, porpoising, creating a lot of spray | None |
| 02/03/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Sei Whale observed 900m off port bow at 18:52 UTC. Crossed ahead of vessel, within 800m of bow. Last seen at 19:19 UTC. | None |
| 03/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed briefly porpoising at bow before traveling away ahead of the vessel. | None |
| 04/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed bowriding for several minutes before swimming away from vessel off port side | None |
| 07/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Dolphins observed off port bow porpoising towards vessel, bowrode briefly before continuing away from vessel off stbd bow | None |

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|------------|---------------------------|--------------------------------|-----------------|--------|---|------|
| 08/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Dolphins observed off stbd bow porpoising towards vessel. Swam along stbd side briefly before traveling away from vessel off stbd bow. | None |
| 08/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | 3 dolphins observed at 13:35, 150m off port bow porpoising towards bow for about a minute. Not seen again | None |
| 09/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed off stbd bow leaping high out of water traveling in the same direction as vessel | None |
| 09/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | large pod of dolphins bow riding, broke up into several groups spread out over several km, some dolphins remained bow riding for ~1.5 hrs | None |
| 09/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed 2km off port bow traveling in an unknown direction. Last seen at 13:08 | None |
| 09/03/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Blow was observed 2km off stbd stern, bearing 40 deg. Later obs 1.6km bearing 25 deg. Last obs 1km at bearing 70 deg. | None |
| 09/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | approx 4 dolphins obs at 14:08 900m off port stern traveling towards vessel. Quickly changed direction to swim away from vessel. Last seen at 14:10 headed away from vessel. | None |
| 10/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed just under surface of water swimming quickly towards vessels bow, observed bowriding for several minutes | None |
| 10/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins leaping high out of water creating large splashes | None |
| 11/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphin swimming away from vessel off stbd bow, rarely breaking surface of water, two animals turned and traveled parallel in the opposite direction | None |
| 11/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins swimming quickly towards port bow of vessel, bowriding for several minutes before swimming away off port stern | None |
| 12/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed briefly swimming towards bow of vessel, not seen again | None |
| 13/03/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | whale originally observed traveling slowly parallel in the opposite direction of the vessel, as vessel passed whale turned and began traveling in same direction. Last seen 700m off port stern | None |
| 14/03/2013 | Unidentified whale | n/a | n/a | Visual | 1-2 blows observed every 3-4 minutes | None |
| 15/03/2013 | Southern right whale | <i>Eubalaena australis</i> | Balaenidae | Visual | surfaced 600m off stbd stern at 10:25, traveling parallel to the vessel, in same direction. Traveled further from vessel, then turned to swim parallel away from vessel. Last obs 10:35 2km away. | None |

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|------------|-------------------------|--------------------------------|-------------|--------|--|------------------|
| 15/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dorsal fins observed 500m off port bow swimming towards vessel. Last observed ~400 m from bow | None |
| 15/03/2013 | Unidentified whale | n/a | n/a | Visual | 1-3 blows observed every 3-6 minutes | None |
| 16/03/2013 | Southern right whale | <i>Eubalaena australis</i> | Balaenidae | Visual | whale originally observed 1.6 km off port bow crossing ahead of the vessel, 8 minutes later whale 100m from bow directly ahead of vessel, last seen just off stbd bow of vessel | None |
| 16/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed bowriding for several minutes before swimming away from vessel off stbd bow | None |
| 17/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | very large active pod feeding, trailing several km, dolphins often observed leaping repeatedly and landing on side, many dolphins bowriding | None |
| 18/03/2013 | Unidentified dolphin | n/a | Delphinidae | Visual | large splashes and dorsal fins briefly sighted | None |
| 21/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed just off stbd bow, bowrode for several minutes before swimming away from vessel directly off bow | None |
| 21/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | 3 dolphins initially obs swimming towards vssl. One swam down along port side, then back twds bow before traveling away. 19:47 total of 6 dolphins observed traveling parallel in the same direction as the vessel at 2km off port. Last seen at 19:51 | None |
| 24/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed swimming quickly towards stern of vessel before turning and swimming quickly away then traveling parallel to vessel, porpoising several times | None |
| 24/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins swam towards bow of vessel then away from vessel off stbd bow, changing course frequently | None |
| 25/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | 8 dolphins observed traveling towards vessel, came within 20m of vessel before turning to travel with pilot whales, 3 dolphins 1200m from vessel traveling with pilot whales, observed leaping landing on side | None |
| 25/03/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | large pod spread out over ~1.5 km, animals closest approached to vessel ~100m, as animals approached floats on streamers they turned and swam to the outside of the gear | Delay soft start |
| 27/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | swimming quickly just under surface creating a lot of spray, one animal briefly porpoising, traveling away from vessel off stbd bow | None |
| 29/03/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed swimming next to vessel for several minutes changing course frequently, swam quickly away from vessel off port bow | None |

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|------------|-------------------------|---------------------------------|-----------------|--------|---|--------------------|
| 01/04/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | whales observed ~4 km off stbd bow, one whale ~1 km away from the other two but joined together as vessel passed, traveling parallel, in opposite direction as vessel, angled away, 2-4 blows observed every several minutes, last seen ~3km off stbd stern | None |
| 01/04/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed porpoising off port bow, one dolphin observed leaping multiple times landing on side | None |
| 01/04/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins observed quickly traveling towards bow, then towards stern along port side. Began swimming in various directions before traveling 1km away | None |
| 02/04/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | 10:54 two blows observed directly ahead of vessel, whale crossing ahead of vessel, 11:05 two blows ~2 km off stbd side, whale now traveling parallel same direction as vessel, 11:07 one blow observed | None |
| 02/04/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | dolphins briefly observed swimming quickly away from vessel off port bow | None |
| 14/04/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | 16:56 UTC a pod of 25 long-finned pilot whales observed porpoising 150m off port bow, behavior consistent during sighting, last seen at 17:09UTC 500m from port guns | None |
| 15/04/2013 | Hourglass dolphin | <i>Lagenorhynchus cruciger</i> | Delphinidae | Visual | Dolphins observed off the port bow porpoising and jumping in a stationary position. Dolphins behavior did not change through the entirety of the sighting. The boat was turning so the dolphins were last sighted off the port beam | None |
| 16/04/2013 | Unidentified Pinniped | n/a | Pinniped | Visual | Pinniped occasionally surfacing for breaths | None |
| 7/05/2013 | Peale's Dolphin | <i>Lagenorhynchus australis</i> | Delphinidae | Visual | Dolphins observed bow-riding, porpoising, tail slapping and jumping around the bow of the ship. | None |
| 19/05/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | consistent blows every 5-7 minutes heading towards the gear | Delayed Soft Start |
| 19/05/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Regular blows from a single whale were seen every 30s-2mins from 1302-1322 UTC. Whale out of sight until 1346 UTC then 3 whales seen off stbd beam blowing and swimming quickly at surface. Last noted 2km off stbd bow at 1406 UTC. | None |
| 27/05/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | A total of five tall, narrow column like blows from a single whale were sighted off the starboard beam ~500m. Whale remained fairly stationary as the vessel moved away from it in the opposite direction. | None |

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|------------|---------------------------|------------------------------|-----------------|--------|--|------|
| 28/05/2013 | Long-finned pilot whale | <i>Globicephala melas</i> | Delphinidae | Visual | Boat approached pod, pod scattered and formed two separate groups around different pieces of gear, surfacing ~2min then never resurfaced after 12:04 | None |
| 28/05/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | 3 whales seen blowing at the surface every 1-2 mins showing the top portion of their bodies & fins from 1531-1545 UTC as they travelled parallel in the opposite direction of the vessel. Re-sighted at 1558 UTC. Last detected at 1602 UTC. | None |
| 30/05/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | 1 whale made a large splash close to boat then appeared 100m from first sighting with head and dorsal fin coming out of the water at the same time, travelling rapidly parallel to vessel | None |
| 31/05/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | A blow was observed 1km off of the forward starboard area and continued to blow every 2-3 minutes with some intermittent longer periods. | None |
| 1/06/2013 | Fin whale | <i>Balaenoptera physalus</i> | Balaenopteridae | Visual | Blows observed off starboard side with two animals, one around 4k and one around 5k off of the boat with consistent blows every 2-5 minutes. Third whale blow observed off starboard stern ~3k with consistent blows every 2-5 minutes, first travelled toward vessel then away. | None |
| 1/06/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | One whale was seen 800m off the stbd stern blowing regularly every 1-2mins. Two additional whales (1 juv.) were seen four mins later 500m off the stbd bow. The juv. approached the bow 200m off. All continued heading N blowing at the surface. | None |
| 2/06/2013 | Sei whale | <i>Balaenoptera borealis</i> | Balaenopteridae | Visual | Whale was seen blowing ~2km off the port bow every 1-3 minutes. The whales crossed ahead of the bow from port to starboard and proceeded down the starboard side of the vessel continuing to blow regularly. | None |
| 3/06/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Whale was seen blowing ~1500m off starboard side ~every 4-7 minutes in about the same area. Normal blows were observed without dorsal or body visible, except brief appearance of body with blow one time | None |
| 3/06/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | Whale was seen behind starboard side of vessel travelling away from vessel blowing approximately every 3-5 minutes | None |
| 4/06/2013 | Unidentified baleen whale | n/a | Balaenopteridae | Visual | A single blow was seen off the starboard bow ~2km followed by disturbances of the water as the whale surfaced. A small portion of dorsal fin was seen just briefly as the whale surfaced. | None |

2.4.2. Fish

Table 5: Fish species observed during visual monitoring.

| Common Name | Class | Order | Family | Genus | Species | Number of Individuals Observed | Number of Days Observed |
|--------------------|----------------|-------|--------|-------|---------|--------------------------------|-------------------------|
| Unidentified shark | Chondrichthyes | n/a | n/a | n/a | n/a | 1 | 1 |

2.4.3. Birds

Table 6: Bird species observed during visual monitoring.

| Common Name | Class | Order | Family | Genus | Species | Number of Individuals Observed | Number of Days Observed |
|-------------------------------------|-------|-------------------|----------------|---------------------|----------------------|--------------------------------|-------------------------|
| Black-browed albatross | Aves | Procellariiformes | Diomedidae | <i>Thalassarche</i> | <i>melanophris</i> | 522 | 121 |
| Unidentified royal albatross | Aves | Procellariiformes | Diomedidae | <i>Diomedea</i> | n/a | 32 | 19 |
| Northern royal albatross | Aves | Procellariiformes | Diomedidae | <i>D. e.</i> | <i>sanfordi</i> | 7 | 4 |
| Southern royal albatross | Aves | Procellariiformes | Diomedidae | <i>D. e.</i> | <i>epomophora</i> | 12 | 8 |
| Wandering albatross | Aves | Procellariiformes | Diomedidae | <i>Diomedea</i> | <i>exulans</i> | 266 | 84 |
| Snowy wandering albatross | Aves | Procellariiformes | Diomedidae | <i>D. e.</i> | <i>exulans</i> | 4 | 2 |
| Unidentified great albatross | Aves | Procellariiformes | Diomedidae | <i>Diomedea</i> | n/a | 80 | 28 |
| Light-mantled sooty albatross | Aves | Procellariiformes | Diomedidae | <i>Phoebetria</i> | <i>palpebrata</i> | 15 | 11 |
| Sooty albatross | Aves | Procellariiformes | Diomedidae | <i>Phoebetria</i> | <i>fusca</i> | 2 | 2 |
| Gray-headed albatross | Aves | Procellariiformes | Diomedidae | <i>Thalassarche</i> | <i>chrysostoma</i> | 3 | 3 |
| Cattle egret | Aves | Ciconiiformes | Ardeidae | <i>Bubulcus</i> | <i>ibis</i> | 56 | 13 |
| Southern fulmar | Aves | Procellariiformes | Procellariidae | <i>Fulmarus</i> | <i>glacialoides</i> | 109 | 21 |
| Antarctic petrel | Aves | Procellariiformes | Procellariidae | <i>Thalassoica</i> | <i>antarctica</i> | 1 | 1 |
| Atlantic petrel | Aves | Procellariiformes | Procellariidae | <i>Pterodroma</i> | <i>lessonii</i> | 63 | 8 |
| Grey petrel | Aves | Procellariiformes | Procellariidae | <i>Procellaria</i> | <i>cinerea</i> | 15 | 7 |
| Cape petrel | Aves | Procellariiformes | Procellariidae | <i>Daption</i> | <i>capense</i> | 805 | 57 |
| Northern giant petrel | Aves | Procellariiformes | Procellariidae | <i>Macronectes</i> | <i>Halli</i> | 80 | 32 |
| Southern giant petrel | Aves | Procellariiformes | Procellariidae | <i>Macronectes</i> | <i>giganteus</i> | 188 | 52 |
| Southern giant petrel (white morph) | Aves | Procellariiformes | Procellariidae | <i>Macronectes</i> | <i>giganteus</i> | 12 | 7 |
| Unidentified giant petrel | Aves | Procellariiformes | Procellariidae | <i>Macronectes</i> | n/a | 331 | 66 |
| Soft-plumaged petrel | Aves | Procellariiformes | Procellariidae | <i>Pterodroma</i> | <i>mollis</i> | 86 | 31 |
| White-chinned petrel | Aves | Procellariiformes | Procellariidae | <i>Procellaria</i> | <i>aequinotialis</i> | 125 | 44 |
| Grey-backed storm-petrel | Aves | Procellariiformes | Hydrobatidae | <i>Oceanites</i> | <i>nereis</i> | 9 | 4 |

| | | | | | | | |
|----------------------------|------|-------------------|----------------|---------------------|--------------------|-----|----|
| Wilson's storm-petrel | Aves | Procellariiformes | Hydrobatidae | <i>Oceanites</i> | <i>oceanicus</i> | 81 | 24 |
| White-bellied storm-petrel | Aves | Procellariiformes | Hydrobatidae | <i>Fregetta</i> | <i>grallaria</i> | 2 | 2 |
| Unidentified storm-petrel | Aves | Procellariiformes | Hydrobatidae | <i>n/a</i> | <i>n/a</i> | 16 | 11 |
| Slender-billed prion | Aves | Procellariiformes | Procellariidae | <i>Pachyptila</i> | <i>belcheri</i> | 3 | 2 |
| Unidentified prion | Aves | Procellariiformes | Procellariidae | <i>Pachyptila</i> | <i>n/a</i> | 241 | 43 |
| Unidentified penguin | Aves | Sphenisciformes | Spheniscidae | <i>n/a</i> | <i>n/a</i> | 1 | 1 |
| Great shearwater | Aves | Procellariiformes | Procellariidae | <i>Puffinus</i> | <i>gravis</i> | 792 | 58 |
| Sooty shearwater | Aves | Procellariiformes | Procellariidae | <i>Puffinus</i> | <i>griseus</i> | 21 | 12 |
| Unidentified shearwater | Aves | Procellariiformes | Procellariidae | <i>Puffinus</i> | <i>n/a</i> | 3 | 3 |
| Snowy sheathbill | Aves | Charadriiformes | Chionidae | <i>Chionis</i> | <i>albus</i> | 116 | 35 |
| Black-chinned siskin | Aves | Passeriformes | Fringillidae | <i>Carduelis</i> | <i>barbata</i> | 1 | 1 |
| Brown (Subantarctic) skua | Aves | Charadriiformes | Stercorariidae | <i>Stercorarius</i> | <i>antarcticus</i> | 24 | 17 |
| Chilean skua | Aves | Charadriiformes | Stercorariidae | <i>Stercorarius</i> | <i>chilensis</i> | 9 | 7 |
| Unidentified skua | Aves | Charadriiformes | Stercorariidae | <i>Stercorarius</i> | <i>n/a</i> | 3 | 3 |
| Unidentified swallow | Aves | Passeriformes | Hirundinidae | <i>n/a</i> | <i>n/a</i> | 1 | 1 |
| Sooty tern | Aves | Charadriiformes | Sternidae | <i>Sterna</i> | <i>fuscata</i> | 8 | 1 |
| Kelp gull | Aves | Charadriiformes | Laridae | <i>Larus</i> | <i>dominicanus</i> | 3 | 2 |
| Peregrine Falcon | Aves | Falconiformes | Falconidae | <i>Falco</i> | <i>peregrinus</i> | 4 | 4 |

3. DATA FORMS

3.1. BASIC DATA FORM

| BASIC DATA FORM | | |
|--|---|------------------|
| Project Number | UOS01285M | |
| Seismic Contractor | PGS Geophysical | |
| Client | Noble Energy Falklands Ltd. | |
| Geographical coordinates of the survey area: | Latitude | Longitude |
| | between | 55.21673°S |
| | and | 56.81693°S |
| | | 52.76204°W |
| | | 51.88380°W |
| Survey Type | 3D | |
| Vessel and/or Rig Name | Ramform Sterling | |
| Permit Number | PON3 | |
| Location / Distance of Airgun Deployment | Astern 700m | |
| Water Depth | Min | 900 |
| | Max | 1700 |
| Dates of project | 29 Nov 2012 | THROUGH |
| Dates included in reporting period | 29 November 2012 through 4 June 2013 | |
| Total time airguns operating – all power levels: | 1515:12 | |
| Amount of time airguns operating at full power: | 1400:22 | |
| Time airguns operating at full power on a survey line: | 1345:37 | |
| Time airguns operating at full power approaching survey line: | 54:45 | |
| Amount of time compliance gun operations: | 00:00 | |
| Amount of time in ramp up: | 87:18 | |
| Number daytime ramp ups: | 153 | |
| Number of night time ramp ups: | 110 | |
| Number of ramp ups from mitigation source: | 0 | |
| Number of ramp ups from PAM system: | 124 | |
| Amount of time conducted in airgun testing: | 27:32 | |
| Duration of visual observations: | 1723:32 | |
| Duration of observations while airguns firing: | 902:03 | |
| Duration of observation during airgun silence: | 821:29 | |
| Duration of acoustic monitoring: | 732:21 | |
| Duration of acoustic monitoring while airguns firing: | 424:42 | |
| Duration of acoustic monitoring during airgun silence: | 307:39 | |
| Visual Observers: | Lynn Henneberger (29 Nov 12- 23 Jan 13) / Heidi Ingram (23 Jan - 3 Apr 2013) / Megan McManus (3 Apr-16 May 13) / Lacey Price (16 May-4 June 13) | |
| | Monica Arancibia-Colgain (29 Nov 12- 23 Jan 13, 16 May-4 June 13) / Tatiana Moreno (23 Jan 13 – 3 Apr 13), Alexandria Denby (3 April-15 May 13) | |
| Acoustic Observers: | Greggo Seward (29 Nov 12- 23 Jan 13, 3 Apr -16 May 13) / Emily Ellis (23 Jan 13 – 3 Apr 13), Jessica Mucci (16 May-4 June 13) | |
| Number of Marine Mammals Visually Detected: | 231 Detections (1620 Individuals) | |
| Number of Marine Mammals Acoustically Detected: | 1 | |
| Number of acoustic detections confirmed by visual sighting: | 0 | |
| Number of visual sighting confirmed by acoustic detection: | 0 | |
| Number of Pinnipeds detected: | 3 | |
| List Mitigation Actions (eg. shutdowns, ramp up delays, turtle pauses) | Ramp up delays (25 Mar & 19 May) | |
| Duration of operational downtime due to mitigation: | None | |
| Flagged Data Forms (list sheet & date) | None | |

3.2. COVER DATA SHEET

| | | | | | |
|--|---|--|--|--|--|
| Regulatory reference number (e.g. DECC no., MMS permit no., OCS lease no., etc.) PON3 | | 4. COUNTRY Falkland Islands, UK | | Ship/ platform name <i>Ramform Sterling</i> | |
| Client Noble Energy Falklands Ltd | | Contractor PGS | | Survey type <input type="checkbox"/> site <input type="checkbox"/> 4C <input type="checkbox"/> 2D <input type="checkbox"/> VSP <input checked="" type="checkbox"/> 3D <input type="checkbox"/> WAZ <input type="checkbox"/> 4D <input type="checkbox"/> other <input type="checkbox"/> OBC | |
| Start date 29 November 2012 | | End date 4 June 2013 | | | |
| Number of source vessels 1 | Type of source (e.g. airguns) Airguns | Number of airguns (only if airguns used) 62 | Source volume (cu. in.) 4130 | | |
| Source depth (metres) 7 | Frequency (Hz) 70 | Intensity (dB re. 1µPa or bar metres) 69 bar meters | Shot point interval (seconds) 10.4 | | |
| Method of soft start <input checked="" type="checkbox"/> increase number of guns <input type="checkbox"/> increase pressure <input type="checkbox"/> increase frequency <input type="checkbox"/> other | | | | | |
| Visual monitoring equipment used (e.g. binoculars, big eyes, etc.) Binoculars | Magnification of optical equipment (e.g. binoculars) 7x50; 10x30 | Height of eye (metres) 22.4 | How was distance of animals estimated? <input checked="" type="checkbox"/> by eye <input type="checkbox"/> with laser rangefinder <input type="checkbox"/> with rangefinder stick/ callipers <input checked="" type="checkbox"/> with reticle binoculars <input checked="" type="checkbox"/> by relating to object at known distance <input type="checkbox"/> other | | |
| Number of dedicated MMOs 2 | Training of MMOs <input checked="" type="checkbox"/> JNCC approved MMO induction course for UK waters <input checked="" type="checkbox"/> PSO training course for the Gulf of Mexico <input type="checkbox"/> MMO training course for Irish waters <input type="checkbox"/> other <input type="checkbox"/> none | | | | |
| Was PAM used? <input checked="" type="checkbox"/> yes <input type="checkbox"/> no | Number of PAM operators 1 | | | | |
| Description of PAM equipment 20 m airgun towed array | | | | | |
| Range of PAM hydrophones from airguns (metres) 15 | Bearing of PAM hydrophones from airguns (relative to direction of travel) 180 | Depth of PAM hydrophones (metres) 6.5 | | | |

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Protected Species Observer Report

Prepared for: PGS

PGS

Ramform Titan

East Falkland Basin (FINA13)

05 November 2013 – 18

February 2014



Project No.: UMS04150

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1 Executive Summary

PGS Geophysical conducted this 3D seismic survey for Noble Energy Falkland Islands Ltd., operated within a permitted area as mentioned in Falklands Islands permit Petroleum Operations Notice 3 (PON3) executed on 22 October 2013. The ensuing report reviews the protected species monitoring activities and seismic survey operations undertaken on the *Ramform Titan* for the period of 5 November 2013 through 18 February 2014. The information supplied in this report is in reference to the mitigation guidelines set forth in the Joint Nature Conservation Committee's August 2010 *JNCC guidelines for minimising the risk of injury and disturbance to marine mammals from seismic surveys*.

Two trained visual marine mammal observers (MMOs), contracted through Vision Project Services, and a dedicated passive acoustic monitoring (PAM) operator, contracted through RPS, were onboard to fulfill the regulatory requirements and reporting mandated by the Joint Nature Conservation Committee. Mitigation measures were implemented to minimize potential impacts to marine mammals throughout the duration of the survey.

Mitigation measures included the use of MMOs during daylight hours and PAM during night-time hours to provide clearance of the 500 meter exclusion zone and implement delays in soft start, if necessary. PAM was also utilized as a mitigation tool to allow for soft starts from source silence during periods of reduced visibility, such as episodes of fog or inclement weather, when visual monitoring for protected species could not be conducted fully as outlined within the PGS Geophysical/Noble Energy Falklands Limited Project Plan section 9.3 the Environmental Management Plan (EMP).

Deployment of seismic equipment began the morning of 1 November 2013. Initial airgun testing commenced on 5 November. Production of Noble Energy Falkland LTD survey began at 14:38 UTC on 6 November 2013. On 24 November 2013 production ceased when inclement weather damaged the spread and the seismic gear was retrieved. The *Ramform Titan* continued at anchor in Berkeley sound until after crew change, 4 December 2013, after which the seismic streamers were redeployed and production resumed on 10 December 2013. After which, there was only minor downtime due to weather during the remainder of the survey.

There were a total of three soft starts of 19 minutes in duration which occurred on 22 January, 16 February and 18 February. On 2 February, from 11:35 through 11:47 there was an episode of unplanned auto-firing. The pre-watch was cleared prior to auto-firing by visual MMOs from 10:35 to 10:40 prior to losing visibility due to fog. Daytime PAM began at 10:50, resulting in a total of unplanned clearance period of 45 minutes in duration through PAM. The entire clearance period prior to auto-firing was 50 minutes in duration.

PAM Operators undertook acoustic watches, accumulating a total of 452 hours and 39 minutes of acoustic monitoring over the course of the survey, 312 hours and 26 minutes of which were completed while the acoustic source was active. There were 14 necessary joint pre-searches which required a combination of passive acoustics and visual protected species observation. In addition to the joint pre-surveys, there was a total of 70 passive acoustic watches conducted prior to the commencement of source operations.

The acoustic monitoring effort detected a project total of eight protected species detection records which included three baleen whales, four Delphinidae and one sperm whale detection. There were no correlated visual and acoustic detections during this project.

Acoustic detections of protected species resulted in no mitigation actions.

2 Introduction

A *Marine Mammal Observer* report was prepared by the Vision Project Services marine mammal observers detailing the Noble Energy Falklands LTD survey operations, with a focus on visual monitoring effort, protected species detections and mitigation actions implemented over the course of the survey. This *Passive Acoustic Monitoring and Mitigation* report was prepared by RPS to inform in detail the operation of the PAM system during the Noble Energy Falkland Island marine 3D seismic survey and source operations during this project. As such, survey specifics not pertaining to the PAM system are not discussed in detail but may be found in the *Marine Mammal Observer* report.

2.1 Project Location and Operation Parameters

The East Falkland Island Basin project was located approximately 230 kilometres northeast of Port Stanley, Falkland Islands (Figure 1), in the Southern Quadrants 30 (block numbers 7, 11-15 and 16-30) 31 (block number 16) and 42 (block numbers 1-4 and 7-8). The prospect covered 8,858 square kilometres in water depths ranging from 1000 to 1650 meters. It consisted of 96 survey lines spaced at 720 meter intervals with line changes varying from three to seven hours. PGS's *Ramform Titan* along with two support vessels, the *Falcon Explorer* and *M/V Fellowship*, executed this survey.



Figure 1: Location of the survey in relation to the Falkland Islands.

The *Ramform Titan* towed two airgun arrays, separated by 60 meters, both sources comprised of three sub-arrays, separated by eight meters and towed at seven meters \pm 1 meter. Each sub-array consisted of ten (1 string) to twelve (2 strings) airguns and all were equipped with one spare airgun. The sources alternated with a shot point interval of 25 meters. The full source included a total of 34 airguns, ranging in volume from 40 in³ to 250 in³, with a maximum source volume of 4130 in³ and a pressure of 2000 psi. Intensity of the source was 69.78 bar meters and the frequency range was a broadband frequency from 0 to 214 hertz. The center of source was 545 meters from the vessel reference point (VRP), which was

2.2 Passive Acoustic Monitoring (PAM) Parameters

A *Seiche Measurements* PAM system was installed by *Seiche Measurements Limited* in dock in Bergen, Norway on 2 October 2013 before the *Ramform Titan* transited to the Falkland Islands. The PAM system was onboard during all operations until the end of the Noble Energy Falklands Limited survey and remained onboard for the following PGS survey.

The PAM system consisted of a 250 meter conventional towed array, 100 meter deck cable, a 20 meter airgun-towed array, a data processing unit with rack mount computer and slide out monitor, net-top computer (for remote monitoring), dual monitors, analysis software program (*Pamguard*), headphones for aural monitoring, various leads and adapters and a complete spare set of aforementioned system. Two additional spare hydrophone towed array cables were received on board the *Ramform Titan* on 4 December 2013.

The liner hydrophone array cable contained four hydrophones and a depth gauged plotted into a 20 meter section. All four elements were broadband, two with a frequency response of 2 to 200 kilohertz and two have frequency responses of 0.2 hertz to 200 kilohertz. Preamplifiers were also embedded into the cable forward of each element. The four-element linear hydrophone array design permitted a broad range for sampling marine mammal vocalizations, from the low frequency moans of baleen whales to the ultra-high frequency clicks of beaked whales.

The distance between the hydrophone array cable and the data processing unit was greater than the remaining hydrophone cable after deployment. To join the hydrophone tow cable to the DPU, a 100 meter deck cable was supplied as an extension and routed from the Lead-in Repair shop to the back deck.

The data processing unit processed the raw data from the hydrophones through two external sound cards, a *National Instruments DAQ* card and an *ASIO Fireface* card (contained within a *RME Fireface 800* unit). The *National Instruments DAQ* sound card sampled raw audio at 500 kilohertz and can be used to detect beaked whale, *Kogia* species, porpoise, and delphinid (echolocation) clicks up to up to 250 kilohertz. The *ASIO Fireface* sound card sampled audio at 48 kilohertz and can be used to detect mysticete, delphinid, and non-delphinid odontocetes (including sperm whale) vocalizations up to 48 kilohertz. The data processing unit also contains a *Measurements and Computing* data logger for the depth gauge, digital signal amplifiers, an *UltraLink Pro* audio mixer, and an *UltraCurve Pro* graphic equalizer.

The two sound cards supplied low and high frequency digital audio feed to a computer that contained a suite of software for monitoring cetacean acoustics. *Pamguard* (Beta version 1.12.05) was the primary software utilized on this survey. The International Federation on Animal Welfare (IFAW) software including *Logger 2000*, *Rainbow Click*, *Whistle*, and *Rainbow Click Porpoise* was the secondary software and was only to be used as back-up software if the *Pamguard* software failed.

The computer received both the raw audio from the *National Instruments DAQ* sound card as well as the raw audio from the *ASIO Fireface* sound card. *Pamguard* modules included both high and low frequency click detectors, a whistle and moan detector, spectrogram, a map with a direct GPGGA GPS feed from the *Ramform Titan's* navigation system, and high and low frequency sound recorders. Raw audio from the *ASIO Fireface* sound card was monitored aurally with *Sennheiser* headphones.

The conventional hydrophone towed array was deployed off lead-in 12 on the port side of the *Ramform Titan*. When deployed there was approximately 181 meters of hydrophone cable was towed astern (Figure 3). Spectra line was paired with the hydrophone array cable, minus the hydrophone section (the final 23 meters), to relieve tension on the cable during deployment, retrieval and severe weather. Forward of the free floating cable the spectra line was attached via a Chinese finger and taped at one to two meter intervals (Figure 4 & 5D). A buoy was fixed to the Spectra line, via a small shackle, 126 meters astern to discern position and help create drag during deployment (Figure 5A). Positioned 50 meters astern the spectra line was affixed to a sliding collar (Figure 5C) on lead-in 12, shackles attached the remaining 50 meters to lead-in 12 to prevent the Spectra line and hydrophone cable from dragging in water (Figure 6).

Prior to deployment, six kilograms of chain weight was added to the Spectra line approximately 11 meters ahead of the first hydrophone element (Figure 5B). The additional weight was used to assist the cable in descending to an appropriate towing depth. Once deployed the cable was approximately 324 meters from the center of source and towed at a depth of approximately six meters +/-1.5 meters. The conventional towed hydrophone array was retrieved and deployed 13 times each during the survey (Table 1).

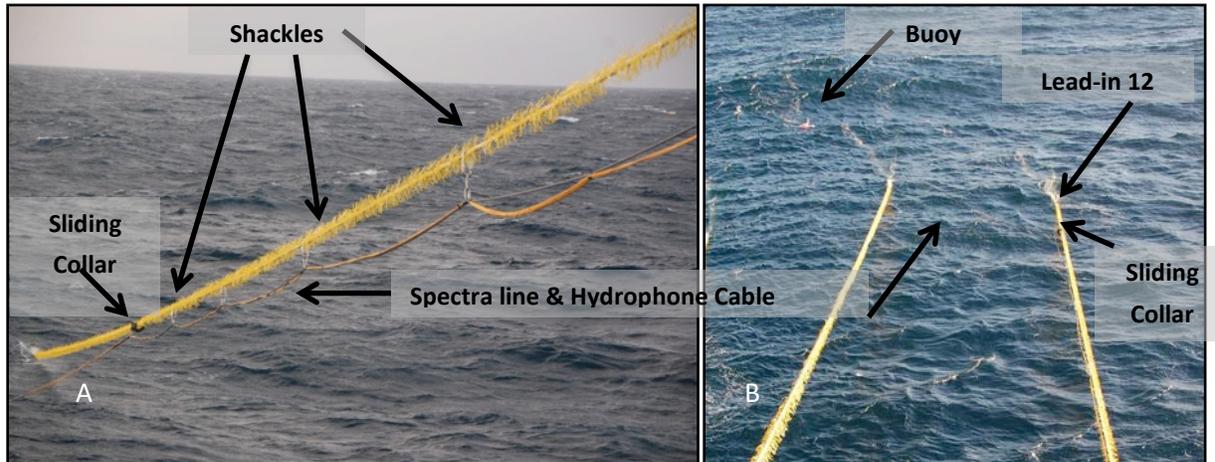


Figure 3: Deployment off port stern of Ramform Titan. A) Sliding collars and shackles B) Buoy and lead-ins.

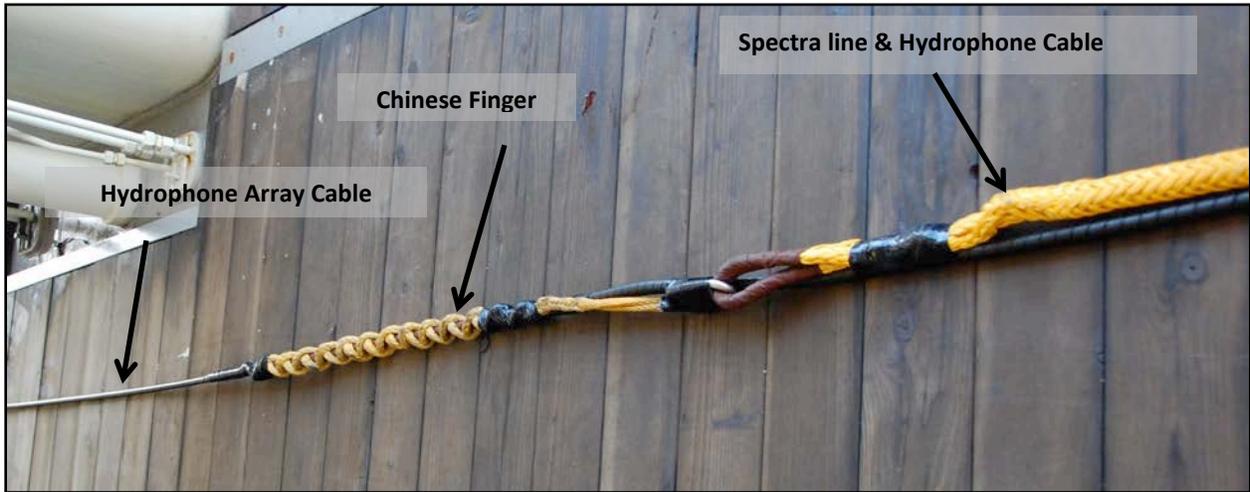


Figure 4: Free floating hydrophone array cable attached to Spectra line via Chinese finger.

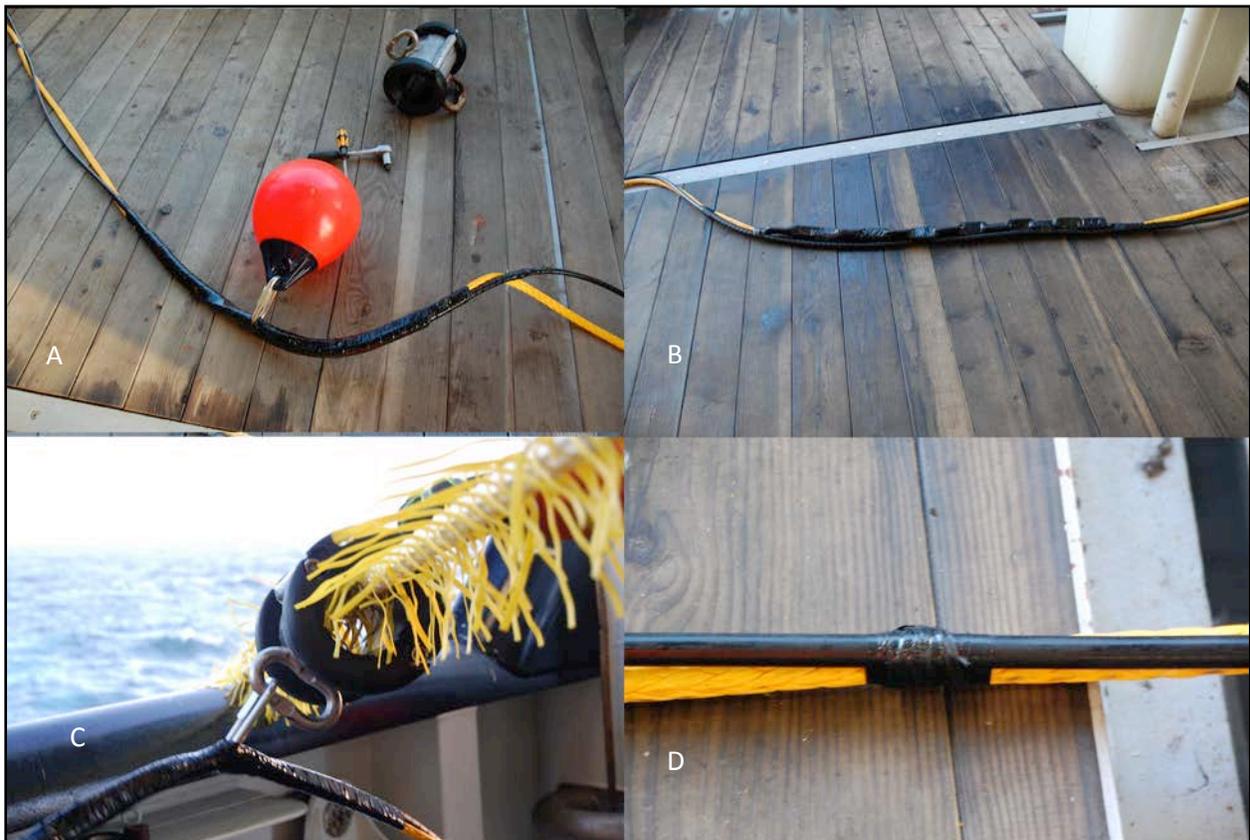


Figure 5: Attachment points, all attachment points are connected directly to Spectra line. A. Buoy B. Weights C. Sliding collar D. Hydrophone towed array cable.

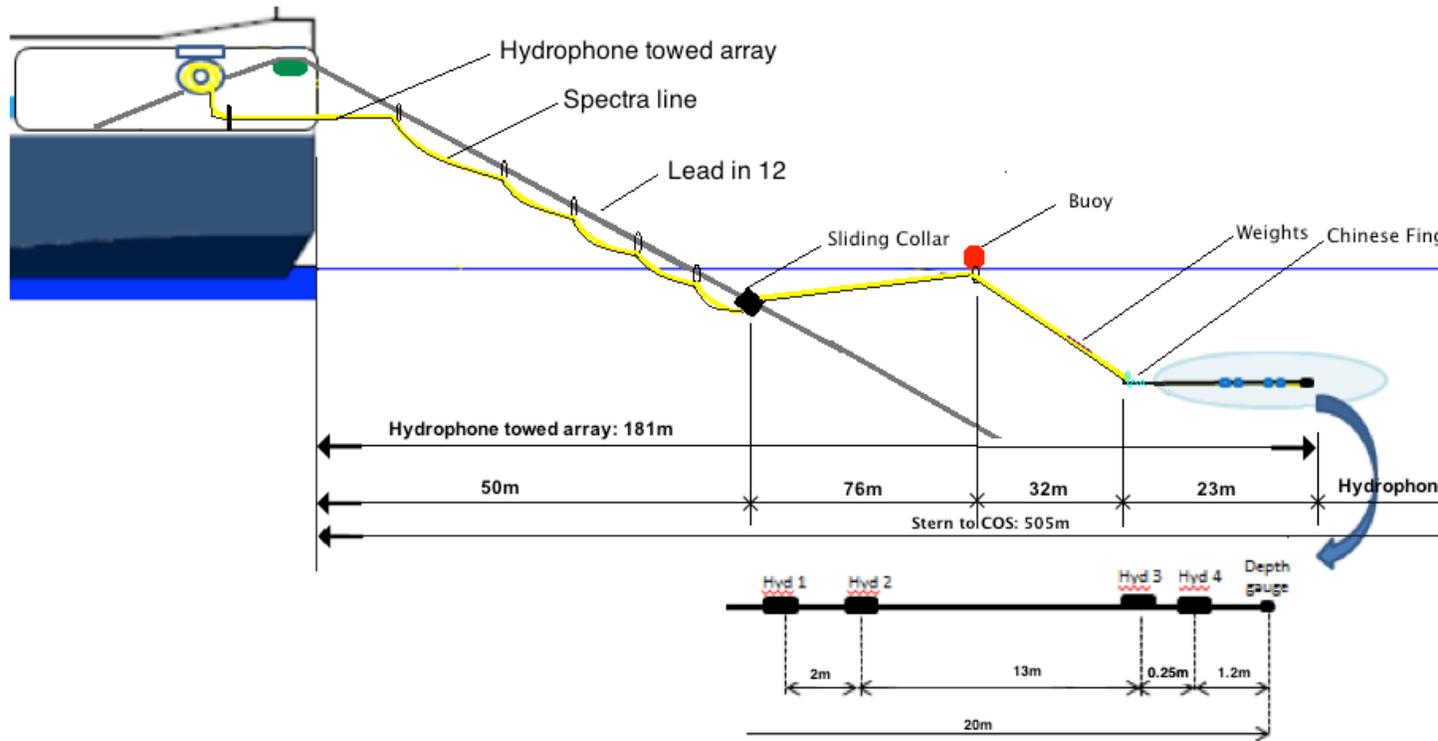


Figure 6: Sketch of the hydrophone deployment on the Ramform Titan.

Table 1: PAM hydrophone cable deployment and retrieval during the East Falkland Basin survey.

| Deployment or Retrieval | Date | Time (UTC) | Reason | Comments |
|--------------------------|---------|------------|--|---|
| Deployment | 02 Nov. | 08:30 | Seismic equipment deployed. Setting up gear for survey. | Deployment off sliding collar on lead in 10, port side. Deployment took one hour due to attaching Chinese fingers to hydrophone cable |
| Retrieval and Deployment | 03 Nov. | 07:30 | Changing deployment set up. Hydrophone depth only four meters. | Retrieval and deployment took one hour due to moving of Chinese fingers. |
| Retrieval and Deployment | 09 Nov. | 00:00 | ~30m of towed hydrophone array cable broke off | Damage cable stored and spare cable briefly deployed. |
| Retrieval | 09 Nov. | 08:30 | Secured on deck for daylight hours | Briefly deployed cable. |
| Deployment | 09 Nov. | 22:30 | Cable deployed temporarily for PAM monitoring. | |
| Retrieval | 10 Nov. | 08:30 | Secured on deck for daylight hours | Briefly deployed cable. |
| Deployment | 10 Nov. | 15:30 | PAM monitoring for fog during daylight hours | |
| Retrieval | 10 Nov. | 17:00 | Set up for new deployment | |
| Deployment | 10 Nov. | 22:00 | New deployment set up. | Deployed 131m off port stern on lead in 12, 50m to sliding collar another 26m to buoy and 55m free |

| | | | | |
|------------|---------|-------|---|--|
| | | | | floating. Deployment took ~2 hours. |
| Retrieval | 15 Nov. | 22:30 | Secure on deck due to up coming weather | |
| Deployment | 17 Nov. | 00:30 | Deployed to resume acoustic monitoring. | Deployment changed to lengthen distance between sliding collar and buoy, making for easier deployment. |
| Retrieval | 20 Nov. | 16:00 | Secured on port winch due to up-coming weather | |
| Deployment | 22 Nov. | 01:00 | Deployed to resume acoustic monitoring | |
| Retrieval | 22 Nov. | 16:00 | Secured on port winch | Streamer maintenance on port side. |
| Deployment | 23 Nov. | 11:30 | Deployed to resume acoustic monitoring | Daytime soft start due to fog. |
| Retrieval | 25 Nov. | 21:00 | All of spread coming on board for repair | |
| Deployment | 10 Dec. | 20:00 | Seismic gear deployed, start of production | PAM hydrophone cable replaced before deployment (old cable being used as spare) |
| Retrieval | 06 Jan. | 10:00 | Picking up seismic gear for crew change | Cable damaged |
| Deployment | 12 Jan. | 18:00 | Deploying gear after crew change | Damaged hydrophone cable SM.3806 was replaced with SM.3439. |
| Retrieval | 05 Feb. | 22:00 | Brought onboard for inspection and poor weather | Cable appeared in good condition. |
| Deployment | 07 Feb. | 08:30 | Deployed to resume acoustic monitoring | |
| Retrieval | 10 Feb. | 13:00 | Picking up seismic gear for crew change | |
| Deployment | 15 Feb. | 20:00 | Deployed to resume acoustic monitoring | |
| Retrieval | 18 Feb. | 20:00 | End of project. | |

2.3 Passive Acoustic Monitoring (PAM) Survey Methodology

This project utilized passive acoustic monitoring (PAM) to acoustically monitor during the night and during times of reduced daytime visibility, in order to clear the exclusion zone prior to soft start when applicable. In addition to inclement weather and fog (including all reduced daytime visibility prior to soft start), the PAM system was monitored on a nightly basis with a start of watch time ranging from 22:30 to 00:25 UTC and an end of watch time from 06:45 to 08:35 UTC. PAM monitoring began on the night of 05 November 2013 and continued through the duration of source operations ending on 18 February 2014.

Passive acoustic monitoring surveys were conducted by the PAM operator in two to four hour shifts separated by one to two hour breaks. Acoustic monitoring for marine mammals was completed aurally with *Sennheiser* headphones and visually with *Pamguard 1.12.05 Beta*. Delphinid whistles, clicks, and burst pulses and sperm whale clicks can be viewed on a spectrogram display within *Pamguard*. In addition, sperm whale, beaked whale, *Kogia* species, and delphinid echolocation clicks can also be viewed on *Pamguard's* low and high frequency click detector displays.

Distances for acoustic detections were primarily based upon a noise or detection score system developed by Gannier *et al* (2002). Gannier *et al* monitored sperm whales (*Physeter macrocephalus*) in the Mediterranean both visually and acoustically. A subjective scale was developed based upon the strength or intensity of the sperm whale clicks at various distances that were measured visually when the sperm whale surfaced. Although the scale is subjective, and sounds produced in marine environments will vary according to local conditions, the scale provided a measure for approximating distances when using a single, linear hydrophone array.

In addition, *Pamguard 1.12.05 Beta* contained a function for calculating an approximate range based upon the least squares fit test. The mathematical function estimated the range to vocalizing animals by calculating the most likely crossing of a series of bearing lines generated from tracked clicks. The clicks were tracked in the click detector module and the bearing lines associated with each click were then plotted on the map display. After several clicks were tracked an estimated range with an associated error was displayed on the map. Several click trains, over a few minutes in duration, were required to obtain a reasonable estimate of range.

During a detection of a vocalizing animal, information regarding position, distance, water depth, and range of animal, if applicable, was recorded, along with any recordings of vocalizations. A detection report was completed, utilizing information gathered from the navigation department, as well as, from the *Pamguard* modules. All information gathered was input into the Detection Report forms and the Daily Passive Acoustic Forms (Appendix B; Appendix C). Details of these detections were submitted in Daily and Weekly Reports in combination with visual MMO data.

2.4 Survey Data

2.4.1 Source Operations Survey Summary

The *Ramform Titan* arrived in Falkland Islands on 29 October 2013. After crew change and vessel fueling, deployment of the seismic gear commenced on 1 November 2013. The seismic source was first fired for testing at 11:56 UTC on 05 November 2013 and production on East Falkland Basin continued until the last shot of the survey at 19:19 on 18 February 2014. Two weeks, 24 November through 10 December, the source was not active due to retrieval of gear, weather and crew change. Once more, from 10 to 15 February the source was silent during seismic gear retrieval for the final crew change.

During the survey, the seismic source was active for a total of 1168 hours and 34 minutes. The majority, 92%, consisted of full volume source operations (1075 hours and 16 minutes); 1039 hours and 22 minutes while in production and 35 hours and 54 minutes during non-production full volume firing. The remaining 8% consisted of soft start, testing operations and a period of auto-firing; 48 hours and 30 minutes in testing, 44 hours and 36 minutes in soft start and 12 minutes of recorded auto-firing (Figure 7). Over the duration of the survey, from the first test shot until the last production shot, there was 1,406 hours and 49 minutes of acoustic silence, resulting in 54.63% silence during the survey.

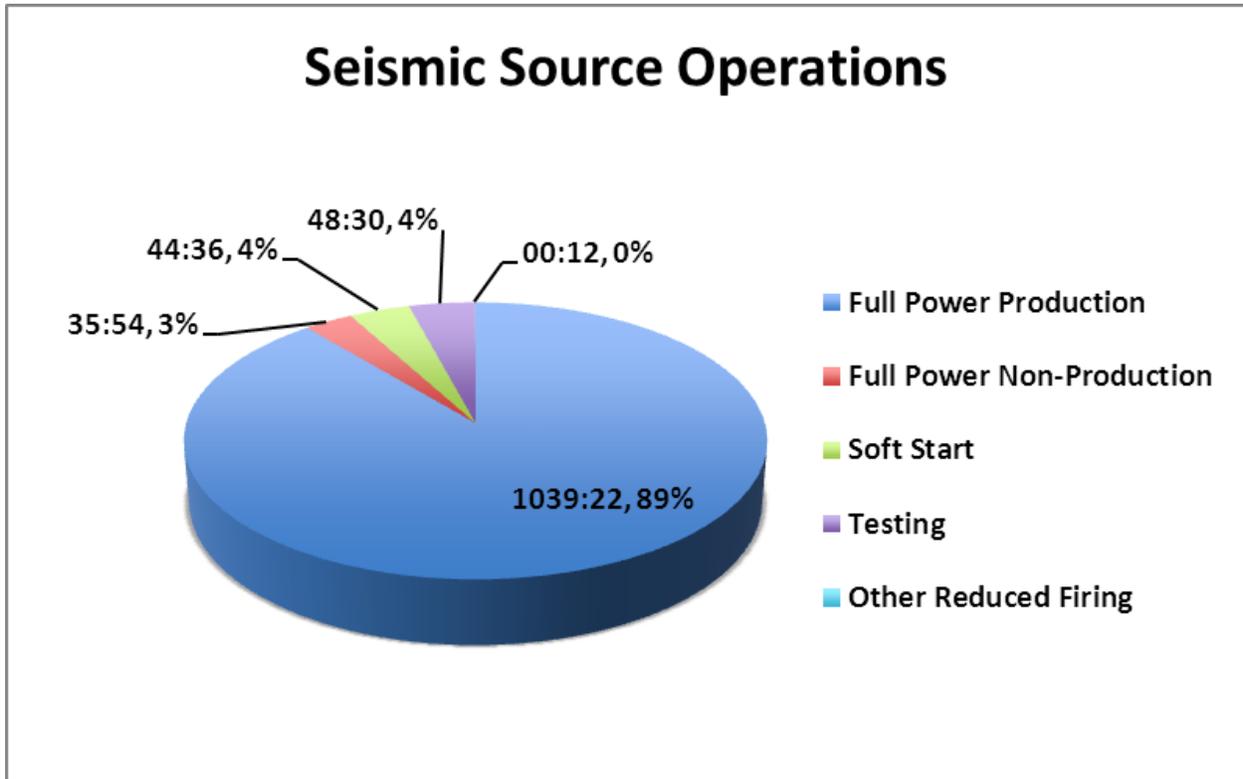


Figure 7: Breakdown of seismic source operations.

There were 130 soft starts performed over the course of the survey, 122 in order to commence full power survey operations and eight prior to acoustic source testing. In addition there was one episode of auto-fire from silence and 45 acoustic source tests that did not reach full volume (and therefore, a soft start was not necessary). Though soft starts were conducted using an automated acoustic source controller program which adds airguns sequentially to achieve full source, there were three episodes of 19 minute soft starts. Prior to soft starts and airgun testing the area was monitored for 60 minutes either visually, during daytime, acoustically, during night-time or periods of limited visibility, or a combination of the two. The only exception was during a time of auto-fire which continuous monitoring was briefly interrupted due to fog. The exclusion zone was cleared of marine mammals and sea turtles prior to the seismic source activity a total of 166 times. The exclusion zone was cleared 82 times through visual monitoring, 70 times through acoustic monitoring and 14 times through a necessary joint effort between visual and acoustic monitoring (Table 2; Figure 8). Of the 82 visual clearances, four were clearances after source silences of less than ten minutes in duration. Furthermore, two acoustic clearances of the total 70 were granted clearance of the seismic source during silences of less than ten minutes during night-time operations. Of the 130 soft starts, 52 occurred during night-time operations.

Table 2: Total acoustic source operations during the East Falkland Basin project.

| Acoustic Source Operations | Number | Duration (hh:mm) |
|---|--------------|------------------|
| Airgun Tests | | 48:30 |
| Soft start | 130 | 44:36 |
| Day-time pre-firing cleared by visual observation | 82 | |
| Day-time pre-firing cleared by PAM | 20 (4 joint) | |
| Night time pre-firing cleared by PAM | 54 | |
| Pre-firing cleared with a combination of PAM and visual | 14 | |
| Full power survey acquisition | | 1039:22 |
| Full power non-production | | 35:54 |
| Total time acoustic source was active | | 1168:34 |

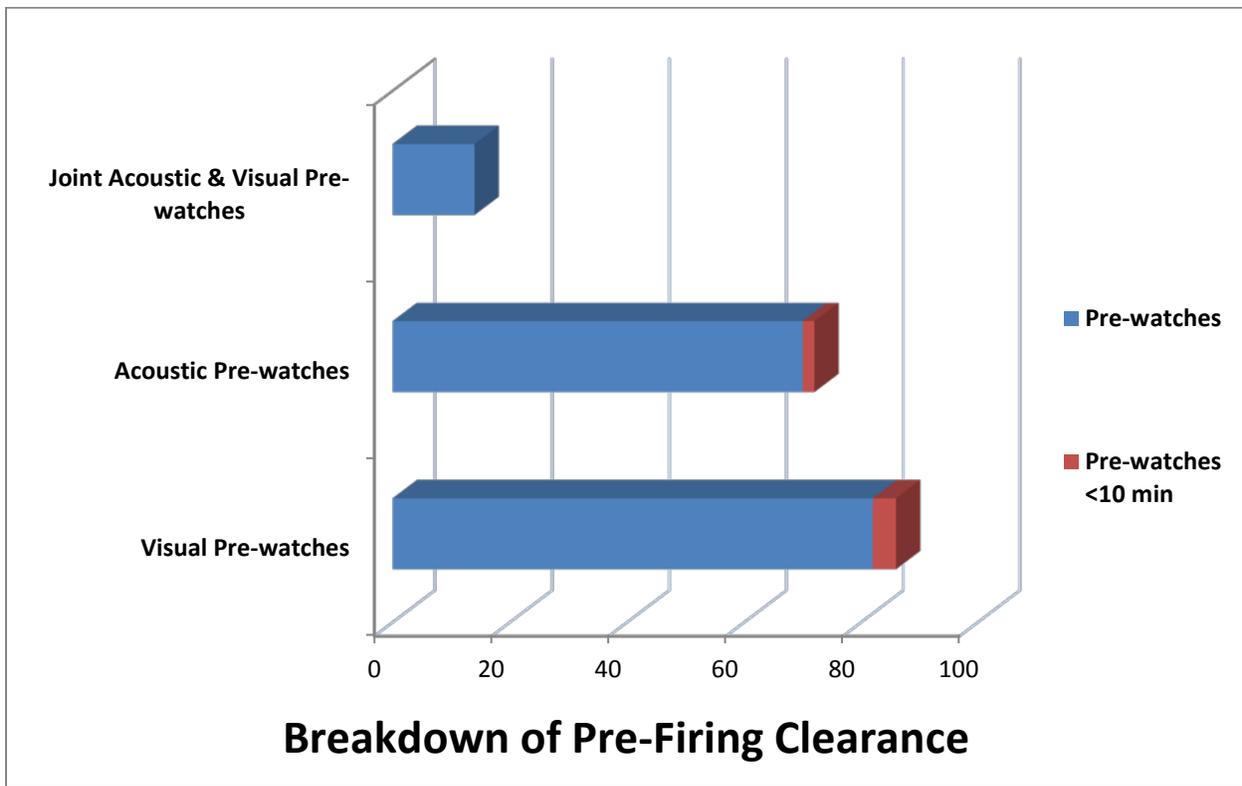


Figure 8: Breakdown of Pre-firing clearance.

2.4.2 Acoustic Monitoring Survey Summary

The RPS PAM operator monitored the hydrophones aurally and visually monitored the *Pamguard* detection software from sunset until sunrise and as needed during reduced visibility (fog or inclement weather). Acoustic monitoring began on 05 November 2013 at 23:00 UTC during a series of airgun tests and continued throughout the East Falklands Basin project. Acoustic monitoring for the project ended at 14:16 UTC on 18 February 2014 when the final survey line was cleared through PAM during a time of

limited visibility due to fog. After the completion of the final survey line, the hydrophone cable was retrieved prior to the retrieval of seismic equipment. The four hydrophone array cables were packed up for demobilization in the next port of call. All other PAM equipment was set to remain onboard.

Over the course of this project, the PAM operators conducted 452 hours and 39 minutes of acoustic monitoring; the seismic source was active during 312 hours 26 minutes (70%) during acoustic monitoring and silent during 140 hours 13 minutes (30%) of acoustic monitoring (Figure 8). Daytime PAM was used to clear the exclusion zone 16 times during this project and overlapped with daytime pre-watches with visual monitoring an additional four times resulting in 20 total periods of passive acoustic monitoring during daytime limited visibility.

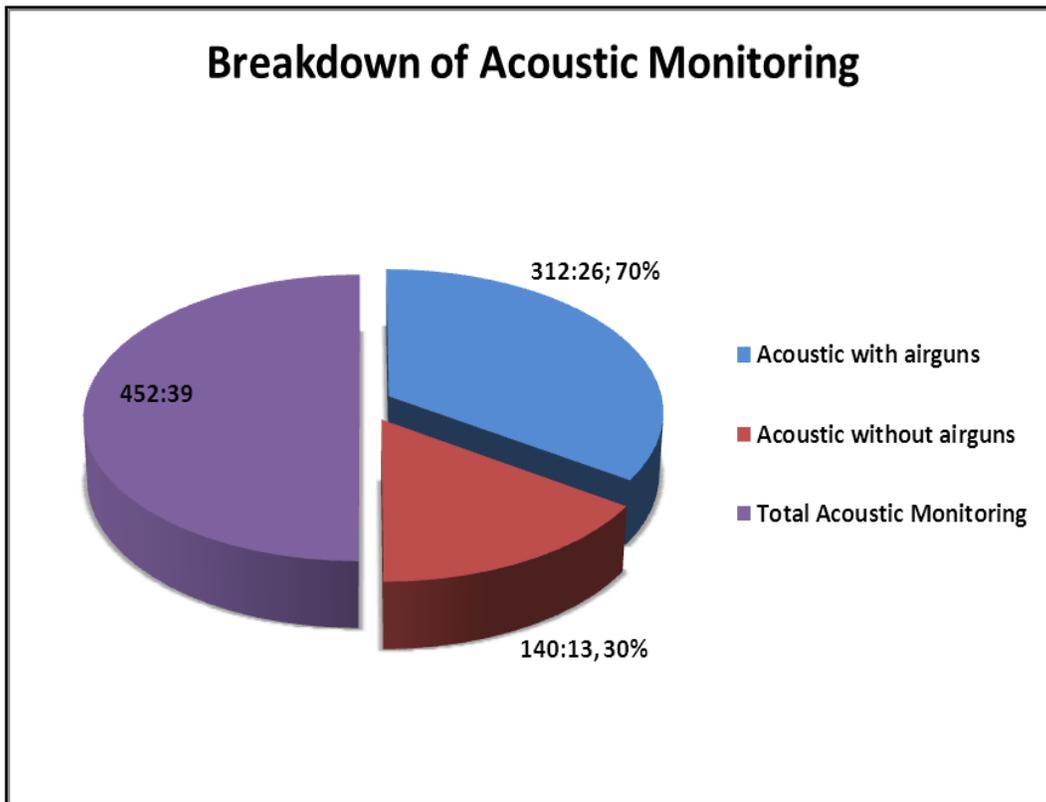


Figure 9: Breakdown of acoustic monitoring

2.5 Mitigation Actions

No mitigation actions were taken as a result of acoustic monitoring during this project.

3 Wildlife Summary

3.1 Acoustic Detections

Acoustic monitoring on the *Ramform Titan* from 05 November 2013 to 18 February 2014 produced eight acoustic detections; three were recorded as unidentified baleen whales, suborder Mysticeti, four from the family Delphinidae and one as a sperm whale (*Physeter macrocephalus*).

Distribution of six of the eight acoustically made detections occurred near the northern section of the prospect, one in the southern end and one mid-prospect (Figure 10). The baleen whales were observed in water depths from 1322 to 1545 meters, the sperm whale 1541 meters and the Delphinidae in 1380 to 1509 meters.

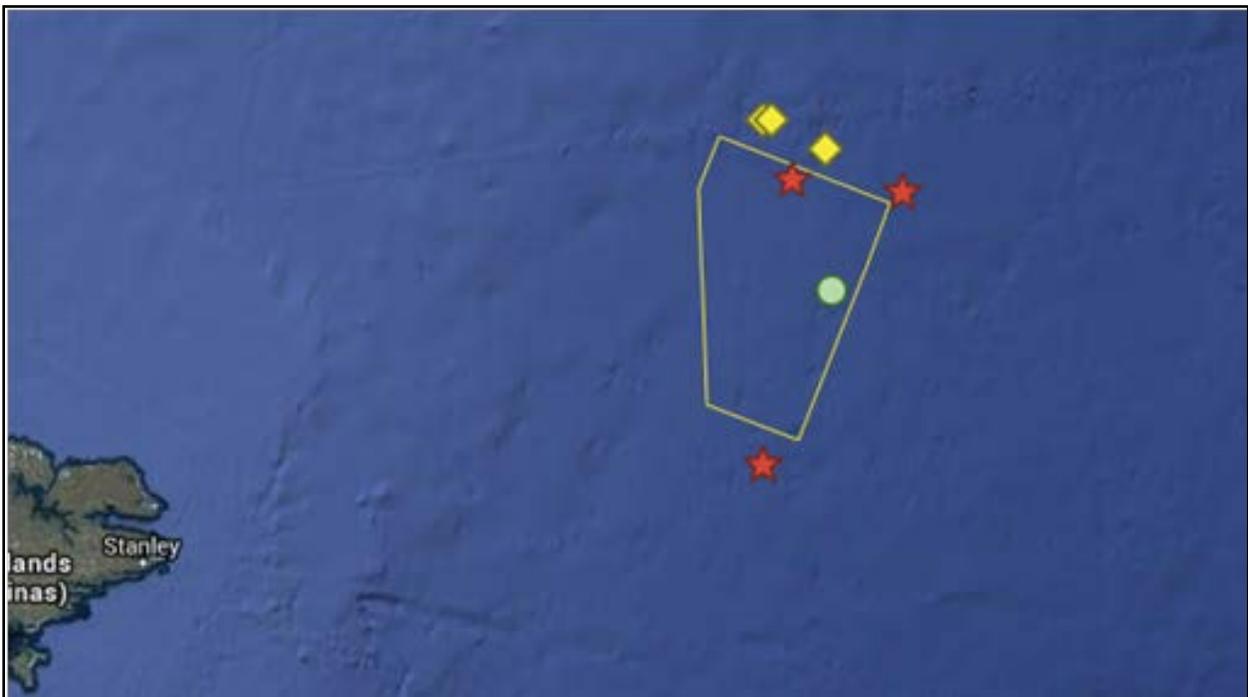


Figure 10: Distribution of acoustic detections mapped on Google Earth: yellow represents the suborder mysticeti, red the family Delphinidae and green sperm whale.

On 11 December 2013 at 06:38 UTC extremely faint double calls were detected by the PAM operator aurally through the headphones and then visually on the *Pamguard LF spectrogram* (acoustic detection 501). Due to the faintness of the calls and that the moans did not register on the *Pamguard Whistle and Moan detector*, distance was determined by the PAM operator as being outside the exclusion zone. The majority of the detection consisted of pairs of 'whoops' with peak energy at 205 hertz (between 165 and 225 hertz). They occurred at regular intervals of about 5.5 seconds between pairs, and 0.6 seconds between calls. The calls themselves about were approximately 0.25 seconds long, tonal and pulse-like (Figure 11).

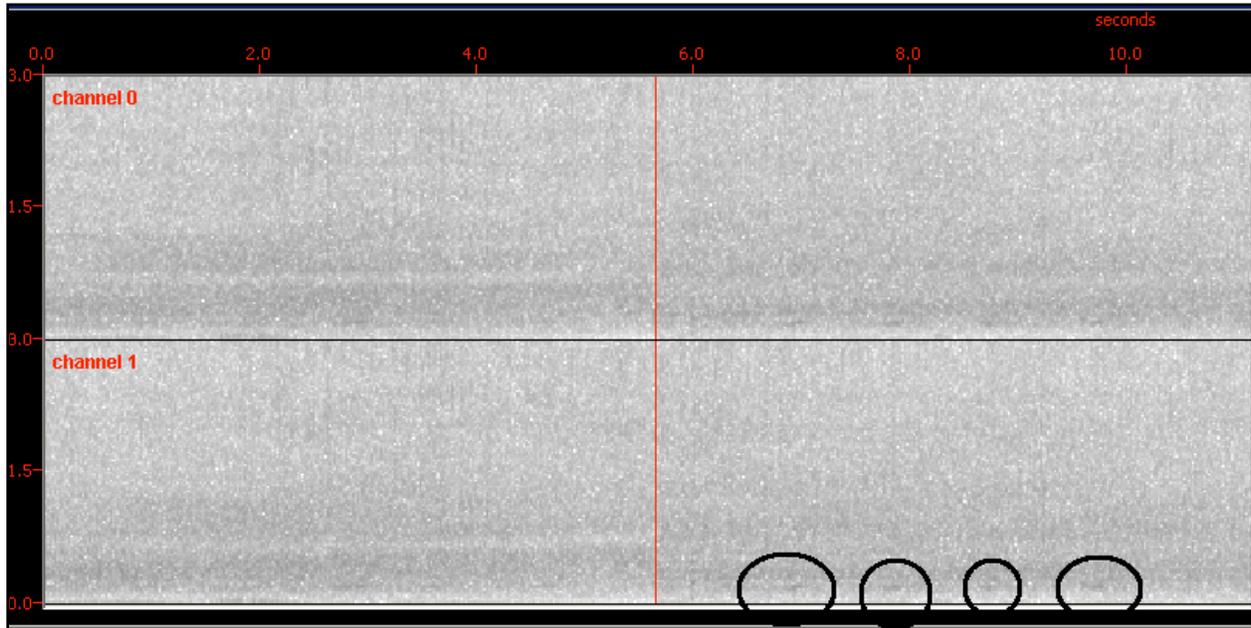


Figure 11: Screen capture of baleen whale detection through *Pamguard* on 11 December 2013 (Acoustic detection 501).

Acoustic detection 501 lasted nine minutes. After a silent period of 17 minutes, at 07:04 UTC the calls were again detected (acoustic detection 502). Double 'whoops' with the same frequency and timing were produced. These call trains of 4 to 6 “whoop’s” lost intensity as the train extended, starting at 120 decibels per $1\mu\text{Pa}$. It is possible that detection 501 and 502 were duplicate detections.

Double calls were once again detected on 23 December 2013 at 7:04 UTC (acoustic detection 503). These calls were detected aurally and on *Pamguard LF spectrogram* (Figure 12) and produced the same signature as previously described detections on 11 December 2013. Moans of 0.5 seconds in length were also detected during this vocalization, upsweeping and tonal. The detection occurred while visual MMO’s were on watch and lasted until 07:47 UTC but a visual correlation was not made.

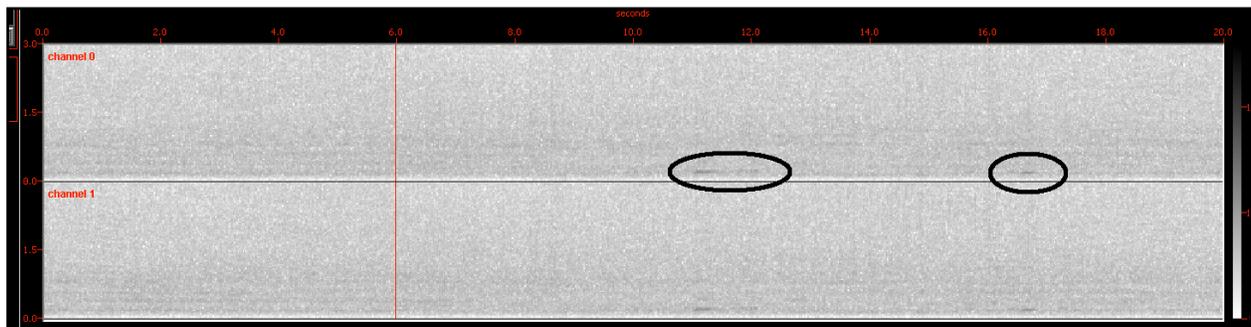


Figure 12: Screen capture of unidentified baleen whale detection through *Pamguard* on 23 December 2013 (acoustic detection 503).

Low frequency (LF) whistles were first detected aurally via headphones then visually on the *Pamguard spectrogram* on 02 January 2014 at 07:18 UTC (acoustic detection 504; Figure 13). Up-sweeping, down-sweeping and tonal single whistles were detected ranging in frequency from 1.6 to 23 kilohertz, in

combination with harmonizing whistles ranging in frequency from 4.4 to 24 kilohertz. Echolocation clicks were also noted during this detection. Via the *Pamguard LF click detector* and the *map module* triangulation of location was determined to be approximately 300 meters from the acoustic source (Figure 14). Intensity of clicks trains and whistles were between 120 to 143 decibels per 1µPa.

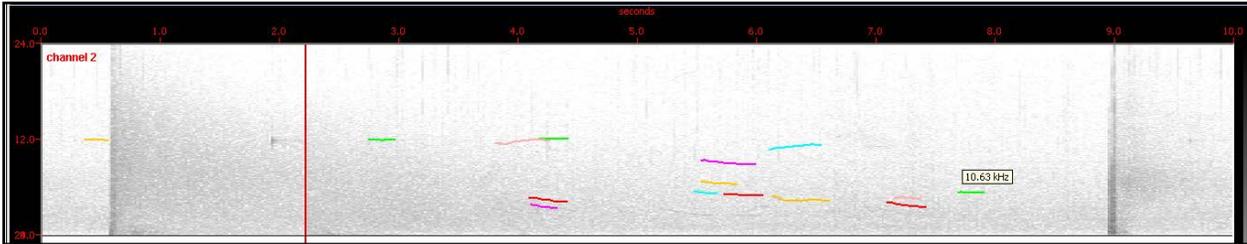


Figure 13: Screen capture of unidentified delphinid whistles detected through *Pamguard* on 02 January 2013 (acoustic detection 504).

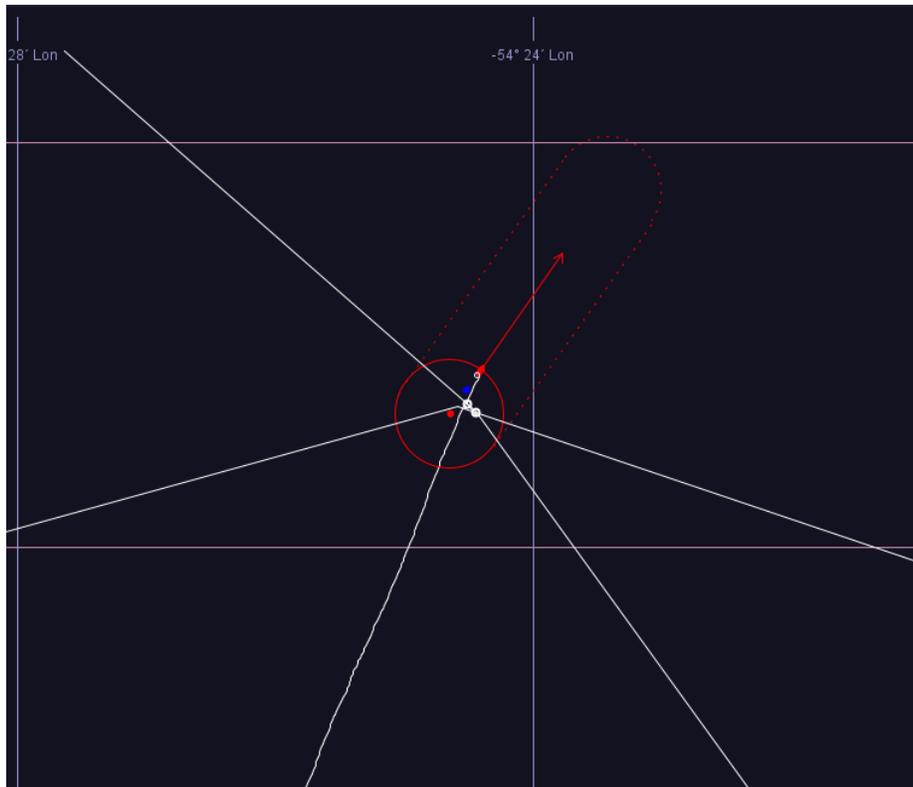


Figure 14: The *Pamguard* map display orientation of echolocation clicks on 02 January 2014 (acoustic detection 504).

On 21 January, acoustic detection 505 occurred on the low frequency system (Figure 15). The sperm whales (two individual click trains side-by-side were observed during the detection) were observed from 06:38 to 07:48 UTC ahead of the vessel. The PAM operator observed the whales ahead, parallel and then behind the vessel during the detection. Echolocation clicks ranged from 2 to 23 kilohertz with peak energy near 2.5 and 11 on various clicks. Click interval was approximately one click per half a second. The distance was estimated to be approximately 830 meters by the majority of cross lines on the map,

as well as, clarity of echolocation clicks aurally observed by the operator. The hydrophone was towing at seven meters during the detection with vessel speed of 4.3 knots. Airguns were firing at full source. Therefore, there was no mitigation action required.

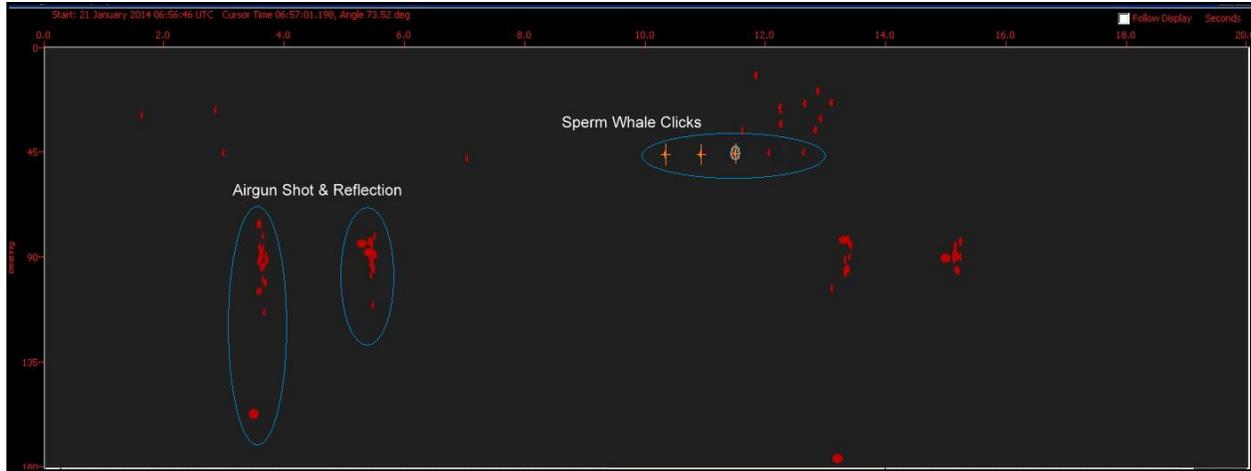


Figure 15: Click trains during acoustic detection 505 and acoustic source activity.

On 26 January, The acoustic detections 506 and 507 of Delphinidae occurred on the low frequency system. Acoustic detection 506 showed low frequency echolocation clicks that appeared split and narrowband, with frequencies between approximately 2.8 to 4 and 7 to 9 kilohertz with peak energy at 3 kilohertz (01:33-01:45 UTC). The inter-click interval appeared to be approximately 0.15 seconds (Figure 16). Bearing was approximately 201 or 335 degrees due to left right ambiguity. It was possible that detection 506 and 507 were the same detection comprised of one or more species since it correlates with acoustic detection 507's bearing off the portside stern as observed when localized. Acoustic detection 507, from 01:46 though 02:27 UTC, exhibited low frequency echolocation clicks from approximately 3 to 22 kilohertz with peak energy between 12 to 14 kilohertz. The inter-click interval appeared to be 0.25 seconds. No whistles or high frequency vocalizations were detected during either detection. Airguns were silent during pre-survey and localization was made outside of the exclusion zone as early as 02:14 UTC approximately 1130 meters off the portside stern. Therefore, there was no mitigation action required when soft start was requested at 02:38 UTC. This soft start was aborted at 02:46 when the vessel went down for weather and all airguns were silenced. The hydrophone was towing at 6.7 meters and the vessel speed was 3.4 knots.

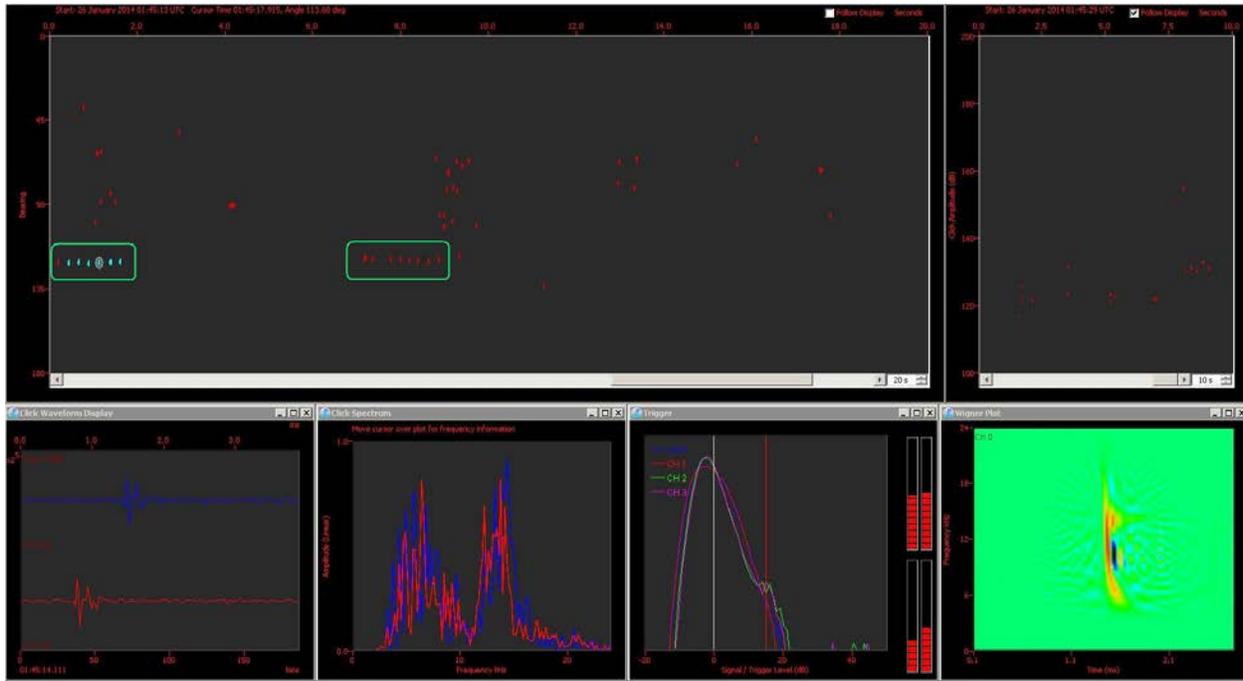


Figure 16: Acoustic detection 506 exhibiting low frequency click trains.

On 18 February, There was one acoustic detection today of unidentified delphinid (AD 508). At 13:12 UTC, the two sinusoidal marks were visually observed but not aurally detected during the pre-watch (Figure 17). Though not determined to be within the exclusion zone, there were no acoustic detections within the final 20 minutes of the pre-watch. Therefore, no mitigation action was required and soft start was cleared at 13:36 UTC. The hydrophone was towing at 6.5 meters and the vessel was traveling at 3.1 knots during the detection.

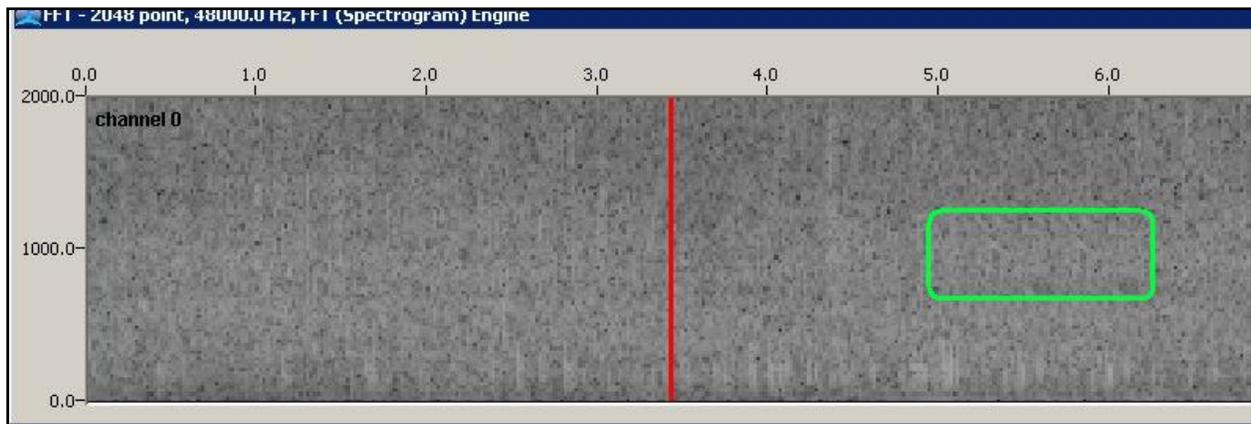


Figure 17: Acoustic detection 508 on 18 February 2014.

05 November 2013 – 18 February 2014

3.2 Wildlife Summary Tables

3.2.1 Protected Species

Table 3: Protected species observed during acoustic monitoring.

| Detection No. | Date | Common Name | Species | Time (UTC) | Name of Recording | Source Activity at Initial Detection | Closest Approach to Source (m) | Mitigation Action |
|---------------|----------|-------------------|-------------------------------|-------------|---|--------------------------------------|--------------------------------|-------------------|
| 501 | 12/11/13 | Baleen whale spp. | Undetermined | 06:38–06:47 | 20131211 Noble PGS Ramform Titan Falklands 063958, 064326 | Silent | >500 | None |
| 502 | 12/11/13 | Baleen whale spp. | Undetermined | 07:04–07:07 | 20131211 Noble PGS Ramform Titan Falklands 070416 | Reduced volume | >500 | None |
| 503 | 12/23/13 | Baleen whale spp. | Undetermined | 07:04-07:47 | 20131223 PGS Noble Ramform Titan Falklands 070459, 070900, 071405, 072650 and 073058 | Silent | >500 | None |
| 504 | 01/02/14 | Delphinid spp. | Undetermined | 07:18-07:28 | 20140102 PGS Noble Ramform Titan Falklands 071826, 072002, 072542 | Full Volume | 300 | None |
| 505 | 01/21/14 | Sperm whale | <i>Physeter macrocephalus</i> | 06:38-07:48 | 20140121 PGS Noble Ramform Titan Falklands: 063807, 065202, 065433, 065745, 070345, 070651, 071238, 072432, 072743, 074340, 074535 & 074825 | Full Volume | 830 | None |
| 506 | 01/26/14 | Delphinid spp. | Undetermined | 01:33-01:45 | 20140126 PGS Noble Ramform Titan Falklands: 013343; 013425; 014550 | Silent | >500 | None |
| 507 | 01/26/14 | Delphinid spp. | Undetermined | 01:46-02:27 | 20140126 PGS Noble Ramform Titan Falklands: 014644; 014707; 015558; 020252; 020352; 020743; 020850; 020907; 021001; 021100 & 021314 | Silent | 1130 | None |
| 508 | 02/18/14 | Delphinid spp. | Undetermined | 13:12-13:13 | 20140218 PGS Noble Ramform Titan Falklands_131208 | Silent | Undetermined | None |

4 Data forms

4.1 Basic Data Form

| BASIC DATA FORM | | | |
|--|-------------------------|---|--------------------------|
| Project Number | | UMS04150 | |
| Seismic Contractor | | PGS | |
| Client | | Noble Energy Falklands Ltd. | |
| Geographical coordinates of the survey area: | | Latitude | Longitude |
| | Northeast Corner | 50°30'20.33"S | 53°53'36.48"W |
| | Southeast Corner | 51°17'59.04"S | 54°23'09.53"W |
| | Southwest Corner | 51°10'41.46"S | 54°52'13.93"W |
| | West mid-point | 50°27'44.02"S | 54°54'45.44"W |
| | Northwest Corner | 50°16'38.27"S | 54°47'44.71"W |
| Survey Type | | 3D dual source | |
| Vessel and/or Rig Name | | <i>Ramform Titan</i> | |
| Permit Number | | PON3 | |
| Location / Distance of Airgun Deployment | | Astern 505m | |
| Water Depth | Min | 1000m | |
| | Max | 1650m | |
| Dates of project | | 05 November 2013 | THROUGH 18 February 2014 |
| Dates included in reporting period | | 05 November 2013 through 18 February 2014 | |
| Total time airguns operating – all power levels: | | 1168:34 | |
| Amount of time airguns operating at full power: | | 1075:16 | |
| Time airguns operating at full power on a survey line: | | 1039:22 | |
| Time airguns operating at full power approaching survey line: | | 35:54 | |
| Amount of time compliance gun/reduced power operations: | | 00:12 recorded auto-fire | |
| Amount of time in soft start: | | 44:36 | |
| Number daytime soft starts | | 78 | |
| Number of night time soft starts | | 52 | |
| Number of soft starts from mitigation source: | | 0 | |
| Number of clearances from PAM system: | | 70 | |
| Number of clearances from PAM and visual: | | 14 | |
| Amount of time conducted in airgun testing: | | 48:30 | |
| Duration of acoustic monitoring: | | 452:39 | |
| Duration of acoustic monitoring while airguns firing: | | 312:26 | |
| Duration of acoustic monitoring during airgun silence: | | 140:13 | |
| Visual Observers: | | Kevin Robinson (05 November 2013 – 08 January 2014), Panagiota Giogli (08 January 2014 – 18 February 2014) | |
| | | Alan Addison (05 November 2013 – 08 January 2014), Rachel Monkhouse (08 January 2014 – 18 February 2014) | |
| Acoustic Observers: | | Breanna Evans (05 November 2013 – 08 January 2014), Lynn Henneberger (08 January 2014 – 18 February 2014) | |
| Number of Marine Mammals Acoustically Detected: | | 8 | |
| Number of acoustic detections confirmed by visual sighting: | | 0 | |
| Number of visual sighting confirmed by acoustic detection: | | 0 | |
| List Mitigation Actions (eg. shutdowns, soft start delays) | | None through PAM | |
| Duration of operational downtime due to mitigation: | | 00:00 | |
| Flagged Data Forms (list sheet & date) | | 22 Jan., 16 Feb. & 18 Feb. 19 minute soft start | |

5 Cover Data Sheet

| RPS | | | | PROTECTED SPECIES RECORDING FORM COVER PAGE | | | |
|---|--|---|---|--|---|--|--|
| Regulatory Reference Number (e.g. DECC no., BOEM permit no., OCS lease no., etc.) PON3 | | Country Falkland Islands | | Project Number UMS04150 | | | |
| Client Noble Energy Falklands LTD | | Seismic Contractor PGS | | Vessel Name Ramform Titan | | | |
| Start Date 2013-11-05 | | End Date 2014-02-18 | | Survey Type 3D | | | |
| Source Vessel(s) Ramform Titan | | Type of Source (e.g. airguns) Airguns | | Number of Airguns (if used) 32 | | | |
| Source Depth (metres) 7 | | Source Volume (cu. in.) 4130 | | Shot Point Interval (metres) 25 | | | |
| Frequency (Hz) 0 - 214 | | Intensity (dB re. 1µPa or bar meters) 140.9 | | Method of Soft Start increase number of guns | | | |
| Visual monitoring equipment used (e.g. binoculars, big eyes, etc.) Binoculars | | Magnification of optical equipment (e.g. binoculars) 12 | Height of eye off water surface (metres) 19 | How was distance of animals estimated? <input checked="" type="checkbox"/> by eye <input type="checkbox"/> with laser rangefinder <input type="checkbox"/> with rangefinder stick / calipers <input checked="" type="checkbox"/> with reticle binoculars <input type="checkbox"/> by relating to object at known distance <input type="checkbox"/> other | | | |
| Marine Mammal Observers Kevin Robinson, Alan Addison, Panagiota Gogli, Rachel Monkhouse | | Training of MMOs <input checked="" type="checkbox"/> JNCC approved MMO induction course for UK waters <input type="checkbox"/> PSO training course for the Gulf of Mexico <input type="checkbox"/> MMO training course for Irish waters <input type="checkbox"/> other <input type="checkbox"/> none | | | | | |
| Was PAM used? yes | | PAM Operator(s) Breanna Evans, Lynn Henneberger | | | | | |
| Description of PAM equipment A 250 meter towed array, deployed off port stern, connected to data processing unit via a 100 meter deck cable. Signals were converted digitally and monitored on Pamguard version 1.12.05 BETA. | | | | | | | |
| Range of PAM hydrophones from airguns (metres) 324 | | Bearing of PAM hydrophones from airguns relative to direction of travel (degrees) 135 | | | Depth of PAM hydrophones (metres) 7 | | |

6 References

Gannier A, Violaine D, Goold JC. 2002. *Distribution and relative abundance of sperm whales in the Mediterranean Sea*. Marine Ecology Progress Series 243:281-291.

Carwardine, Mark. *Whales, Dolphins and Porpoises*. Smithsonian Handbooks. Ed. Evans, P., and Weinrich, M., Illustrated by Camm, M., Dorling Kindersley Publishing, Inc. (1995).

Joint Nature Conservation Committee, (2010). JNCC Guidelines for minimizing the risk of disturbance and injury to marine mammals from seismic surveys. JNCC Aberdeen, United Kingdom.

Petroleum Operations Notice 3 Notification of Geophysical Surveys Form. Dept. of Mineral Resources, Stanley, Falkland Islands.

Project Plan PGS Geophysical Ramform Titan Project No 2013063 and Noble Energy Falklands Limited East Falkland Basin version 1.2. PGS Geophysical, Norway.

**APPENDIX A: JNCC GUIDELINES FOR MINIMIZING THE
RISK OF INJURY AND DISTURBANCE TO MARINE
MAMMALS FROM SEISMIC SURVEYS**



JNCC guidelines for minimizing the risk of injury and disturbance to marine mammals from seismic surveys

August 2010

To find out more about seismic surveys visit <http://www.incc.gov.uk/page-1534>
To learn more about JNCC visit <http://www.incc.gov.uk/page=1729>

JNCC guidelines for minimising the risk of injury and disturbance to marine mammals from seismic surveys

August 2010

Introduction

The guidelines have been written for activities on the United Kingdom Continental Shelf (UKCS) and are aimed at reducing the risk of injury to negligible levels and can also potentially reduce the risk of disturbance from seismic surveys to marine mammals including seals, whales, dolphins and porpoises. Whilst there are no objections to these guidelines being used elsewhere JNCC would encourage all operators to determine if any special or local circumstances pertain, as we would not wish these guidelines to be used where a local management tool has already been adopted (for instance in the Gulf of Mexico OCS Region). In this context, JNCC notes that other protected fauna, for example turtles, will occur in waters where these guidelines may be used, and would suggest that, whilst the appropriate mitigation may require further investigation, the soft-start procedures for marine mammals would also be appropriate for marine turtles and basking sharksⁱ.

The guidelines require the use of trained Marine Mammal Observers (MMOs) whose role is to advise on the use of the guidelines and to conduct pre-shooting searches for marine mammals before commencement of any seismic activity. A further duty is to ensure that the JNCC reporting forms are completed for inclusion in the MMO report. In addition to the visual mitigation provided by MMOs, if seismic surveys are planned to start during hours of darkness or low visibility it is considered best practice to deploy Passive Acoustic Monitoring (PAM).

The 2010 version of the JNCC seismic guidelines reflects amendments (2007 and 2009 amendments) to the Conservation (Natural Habitats &c.) Regulations 1994 (Habitat Regulations, HR) for England and Walesⁱⁱ and the Offshore Marine Conservation (Natural Habitats, &c.) Regulations 2007 (Offshore Marine Regulations, OMR, as amended in 2009 and 2010). Both regulations have revised the definition of deliberate disturbance of 'European Protected Species' (EPS), which now excludes

ⁱ Basking sharks are protected from intentional capture or disturbance in British waters (up to 12 miles offshore) under a 1998 listing on the Wildlife and Countryside Act (1981), Schedule 5.

ⁱⁱ In 2010 a consolidated version of the regulations came into force: The Conservation of Habitats and Species Regulations 2010.

trivial disturbance from the offence. Both regulations now also include the offence of deliberate injury. European Protected Species include cetaceans and turtles.

It has been recognised that sound generated from seismic sources has the potential to cause injury and possibly also disturbance to marine mammals. Seismic surveys have therefore the potential to cause a deliberate injury offence as defined under regulations 41(1)(a) and 39(1)(a) and a deliberate disturbance offence as in 41(1)(b) and 39(1)(b) of the HR and OMR, respectively. The JNCC seismic guidelines reflect best practice for operators to follow during the planning, operational and reporting stages. **It is considered that compliance with the recommendations in these guidelines will reduce the risk of injury to EPS to negligible levels.**

Please note that the mitigation measures recommended in the existing guidelines are more relevant to the prevention of injury rather than disturbance as defined in regulations 41(2) and 39(1A), of the HR and OMR, respectively. The onus should be on the entity responsible for the activity to assess whether a disturbance offence is likely to occur. Guidance on how to carry out such risk assessment is provided in the JNCC, NE and CCW document ‘The protection of marine European Protected Species from injury and disturbance’.

In relation to oil and gas seismic surveys in the UKCS, it is a requirement of the consent issued under regulation 4 of the Petroleum Activities (Conservation of Habitats) Regulations 2001 (& 2007 Amendments) by the Department for Energy Climate Change (DECC), that the JNCC Seismic Guidelines must be followed, and the elements of the guidelines that are relevant to a particular survey are incorporated into the legally-binding condition of consent. It should be noted that it is the responsibility of the company issued consent by DECCⁱⁱⁱ, referred to in these guidelines as the ‘applicant’, to ensure that these guidelines are followed, and it is recommended that a copy of the JNCC guidelines are available onboard all vessels undertaking seismic activities in UK waters. Where relevant, when the survey is completed a MMO report must be submitted to the JNCC.

ⁱⁱⁱ Department of Energy and Climate Change was formerly known as Department for Business and Regulatory Reform (BERR)

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Terminology

Marine European Protected Species: These are marine species in Annex IV(a) of the Habitats Directive that occur naturally in the waters of the United Kingdom. These consist of several species of cetaceans (whales, dolphins and porpoises), turtles, and the Atlantic Sturgeon.

Marine Mammal Observer (MMO): Individual responsible for conducting visual watches for marine mammals. For some seismic surveys it may be requested that observers are trained, dedicated and / or experienced. The MMO may also be a PAM operative if trained.

- **Trained MMO:** Has been on a JNCC recognised course
- **Dedicated MMO:** Trained observer whose role on board is to conduct visual watches for marine mammals (although it could double up as a PAM operative)
- **Experienced MMO:** Trained observer with 3 years of field experience observing for marine mammals, and practical experience of implementing the JNCC guidelines
- **PAM Operative:** Person experienced in the use of PAM software and hardware and marine mammal acoustics

Mitigation Zone: The area where a Marine Mammal Observer keeps watch for marine mammals (and delays the start of activity should any marine mammals be detected).

Passive Acoustic Monitoring (PAM): Software system that utilises hydrophones to detect the vocalisations of marine mammals.

Seismic Survey: Any survey that uses airguns, including 2D/3D/4D and OBC (On-Bottom Cabling) surveys and any similar techniques that use airguns. Surveys using multibeam systems and sub-bottom profiling equipment such as boomers, pingers etc are not considered in these guidelines. However, the guidelines can be adapted and applied to the operation of such systems if considered appropriate.

Shot Point Interval (SPI): Interval between firing of the airgun or airguns.

Site Survey: Seismic survey of a limited area proposed for drilling, infrastructure emplacement etc (typically with source size of 180 cubic inches or less).

Soft-Start: Turning on the airguns at low power and gradually and systematically increasing the output until full power is achieved (usually over a period of 20 minutes). The appropriate soft-start method is dependant upon the type of seismic survey and is discussed in section 3.

United Kingdom Waters: Parts of the sea in or adjacent to the United Kingdom from the low water mark up to the limits of the United Kingdom Continental Shelf.

Vertical Seismic Profiling (VSP) or Borehole Seismic: Seismic survey undertaken 'down hole' in connection with well operations (typically with a source size of 500 cubic inches).

Section 1 – Assessing and minimising the risk of injury

1.1 The Planning Stage

When a seismic survey is being planned, the applicant should consider the following recommendations and best practice advice:

- Determine what marine mammal species are likely to be present in the survey area and assess if there are any seasonal considerations that need to be taken into account, for example periods of migration, breeding, calving or pupping. For UKCS activities the '[Atlas of cetacean distribution in north-west European waters](#)' (Reid *et al.* 2003) is a useful starting point.
- Consult the latest relevant regulatory guidance notes; in the UK, DECC issues guidance notes for oil and gas seismic activities.
- As part of the environmental impact assessment, assess the likelihood of injuring or disturbing a European Protected Species. In the UK, it will be necessary to assess the likelihood of committing an offence as defined in the HR and in the OMR.
- Consult the JNCC, NE and CCW guidance on 'The protection of marine European Protected Species from injury and disturbance' to assist in the environmental impact assessment. To obtain a copy of the latest draft version of the guidance please contact JNCC.

The operator should whenever possible implement the following best practice measures:

- If marine mammals are likely to be in the area, only commence seismic activities during the hours of daylight when visual mitigation using Marine Mammal Observers (MMOs) is possible.
- Only commence seismic activities during the hours of darkness, or low visibility, or during periods when the sea state is not conducive to visual mitigation, if a Passive Acoustic Monitoring (PAM) system is in use to detect marine mammals likely to be in the area, noting the limitations of available PAM technology (seismic surveys that commence during periods of darkness, or low visibility, or during periods when the observation conditions are not conducive to visual mitigation, could pose a risk of committing an injury offence).
- Plan surveys so that the timing will reduce the likelihood of encounters with marine mammals. For example, this might be an important consideration in certain areas/times, e.g. during seal pupping periods near Special Areas of Conservation for common seals or grey seals.
- Provide trained MMOs to implement the JNCC guidelines.
- Use the lowest practicable power levels to achieve the geophysical objectives of the survey.
- Seek methods to reduce and/or baffle unnecessary high frequency noise produced by the airguns (this would also be relevant for other acoustic energy sources).

Section 2 - Marine Mammal Observers

2.1. Role of an MMO

The primary role of an MMO is to act as an observer for marine mammals and to recommend a delay in the commencement of seismic activity should any marine mammals be detected. In addition, a MMO should be able to advise the crew on the procedures set out in the JNCC guidelines and to provide advice to ensure that the survey programme is undertaken in accordance with the guidelines. Before the survey commences it is important to attend any pre-mobilisation meetings to discuss the working arrangements that will be in place, and to request a copy of the survey consent issued by DECC (if applicable). An MMO may also work closely with Passive Acoustic Monitoring operatives. As the MMO role in relation to the vessel and survey operations is purely advisory, it is important to be aware of the command hierarchy and communication channels that will be in place, and determine who the main MMO / PAM operative contacts should be.

In a typical vessel based seismic survey, the MMO / PAM operative may pass advice to the party chief and client's representative through the navigators or seismic observers, and it is important to establish what the working arrangements are, as this may vary from one survey to the other. The MMOs should consider themselves as part of the crew and respect the chain of command that is in place.

MMOs should make certain that their efforts are concentrated on the pre-shooting search before the soft-start. These guidelines cannot be interpreted to imply that MMOs should keep a watch during all daylight hours, but JNCC would encourage all MMOs to manage their time to ensure that they are available to carry out a watch to the best of their ability during the crucial time - the 30 minutes before commencement of the firing of the seismic source (or 60 minutes if surveying where deep diving marine mammals are likely to be present). Whilst JNCC appreciates the efforts of MMOs to collect data at other times, this should be managed to ensure that those observations are not detrimental to the ability to undertake a watch prior to a soft-start. Where two MMOs are onboard a seismic vessel, JNCC would encourage collaboration to ensure that cetacean monitoring is always undertaken during all daylight hours.

2.2. Training requirements for MMOs

A prerequisite for an MMO to be classified as a 'trained MMO' is that they must have received formal training on a JNCC recognised course. (Further information on MMO course providers is available at: <http://www.jncc.gov.uk/page-4703>)

2.3. MMO equipment and reporting forms

MMOs should be equipped with binoculars, a copy of the JNCC guidelines and the 'Marine Mammal Recording Form' which is an Excel spreadsheet and has embedded worksheets named: 'Cover Page', 'Operations', 'Effort' and 'Sightings'. A Word document named 'Deckforms' is also available, and MMOs may prefer to use this when observing before transferring the details to the Excel spreadsheets.

The ability to determine range is a key skill for MMOs to have, and a useful tool to perform this function is a range finding stick.

All MMO forms, including a guide to completing the forms, and instructions on how to make and use a range finding stick are available on the JNCC website.

2.4. Reporting requirements – the MMO report

A report, the 'MMO report', should be sent to the JNCC after the survey has been completed. It is the responsibility of the consent holder to ensure that the MMO report is sent to JNCC. Ideally the MMO report should be sent via e-mail to seismic@jncc.gov.uk, or it can be posted to the address on the front page of these guidelines. Reports should include completed JNCC marine mammal recording forms and contain details of the following:

- The seismic survey reference number provided to the applicant by DECC.
- Date and location of survey.
- Total number and volume of the airguns used.
- Nature of airgun array discharge frequency (in Hz), intensity (in dB re. 1µPa or bar metres) and firing interval (seconds), and / or details of any other acoustic energy used.
- Number and types of vessels involved in the survey.
- A record of all occasions when the airguns were used.
- A record of the watches made for marine mammals, including details of any sightings and the seismic activity during the watches.
- Details of any problems encountered during the seismic survey including instances of non-compliance with the JNCC guidelines.

If there are instances of non-compliance with the JNCC guidelines that constitute a breach of the survey consent conditions, JNCC will copy the report, and their comments on the potential breach to DECC. It is therefore essential that MMO reports are completed as soon as possible after the survey has been completed.

Section 3 – Guidance before and during seismic activity

All observations should be undertaken from the source vessel (where the airguns are being deployed from), unless alternative arrangements have been agreed with DECC. The MMO should be positioned on a high platform with a clear unobstructed view of the horizon, and communication channels between the MMO and the crew should be in place before commencement of the pre-shooting search (this may require portable VHF radios). The MMO should be aware of the timings of the proposed operations, so that there is adequate time to conduct the pre-shooting search. Figure 1 illustrates a typical seismic survey with decision making pathways in the event a marine mammal is detected.

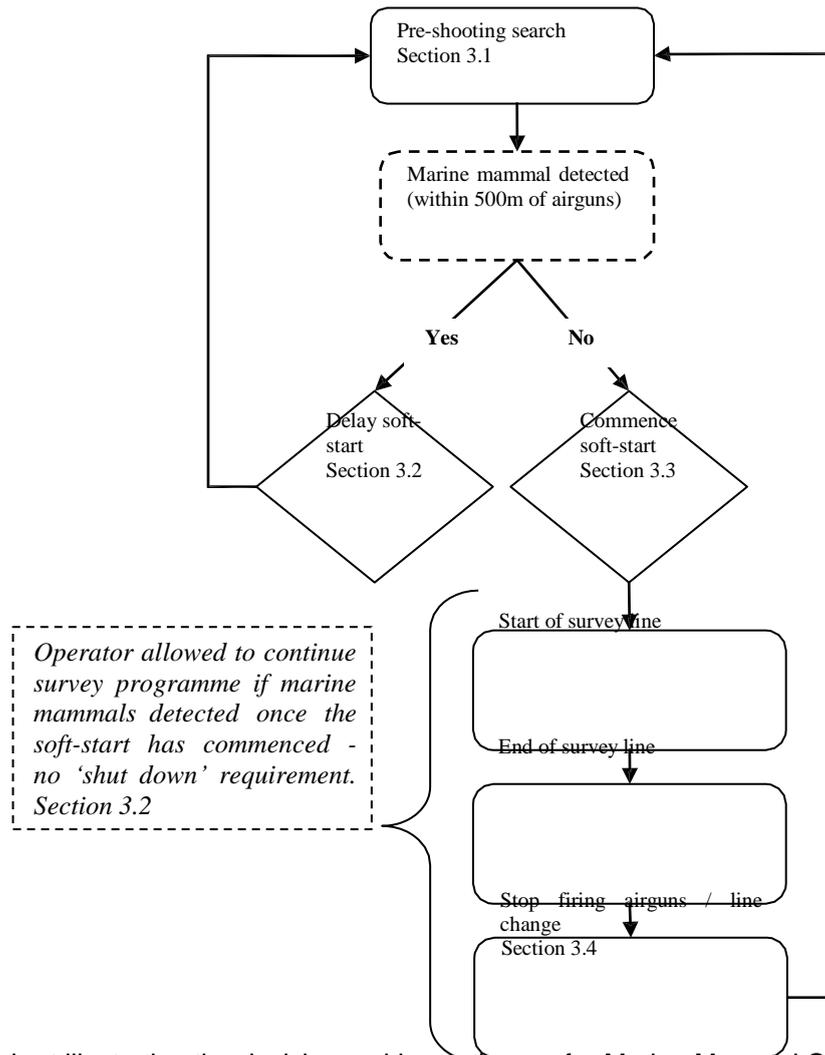


Figure 1. Flowchart illustrating the decision making pathway of a Marine Mammal Observer during a seismic survey.

3.1 Pre-shooting search

The pre-shooting search should normally be conducted over a period of 30 minutes before commencement of any use of the airguns. The MMO should make a visual assessment to determine if any marine mammals are within 500 metres of the centre of the airgun array.

In deep waters (>200m) the pre-shooting search should extend to 60 minutes as deep diving species (e.g. sperm whale and beaked whale) are known to dive for longer than 30 minutes. A longer search time in such areas is likely to lead to a greater detection and tracking of deep diving marine mammals.

To facilitate more effective timing of proposed operations when surveying in deeper waters, the searches for marine mammals can commence before the end of the survey line (whilst the airguns are still firing); this condition may be necessary for surveys which have relatively fast line turn times. If any marine mammals are

detected whilst the airguns are still firing, then no action is required other than for the MMO to monitor and track any marine mammals. The commencement of the soft-start for any subsequent survey lines should be delayed for at least 20 minutes if marine mammals are detected when the airguns have ceased firing.

If PAM is used in conjunction with visual monitoring the PAM operatives should ensure the system is deployed and being monitored for vocalisations during each designated pre-shooting period.

3.2 Delay if marine mammals are detected within the mitigation zone (500 metres)

If marine mammals are detected within 500 metres of the centre of the airgun array during the pre-shooting search, the soft-start of the seismic sources should be delayed until their passage, or the transit of the vessel, results in the marine mammals being more than 500 metres away from the source. In both cases, there should be a 20 minute delay from the time of the last sighting within 500 metres of the source to the commencement of the soft-start, in order to determine whether the animals have left the area. If PAM is used it is the responsibility of the PAM operatives to assess any acoustic detections and determine if there are likely to be marine mammals within 500 metres of the source. If the PAM operatives consider marine mammals are present within that range then the start of the operation should be delayed as outlined above.

If marine mammals are detected within 500 metres of the centre of the airgun array whilst the airguns are firing, either during the soft-start procedure or whilst at full power, there is no requirement to stop firing the airguns.

In situations where seal(s) are congregating around a drilling or production platform that is within the survey area, it is recommended that the soft-start should commence at a location at least 500 metres from the platform.

3.3 The soft-start

The soft-start is defined as the time that airguns commence shooting till the time that full operational power is obtained. Power should be built up slowly from a low energy start-up (e.g. starting with the smallest airgun in the array and gradually adding in others) over at least 20 minutes to give adequate time for marine mammals to leave the area. This build up of power should occur in uniform stages to provide a constant increase in output. There should be a soft-start every time the airguns are used, the only exceptions being for certain types of airgun testing (section 3.3.2), and the use of a 'mini-airgun' (single gun volume less than 10 cubic inches), these are used on site-surveys (section 3.3.1). The duration of the pre-shooting search (at least 30 minutes) and the soft-start procedure (at least 20 minutes) should be factored into the survey design.

General advice to follow for soft-starts:

- To minimise additional noise in the marine environment, a soft-start (from commencement of soft-start to commencement of the line) should not be significantly longer than 20 minutes (for example, soft-starts greater than 40

minutes are considered to be excessive, and an explanation should be provided within the MMO report).

- Where possible, soft-starts should be planned so that they commence within daylight hours.
- Once the soft-start has been performed and the airguns are at full power the survey line should start immediately. Operators should avoid unnecessary firing at full power before commencement of the line.
- If, for any reason, firing of the airguns has stopped and not restarted for at least 10 minutes, then a pre-shooting search and 20 minute soft-start should be carried out (the requirement for a pre-shooting search only applies if there was no MMO on duty and observing at this time, and if the break in firing occurred during the hours of daylight). After any unplanned break in firing for less than 10 minutes the MMO should make a visual assessment for marine mammals (not a pre-shooting search) within 500 metres of the centre of the airgun array. If a marine mammal is detected whilst the airguns are not firing the MMO should advise to delay commencement, as per the pre-shooting search, delay and soft start instructions above. If no marine mammals are present then they can advise to commence firing the airguns.
- When time-sharing, where two or more vessels are operating in adjacent areas and take turns to shoot to avoid causing seismic interference with each other, the soft-start and delay procedures for each vessel should be communicated to, and applied on, all the vessels involved in the surveying.

3.3.1 Soft-start requirements for site survey or Vertical Seismic Profiling (VSP)

Surveys should be planned so that, whenever possible, the soft-start procedures for site surveys and Vertical Seismic Profiles (VSP's) commence during daylight hours. Whilst it is appreciated that high resolution site surveys / VSP operations may produce lower acoustic output than 2D or 3D surveys it is still considered desirable to undertake a soft-start to allow for marine mammals to move away from the seismic source.

For ultra high resolution site surveys that only use a 'mini-airgun' (single airgun with a volume of less than 10 cubic inches) there is no requirement to perform a soft-start, however, a pre-shooting search should still be conducted before its use.

For site surveys and VSPs, a number of options are available to effect a soft-start.

- The standard method, where power is built up slowly from a low energy start-up (e.g. starting with the smallest airgun in the array and gradually adding in others) over at least 20 minutes to give adequate time for marine mammals to leave the vicinity.
- As the relationship between acoustic output and pressure of the air contained in the airgun is close to linear and most site surveys / VSP operations use only a small number of airguns and a soft-start can be achieved by slowly increasing the air pressure in 500 psi steps. From our understanding, the minimum air pressure which the airgun array can be set to will vary, as this is dependent on the make and model of the airgun being used. The time from initial airgun start up to full power should be at least 20 minutes.

- Over a minimum time period of 20 minutes the airguns should be fired at an increasing frequency (by decreasing the Shot Point Interval (SPI)) until the desired firing frequency is reached.

3.3.2 Soft-starts and airgun testing

Airgun tests may be required before a survey commences, or to test damaged or misfiring guns following repair, or to trial new arrays. Individual airguns, or the whole array may need testing, and the airguns may be tested at varying power levels. The following guidance is provided to clarify when a soft-start is required:

- If the intention is to test all airguns at full power then a 20 minute soft-start is required.
- If the intention is to test a single airgun on low power then a soft-start is not required.
- If the intention is to test a single airgun, or a number of guns on high power, the airgun or airguns should be fired at lower power first, and the power then increased to the level of the required test; this should be carried out over a time period proportional to the number of guns being tested and ideally not exceed 20 minutes in duration.

MMOs should maintain a watch as outlined in the pre-shooting search guidance (section 3.1) before any instances of gun testing.

3.4 Line Change

Seismic data is usually collected along predetermined survey lines. Line change is the term used to describe the activity of turning the vessel at the end of one line prior to commencement of the next line. Depending upon the type of seismic survey being undertaken, the time for a line change can vary. Line changes are not necessary for all types of seismic surveys, for example, in certain regional surveys where there is a significant distance between the lines, and for VSP operations.

The guidance relating to line change depends upon the airgun volume.

3.4.1 Seismic surveys with an airgun volume of 500 cubic inches or more

- If the line change time is expected to be greater than 20 minutes, airgun firing should be terminated at the end of the line and a full 20 minute soft-start undertaken before the next line. A pre-shooting search should also be undertaken during the scheduled line change, and the soft-start delayed if marine mammals are seen within 500 metres of the centre of the airgun array.

3.4.2 Seismic surveys with an airgun volume of 180 cubic inches or less (site surveys)

- If the line change time is expected to be greater than 40 minutes, airgun firing should be terminated at the end of the line and a full 20 minute soft-start undertaken before the next line. The pre-shooting search should also be

- undertaken during the scheduled line change, and the soft-start delayed if marine mammals are seen within 500 metres of the centre of the airgun array.
- If the line change time is expected to be less than 40 minutes, airgun firing can continue during the turn, but the Shot Point Interval (SPI) should be increased (longer duration between shots). Ideally, the SPI should not exceed 5 minutes during the turn.

Depending upon the duration of the line turns and the nature of seismic survey it may be necessary to vary the soft-start procedures. If an applicant determines that an effective line change can not be achieved using the above methods please contact JNCC at the earliest possible opportunity to discuss the proposed alternative, and include the details of the agreed procedure and the consultation with the JNCC in the application for survey consent.

3.5 Undershoot operations

During an undershoot operation, one vessel is employed to tow the seismic source and a second vessel used to tow the hydrophone array, although the main vessel will still tow the hydrophone array. This procedure is used to facilitate shooting under platforms or other obstructions. The MMO may be too far away from the airguns to effectively monitor the mitigation zone, and it is therefore recommended to place the MMO on the source vessel. If this is not possible, for example for logistical reasons, or the health and safety implications of transferring personnel from one vessel to another, the application should explain that the recommended procedure cannot be followed in the application for the survey consent, or the application for a variation of that consent. Irrespective of the MMO location agreed with DECC, the pre-shooting search and soft-start procedures should still be followed prior to undertaking an undershoot operation.

Section 4 - Acoustic Monitoring

Visual observation is an ineffective mitigation tool during periods of darkness or poor visibility (such as fog), or during periods when the sea state is not conducive to visual mitigation, as it will not be possible to detect marine mammals in the vicinity of airgun sources. Under such conditions, PAM is considered to be the only currently available mitigation technique that can be used to detect marine mammals. Current PAM systems can be particularly helpful in detecting harbour porpoises within the 500 metre mitigation zone, although the systems have their limitations and can only be used to detect vocalising species of marine mammals.

PAM systems consist of hydrophones that are deployed into the water column, and the detected sounds are processed using specialised software. PAM operatives are needed to set up and deploy the equipment and to interpret the detected sounds.

4.1 Use of PAM as a mitigation tool

PAM can provide a useful supplement to visual observations undertaken by MMOs and JNCC may recommend that it is used as a mitigation tool when commenting on applications for survey consents. However, in many cases it is not as accurate as

visual observation for determining range, and this will mean that the mitigation zone will reflect the range accuracy of the system. For example, if the range accuracy of a system is estimated at +/-300 metres, animals detected and calculated to be within 500 metres from the source could, in reality, be $500 + 300 = 800$ metres, but their detection would still lead to a delay in the soft-start. Although, at present it is not possible to express the range accuracy of most PAM systems in numerical terms, this example serves to illustrate that it is in the operator's best interests to use the most accurate system available, and for the PAM operative to factor in a realistic estimate of the range accuracy.

Some PAM systems do not have a reliable range determination facility or can only calculate the range for some species. In such cases, the detection of a confirmed cetacean vocalisation should still be used to initiate postponement of the soft-start if the PAM operator is able to make a judgement about the range of the animals from the airgun source, because of their experience gained in differentiating between distant and close vocalisations. In the absence of PAM systems capable of range determination, this expert judgement will constitute the basis for deciding whether an area is free from cetaceans prior to the soft-start.

In all cases where PAM is employed, a brief description of the system and an explanation of how the applicant intends to deploy PAM to greatest effect should be included in the application for survey consent.

In the last few years, software that processes and analyses cetacean sounds has been developed. An example of this is PAMGuard, an open source software that has been developed as part of the International Association of Oil and Gas Producers Joint Industry Project (JIP). JNCC recognises that PAMGuard is currently in a transition period between use as a research tool and widespread adoption as a monitoring technique. Moreover, JNCC recognises the need to balance proactive implementation of PAM with the need to further develop its capability, for example to include species recognition and baleen whale detection, and therefore encourages users of these systems to actively contribute to their development and refinement.

Section 5 – Requirements for MMOs and PAM

Any survey application or consultation received by JNCC will be considered on a case-by-case basis, and the mitigation measures advised to DECC will reflect the particulars of the survey and the importance of the survey area for marine mammals. The following paragraphs are provided as a guide to the advice applicants are likely to receive following submission of an application with JNCC.

For areas that are currently considered particularly important for marine mammals, for example in the UK this includes areas West of Scotland, the Moray Firth and Cardigan Bay, JNCC may recommend that:

- The MMOs should be experienced MMOs, and that PAM should be used.
- The PAM system should be used to supplement visual observations, or as the main mitigation tool if the seismic survey activity commences during periods of

darkness or poor visibility, or during periods when the sea state is not conducive to visual mitigation.

JNCC will advise that two marine mammal observers should be used when daylight hours exceed approximately 12 hours per day (between 1st April and 1st October north of 57^o latitude), or the survey is in an area considered particularly important for marine mammals.

When a non-dedicated MMO is recommended by JNCC (e.g. for VSPs and certain site-surveys), and the recommendation is incorporated into the conditions of the survey consent, a member of the rig's or vessels crew can perform the duties providing the crew member is a trained MMO.

When a dedicated MMO is recommended and this is a condition of the survey consent, the MMO should be employed solely for the purpose of monitoring the implementation of the guidelines and undertaking visual observations to detect marine mammals during periods of seismic activity.

When two dedicated MMOs are requested and this is a condition of the survey consent, both should be employed solely for the purposes of monitoring the implementation of the guidelines and undertaking visual observations, and the use of a crew member with other responsibilities as the second observer is not considered to be an adequate substitute for a dedicated MMO, or to be in compliance with the conditions of the survey consent.

Section 6 - Background Information

These guidelines were originally prepared by a Working Group convened by the Department of the Environment, and were developed from a draft prepared by the Sea Mammal Research Unit (SMRU). The guidelines have subsequently been reviewed three times by the Joint Nature Conservation Committee, following consultation with interested parties.

6.1. Existing protection to cetaceans

Section 9 of the Wildlife and Countryside Act 1981 (CRoW amended) prohibits the intentional or reckless killing, injuring or disturbance of any cetacean. The UK is also a signatory to the Agreement on the Conservation of Small Cetaceans of the Baltic and North Seas (ASCOBANS) and has applied its provisions in all UK waters. Amongst other actions required to conserve and manage populations of small cetaceans, ASCOBANS requires range states to "work towards...the prevention of ...disturbance, especially of an acoustic nature".

Reflecting the requirements of the Convention on the Conservation of European Wildlife and Habitats (the Bern Convention) and Article 12 of the EC Habitats and Species Directive (92/43/EEC), the UK has the following legislation in place:

- The Conservation of Habitats and Species Regulations 2010
- The Conservation (Natural Habitats, &c.) Regulations 1995 (Northern Ireland) (and 2009 amendments)

- The Conservation (Natural Habitats, &c.) Amendment (No. 2) Regulations 2008 (Scotland) (and 2009 amendments)
- The Offshore Petroleum Activities (Conservation of Habitats) Regulations 2001 (and 2007 amendments),
- The Offshore Marine Conservation (Natural Habitats, &c.) Regulations 2007 (and 2009 and 2010 amendments) (beyond 12 nautical miles UKCS)

Section 7 – References and contacts

Further information on DECC's survey consent procedure can be found at:

<http://www.og.decc.gov.uk/>.

A copy of these guidelines, the standard forms (electronic and hard copy) and further background information is available from the above address, or can be found on the JNCC website at: <http://www.jncc.gov.uk/page-1534>

Reid, J.B., Evans, P.G.H., & Northridge, S.P. (2003). '[Atlas of cetacean distribution in north-west European waters](http://www.jncc.gov.uk/page-2713)' (Online).
<http://www.jncc.gov.uk/page-2713>

If you have any comments or questions relating to these guidelines, or suggestions on how they may be improved, please email seismic@jncc.gov.uk

APPENDIX B: MARINE MAMMAL RECORDING FORMS - ACOUSTIC DETECTIONS

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | |
|---|--|---|--|--|-----|---|
| Regulatory reference number | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 501 for first detection of survey) | | |
| PON3 | Ramform Titan | | | 501 | | |
| Date | | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC, 24hr clock) | | |
| 11/12/2013 | | | 6:38 | 6:47 | | |
| Were animals detected visually and/ or acoustically? | | How were the animals first detected? | | | | |
| | | visually detected by observer keeping a continuous watch | | | | |
| visual | | visually spotted incidentally by observer or someone else | | | | |
| X | acoustic | X | acoustically detected by PAM | | | |
| both | | both visually and acoustically before operators/ observers informed each other | | | | |
| Observer's/ operator's name | | Position (latitude and longitude) | | Water depth (metres) | | |
| Breanna Evans | | 50°13.273 54°34.235 | | 1322 | | |
| Species/ species group | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | |
| Unidentified baleen whale | | | Double calls mainly flat with occasional down-sweeping and upsweeping moans, between 165 and 315 Hz, with peak energy at 205 Hz. | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | | | |
| Unknown | | >500 | | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | Number of calves (visual sightings only) | Photograph taken | | |
| n/a | n/a | n/a | n/a | | Yes | X No |
| Behaviour (visual sightings only) | | | | | | |
| n/a | | | | | | |
| Direction of travel (relative to ship) | | | | Direction of travel (compass points) | | |
| | towards ship | | variable | | N | SW |
| | away from ship | | milling | | NE | W |
| | parallel to ship in same direction as ship | | stationary | | E | NW |
| | parallel to opposite direction to ship | | other | | SE | variable |
| | crossing perpendicular ahead of ship | X | unknown | | S | stationary |
| Airgun (or other source) activity when animals first detected | | Airgun (or other source) activity when last detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) | | X unknown |
| | full power | | full power | n/a | | Time animals left mitigation/ exclusion zone (UTC) |
| X | not firing | | not firing | | | n/a |
| | soft start | | soft start | Closest distance of animals from airguns (or other source) (metres) | | Time of closest approach (UTC) |
| | reduced power (other than soft start) | X | reduced power (other than soft start) | 0 | | 0:00 |
| If seen during soft start give: | | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | Length of power-down and/ or shut-down) | | Estimated loss of production (if relevant) due to mitigating actions (km) |
| First distance: | | n/a | | | | |
| Closest distance: | | n/a | X none required | | | |
| Last distance: | | n/a | delay start of firing | | | |
| during soft start (metres) | | | shut-down of active source | | | |
| Other notes or comments | | | power-down of active source | | | |
| It was determined by PAM op that the animal was outside the exclusion zone, exact distance could not be determined. Number of animals, direction of travel, and any fields left blank could not be determined due to the type of detection. | | | power-down then shut-down of active source | n/a | | n/a |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | | |
|---|--|---|--|--|------------|--|-----|
| Regulatory reference number | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 501 for first detection of survey) | | | |
| PON3 | Ramform Titan | | | 502 | | | |
| Date | | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC, 24hr clock) | | | |
| 11/12/2013 | | | 7:04 | 7:07 | | | |
| Were animals detected visually and/ or acoustically? | | How were the animals first detected? | | | | | |
| | | visually detected by observer keeping a continuous watch | | | | | |
| visual | | visually spotted incidentally by observer or someone else | | | | | |
| X | acoustic | X | acoustically detected by PAM | | | | |
| both | | both visually and acoustically before operators/ observers informed each other | | | | | |
| Observer's/ operator's name | | Position (latitude and longitude) | | Water depth (metres) | | | |
| Breanna Evans | | 50°13.11 54°31.41 | | 1322 | | | |
| Species/ species group | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | | |
| Unidentified baleen whale | | | | | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | |
| Unknown | | >500 | | Double calls mainly flat with occasional down-sweeping and upsweeping moans, between 165 and 315 Hz with peak energy at 205 Hz. Presumably the same animal in both detections. | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | Number of calves (visual sightings only) | Photograph taken | | | |
| n/a | n/a | n/a | n/a | Yes | X | No | |
| Behaviour (visual sightings only) | | | | | | | |
| n/a | | | | | | | |
| Direction of travel (relative to ship) | | | Direction of travel (compass points) | | | | |
| towards ship | | | variable | N | SW | | |
| away from ship | | | milling | NE | W | | |
| parallel to ship in same direction as ship | | | stationary | E | NW | | |
| parallel to opposite direction to ship | | | other | SE | variable | | |
| crossing perpendicular ahead of ship | | | X unknown | S | stationary | | |
| Airgun (or other source) activity when last detected | | Airgun (or other source) activity when animals first detected | | Time animals entered mitigation/ exclusion zone (UTC) | | X unknown | |
| full power | | full power | | n/a | | Time animals left mitigation/ exclusion zone (UTC) | |
| not firing | | not firing | | n/a | | n/a | |
| soft start | | soft start | | Closest distance of animals from airguns (or other source) (metres) | | Time of closest approach (UTC) | |
| X | reduced power (other than soft start) | X | reduced power (other than soft start) | 0 | | 0:00 | |
| If seen during soft start give: | | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | | | | |
| First distance: | n/a | X none required | | | | | |
| Closest distance: | n/a | | | | | | |
| Last distance: | n/a | | | | | | |
| during soft start (metres) | | shut-down of active source | | | | | |
| Other notes or comments | | power-down of active source | | | | | |
| It was determined by PAM op that the animal was outside the exclusion zone, exact distance could not be determined. Number of animals, direction of travel, and any fields left blank could not be determined due to the type of detection. | | power-down then shut-down of active source | | | n/a | | n/a |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | | |
|---|--|---|--|---|---|---------|--|
| Regulatory reference number | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 501 for first detection of survey) | | | |
| PON3 | Ramform Titan | | | 503 | | | |
| Date | | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC) | | | |
| 23/12/2013 | | | 7:04 | 7:47 | | | |
| Were animals detected visually and/ or acoustically? | | How were the animals first detected? | | | | | |
| | | visually detected by observer keeping a continuous watch | | | | | |
| visual | | visually spotted incidentally by observer or someone else | | | | | |
| X | acoustic | X | acoustically detected by PAM | | | | |
| both | | both visually and acoustically before operators/ observers informed each other | | | | | |
| Observer's/ operator's name | | Position (latitude and longitude) | | Water depth (metres) | | | |
| Breanna Evans | | 50°19.212 54°14.349 | | 1545 | | | |
| Species/ species group | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | | |
| Unidentified baleen whale | | | | | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | Double "whoop-whoop" calls, first up-sweeping second with less intensity and more tonal. Occasional single moans 0.5 seconds in duration. Trains of 3 to 6 also detected slowly losing intensity with each call. Frequency range of 178 to 373Hz. | | | |
| Unknown | | >500 | | | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | Number of calves (visual sightings only) | Photograph taken | | | |
| n/a | n/a | n/a | n/a | Yes | X | No | |
| Behaviour (visual sightings only) | | | | | | | |
| n/a | | | | | | | |
| Direction of travel (relative to ship) | | | Direction of travel (compass points) | | | | |
| towards ship | | | variable | N | SW | | |
| away from ship | | | milling | NE | W | | |
| parallel to ship in same direction as ship | | | stationary | E | NW | | |
| parallel to opposite direction to ship | | | other | SE | variable | | |
| crossing perpendicular ahead of ship | | | X unknown | S | stationary | | |
| Airgun (or other source) activity when last detected | | Airgun (or other source) activity when animals first detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) | X | unknown | |
| full power | | full power | | n/a | Time animals left mitigation/ exclusion zone (UTC) | | |
| X | not firing | X | not firing | | n/a | | |
| soft start | | soft start | | Closest distance of animals from airguns (or other source) (metres) | Time of closest approach (UTC) | | |
| reduced power (other than soft start) | | reduced power (other than soft start) | | | 0 0:00 | | |
| If seen during soft start give: | | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | | | | |
| First distance: | n/a | | | Length of power-down and/ or shut-down (if relevant) (length of time until subsequent soft start, in minutes) | Estimated loss of production (if relevant) due to mitigating actions (km) | | |
| Closest distance: | n/a | X | none required | | | | |
| Last distance: | n/a | delay start of firing | | | | | |
| during soft start (metres) | | shut-down of active source | | n/a | n/a | | |
| Other notes or comments | | power-down of active source | | | | | |
| It was determined by PAM op that the animal was outside the exclusion zone, exact distance could not be determined. Number of animals, direction of travel, and any fields left blank could not be determined due to the type of detection. | | power-down then shut-down of active source | | | | | |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | |
|---|---|--|---|
| Regulatory reference number | Ship/ platform name | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 501 for first detection of survey) |
| PON3 | Ramform Titan | | 504 |
| Date | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC, 24hr clock) |
| 02/01/2014 | | 7:18 | 7:28 |
| Were animals detected visually and/ or acoustically? | How were the animals first detected? | | |
| | visually detected by observer keeping a continuous watch | | |
| | visually spotted incidentally by observer or someone else | | |
| X | visual | acoustically detected by PAM | |
| | acoustic | X | both visually and acoustically before operators/ observers informed each other |
| | both | | |
| Observer's/ operator's name | Position (latitude and longitude) | | Water depth (metres) |
| Breanna Evans | 50°25.431 54°24.628 | | 1380 |
| Species/ species group | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | |
| Unidentified dolphin spp. | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | Range to animal (when first seen or heard) (metres) | | Single LF up-sweeping, down-sweeping and tonal whistles from 1.6 to 23kHz. Harmonizing whistles between 4 - 24 kHz. Echolocation clicks bearing 36 (as pod moved further astern vessel) final 106. Intensity between 120 and 143 dB per 1µPa. |
| 36 | 300 | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | Number of calves (visual sightings only) |
| n/a | n/a | n/a | n/a |
| Photograph taken | | | |
| Yes X No | | | |
| Behaviour (visual sightings only) | | | |
| n/a | | | |
| Direction of travel (relative to ship) | | Direction of travel (compass points) | |
| | towards ship | variable | N SW |
| X | away from ship | milling | NE W |
| | parallel to ship in same direction as ship | stationary | E NW |
| | parallel to opposite direction to ship | other | SE variable |
| | crossing perpendicular ahead of ship | unknown | X S stationary |
| Airgun (or other source) activity when last detected | Airgun (or other source) activity when animals first detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) |
| | | | unknown |
| | full power | full power | Time animals left mitigation/ exclusion zone (UTC) |
| | not firing | not firing | 7:28 |
| | soft start | soft start | |
| | reduced power (other than soft start) | reduced power (other than soft start) | Closest distance of animals from airguns (or other source) (metres) |
| | | | 300 |
| | | | Time of closest approach (UTC) |
| | | | 7:27 |
| If seen during soft start give: | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | |
| First distance: | n/a | none required | |
| Closest distance: | n/a | delay start of firing | |
| Last distance: | n/a | shut-down of active source | |
| during soft start (metres) | | power-down of active source | |
| Other notes or comments | | power-down then shut-down of active source | |
| It is thought that the pod was traveling away from the vessel, but the change in bearings could have also come from the movement of the vessel. | | n/a | n/a |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | |
|---|---|--|--|---|--------------------------------|----|
| Regulatory reference number | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 601 for first detection of survey) | | |
| PON3 | Ramform Titan | | | 505 | | |
| Date | | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC, 24hr clock) | | |
| 21/01/2014 | | | 6:38 | 7:48 | | |
| Were animals detected visually and/ or acoustically? | How were the animals first detected? | | | | | |
| | visually detected by observer keeping a continuous watch | | | | | |
| | visual | visually spotted incidentally by observer or someone else | | | | |
| X | acoustic | X | acoustically detected by PAM | | | |
| | both | both visually and acoustically before operators/ observers informed each other | | | | |
| Observer's/ operator's name | | Position (latitude and longitude) | | Water depth (metres) | | |
| Lynn Henneberger | | 50°47.441 54°12.141 | | 1541 | | |
| Species/ species group | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | |
| Sperm whale | | | | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | multiple LF click trains (~2 to 23kHz peak energy near 2.5 and 11), click interval was ~0.5 sec | | |
| 155 at 07:26 | | 830 | | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | Number of calves (visual sightings only) | Photograph taken | | |
| 2 | n/a | n/a | n/a | Yes | X | No |
| Behaviour (visual sightings only) | | | | | | |
| Was first observed ahead, then 90 degrees, then behind the hydrophones as the detection progressed. | | | | | | |
| Direction of travel (relative to ship) | | | Direction of travel (compass points) | | | |
| towards ship | | | variable | N | SW | |
| away from ship | | | milling | NE | W | |
| parallel to ship in same direction as ship | | | stationary | E | NW | |
| parallel to opposite direction to ship | | | other | SE | variable | |
| crossing perpendicular ahead of ship | | | X | unknown | stationary | |
| Airgun (or other source) activity when animals first detected | Airgun (or other source) activity when last detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) | X | unknown | |
| full power | full power | | n/a | Time animals left mitigation/ exclusion zone (UTC) | | |
| not firing | not firing | | | n/a | | |
| soft start | soft start | | | Closest distance of animals from airguns (or other source) (metres) | Time of closest approach (UTC) | |
| reduced power (other than soft start) | reduced power (other than soft start) | | 830 | 7:26 | | |
| If seen during soft start give: | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | Length of power-down and/ or shut-down (if relevant) (length of time until subsequent soft start, in minutes) | Estimated loss of production (if relevant) due to mitigating actions (km) | | |
| First distance: | n/a | none required | | | | |
| Closest distance: | n/a | | | | | |
| Last distance: | n/a | | | | | |
| during soft start (metres) | shut-down of active source | | n/a | n/a | | |
| Other notes or comments | power-down of active source | | | | | |
| outside the exclusion zone | power-down then shut-down of active source | | | | | |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | | |
|--|--|---|-----------------------------------|--|------------------|---|----|
| Regulatory reference number | | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | | Acoustic detection number (start at 701 for first detection of survey) | |
| PON3 | | Ramform Titan | | | | 506 | |
| Date | | | | Time at start of encounter (UTC, 24hr clock) | | Time at end of encounter (UTC) | |
| 26/01/2014 | | | | 1:33 | | 1:45 | |
| Were animals detected visually and/ or acoustically? | | How were the animals first detected? | | | | | |
| | | visually detected by observer keeping a continuous watch | | | | | |
| visual | | visually spotted incidentally by observer or someone else | | | | | |
| X | | X | | acoustically detected by PAM | | | |
| both | | both visually and acoustically before operators/ observers informed each other | | | | | |
| Observer's/ operator's name | | | Position (latitude and longitude) | | | Water depth (metres) | |
| Lynn Henneberger | | | 51°22.453 | | 54°34.021 | 1509 | |
| Species/ species group | | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | |
| Delphinidae | | | | split, narrowband echolocation clicks from 2.8 to 4 and 7 to 9 kHz, peak energy at 3; inter-click interval 0.15s | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | | | | |
| 201 or 335 | | >500 | | | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | | Number of calves (visual sightings only) | Photograph taken | | |
| 1 | n/a | n/a | | n/a | Yes | X | No |
| Behaviour (visual sightings only) | | | | | | | |
| n/a | | | | | | | |
| Direction of travel (relative to ship) | | | | Direction of travel (compass points) | | | |
| towards ship | | | | variable | N | SW | |
| away from ship | | | | milling | NE | W | |
| parallel to ship in same direction as ship | | | | stationary | E | NW | |
| parallel to opposite direction to ship | | | | other | SE | variable | |
| crossing perpendicular ahead of ship | | X | | unknown | S | stationary | |
| Airgun (or other source) activity when animals first detected | | Airgun (or other source) activity when last detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) | | X unknown | |
| full power | | full power | | n/a | | Time animals left mitigation/ exclusion zone (UTC, 24hr clock) | |
| not firing | | not firing | | | | n/a | |
| soft start | | soft start | | Closest distance of animals from airguns (or other source) (metres) | | Time of closest approach (UTC, 24hr clock) | |
| reduced power (other than soft start) | | reduced power (other than soft start) | | >500 | | 0:00 | |
| If seen during soft start give: | | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | | | | |
| First distance: | n/a | | | Length of power-down and/ or shut-down (if relevant) (length of time until subsequent soft start, in minutes) | | Estimated loss of production (if relevant) due to mitigating actions (km) | |
| Closest distance: | n/a | X none required | | | | | |
| Last distance: | n/a | delay start of firing | | | | | |
| during soft start (metres) | | shut-down of active source | | | | | |
| Other notes or comments | | power-down of active source | | n/a | | n/a | |
| outside the exclusion zone | | power-down then shut-down of active source | | | | | |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | |
|--|--|---|---|---|----|---|
| Regulatory reference number | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 801 for first detection of survey) | | |
| PON3 | Ramform Titan | | | 507 | | |
| Date | | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC, 24hr clock) | | |
| 26/01/2014 | | | 1:46 | 2:27 | | |
| Were animals detected visually and/ or acoustically? | | How were the animals first detected? | | | | |
| | | visually detected by observer keeping a continuous watch | | | | |
| visual | | visually spotted incidentally by observer or someone else | | | | |
| X | acoustic | X | acoustically detected by PAM | | | |
| both | | both visually and acoustically before operators/ observers informed each other | | | | |
| Observer's/ operator's name | | Position (latitude and longitude) | | Water depth (metres) | | |
| Lynn Henneberger | | 51°22.453 54°34.021 | | 1509 | | |
| Species/ species group | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks | | | |
| Delphinidae | | | clicks from 3 to 22kHz, peak from 12-14kHz; inter-click interval ~0.25s | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | | | |
| 335 | | 1130 | | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | Number of calves (visual sightings only) | Photograph taken | | |
| 2 | n/a | n/a | n/a | Yes | X | No |
| Behaviour (visual sightings only) | | | | | | |
| n/a | | | | | | |
| Direction of travel (relative to ship) | | | | Direction of travel (compass points) | | |
| towards ship | | | | variable | N | SW |
| away from ship | | | | milling | NE | W |
| parallel to ship in same direction as ship | | | | stationary | E | NW |
| parallel to opposite direction to ship | | | | other | SE | variable |
| crossing perpendicular ahead of ship | | | | X unknown | S | stationary |
| Airgun (or other source) activity when animals first detected | | Airgun (or other source) activity when last detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) | | X unknown |
| full power | | full power | | n/a | | Time animals left mitigation/ exclusion zone (UTC) |
| not firing | | not firing | | | | n/a |
| soft start | | soft start | | Closest distance of animals from airguns (or other source) (metres) | | Time of closest approach (UTC) |
| reduced power (other than soft start) | | reduced power (other than soft start) | | 1130 | | 2:14 |
| If seen during soft start give: | | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | | | |
| First distance: | n/a | | | Length of power-down and/ or shut-down (if relevant) (length of time until subsequent soft start, in minutes) | | Estimated loss of production (if relevant) due to mitigating actions (km) |
| Closest distance: | n/a | X | none required | | | |
| Last distance: | n/a | delay start of firing | | | | |
| during soft start (metres) | | shut-down of active source | | | | |
| Other notes or comments | | power-down of active source | | n/a | | n/a |
| outside the exclusion zone | | power-down then shut-down of active source | | | | |

**PROTECTED SPECIES RECORDING FORM
DETECTION**

| | | | | | | |
|--|--|---|--|---|------------------|--|
| Regulatory reference number | Ship/ platform name | | Sighting number (start at 1 for first sighting of survey) | Acoustic detection number (start at 901 for first detection of survey) | | |
| PON3 | Ramform Titan | | | 508 | | |
| Date | | | Time at start of encounter (UTC, 24hr clock) | Time at end of encounter (UTC, 24hr clock) | | |
| 18/02/2014 | | | 13:12 | 13:13 | | |
| Were animals detected visually and/ or acoustically? | | How were the animals first detected? | | | | |
| | | visually detected by observer keeping a continuous watch | | | | |
| visual | | visually spotted incidentally by observer or someone else | | | | |
| X | acoustic | X | acoustically detected by PAM | | | |
| both | | both visually and acoustically before operators/ observers informed each other | | | | |
| Observer's/ operator's name | | Position (latitude and longitude) | | Water depth (metres) | | |
| Lynn Henneberger | | 50°27.77 53°49.852 | | 1213 | | |
| Species/ species group | | | Description (include features such as overall size; shape of head; colour and pattern; size, shape and position of dorsal fin; height, direction and shape of blow; characteristics of whistles/ clicks) | | | |
| Delphinidae | | | | | | |
| Bearing to animal (when first seen or heard) (bearing from true north) | | Range to animal (when first seen or heard) (metres) | | Two sinusoidal marks from 1.027 to 1.05 kilohertz. The sounds were not aurally detected or high-lighted by the program. | | |
| Unknown | | Unknown | | | | |
| Total number | Number of adults (visual sightings only) | Number of juveniles (visual sightings only) | | Number of calves (visual sightings only) | Photograph taken | |
| 1 | n/a | n/a | | n/a | Yes | X No |
| Behaviour (visual sightings only) | | | | | | |
| n/a | | | | | | |
| Direction of travel (relative to ship) | | | | Direction of travel (compass points) | | |
| towards ship | | variable | | N | SW | |
| away from ship | | milling | | NE | W | |
| parallel to ship in same direction as ship | | stationary | | E | NW | |
| parallel to opposite direction to ship | | other | | SE | variable | |
| crossing perpendicular ahead of ship | | X | unknown | S | stationary | |
| Airgun (or other source) activity when animals first detected | | Airgun (or other source) activity when last detected | | Time animals entered mitigation/ exclusion zone (UTC, 24hr clock) | | X unknown |
| full power | | full power | | n/a | | Time animals left mitigation/ exclusion zone (UTC) |
| not firing | | not firing | | | | n/a |
| soft start | | soft start | | Closest distance of animals from airguns (or other source) (metres) | | Time of closest approach (UTC) |
| reduced power (other than soft start) | | reduced power (other than soft start) | | 0 | | 13:12 |
| If seen during soft start give: | | What action was taken?(according to requirements of guidelines/ regulations in country concerned) | | | | |
| First distance: | n/a | X | none required | | | |
| Closest distance: | n/a | | | | | |
| Last distance: | n/a | delay start of firing | | | | |
| during soft start (metres) | | shut-down of active source | | | | |
| Other notes or comments | | power-down of active source | | | | |
| Low frequency that was not aurally detected. Not localized | | power-down then shut-down of active source | | | | |
| | | | | n/a | | n/a |